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INDEX-CATALOGUE OF MEDICAL AND VETERINARY ZOOLOGY

SUPPLEMENT 22, PART 2

PARASITE-SUBJECT CATALOGUE
PARASITES: PROTOZOA

UNITED STATES
DEPARTMENT OF
AGRICULTURE

PREPARED BY
SCIENCE AND
EDUCATION
ADMINISTRATION

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SUPPLEMENT 22, PART 2

PARASITE-SUBJECT CATALOGUE
PARASITES: PROTOZOA

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PREFACE

The Index-Catalogue of Medical and Veterinary Zoology is an index to the world's literature on animal parasites of animals, including man. The Catalogue is distributed to qualified individuals and libraries throughout the world without charge. It has been maintained in cumulative files since 1892. Only the Author Catalogue has been published in its entirety. A revision of the Author Catalogue of the Index-Catalogue of Medical and Veterinary Zoology, consisting of Parts 1 to 18, was published during the period 1932-52. Beginning in 1953, a series of supplements designed to publish the backlog was initiated. This was completed with Supplement 6, published in 1956. From 1956 to 1964, supplements covering authors A to Z were issued on an annual basis.

Beginning with Supplement 15, the Parasite-Subject Catalogues, containing indices to the author references, have been issued. The Author Catalogues of Supplements 15-21 continued the format of previous supplements. Users should note that for each reference in the Author Catalogues of these supplements the author(s) plus the date and letter (e.g., Smith, J.; and Doe, L., 1978 b) are the key to all items in the Parasite-Subject Catalogues derived from that reference. In other words, when using the Parasite-Subject Catalogues of Supplements 15-21, it is necessary to consult the Author Catalogue of the corresponding supplement for complete bibliographic information.

Commencing with Supplement 22, basic bibliographic information is included with each entry in Parts 2-7. It should be emphasized, however, that it will still be useful to consult the Author Catalogue for a variety of other information that may be found there: Title of the reference, translated title, language of text and summaries, issue date, library from which the original may be obtained, published corrections, related references by the same author, and other miscellaneous information.

Each supplement consists of the following parts:

Part 1, Authors: A-Z
Part 2, Parasite-Subject Catalogue: Parasites: Protozoa
Part 3, Parasite-Subject Catalogue: Parasites: Trematoda and Cestoda
Part 4, Parasite-Subject Catalogue: Parasites: Nematoda and Acanthocephala
Part 5, Parasite-Subject Catalogue: Parasites: Arthropoda and Miscellaneous Phyla
Part 6, Parasite-Subject Catalogue: Subject Headings and Treatment
Part 7, Parasite-Subject Catalogue: Hosts

Users should bear in mind that this is an Index-Catalogue, not a treatise, and should not expect to find reasons for any given entry. Nor does citing of synonymy mean that it is necessarily correct. The same statement holds for hosts, locations, localities, authorship of taxa, designation of new taxa, etc. These items are cited as given by the author(s) of the publication being indexed.

The information included in any given supplement represents only the publications that have been indexed in that supplement; and therefore, exclusion of, or limited entries for, any given author or parasite has no significance. No pretension is made for completeness, and assistance in correcting errors or obtaining additional information is appreciated. Reprints of papers on parasitology are requested.

recorded after each host name; when the hosts of a parasite are all from one locality, they are recorded as "all from" this locality.

(7) **Other Subject Matter:** Phrases indicate other subject matter discussed (e.g., immunity, metabolism, morphology, etc.).

Subject Headings Catalogue

The Subject Headings Catalogue (the first section of Part 6 of each supplement) is an alphabetic arrangement of entry terms from a controlled list of subject headings. Each entry consists of the subject heading, bibliographic information, and a subheading reflecting the information contained in the paper. Subject headings with numerous entries are separated into alphabetized subdivisions, e.g.,

Immunity
Immunity, Agglutination
Immunity, Allergy

Treatment Catalogue

In the Treatment Catalogue (a section of Part 6 of each supplement), all entries referring to one antiparasitic agent are grouped under one heading (regardless of the name used by the investigator) and are then listed alphabetically by author. Other names for the same agent are cross-referenced to the name used for filing. When generic and chemical names are available, preference is given to those names as headings rather than to trade names or code numbers and letters. Code number designations for compounds are entered in the Number Index in numerical order and cross-referenced to the name under which they are listed in the alphabetical section. Salts of a compound are usually grouped together, e.g., piperazine adiphate, piperazine citrate, etc., are all listed under Piperazine. Sometimes verifying synonymy of drug names is impossible; consequently, groupings and cross-references are not always authenticated although as many as possible have been checked with reliable sources. In some instances, the cross-references are based entirely on information in papers indexed and verification was not possible. Foreign language terminology has been anglicized where feasible. Chemosterilants, Molluscicides, and Repellents are entered under these three collective headings and not under the individual chemical. The format is the same as the parasite entries: Entry term (in this case, drug name), bibliographic information, and subheading.

Host Catalogue

The Host Catalogue (Part 7 of each supplement) is arranged alphabetically by genera, common names, and higher taxa and then alphabetically by species within genera. Nominate subspecies are interfiled with the species. Entries under each heading are in turn arranged alphabetically by author(s) and then chronologically for each author. The format is the same as in the other Catalogues, i.e., entry term (in this case, host name), bibliographic information, and subheading. Indented beneath the author line(s) of each host entry are all the parasites of a particular phylum that were reported from this host in the paper in question. Body locations of these parasites will be found in parentheses ( ) either in the subheading or with the host name. Experimental infection is reported as such. When there are multiple parasites and geographic localities, the appropriate locality is recorded after each parasite name; when the parasites from this host are all from one locality, they are recorded as "all from" this locality. When authors use only common names of hosts, scientific names are cautiously supplied from authoritative sources after careful consideration. Cross-references from the common name used by the author to the scientific name supplied by the Index-Catalogue are filed among the host entries. Such supplied names are given in square brackets [ ]. If a scientific name cannot be supplied, English common names are used. Scientific names or English common names are always supplied for common names in Cyrillic alphabet languages, and no cross-references are made. Surveys of parasites of humans and domestic animals are often indexed under geographic headings and entered in Part 6, Subject Headings, in addition to appearing in the Host Catalogue. In this case, all parasite phyla are grouped under the same host entry, and individual parasite entries are not included in the Parasite Catalogue.

EXPLANATORY NOTE

Author Catalogue

The Author Catalogue (Part 1 of each supplement) contains full bibliographic information for each publication indexed during the compilation of that supplement. A symbol for the library from which the original publication may be obtained is given at the end of each entry, e.g., Wa, Wm, Wc, etc. A key to these library symbols may be found in Supplements 10 and 20. A list of serial abbreviations new to our files is published at the beginning of each Author Catalogue.

Parasite Catalogues

The Parasite Catalogues (Parts 2-5 of each supplement) are divided by parasite phyla (Protozoa, Trematoda, etc.). They are arranged alphabetically by genera, parasitic diseases, and higher taxa and then alphabetically by species within genera. Entries under each heading are in turn arranged alphabetically by authors and then chronologically for each author. Each entry consists of the name of the parasite or parasitic disease, the author(s) of the publication, date, abbreviated title of the publication, volume, number, inclusive pages, and a subheading. Illustrations of parasites are indicated by the word illus. following the name of the parasite.

►Fasciola hepatica, illus.
►Smith, J.; and Doe, L., 1978, J. Parasitol., v. 64 (1), 30-38
►Fasciola hepatica, white mice, successful vaccination with culture incubate antigens and antigens from sonic disruption of immature worms
Subheading
Bibliographic information (author line(s))
Entry term

A variety of information is found indented beneath the author line(s) of each entry: Classification, hosts, synonymy, keys, treatment, etc. Subheadings are guides to the subject matter of the publication.

(1) **Classification:** In entries based on systematic articles, the subheading may give the higher taxa in which the taxon has been placed or it may list the lower taxa included in a higher taxon.

(2) **Hosts:** The only hosts recorded are those that pertain directly to the author's own work. Scientific host names are used unless the author gives only common names, in which case the host names are given exactly as in the original publication.

However, when host common names are in Cyrillic alphabet languages, host Latin names are assigned and listed instead of the common name; these are in square brackets [ ].

Locations of parasites in or on hosts are given in parentheses ( ). Where a host-parasite relationship is well known, a host may be given under a parasite name and not in the Host Catalogue; this applies particularly to parasites of medical and veterinary importance and of worldwide distribution. A + before the host name on the parasite entry means that no host entry was made for this particular reference.

(3) **Synonymy:** Usually only those synonyms which the author indicates as new, or which are new to the files of the Index-Catalogue of Medical and Veterinary Zoology, are given.

(4) **Keys:** The subheading "key" indicates that the name is included in a taxonomic key.

(5) **Treatment:** When there are several antiparasitic agents mentioned in a publication, a general term is used in the subheading, e.g., anthelmintics, insecticides, protozoacides. However, in the Treatment Catalogue, all agents tested by the investigator(s) are listed.

(6) **Geographic Distribution:** When there are multiple hosts and geographic localities, the appropriate locality is

Visitors are welcome to come to the Animal Parasitology Institute to use the cumulative files. Arrangements should be made in advance for lengthy visits.

All correspondence should be addressed to:

Index-Catalogue of Medical and Veterinary Zoology
Animal Parasitology Institute
USDA, SEA-AR, BARC-East, Building 1180
Beltsville, Maryland 20705 U.S.A.

It is hoped that these Catalogues will serve as a useful tool to workers in the field of parasitology. Users are requested to preserve the Catalogues, since they are not designed for general distribution and the edition is limited.

The compilers thank the staffs of the Technical Information Systems of the Science and Education Administration, the National Library of Medicine, and all other libraries who have aided us invaluably by making publications available to us.

Trade names are used in this publication solely for the purpose of providing specific information. Mention of a trade name does not constitute a guarantee or warranty of the product by the U.S. Department of Agriculture or an endorsement by the Department over other products not mentioned.

Acanthamoeba
Barrett, R. A.; and Alexander, M., 1977, Applied and Environment. Microbiol., v. 33 (3), 670-674
Acanthamoeba, susceptibility of cysts to microbial and enzymatic degradation in soil and in vitro

Acanthamoeba
Culbertson, C. G.; and Harper, K., 1977, Am. J. Clin. Path., v. 68 (4), 529-530 [Letter]
E[ntamoeba] histolytica, immunoperoxidase staining in formalin-fixed tissue, diagnostic differentiation from Acanthamoeba and other soil amoebas

Acanthamoeba-Hartmannella group
Lyons, T. B. III.; and Kapur, R., 1977, Applied and Environment. Microbiol., v. 33 (3), 551-555
limax amoebae in public swimming pools, concentration, correlations, and public health significance, preliminary tests for the human pathogen Naegleria fowleri were inconclusive: Albany, Schenectady, and Rensselaer counties, New York

Acanthamoeba
Sawyer, T. K.; Visvesvara, G. S.; and Harke, B. A., 1977, Science (4296), v. 196, 1324-1325
isolation of pathogenic Acanthamoeba from oceanic and brackish-water bottom sediments suggests that caution should be exercised in dumping of sewage and dredge spoils in previously unaffected areas: New York Bight; Baltimore Harbor, Maryland

Acanthamoeba sp., illus.
Martinez, A. J.; Fultz, D. G.; and Armstrong, C. E. V., 1976, Proc. 34. Ann. Meet. Electron Microsc. Soc. America (Miami Beach, Florida, Aug. 9-13), 226-227
amebic meningoencephalitides, electron microscopy, differential diagnosis, embedding and staining technique

Acanthamoeba sp., illus.
Ringsted, J.; et al., 1976, Am. J. Clin. Path., v. 66 (4), 723-730
disseminated amebic meningoencephalitis probably due to Acanthamoeba sp., visceral and subcutaneous amebic granulomas discovered at autopsy, child: Korea

Acanthamoeba sp.
Sawyer, T. K., 1975, Tr. Am. Micr. Soc., v. 94 (3), 437 [Abstract]
description of a heat-tolerant species from ocean sediment (except for demonstrable animal pathogenicity yet to be tested, the amoebae have all of the characteristics of pathogenic Acanthamoeba): southeast of Ambrose Light, New York Bight

Acanthamoeba sp. Lilly A-1 strain
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
as syn. of Acanthamoeba culbertsoni

Acanthamoeba sp.
Visvesvara, G. S.; Healy, G. R.; and Jones, D. B., 1976, J. Protozool., v. 23 (2), 18A [Abstract]
Acanthamoeba spp., in vitro activity of several chemotherapeutic agents

Acanthamoeba [sp.]
Warhurst, D. C.; Stamm, W. P.; and Phillips, E. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 279 [Demonstration]
Acanthamoeba isolated from eye of patient with corneal ulcer, characterization, chemotherapeutic tests, pathogenicity in mice, isolation from mouse brain

Acanthamoeba spp.
Warhurst, D. C.; and Thomas, S. C., 1975, J. Protozool., v. 22 (3), 57A [Abstract]
Acanthamoeba spp. isolated from corneal ulcers of a schoolteacher and a farmer

Acanthamoeba spp.
Willaert, E.; and Stevens, A. R., 1976, Path. Biol., v. 24 (2), 89-91
Acanthamoeba castellani, purified plasma membranes, elicited antisera assayed by immunoprecipitation and immunofluorescence methods, cross reaction with other species of Acanthamoeba; plasma membrane antisera may allow identification of species or even strains

Acanthamoeba astronyxis (Ray & Hayes)
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
Acanthamoeba, Naegleria, Hartmannella, comparative studies on free-living and pathogenic amebae: cyst structure; nutrition; protein composition; immunology; cell free plaques and other cytopathic effects; phospholipase liberation; sensitivity to amphotericin B

Acanthamoeba castellanii
Cursons, R. T. M.; and Brown, T. J., 1976, N. Zealand J. Marine and Freshwater Research, v. 10 (2), 245-262
pathogenic free-living amebae, identification, classification, agents of primary amebic meningo-encephalitis

Acanthamoeba castellanii
Duma, R. J.; and Finley, R., 1976, Antimicrob. Agents and Chemotherapy, v. 10 (2), 370-376
Naegleria fowleri, Acanthamoeba castellanii, A. polyphaga, in vitro susceptibility to variety of therapeutic agents, results suggest that amphotericin B is most effective agent against Naegleria but few agents are effective against Acanthamoeba

Acanthamoeba castellanii, illus.
Martinez, A. J.; Markowitz, S. M.; and Duma, R. J., 1975, J. Infect. Dis., v. 131 (6), 692-699
electronmicroscopic and clinicopathologic features of pneumonia and encephalitis caused by Acanthamoeba castellanii and Acanthamoeba polyphaga in exper. mice, comparisons with infections caused by Naegleria

Acanthamoeba castellanii (Ryan and Gresham strains)
Nagington, J.; and Richards, J. E., 1976, J. Clin. Path., v. 29 (7), 648-651
in vitro testing of 24 compounds (9 found promising) for possible cysticidal activity against Acanthamoeba polyphaga and A. castellanii (2 strains), comparison with results with Naegleria gruberi

Acanthamoeba castellanii, illus.
Stevens, A. R.; et al., 1977, J. Protozool., v. 24 (2), 316-324
Acanthamoeba castellanii, A. culbertsoni, isolation and purity of plasma membrane antigens (electron microscopy, assays of marker enzymes), antisera raised against these antigens tested against homologous and heterologous Acanthamoeba spp. in agglutination and immunofluorescence tests, results strongly indicate value of plasma membrane antisera for immunotaxonomy and immunodiagnosis of Acanthamoeba

Acanthamoeba castellanii
Stevens, A. R.; and Stein, S., 1977, J. Parasitol., v. 63 (1), 151-152
Acanthamoeba, Naegleria, pathogenic and nonpathogenic species (strains), susceptibility to lectin-induced agglutination

Acanthamoeba castellanii (Douglas), illus.
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
Acanthamoeba, Naegleria, Hartmannella, comparative studies on free-living and pathogenic amebae: cyst structure; nutrition; protein composition; immunology; cell free plaques and other cytopathic effects; phospholipase liberation; sensitivity to amphotericin B
Syn.: Acanthamoeba terricola Pussard

Acanthamoeba castellanii
Willaert, E.; and Stevens, A. R., 1976, J. Protozool., v. 23 (4), 22A [Abstract]
Acanthamoeba castellanii, A. culbertsoni, use of antibodies directed against cell surface membranes to distinguish pathogenic from nonpathogenic species

Acanthamoeba castellani
Willaert, E.; and Stevens, A. R., 1976, Path. Biol., v. 24 (2), 89-91
Acanthamoeba castellani, purified plasma membranes, elicited antisera assayed by immunoprecipitation and immunofluorescence methods, cross reaction with other species of Acanthamoeba; plasma membrane antisera may allow identification of species or even strains

Acanthamoeba castellanii
Willaert, E.; and Stevens, A. R., 1976, Path. Biol., v. 24 (8), 545-547
Acanthamoeba castellanii, A. culbertsoni, humans, fatal meningoencephalitis, case reports, indirect immunofluorescence diagnosis at post-mortems: Texas; Peru; Venezuela

Acanthamoeba comandoni Pussard
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
Acanthamoeba, Naegleria, Hartmannella, comparative studies on free-living and pathogenic amebae: cyst structure; nutrition; protein composition; immunology; cell free plaques and other cytopathic effects; phospholipase liberation; sensitivity to amphotericin B

Acanthamoeba culbertsonii
Allen, D. J., 1976, Proc. North Dakota Acad. Sc., v. 30, Part I, 1
Acanthamoeba culbertsonii, scanning electron microscopy, comparison of culture specimens with cells from infection, surface micro appendages, possible use of SEM in diagnosis

Acanthamoeba culbertsoni
Cursons, R. T. M.; and Brown, T. J., 1976, N. Zealand J. Marine and Freshwater Research, v. 10 (2), 245-262
pathogenic free-living amebae, identification, classification, agents of primary amebic meningo-encephalitis

Acanthamoeba culbertsoni
Dunnebacke, T. H.; and Schuster, F. L., 1974, J. Protozool., v. 21 (2), 327-329
presence of cytopathogenic agent that is filterable and passageable only in strains of Naegleria (free-living gruberi as well as pathogenic fowleri) not in Acanthamoeba (pathogenic or free-living)

Acanthamoeba culbertsoni, illus.
Martinez, A. J.; Fultz, D. G.; and Armstrong, C. E. V., 1976, Proc. 34. Ann. Meet. Electron Microsc. Soc. America (Miami Beach, Florida, Aug. 9-13), 226-227
amebic meningoencephalitides, electron microscopy, differential diagnosis, embedding and staining technique

Acanthamoeba culbertsoni, illus.
Proca-Ciobanu, M.; and Ionescu, M. D., 1975, Arch. Roumaines Path. Exper. et Microbiol., v. 34 (4), 329-340
Acanthamoeba culbertsoni, ultrastructural comparisons of strain grown in brain of experimentally infected mouse and same strain grown in axenic culture

Acanthamoeba culbertsoni, illus.
Sawyer, T. K.; Visvesvara, G. S.; and Harke, B. A., 1977, Science (4296), v. 196, 1324-1325
isolation from ocean sediment from sewage-spoil dump site, fatal to laboratory mice after infection by intranasal route: New York Bight apex near Ambrose light

Acanthamoeba culbertsoni, illus.
Stevens, A. R.; et al., 1977, J. Protozool., v. 24 (2), 316-324
Acanthamoeba castellanii, A. culbertsoni, isolation and purity of plasma membrane antigens (electron microscopy, assays of marker enzymes), antisera raised against these antigens tested against homologous and heterologous Acanthamoeba spp. in agglutination and immunofluorescence tests, results strongly indicate value of plasma membrane antisera for immunotaxonomy and immunodiagnosis of Acanthamoeba

Acanthamoeba culbertsoni
Stevens, A. R.; and Stein, S., 1977, J. Parasitol., v. 63 (1), 151-152
Acanthamoeba, Naegleria, pathogenic and nonpathogenic species (strains), susceptibility to lectin-induced agglutination

Acanthamoeba culbertsoni
Taylor, P. W., 1977, J. Parasitol., v. 63 (2), 232-237
failure to produce exper. infections in fresh-water fish

Acanthamoeba culbertsoni
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
Acanthamoeba, Naegleria, Hartmannella, comparative studies on free-living and pathogenic amebae: cyst structure; nutrition; protein composition; immunology; cell free plaques and other cytopathic effects; phospholipase liberation; sensitivity to amphotericin B
Syn.: Acanthamoeba sp. Lilly A-1 strain

Acanthamoeba culbertsoni
Visvesvara, G. S.; Healy, G. R.; and Jones, D. B., 1976, J. Protozool., v. 23 (2), 18A [Abstract]
Acanthamoeba spp., in vitro activity of several chemotherapeutic agents

Acanthamoeba culbertsoni
Willaert, E.; and Stevens, A. R., 1976, J. Protozool., v. 23 (4), 22A [Abstract]
Acanthamoeba castellanii, A. culbertsoni, use of antibodies directed against cell surface membranes to distinguish pathogenic from nonpathogenic species

Acanthamoeba culbertsoni
Willaert, E.; and Stevens, A. R., 1976, Path. Biol., v. 24 (8), 545-547
Acanthamoeba castellanii, A. culbertsoni, humans, fatal meningoencephalitis, case reports, indirect immunofluorescence diagnosis at post-mortems: Texas; Peru; Venezuela

Acanthamoeba culbertsoni
Wong, M. M.; Karr, S. L., jr.; and Balamuth, W. B., 1975, J. Parasitol., v. 61 (4), 682-690
Acanthamoeba culbertsoni, susceptibility in Old World monkeys, no disease resulting from intranasal or intravenous inoculation, fatal meningoencephalitis in 4 of 7 monkeys inoculated intrathecally, virulence and pathogenicity vary with cultural age of inoculum, size of inoculum, passages through animal tissues, and host-immune competence
Cercocebus atys (exper.)
Macaca nemestrina (exper.)
M. arctoides (exper.)
M. mulatta (exper.)

Acanthamoeba hatchetti n. sp., illus.
Sawyer, T. K.; Visvesvara, G. S.; and Harke, B. A., 1977, Science (4296), v. 196, 1324-1325
isolation from brackish-water sediments, fatal to laboratory mice after infection by intranasal route: Brewerton Channel, Baltimore Harbor, Maryland

Acanthamoeba polyphaga
Duma, R. J.; and Finley, R., 1976, Antimicrob. Agents and Chemotherapy, v. 10 (2), 370-376
Naegleria fowleri, Acanthamoeba castellanii, A. polyphaga, in vitro susceptibility to variety of therapeutic agents, results suggest that amphotericin B is most effective agent against Naegleria but few agents are effective against Acanthamoeba

Acanthamoeba polyphaga, illus.
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electronmicroscopic and clinicopathologic features of pneumonia and encephalitis caused by Acanthamoeba castellanii and Acanthamoeba polyphaga in exper. mice, comparisons with infections caused by Naegleria

Acanthamoeba polyphaga
Nagington, J.; and Richards, J. E., 1976, J. Clin. Path., v. 29 (7), 648-651
in vitro testing of 24 compounds (9 found promising) for possible cysticidal activity against Acanthamoeba polyphaga and A. castellanii (2 strains), comparison with results with Naegleria gruberi

Acanthamoeba polyphaga
Stevens, A. R.; et al., 1977, J. Protozool., v. 24 (2), 316-324
Acanthamoeba castellanii, A. culbertsoni, isolation and purity of plasma membrane antigens (electron microscopy, assays of marker enzymes), antisera raised against these antigens tested against homologous and heterologous Acanthamoeba spp. in agglutination and immunofluorescence tests, results strongly indicate value of plasma membrane antisera for immunotaxonomy and immunodiagnosis of Acanthamoeba

Acanthamoeba polyphaga, illus.
Taylor, P. W., 1977, J. Parasitol., v. 63 (2), 232-237
isolation and experimental infection of free-living amebae in freshwater fishes, only Acanthamoeba polyphaga elicited pathological lesions
Micropterus coosae (spleen, urinary bladder, gall bladder): Lake Chinabee, Alabama
Micropterus salmoides (spleen): Lake Chinabee, Alabama; Potomac River, Maryland
Morone saxatiles (gill): Auburn University Experimental Station
Ictalurus punctatus (gill): Auburn University Experimental Station
Carassius auratus (gill): Farmer, Kentucky
Salmo gairdneri (gill): Huntsville, Alabama
Cyprinus carpio (nat. and exper.) (blood, gills, spleen, liver, intestine, kidney, brain): Wetumpka, Alabama
Sarotherodon aureus (Tilapia aurea) (nat. and exper.) (gills, spleen, liver, intestine, kidney, brain): Lake Parker, Florida

Acanthamoeba polyphaga (Puschkarew)
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
Acanthamoeba, Naegleria, Hartmannella, comparative studies on free-living and pathogenic amebae: cyst structure; nutrition; protein composition; immunology; cell free plaques and other cytopathic effects; phospholipase liberation; sensitivity to amphotericin B

Acanthamoeba polyphaga
Visvesvara, G. S.; Healy, G. R.; and Jones, D. B., 1976, J. Protozool., v. 23 (2), 18A [Abstract]
Acanthamoeba spp., in vitro activity of several chemotherapeutic agents

Acanthamoeba rhysodes
Stevens, A. R.; et al., 1977, J. Protozool., v. 24 (2), 316-324
Acanthamoeba castellanii, A. culbertsoni, isolation and purity of plasma membrane antigens (electron microscopy, assays of marker enzymes), antisera raised against these antigens tested against homologous and heterologous Acanthamoeba spp. in agglutination and immunofluorescence tests, results strongly indicate value of plasma membrane antisera for immunotaxonomy and immunodiagnosis of Acanthamoeba

Acanthamoeba terricola Pussard
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
as syn. of Acanthamoeba castellanii (Douglas)

Acanthodiophrya n.gen.
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Hoplitophryidae, Radiophryinae; tod: A. almae n.g., n.sp.

Acanthodiophrya de Puytorac et Dragesco, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Acanthodiophryinae n. s. fam.

Acanthodiophrya almae n.g., n.sp., illus. (tod)
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Alma emini (tube digestif): Gabon

Acanthodiophrya almae de Puytorac et Dragesco, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Acanthodiophryinae n. s. fam.
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam.
includes: Acanthodiophrya

Acanthophrya Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Maupasellidae, Acanthophryinae n. s. fam.

Acanthophrya inermis (Stein, 1859) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Helobdella stagnalis

Acanthophryinae n. s. fam.
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Maupasellidae
includes: Acanthophrya

Acanthospora polymorpha Leger 1897, illus.
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
description
Hydrochara caraboides (region moyenne du mesenteron): Loire-Atlantique (Ouest de la France)

Actinocephalidae [sp.], indetermine, illus.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Taraxides laevigatus
T. moerens
all from Gabon

Actinocephalus dujardini Schneider, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
brief description
Lithobius forficatus (intestine): Poland

Actinocephalus echinatus, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Pterostichus niger

Actinocephalus permagnus
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Carabus cancellatus

Actinocephalus stelliformis Schneider, 1875
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Ancyrophora stelliformis (Schneider)

Actinocephalus tipulae
Carter, J. B., 1976, J. Applied Ecol., v. 13 (1), 103-122
Tipula paludosa
T. oleracea
T. czizeki
T. luna
T. unca
T. vernalis
T. variipennis
T. marmorata or alpium?
T. truncorum
all from north-east England

Acutispora macrocephala, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Lithobius forficatus

Adelina sp., illus.
Courtney, C. H.; et al., 1975, J. Parasitol., v. 61 (4), 695-699
description, "species found in the feces of sandhill cranes is probably a spurious parasite, and we consider it inappropriate to describe this parasite as a new species without knowing the identity of the true host."
Grus canadensis pratensis
G. canadensis tabida
(feces of all): all from Florida

Adelina sp.
Forrester, D. J.; Bush, A. O.; and Williams, L. E., jr., 1975, J. Parasitol., v. 61 (3), 547-548
Grus canadensis pratensis (feces): Florida

Adelina rayi n. sp., illus.
Narasimhamurti, C. C., 1977, Proc. Indian Acad. Sc., Sect. B, v. 86 (4), 239-244
Adelina rayi n. sp., sporogony
Rhysida longipes (gut): University Campus at Waltair

Adelina tribolii Bhatia, 1937
Zizka, Z., 1977, J. Protozool., v. 24 (2), Suppl., 17A-18A [Abstract]
Adelina tribolii microgametocytes, fine structure

Aegyptianella
Kreier, J. P.; and Gothe, R., 1976, Vet. Parasitol., v. 2 (1), 83-95
Aegyptianella, Eperythrozoon, Grahamella, Haemobartonella, morphology, host-parasite interactions, review

Aegyptianella
Kreier, J. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 24-28
Rickettsiales, Anaplasmataceae

Aegyptianella [sp.]
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Anas platyrhynchus (blood): Tanzania

Aegyptianella sp., illus.
Peirce, M. A.; and Bevan, B. J., 1977, Vet. Rec., v. 100 (14), 282-283
Amazona aestiva (blood): imported to London from Indonesia

Aegyptianella anseris
Peirce, M. A., 1973, J. Protozool., v. 20 (5), 543-546
Syn.: Babesiosoma anseris

Aegyptianella pullorum
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Aggregata eberthi
Vivier, E.; and Porchet, E., 1976, J. Protozool., v. 23 (4), 22A
Aggregata eberthi, Hepatozoon domerguei, Dehornia sthenelais, crystalline inclusions

Aikinetocystidae Bhatia, 1930
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
diagnosis; checklist of genera and species of 6 aseptate gregarine families

Aikinetocystis Gates, 1926
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Aikinetocystidae
diagnosis; checklist of species

Akiba Bennett, Garnham & Fallis, 1965
Bennett, G. F.; et al., 1975, J. Protozool., v. 22 (1), 24-30
extensive remarks on taxonomic and nomenclatural status of Leucocytozoon, classification (Haemosporina, Leucocytozoidae), emended generic diagnosis, proposal that authorship be credited to Sambon 1908 and that L. majoris (Laveran, 1902) be designated as type species (L. danilewskyi being a nomen nudum), rejection of recent synonymy of Akiba and Saurocytozoon with Leucocytozoon

Akiba Bennett, Garnham & Fallis, 1965
Hsu, C.-K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon Berestnev, 1904

Akiba [sp.]
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Basileuterus rivularis
Thryothorus nigricapillus
(blood of all): all from Colombia

Akiba or Leucocytozoon, unidentified sp., illus.
Walker, D.; and Garnham, P. C. C., 1972, Vet. Rec., v. 91 (3), 70, 71, 72 [Letter]
cysts in heart and gizzard of parakeets resembling megaloschizonts of Leucocytozoon or Akiba, description, clinical signs of disease, lesions, pathology, case report
Neophema pulchella: Surrey, England
Platycercus adelidae: Oxon., England
Alisterus scapularis: Cornwall, England
Polytelis anthopeplus: Glos., England
(heart of all)

Akiba caulleryi (Mathis & Leger, 1909) Bennett, Garnham & Fallis, 1965
Hsu, C.-K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon (Akiba) caulleryi Mathis & Leger, 1909

Akiba caulleryi
Manuel, M. F.; and Morales, E. G., 1975, Philippine J. Vet. Med., v. 13 (1-2), 1974, 147-155
Akiba caulleryi, chickens, clinical signs, infectivity and mortality rates, prophylactic value of sulfamonomethoxine administered in feed, good results; drug not effective against E[imeria] tenella

Akiba caulleryi
Skippon, J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 3 [Demonstration]
domestic chickens (blood): vicinity of Kota Kinabalu, Borneo

Akidodes Lom, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Jirovecellinae n. s. fam.

Akidodes henleae Lom, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Henlea nasuta

Akidodes symmetricus Lom, 1959, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Mesenchytraeus flavidus
Enchytraeus argenteus
Marionina sphagnetorum

Allantocystidae Bhatia, 1930
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
diagnosis; checklist of genera and species of 6 aseptate gregarine families

Allantocystis Keilin, 1920
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Allantocystidae
diagnosis; checklist of species

Amoeba coli, illus.
Loesch, F. A., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 383-392
first description of Amoeba coli with report of clinical aspects of recurrent human intestinal amoebiasis, English translation of German article published in 1875

Amoebae, resembling Acanthamoeba
Casemore, D. P., 1977, Lancet, London (8047), v. 2, 1078 [Letter]
amoebae resembling Acanthamoeba present in both main water supply and in kidney hemodialysis unit of man suffering from unexplained episodes of pyrexia with rigors, discussion as possible pathogen

Amoebae
Low, W. A., 1976, Canad. Field-Naturalist, v. 90 (2), 189-191
Rangifer tarandus caribou (feces): Tweedsmuir Provincial Park, British Columbia

Amoebae, limax
Lyons, T. B. III.; and Kapur, R., 1977, Applied and Environment. Microbiol., v. 33 (3), 551-555
limax amoebae in public swimming pools, concentration, correlations, and public health significance, preliminary tests for the human pathogen Naegleria fowleri were inconclusive: Albany, Schenectady, and Rensselaer counties, New York

Amoebae
Stamm, W. P.; and Parelkar, S. N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 10 [Demonstration]
amoebae in tissues can be specifically stained by indirect immunofluorescent technique, in some cases years after original fixation

Amoebae, possibly Hartmanellidae, illus.
Voelker, F. A.; et al., 1977, Vet. Path., v. 14 (3), 247-255
ultrastructure
Carassius auratus (kidneys, coelomic cavity, liver, skeletal muscle, meninges, swim bladder)

Amoebiasis
Alora, B. D.; Ramon, F. J.; and Tan-Alora, A., 1972, Philippine J. Int. Med., v. 10 (1), 40-47
human amoebiasis, radiologic visualization of hepatic abscess cavity after aspiration in order to assess size of cavity and response to therapy

Amoebiasis
Anderson, R. E.; and Varma, V. M., 1972, J. Nuclear Med., v. 13 (3), 217-218
human amoebiasis complicated by amoebic pericarditis, diagnosis by radioisotopes, case report: Louisiana (Vietnam veteran)

Amoebiasis
Andre, L. J., 1973, Medecine et Armees, v. 1 (6), 51-54
human amoebiasis, current treatment methods and clinical aspects, metronidazole, seconidazole

Amoebiasis
Andre, L. J.; and Charmot, G., 1973, Medecine Trop., v. 33 (3), 311-315
human amoebiasis, clinical trials with seconidazole and comparison with metronidazole, suitable for mass therapy in developing nations

Amoebiasis
Andre, L. J.; Pieri, F.; and Abed, L., 1973, Medecine Afrique Noire, v. 20 (12), 991-993
human amoebiasis, indications for use of metronidazole

Amoebiasis
Angate, Y.; et al., 1974, Medecine Afrique Noire, v. 21 (1), 61-65
symptoms of acute abdomen resulting from human intestinal parasites, medical and surgical care, case reports: Abidjan, Ivory Coast

Amoebiasis
Areekul, S.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 107 [Demonstration]
comparison of serum vitamin B_{12} levels in patients with hepatic amoebic abscess, opisthorchiasis or hookworm infections

Amoebiasis
Areekul, S.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 375-379
amoebiasis, opisthorchiasis, humans, changes in serum vitamin B_{12} in presence of infections

Amoebiasis
Areekul, S.; et al., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 154
humans, changes in ^{131}I-fibrinogen metabolism and fibrinolytic activity in patients with hepatic abscess

Amoebiasis
Areekul, S.; Devakul, K.; and Kanakakorn, K., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 414 [Demonstration]
hookworm, amoebiasis, humans, effects of anemia on iron absorption compared in infected and control subjects

Amoebiasis
Benex, J., 1972, Medecine et Malad. Infect., v. 2 (10), 351-357
quantitative immunofluorescence in serologic diagnosis of human parasitoses, guidelines for use

Amoebiasis
Berthoud, S.; Rime, F.; and Buffle, P., 1976, Schweiz. Med. Wchnschr., v. 106 (24), 828-830
hepatic amoebic abscess presenting with systemic symptoms in man who had traveled extensively in tropics, abscess masked by previous treatment of intestinal symptoms with iodoquinolines, diagnosed by liver scan and cured with metronidazole: Switzerland

Amoebiasis
Bertrand, E.; and Cornet, L., 1970, Medecine Afrique Noire, v. 17 (6), 435-441
human hepatic amoebiasis, use of liver puncture in treatment and diagnosis

Amoebiasis
Bhasin, A. S.; et al., 1977, J. Trop. Med. and Hyg., v. 80 (8), 169-172
human hepatic amoebic abscesses, use of micro-opaque barium sulfate to assess resolution time of abscesses, favorable comparison with scintiscanning

Amoebiasis
Bos, H. J.; van den Eijk, A. A.; and van der Kaay, H. J., 1977, Trop. and Geogr. Med., v. 29 (2), 204 [Abstract]
human amoebiasis, comparative evaluation of usefulness of enzyme-linked immunosorbent assay and counter immunoelectrophoresis in diagnosis

Amoebiasis
Bowry, T. S.; and Cameron, H. M., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 439-443
schistosomiasis, amoebiasis, causes of liver pathology in early childhood in Kenya

Amoebiasis
Brumpt, L., 1971, Medecin Paris (6), 7-10
clinical review of human intestinal amoebiasis, medical management and treatment

Amoebiasis
Bunnag, D.; et al., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 99-102
human hepatic amoebic abscess, successful clinical trials combining a single course of metronidazole with abscess aspirations: Bangkok, Thailand

Amoebiasis
Capron, A.; Niel, G.; and Bouvry, M., 1970, Medecine Afrique Noire, v. 17 (6), 457-461
review of immunological techniques used in diagnosing human amoebiasis

Amoebiasis
Carayon, A., 1975, Medecine Trop., v. 35 (2), 119-122
human severe acute amoebic colitis, amoebomas, surgical complications, clinical management

Amoebiasis
Carayon, A., 1975, Medecine Afrique Noire, v. 22 (3), 243-251
human severe amoebic colitis, complications, clinical management and treatment

Amoebiasis
Carayon, A.; and Bezes, H., 1975, Medecine Trop., v. 35 (2), 123-130
acute necrotic amoebic colitis, pathology, clinical management, surgical measures

Amoebiasis
Carayon, A.; and Gargaratchi, 1975, Médecine Trop., v. 35 (2), 169-173
pathologic sequellae after recovery from severe human amoebic colitis, case reports

Amoebiasis
Chmielewski, J.; Jaremin, B.; and Zdrojewski, J., 1974, Polskie Arch. Med. Wewn., v. 51 (1), 91-94
human amoebiasis, hepato-pulmonary-pleural form, case report, diagnosis based on gel-precipitation test, combined therapy of metronidazole, dihydroemetine and oxytetracycline successful: Hindu sailor in Poland

Amoebiasis
Cordero A., O., 1974, Neumol. y Cirug. Torax, v. 35 (2), 83-89
human amoebiasis, thoracic complications of hepatic abscess, clinical aspects, surgical management, diagnosis

Amoebiasis
Cornet, L.; et al., 1975, Medecine Trop., v. 35 (2), 135-147
human malignant amoebic colitis, case reports, clinical and pathologic findings, surgical intervention

Amoebiasis
Dale, D. C.; and Petersdorf, R. G., 1973, Med. Clin. North Am., v. 57 (5), 1277-1287
parasitic infections in the immuno-suppressed steroid-treated patient, review

Amoebiasis
Darmawan, S.; and Sudjarwo, S. R., 1971, Paediat. Indonesiana, v. 11 (4), 167-172
amoebiasis, fatal intestinal perforation due to infection in 8-month-old child, case report: Indonesia

Amoebiasis
Devic, J.; and Dosen, H., 1974, Med. Pregl., v. 27 (1-2), 79-83
human intestinal amoebiasis, clinical trials comparing 2-dehydroemetine with resotren in treatment: Yugoslavia

Amoebiasis
Dharmalingam, S. K.; and Mahadev, V., 1970, Med. J. Malaya, v. 25 (2), 83-90
radioisotope color scanning of the liver in diagnosis of human hepatic amoebic abscess

Amoebiasis
Diaz M., G. S.; et al., 1971, Neumol. y Cirug. Torax, v. 32 (6), 393-403
human hepatic amoebic abscess with resulting amoebic pericarditis, need for early diagnosis stressed: Mexico

Amoebiasis
Donaldson, M.; et al., 1975, Am. J. Vet. Research, v. 36 (6), 807-817
reptilian amebiasis, fatal epizootic, possible cause of many previous snake deaths, pathologic changes, probable mode of introduction and spread, treatment (metronidazole successful) and prophylaxis, amebae identified as or presumed to be Entamoeba invadens: Steinhart Aquarium, California Academy of Sciences, Golden Gate Park, San Francisco

Amoebiasis
Douetteau, C.; Attia, Y.; and Bouvry, M., 1970, Medecine Afrique Noire, v. 17 (6), 463-465
clinical use of bromosulphaphthalene test in course of human hepatic amoebiasis, test findings

Amoebiasis
Dournovo, P.; et al., 1976, Nouv. Presse. Med., v. 5 (34), 2237-2239
human hepatic amoebic abscess, clinical cases, current status of therapy

Amoebiasis
Doust, B. D., 1976, Gastroenterology, v. 70 (4), 602-610
use of ultrasound to diagnose and assess healing process of human amoebic liver abscess

Amoebiasis
Dritz, S. K.; et al., 1977, Lancet, London (8027), v. 2, 3-4
human amoebiasis, increasing incidence of sexually transmitted enteric infections in persons with no common food source, public health efforts to control spread and treat infections have been unsuccessful: San Francisco, California

Amoebiasis
Duflo, B., 1975, Medecine Interne, v. 10 (10), 447-453
human cardiac complications of tropical parasitoses, pathologic findings

Amoebiasis
Durosoir, J. L.; et al., 1975, Medecine Trop., v. 35 (6), 457-462
use of peroxidase labelled antiglobulins in serological diagnosis of human amoebiasis, schistosomiasis, toxoplasmosis

Amoebiasis
Durosoir, J. L.; Thabaut, A.; and Laverdant, C., 1974, Nouv. Presse Med., v. 3 (23), 1507 [Letter]
amoebiasis, toxoplasmosis, schistosomiasis in humans, application of enzyme labelling with peroxidase for immunodiagnosis

Amoebiasis
Felgner, P.; and Lederer, I., 1976, Tropenmed. u. Parasitol., v. 27 (2), 165-168
comparative study using complement fixation, indirect agglutination and latex agglutination tests in immunodiagnosis of amoebiasis (differently prepared antigens, specific recommendations for latex agglutination, long-term storage of sensitized sheep red cells and temperature variations)

Amoebiasis
Felix, H.; Depinay, R.; and Maillard, A., 1970, Medecine Afrique Noire, v. 17 (6), 443-447
oral administration of dehydroemetine in treatment of human amoebiasis, extensive clinical trials

Amoebiasis
Flores Barroeta, F.; et al., 1975, Patologia, v. 13 (1), 17-35
human amoebiasis and cysticercosis, value and statistics of postmortem studies for previously undiagnosed infections: Mexico

Amoebiasis
Franco Ramirez, G.; Perez Norzagaray, J.; and de Saade, M. T., 1970, Pediatria, Bogota, v. 11 (4), 379-384
amoebiasis in nursing infants causing acute diarrhea, established as clinically distinct form from that of adults, diagnosis, pathology, clinical aspects: Colombia

Amoebiasis
Gastaut, J. A.; Ranque, P.; and Quilici, M., 1972, Marseille Med., v. 109 (10), 627-629
amoebiasis, human, diagnosis using immunofluorescence or agglutination, comparison trials

Amoebiasis
Gaxiola, R., 1969, Prensa Med. Mexicana, v. 34 (11-12), 427-437
human hepatic amoebic abscess, historical review of incidence in Mexico

Amoebiasis
Geslien, G. E.; Thrall, J. H.; and Johnson, M. C., 1974, J. Nuclear Med., v. 15 (7), 561-563
human hepatic amoebic abscess, gallium scanning to assess size and resolution of acute lesions

Amoebiasis
Goulston, S. J. M.; and McGovern, V. J., 1972, Med. J. Australia, v. 1 (24), 1234-1238
use of rectal biopsy in differential diagnosis of human intestinal amoebiasis

Amoebiasis
Gruet, M.; et al., 1973, Medecine et Armees, v. 1 (3), 5-10
human hepatic amoebiasis, use of scintigraphy to diagnose amoebic abscess and evaluate need for surgical intervention

Amoebiasis
Haff, R. C.; and Norgaard, R. P., 1974, Mil. Med., v. 139 (3), 192-195
human amoebiasis, metronidazole treatment of cecal amebomas and hepatic abscesses after locating by liver scan, military personnel returning from duty in Southeast Asia

Amoebiasis
Haider, Z.; Rasul, A.; and un-Din, F., 1974, Pakistan J. Med. Research, v. 13 (2-3), 39-42
human hepatic and intestinal amoebiasis, diagnosis using latex agglutination test

Amoebiasis
Ibarra-Perez, C., 1972, Prensa Med. Mexicana, v. 37 (7-8), 242-245
human hepatic amoebic abscess, case reports, of abscess rupture into pericardial cavity, diagnostic problems, clinical aspects: Mexico

Amoebiasis
Jarpa, A., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 96-102
statistical analysis of clinical records of 85 patients with hepatic amoebiasis abscesses: Santiago, Chile

Amoebiasis
Jo Kian Tjaij; Raid, N.; and Sutanto, A. H., 1970, Paediat. Indonesiana, v. 10 (4), 139-148
amoebic dysentery in children, clinical trials evaluating dehydroemetine

Amoebiasis
Jo Kian Tjaij; Raid, N.; and Sutanto, A. H., 1971, Paediat. Indonesiana, v. 11 (1), 1-12
intestinal amoebiasis in children, metronidazole the drug of choice: North Sumatra

Amoebiasis
Jo Kian Tjaij; Sutanto, A. H.; and Simatupang, J., 1976, Paediat. Indonesiana, v. 16 (9-10), 412-414
amoebic dysentery in children, clinical trials with metronidazole, recommendations for single dose therapy especially in poor, developing areas after evaluation of long-term results: Indonesia

Amoebiasis
Kaude, J.; and DeLand, F., 1975, Med. Clin. North Am., v. 59 (1), 145-167
X-ray and scintigraphy diagnosis of human hepatomegaly resulting from amoebiasis

Amoebiasis
Kim, C. W., 1975, Progr. Clin. Path., v. 6, 267-288
extensive review of techniques used to diagnose human parasitic diseases

Amoebiasis
Lamont, A. C.; and Wicks, A. C. B., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 302-305
amoebic liver abscess, review of 88 patients, clinical aspects including some atypical presentations, laboratory investigations, need for diagnostic awareness in young age groups, metronidazole drug of choice with repeated aspirations for large abscesses: Rhodesia

Amoebiasis
Lane, D., 1969, Med. J. Australia, v. 1 (15), 764-767
differential diagnosis of human hepatic amoebic abscess from primary hepatic carcinoma

Amoebiasis
Leuschner, U., 1972, Medicina Alemana, v. 13 (6), 903-911
clinical review of diagnosis and treatment of human hepatic amoebic abscess

Amoebiasis
Lin, C. S.; and Schwartz, I. S., 1976, N. York State J. Med., v. 76 (4), 574-578
human invasive amoebiasis, clinical case review of recent cases diagnosed in non-endemic area, need for prompt diagnosis and treatment: New York City

Amoebiasis
McCort, J. J., 1976, West. J. Med., San Francisco, v. 124 (5), 426-428
diagnosis of human hepatic amoebic abscess using total body opacification and tomography

Amoebiasis
Makoni, S. H. S.; and Sugden, J. K., 1976, Pharm. Acta Helvetiae, v. 51 (6), 153-158
discussion and classification of antiamoebic agents on pharmaceutical basis

Amoebiasis
Maldonado Ramirez, H.; et al., 1972, Neumol. y Cirug. Torax, v. 33 (1), 19-24
human amoebic hepatic abscess with associated amoebic pericarditis and tamponage, surgical management of 3 cases: Mexico

Amoebiasis
Oropeza Martinez, G.; and Maldonado Campos, F., 1976, Neumol. y Cirug. Torax, v. 37 (2), 103-109
human amoebic pericarditis with resulting cardiac tamponage, clinical case successfully treated surgically: Mexico

Amoebiasis
Mannweiler, E.; et al., 1976, Deutsche Med. Wchnschr., v. 101 (52), 1915-1919
human amoebic hepatic abscesses, comparative seroimmunologic tests for antibody presence in infected persons, controls and persons from endemic areas show multiple tests needed for accurate diagnosis

Amoebiasis
Mansueto, S.; and Cucchiara, E., 1974, Policlin., Roma. Sez. Med., v. 81 (2), 70-84
human hepatic amoebic abscess with transdiaphragmatic fistula into lung, clinical case report: Italy

Amoebiasis
Manzano, C.; and Barrera, J. L., 1977, Pediat. Radiol., v. 5 (3), 148-153
intestinal amoebiasis in young child, associated invasion of skin and perineum with resulting stenosis of the anus and secondary megacolon, diagnosis by X-ray

Amoebiasis
Meeroff, M., 1968, Bol. Chileno Parasitol., v. 23 (1-2), 45-47
human intestinal amoebiasis, clinical trials using erythromycin stearate in varying doses

Amoebiasis
Miyamoto, A. T.; Thadepalli, H.; and Mishkin, F. S., 1974, N. England J. Med., v. 291 (25), 1363 [Letter]
human hepatic amoebic abscess, ^{67}gallium images of gallium citrate scan used in diagnosis

Amoebiasis
Mohr, W., 1976, Med. Klin., Berlin, v. 71, 1204-1209
human protozoal infections, value of immunoserologic techniques in diagnosis, comparison with results of direct blood examination and culture methods

Amoebiasis
Monteiro, B.; and Deroeck, E., 1975, Medecine Afrique Noire, v. 22 (8-9), 601-602
woman, amoebic pleuro-pneumonia, case report: Kinshasa, Zaire

Amoebiasis
Morales Mareles, P.; Suarez Sanchez, F.; and Boom, R. A., 1975, Prensa Med. Mexicana, v. 40 (3-4), 124-126
human hepatic amoebic abscess, double blind clinical trials comparing therapy with intravenous metronidazole and intramuscular emetine: Mexico

Amoebiasis
Most, H., 1972, N. England J. Med., v. 287 (10), 495-498; (14), 698-702
common parasitic infections of man encountered in the United States, recommendations for treatment, review

Amoebiasis
Moulin, F.; Riviere, C.; and Grisez, J., 1975, Medecine Trop., v. 35 (2), 149-156
acute amoebic necrotic colitis, case reports, possible evolution, pathology, clinical management: Brazzaville

Amoebiasis
Munabe, K. K.; et al., 1975, Mater. Med. Pol. (22), v. 7 (1), 41-46
amoebiasis, locations of human hepatic abscess, clinical picture and differentiation from various other hepatic pathologies

Amoebiasis
Nava, C.; Metlich, M. A.; and Marti, M., 1973, Medicina, Mexico (1160), an. 54, v. 53, 351-353
successful treatment of 22 persons with amoebic colitis using tinidazole, few side effects: Mexico

Amoebiasis
Nordlinger, B., 1975, Medecine Trop., v. 35 (2), 131-138
acute severe amoebic colitis, clinical and pathologic review, surgical aspects

Amoebiasis
Ognibene, A. J., 1970, Med. Ann. Dist. Columbia, v. 39 (7), 352-355
emerging medical problems associated with U.S. army troops, clinical management and prophylaxis: Vietnam

Amoebiasis
Palinacci, C.; Ramiara, Y.; and Laroche, R., 1972, Medecine Trop., v. 32 (6), 755-757
appendectomy in man with amoebiasis resulting in cecal fistula and cutaneous amoebiasis, case report, cure with metronidazole

Amoebiasis
Parrish, R. A., jr.; and Still, J., 1970, Med. Times, New York, v. 98 (2), 157-160
amoebic abscess of liver in children, diagnostic problems, clinical management: Georgia

Amoebiasis
Pastore, R. A., 1970, Mil. Med., v. 135 (6), 476-478
human amoebic hepatic abscess, use of percutaneous catheter drainage, case report: Denver (soldier returned from Vietnam)

Amoebiasis
Payet, M., 1975, Medecine Trop., v. 35 (2), 105-110
human malignant amoebic colitis, clinical aspects, pathology, diagnosis, treatment with dehydroemetine or surgery

Amoebiasis
Payet, M.; Sankale, M.; and Ancelle, J. P., 1970, Medecine Afrique Noire, v. 17 (3), 219-224
medical management of amoebic hepatic abscesses, case reports, evaluation of treatment using scintography

Amoebiasis
Pene, P.; Bouvry, M.; and Pignol, F., 1970, Medecine Afrique Noire, v. 17 (3), 213-217
comparison of dehydroemetine, niridazole and metronidazole in treatment of human amoebiasis

Amoebiasis
Pignol, F.; et al., 1970, Medecine Afrique Noire, v. 17 (6), 449-455
use of indirect immunofluorescence in diagnosis of human thoracic amoebiasis, review

Amoebiasis, illus.
Pignol, F.; Colonna, J.; and Gallais, H., 1975, Nouv. Presse. Med., v. 4 (42), 3007 [Letter]
human intestinal amoebiasis, case report of chronic malignant colitis cured by right hemicolectomy with resection of terminal ileus: France (had resided in Morocco)

Amoebiasis
Pignol, F.; Colonna, J.; and Gallais, H., 1975, Medecine et Malad. Infect., v. 5 (12), 579-582
human malignant amoebic colitis, diagnostic difficulties, case report, clinical aspects

Amoebiasis
Radermecker, M.; et al., 1974, Internat. Arch. Allergy and Applied Immunol., v. 47 (2), 285-295
various human helminthic or protozoal infections, serum IgE concentration, IgE level often raised in parasitosis with prominent tissue phases and remains normal with helminths restricted to lumen of digestive tract, IgE level tends to increase significantly and rapidly following specific treatment and then to decrease slowly and return to normal in a few months

Amoebiasis
Ramachandran, S., 1974, Postgrad. Med. J., London (589), v. 50, 689-693
human hepatic amoebiasis, radiologic changes in left lobe amoebic liver abscess provides adequate diagnostic confirmation

Amoebiasis
Ramanathan, P.; Ganatra, R. D.; and Blau, M., 1974, J. Nuclear Med., v. 15 (11), 1021-1024
human hepatic echinococcosis and amoebiasis, dynamic bloodflow studies using scintillation cameras, differential diagnosis of benign and malignant lesions

Amoebiasis
Ramirez Sanchez, C.; and de Cisneros Santos, T. A., 1976, Medicina, Mexico (1212), an. 57, v. 56, 279-282
amoebiasis, human, surgical treatment of hepatic abscesses that had responded poorly to previous metronidazole and emetine therapy: Puebla, Mexico

Amoebiasis
Rao, A. P.; and Rao, H. K., 1974, J. Mysore Med. Ass., v. 39, 1-5
human infection, statistical incidence survey: Mangalore

Amoebiasis
Ravi, V. V.; et al., 1975, Indian J. Med. Research, v. 63 (12), 1732-1736
titers of indirect hemagglutination test and complement levels compared using sera and abscess pus from patients infected with hepatic amoebic abscesses, sero-negative and sero-positive cases investigated for possible differences in immune responses

Amoebiasis
Reynaud, R., 1973, Maroc Med. (570), v. 53, 421-424
human infection, review of clinical forms, medical and surgical management

Amoebiasis
Rieck, M.; and Halter, F., 1972, Minerva Med., v. 63 (58), 3166-3168
human rectal amoebiasis presenting as solitary ulcer, differential diagnosis

Amoebiasis
Rivera Reyes, H. H.; Rodriguez Trapala, A.; and Nieto de Pascual, R. H., 1975, Neumol. y Cirug. Torax, v. 36 (2), 91-98
human amoebic pericarditis, case report, clinical aspects followed by electrocardiograph: Mexico

Amoebiasis
Rodriguez Diaz, E.; and Renteria Cardenas, A. A., 1974, Bol. Med. Hosp. Inf., v. 31 (2), 321-331
hepatic amoebiasis, child, complicated by portal hypertension, case report: Mexico

Amoebiasis
Rungs, H., 1969, Maroc Med. (521), v. 49, 113-115
human amoebic chronic colitis, malabsorption and avitaminosis, clinical review

Amoebiasis
Sankale, M., 1973, Medecine Afrique Noire, v. 20 (12), 959-973
human amoebiasis, clinical indications for therapy, drugs in current use, review

Amoebiasis
Sankale, M.; et al., 1970, Medecine Afrique Noire, v. 17 (6), 479-489
use of scintigraphy in diagnosis and evaluation of course of human hepatic amoebiasis

Amoebiasis
Sankale, M.; et al., 1975, Medecine Trop., v. 35 (2), 111-117
pathological and clinical study of human malignant amoebic colitis

Amoebiasis
Sankale, M.; Diop, B.; and Diouf, S., 1970, Medecine Afrique Noire, v. 17 (6), 467-477
statistical review of 223 cases of hepatic amoebiasis: Dakar

Amoebiasis
Saugrain, J., 1971, Medecine Trop., v. 31 (2), 233-236
filariasis, Angiostrongylus cantonensis, amoebiasis, human parasitic diseases of medical interest and their etiology: French Polynesia

Amoebiasis
Savanat, T.; et al., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (2), 149-154
determinations of total serum IgE levels in humans with amoebic liver abscess or other parasitic infections

Amoebiasis
Selman Lama, M.; et al., 1976, Prensa Med. Mexicana, v. 41 (7-8), 223-225
human hepatic amoebic abscesses, pleural and thoracic complications, case reviews: Mexico

Amoebiasis
Shozawa, T.; et al., 1972, Nettai Igaku (Trop. Med.), v. 14 (4), 165-175
human amoebiasis, pathologic and clinical findings, correlation of results of 10 autopsies: Japan and Kenya

Amoebiasis
Stamm, W. P.; Ashley, M. J.; and Bell, K., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 49-53
low endemicity area of human amoebiasis, assessment of indirect haemagglutination, latex agglutination, indirect fluorescent antibody test and gel diffusion precipitin test for diagnostic serology

Amoebiasis
Stamm, W. P.; Ashley, M. J.; and Parelkar, S. N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 211-213
amoebiasis, human, diagnosis, commercially available latex agglutination test marketed in kit form as Serameba, compared with gel diffusion precipitin, fluorescent antibody, and indirect haemagglutination tests

Amoebiasis
Szarfman, A.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 19-24
evaluation of specificity of EVI factor observed in Trypanosoma cruzi cases using indirect fluorescent antibody test on sera from patients with other parasitic diseases, immunofluorescence of anti-skeletal muscle antibody from leishmaniasis

Amoebiasis
Tubis, M.; et al., 1975, Nuclear-Med., v. 14 (2), 163-171
human amoebiasis, labeled metronidazoles (bromometronidazole and technetium-penicillamine-flagyl complex) as potential agents for scintigraphic visualization of hepatic abscess

Amoebiasis
Valdez Ochoa, S.; et al., 1976, Prensa Med. Mexicana, v. 41 (5-6), 159-162
human hepatic amoebic abscess complicated by erosion into bronchus or pleural cavity, clinical management: Mexico

Amoebiasis
Weise, H. J., 1976, Munchen. Med. Wchnschr., v. 118 (35), 1061-1068
human exotic parasitic diseases, risks to tourists travelling in tropical countries, prophylactic and control measures, need for improved diagnosis

Amoebiasis
Woodruff, A. W., 1971, Brit. J. Clin. Pract., v. 25 (12), 529-535
achievements in tropical medicine during past 25 years, control, prophylaxis, treatment

Amoebiasis
Zerolo, J.; et al., 1974, Marseille Med., v. 111 (3), 153-156
man, hepatic hydatid cyst in association with hepatic amoebiasis, case report, clinical aspects: France

Amoebiasis
Zuberi, S. J.; and Panjvani, Z., 1973, Pakistan J. Med. Research, v. 12 (3), 10-14
human amoebiasis, differential diagnosis of causes of diarrhea with and without fecal blood, comparison study of direct stool examination, sigmoidoscopy and rectal biopsy

Anaplasma
Buening, G. M., 1977, Am. J. Vet. Research, v. 38 (1), 137-138
Anaplasma, 2-mercaptoethanol enhances in vitro response of sensitized bovine peripheral blood lymphocytes to Anaplasma antigen

Anaplasma
Kreier, J. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 24-28
Rickettsiales, Anaplasmataceae

Anaplasma, illus.
Ristic, M., 1976, Vet. Parasitol., v. 2 (1), 31-47
intracellular blood protista: intraerythrocytic behavior, transfer, and circulatory clearance; survival and development within macrophages; persistence of organism in immunologically hostile host; immune responses and protection; serodiagnosis; vaccination

Anaplasma [sp.]
Loehr, K. F.; and Meyer, H., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 192-197
Anaplasma organisms isolated in splenectomized calves following blood inoculation from Connochaetes taurinus, Alcelaphus buselaphus cokii, and Gazella thomsonii, morphologically indistinguishable from Aepyceros melampus, mild reaction of receptor calves to antelope-derived infection and severe reaction to subsequent A. melampus challenge, indirect fluorescent antibody results established close antigenic relationship between antelope anaplasms and A. melampus and less so to Anaplasma centrale: Kenya

Anaplasma [sp.]
Mohammed, A. N.; and Leeflang, P., 1975, Samaru Agric. Newsletter, v. 17 (2), 75-77
cattle (blood): Shika, Nigeria

Anaplasma centrale
Callow, L. L., 1976, World Animal Rev. (18), 9-15
Anaplasma centrale, Babesia argentina, B. bigemina, cattle, factors in transmission by Boophilus microplus, immunization with strains from splenectomized calves, review: Australia

Anaplasma centrale, illus.
Gallo, C.; and Balbo, S. M., 1976, Riv. Zootecn. e Vet. (3), 229-245
classification, morphology, epizootiology, transmission, vectors, pathogenicity, immunity, diagnosis, prophylaxis, review

Anaplasma centrale
Kuttler, K. L.; and Todorovic, R. A., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 106-112
Anaplasma marginale, cattle, premunization, Gloxazone and Imidocarb superior to oxytetracycline in moderating premunizing infection, possible factors affecting success (age of host; virulence, size, and potency of premunizing inoculum; strain or size of challenge exposure; temperature and altitude)

Anaplasma centrale
Loehr, K. F.; and Meyer, H., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 192-197
Anaplasma organisms isolated in splenectomized calves following blood inoculation from Connochaetes taurinus, Alcelaphus buselaphus cokii, and Gazella thomsonii, morphologically indistinguishable from Aepyceros melampus, mild reaction of receptor calves to antelope-derived infection and severe reaction to subsequent A. melampus challenge, indirect fluorescent antibody results established close antigenic relationship between antelope anaplasms and A. melampus and less so to Anaplasma centrale: Kenya

Anaplasma centrale
Loehr, K. F.; Ross, J. P. J.; and Meyer, H., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 86-95
Anaplasma marginale, A. centrale, cattle, homologous and heterologous indirect fluorescent antibody and capillary tube agglutination responses to primary infection and reinfection and to cross-infection with the heterologous organisms, clinical reactions; A. centrale carrier animals showed high degree of premunity to severe challenge with A. marginale

Anaplasma centrale
Mehlitz, D.; and Ehret, R., 1974, Tropenmed. u. Parasitol., v. 25 (1), 3-10
Anaplasma, Babesia, Theileria, cattle, serological survey, capillary-tube agglutination test, indirect fluorescent antibody test, and complement fixation test: Botswana

Anaplasma centrale
Parker, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 387-390
Babesia argentina, direct counting technique for estimating high parasitaemias, also suitable for B. bigemina and Plasmodium berghei but not for B. rodhaini or Anaplasma centrale

Anaplasma centrale
Ranatunga, P.; and Wanduragala, L., 1972, Brit. Vet. J., v. 128 (1), 9-18
Babesia bigemina, Anaplasma centrale, premunization of Jersey cattle imported to Ceylon from New Zealand, reactions and haematology

Anaplasma centrale
Weilgama, D. J.; Jayasekara, M. U.; and Hussain, M., 1975, Ceylon Vet. J., v. 23 (3-4), 49-53
Babesia bigemina, B. argentina, Anaplasma centrale, A. marginale, three breeds of cattle (Ayrshire, Friesian and Shorthorn), haematological changes including parasitaemia and temperature reactions following premunisation with blood from infected cattle: Sri Lanka, imported from New Zealand

Anaplasma marginale
Amerault, T. E., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 133-135
Anaplasma marginale, cattle, diagnosis, review of card test (nature and production of test antigen; card test procedure for plasma and serum; comparison of card test, complement fixation test, and calf inoculation results)

Anaplasma marginale
Amerault, T. E.; and Roby, T. O., 1977, World Animal Rev. (22), 34-38
card test as an accurate and simple procedure for detecting Anaplasma marginale in cattle, possible use for determining wildlife reservoirs of anaplasmosis and detecting babesiosis, review

Anaplasma marginale
Amerault, T. E.; Roby, T. O.; and Sealock, R. L., 1975, Am. J. Vet. Research, v. 36 (10), 1515-1519
Anaplasma marginale, ultrastructure after freeze-fracturing, comparison with thin sectioning and staining

A[naplasma] marginale
Artemenko, L. P.; and Ponomarenko, V. Ia., 1975, Veterinariia, Kiev (41), 68-71
A[naplasma] marginale, seasonal distribution of acute disease in cattle (May to June) coincides with high incidence of mosquitoes biting cattle, possible vectors: Western Polesia, Ukraine

Anaplasma marginale
Bauer, F.; and Hochheimer, N., 1975, Cahiers Bleus Vet. (25), 156-158
Anaplasma marginale, cows, Berenil

Anaplasma marginale
Bear, P. D., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 16-18
Anaplasma marginale, biochemical and immunological nature, brief review: nucleic acids; opsonins and autoimmunity

Anaplasma marginale, illus.
Bear, P. D., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 21-23
Anaplasma marginale, observations of "tailed" forms

Anaplasma marginale
Bedell, D. M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 59-60
Anaplasma marginale, pathogenesis, brief review of recent work

Anaplasma marginale
Billups, M. E.; Johns, R. W.; and Dimopoullos, G. T., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 19-21
Anaplasma marginale-infected calf erythrocytes, amino acid and nucleic acid metabolism

Anaplasma marginale
Boulanger, P.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 129-133
Anaplasma marginale, comparison of 2 complement fixation test methods (U.S. Dept. Agriculture vs. Canadian Animal Dis. Research Inst.), survey of cattle population of Southern Alberta in 1966 revealed no active infection, control of first Canadian outbreak in Manitoba in 1968 by serological identification and slaughter, surveillance exercised since 1970: Canada

Anaplasma marginale, illus.
Budden, J. R.; and Dimopoullos, G. T., 1977, Am. J. Vet. Research, v. 38 (5), 633-636
Anaplasma marginale, finite purification, antibody in anaplasmosis detected by agglutination tests is directed against erythrocytic stromata, not against finitely purified anaplasma bodies

Anaplasma marginale
Buening, G. M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 71-76
Anaplasma marginale, calves (intact, splenectomized, vaccinated, unvaccinated), comparison of results in leukocyte migration inhibition test, complement fixation test, and intradermic skin tests

Anaplasma marginale
Buening, G. M., 1976, Am. J. Vet. Research, v. 37 (10), 1215-1218
Anaplasma marginale, cell-mediated immune response in splenectomized and intact calves, micro cell-mediated cytotoxicity assay and leukocyte migration-inhibiton test compared, no correlation between in vitro assay systems

Anaplasma marginale
Callow, L. L.; Quiroga, Q. C.; and McCosker, P. J., 1976, Internat. J. Parasitol., v. 6 (4), 307-310
Babesia argentina, Anaplasma marginale, comparison of strains of each from Australia vs. Bolivia with indirect fluorescent antibody test showed serological identity of the two strains of each parasite, implications for vaccination; since earlier study showed serological identity between B. bovis and B. argentina, the small Babesia of Australia and South America should by priority be called B. bovis

Anaplasma marginale Theiler, 1910
Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
Anaplasma marginale, incidence in Syncerus caffer (blood), level of parasitemia in different age groups: Ngamiland, Botswana

Anaplasma marginale
Carson, C. A.; Sells, D. M.; and Ristic, M., 1976, Am. J. Vet. Research, v. 37 (9), 1059-1063
Anaplasma marginale, effect of blood group substances vs. parasitic components on induction of delayed cutaneous hypersensitivity and production of isoagglutinins in cattle injected with live or inactivated parasites in ovine or bovine erythrocytes, results indicate that inactivated sheep origin vaccine may avoid eliciting neonatal isoerythrolysis syndrome in calves from vaccinated dams

Anaplasma marginale
Carson, C. A.; Sells, D. M.; and Ristic, M., 1976, Vet. Parasitol., v. 2 (1), 75-81
Anaplasma marginale, cattle, cell-mediated immunity and correlation with protection induced by vaccination, review

Anaplasma marginale
Carson, C. A.; Sells, D. M.; and Ristic, M., 1977, Am. J. Vet. Research, v. 38 (2), 173-179
Anaplasma marginale, cell-mediated immune response in cattle given virulent, attenuated and inactivated preparations, measured by leukocyte migration-inhibition test and lymphocyte transformation of blood leukocytes

Anaplasma marginale
Carson, C. A.; Sells, D. M.; and Ristic, M., 1977, Am. J. Vet. Research, v. 38 (8), 1167-1172
Anaplasma marginale, cows (exper.), cell-mediated immunity and severity of clinical symptoms in response to challenge infection after inoculation with virulent, live attenuated, or killed A. marginale, effect of chemosterilization on residual immunity

Anaplasma marginale
Castro, E. R.; and Canabez, F., 1968, Bol. Chileno Parasitol., v. 23 (1-2), 23-25
Anaplasma marginale, splenectomized and normal bovines, infectivity of whole blood not destroyed by treatment with gamma ray radiation

Anaplasma marginale, illus.
Corbridge, M. H.; Trueblood, M. S.; and Bear, P. D., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 29-32
Anaplasma marginale, nucleic acids, composition, partial characterization

Anaplasma marginale
Corrier, D. E.; and Adams, L. G., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 60-65
Anaplasma marginale, Babesia bigemina, concurrent infections in calves (exper.), clinical course, serological response, pathological manifestations

Anaplasma marginale
Decia, C.; and Castro, E. R., 1968, Bol. Chileno Parasitol., v. 23 (1-2), 19-23
Anaplasma marginale in bovines (exper.), changes in serum protein values during infection

Anaplasma marginale
Decker, J. L.; Hart, L. T.; and Larson, A. D., 1976, Abst. Ann. Meet. Am. Soc. Microbiol., 135
Anaplasma marginale, technique for separating from infected bovine red blood cells

Anaplasma marginale
Dimopoullos, G. T., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 14-15
aspirations and pitfalls in anaplasmosis research, brief review

Anaplasma marginale
Dimopoullos, G. T.; and Finerty, J. F., 1976, Am. J. Vet. Research, v. 37 (6), 693-695
Anaplasma marginale, Plasmodium lophurae, P. berghei, cross reactivity in passive haemagglutination tests

Anaplasma marginale
Erp, E.; and Fahrney, D., 1975, Am. J. Vet. Research, v. 36 (5), 707-709
Anaplasma marginale, evidence that marginal bodies may leave bovine red blood cells without concomitant lysis, supports proposed endocytosis-exocytosis mechanism of host cell entry and exit in anaplasmosis

Anaplasma marginale
Erp, E. E.; and Fahrney, D., 1976, Am. J. Vet. Research, v. 37 (5), 607-609
Anaplasma marginale, splenectomized cattle, increases in serum concentration of phagocytosis-stimulating factor and acceleration of erythrophagocytosis; factor may be new hormone

Anaplasma marginale
Galhotra, A. P.; Gautam, O. P.; and Banerjee, D. P., 1977, Indian Vet. J., v. 54 (7), 522-527
Anaplasma marginale, calves (nat. and exper.), treatment trials with various drugs (terramycin, steclin, achromycin, reverin, gloxazone, reverin + gloxazone, steclin + gloxazone) during early and late stages of infection, reverin alone and combined therapy of steclin or reverin with gloxazone were most effective

Anaplasma marginale, illus.
Gallo, C.; and Balbo, S. M., 1976, Riv. Zootecn. e Vet. (3), 229-245
classification, morphology, epizootiology, transmission, vectors, pathogenicity, immunity, diagnosis, prophylaxis, review

Anaplasma marginale
Gonzalez, E. F.; Todorovic, R. A.; and Thompson, K. C., 1976, Tropenmed. u. Parasitol., v. 27 (4), 427-437
Anaplasma marginale, Babesia argentina, and B. bigemina in calves (exper.), immunization, economical method using minimum infective doses of parasite stabilates produced mild post-immunization reactions but conferred complete protection against blood-borne challenge, evaluation under laboratory conditions shows promise

Anaplasma marginale
Hidalgo, R. J., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 46-48
Anaplasma marginale, in vitro culture, review of recent research including personal communications of negative results

Anaplasma marginale, illus.
Hidalgo, R. J., 1975, Am. J. Vet. Research, v. 36 (5), 635-640
Anaplasma marginale, propagation in bovine lymph node cell culture, direct fluorescent antibody technique used for detection of organism in culture and combined with standard microscopic count procedure to obtain numerical estimates of organism as criteria of growth, effect of oxytetracycline HCl

Anaplasma marginale
Hines, H. C.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 82-85
Anaplasma marginale, presence of bovine blood group antigens in Anaplaz, ability to elicit antibody response, effects upon calves of vaccination of their dams

Anaplasma marginale
Howarth, J. A.; and Hokama, Y., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 117-120
Anaplasma marginale, laboratory transmission trials using Ixodes pacificus (transstadial transmission trial), Dermacentor occidentalis (transstadial and transovarian transmission trials), and D. andersoni (injection of macerated ticks; injection of tick saliva)

Anaplasma marginale
Howarth, J. A.; Hokama, Y.; and Amerault, T. E., 1976, J. Wildlife Dis., v. 12 (3), 427-434
Anaplasma marginale, detection in wild Odocoileus hemionus columbianus using modified card agglutination test, accuracy confirmed by calf inoculation with deer blood: California

Anaplasma marginale
Howarth, J. A.; and McNeal, D. S., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 123-126
occurrence of Anaplasma marginale in susceptible cattle subjected to conditions of natural exposure where other cattle were excluded as source of infection, no infection occurred in cows placed on tick-proof platform in same pasture, findings give proof that persistent non-bovine reservoir exists (probably deer) and that arthropod transmission readily occurs: California

Anaplasma marginale
Kingston, N.; and Trueblood, M. S., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 42-45
Anaplasma marginale, attempted infection of small laboratory animals gave equivocal or negative results, organ cultures prepared from exposed Dermacentor andersoni failed to maintain infection, attempts to grow in chick embryos by direct inoculation gave negative results
Oryctolagus cunniculus (exper., "maintained . . . 28 days")
Cavia porcellus (exper., "transient infection")

Anaplasma marginale
Kliewer, I. O.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 39-41
Anaplasma marginale, preservation by freezing in liquid nitrogen, length of storage, minimum infective dose

Anaplasma marginale
Kumar, B. J.; Muraleedharan, K.; and Rajasekharan, D., 1977, Mysore J. Agric. Sc., v. 11 (2), 214-216
Anaplasma marginale, crossbred cows (blood), case report, oxytetracycline hydrochloride, good results: Bangalore, India

Anaplasma marginale
Kuttler, K. L., 1972, Research Vet. Sc., v. 13 (6), 536-539
Anaplasma marginale, splenectomized calves, evaluation of 12 treatment schedules combining oxytetracycline and gloxazone

Anaplasma marginale
Kuttler, K. L., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 93-97
Anaplasma marginale, cattle, treatment of acute and chronic forms, review

Anaplasma marginale
Kuttler, K. L.; and Adams, L. G., 1977, Am. J. Vet. Research, v. 38 (9), 1327-1330
Anaplasma marginale in splenectomized carrier calves, recrudescent infection resulted from dexamethasone treatment, serum and blood changes during and after treatment suggest blockage of cell-mediated immune system

Anaplasma marginale
Kuttler, K. L.; and Craig, T. M., 1975, Am. J. Vet. Research, v. 36 (3), 323-325
splenectomized calves inoculated with pooled blood samples from eastern Texas cattle developed dual infections of Anaplasma marginale and a Theileria resembling Theileria mutans, treatment with gloxazone and imidocarb selectively eliminated Anaplasma, pure Theileria infections only mildly pathogenic

Anaplasma marginale
Kuttler, K. L.; and Johnson, L. W., 1977, Vet. Med. and Small Animal Clin., v. 72 (8), 1354, 1356-1359
Anaplasma marginale, Babesia bigemina, B. argentina, Holstein heifers destined for shipment to Nicaragua, premunition by inoculation with organisms, oxytetracycline given to heifers administered attenuated or non-attenuated A. marginale; apparently resistant to field challenge in Nicaragua

Anaplasma marginale
Kuttler, K. L.; and Todorovic, R. A., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 106-112
Anaplasma marginale, cattle, premunization, Gloxazone and Imidocarb superior to oxytetracycline in moderating premunizing infection, possible factors affecting success (age of host; virulence, size, and potency of premunizing inoculum; strain or size of challenge exposure; temperature and altitude)

Anaplasma marginale
Leatch, G., 1973, Austral. Vet. J., v. 49 (1), 16-19
Anaplasma marginale experimentally transmitted by transfer of various stages of Boophilus microplus from an infected calf to uninfected calves; transovarial transfer of infection did not occur

Anaplasma marginale
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Anaplasma marginale
Loehr, K. F.; and Meyer, H., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 192-197
Anaplasma organisms isolated in splenectomized calves following blood inoculation from Connochaetes taurinus, Alcelaphus buselaphus cokii, and Gazella thomsonii, morphologically indistinguishable from Aepyceros melampus, mild reaction of receptor calves to antelope-derived infection and severe reaction to subsequent A. melampus challenge, indirect fluorescent antibody results established close antigenic relationship between antelope anaplasms and A. melampus and less so to Anaplasma centrale: Kenya

Anaplasma marginale
Loehr, K. F.; Ross, J. P. J.; and Meyer, H., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 86-95
Anaplasma marginale, A. centrale, cattle, homologous and heterologous indirect fluorescent antibody and capillary tube agglutination responses to primary infection and reinfection and to cross-infection with the heterologous organisms, clinical reactions; A. centrale carrier animals showed high degree of premunity to severe challenge with A. marginale

Anaplasma marginale
Loehr, K. F.; Ross, J. P. J.; and Meyer, H., 1974, Tropenmed. u. Parasitol., v. 25 (2), 217-226
sera of 1505 game animals of 19 different species screened for antibodies to Anaplasma marginale, Babesia bigemina, and Theileria parva, capillary tube agglutination and indirect fluorescent antibody tests, antibodies more prevalent in sera of antelopes grazing in vicinity of non-dipped cattle than in areas where cattle are either dipped regularly or are not present at all, need for studies on transmission of these organisms from game to cattle and vice versa: Kenya; Tanzania; Uganda; Zambia

Anaplasma marginale
Love, J. N.; McEwen, E. G.; and Rubin, R. M., 1976, Am. J. Vet. Research, v. 37 (7), 857-858
infectivity of cryogenically preserved Anaplasma marginale-infected erythrocytes which were thawed and then maintained for periods from 2 to 96 hours and 5 to 19 days at 25 C., calves

Anaplasma marginale
McCallon, B. R., 1976, J. Dairy Sc., v. 59 (6), 1171-1174
Anaplasma marginale, cattle, etiology, economic aspects, control, review

Anaplasma marginale
McHardy, N., 1974, Ann. Trop. Med. and Parasitol., v. 68 (1), 51-57
Anaplasma marginale, calves, beneficial effects of injecting anti-erythrocyte serum more pronounced in intact than splenectomized animals, no differences in immune responses noticed

Anaplasma marginale, illus.
McHardy, N.; and Gilson, C., 1974, Tropenmed. u. Parasitol., v. 25 (1), 11-21
Anaplasma marginale, differences in ultrastructure of agglutinating and complement-fixing antigens, relation to structure of complete organism

Anaplasma marginale
McHardy, N.; and Simpson, R. M., 1973, Trop. Animal Health and Prod., v. 5 (3), 166-173
Anaplasma marginale (South African and Kenyan strains), attempted immunization of cattle using a killed vaccine with saponin as adjuvant

Anaplasma marginale
McHardy, N.; and Simpson, R. M., 1975, Trop. Animal Health and Prod., v. 7 (3), 139-148
Anaplasma marginale, non-splenectomized experimentally infected cattle, treated with single dose of imidocarb dihydrochloride followed by second dose at onset of relapse which delayed and reduced its severity (second dose administered before onset of relapse only delayed but did not reduce the severity of relapse), mechanism of relapse

Anaplasma marginale
Marble, D. W.; and Hanks, M. A., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 49-50
Anaplasma marginale in primary rabbit bone marrow culture

Anaplasma marginale
Marble, D. W.; and Hanks, M. A., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 53-54
Anaplasma marginale, growth in stable rabbit bone marrow cell line vs. primary cell line

Anaplasma marginale
Martin, W. H.; and Ritchie, W. H., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 141-142
Anaplasma marginale, cattle, diagnosis, microtiter complement fixation technique

Anaplasma marginale, illus.
Mazzola, V.; Amerault, T. E.; and Roby, T. O., 1976, Am. J. Vet. Research, v. 37 (8), 987-989
Anaplasma marginale, survival in cell line cultures from Aedes albopictus

Anaplasma marginale
Mehlitz, D.; and Ehret, R., 1974, Tropenmed. u. Parasitol., v. 25 (1), 3-10
Anaplasma, Babesia, Theileria, cattle, serological survey, capillary-tube agglutination test, indirect fluorescent antibody test, and complement fixation test: Botswana

Anaplasma marginale
Miller, R. M.; Price, M. A.; and Kuttler, K. L., 1976, Southwest Entom., v. 1 (3), 107-110
Amblyomma americanum, A. cajennense, nymphs feeding on Anaplasma marginale and Theileria infected cattle, attempted transstadial transmission to splenectomized calves, unsuccessful

Anaplasma marginale
Norman, B. B., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 103-106
Anaplasma marginale, cattle, current status of premunization and vaccination for control

Anaplasma marginale
Osorno, B. M.; et al., 1975, Am. J. Vet. Research, v. 36 (5), 631-633
Anaplasma marginale, young cattle, attenuated vaccine, evaluation of protection induced against challenge exposure of naturally transmitted anaplasmosis in enzoötic areas of Mexico, concluded that vaccine provided means for safe adaptation of high-quality young cattle to tropics

Anaplasma marginale, illus.
Osorno, B. M.; and Ristic, M., 1977, Veterinaria, Mexico, v. 8 (3), 85-98
Anaplasma marginale, bovine, control, diagnosis, distribution, use of attenuated vaccine of A. marginale, review: Mexico

Anaplasma marginale
Osorno, B. M.; Solana, M. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 113-116
Anaplasma marginale, cattle, safety and efficacy of attenuated University of Illinois vaccine in inducing protection against artificial challenge with virulent Mexican Anaplasma strain of known potency

Anaplasma marginale
Paine, G. D.; and Miller, A. S., 1977, Vet. Rec., v. 100 (3), 58 [Letter]
Anaplasma marginale, newborn calf, possible prenatal infection

Anaplasma marginale
Peterson, K. J., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 11-14
Anaplasma marginale, cattle, epizootiology: Oregon

Anaplasma marginale
Peterson, K. J.; et al., 1977, Am. J. Vet. Research, v. 38 (3), 351-354
Anaplasma marginale, latent-infected and anaplasmosis-susceptible cattle maintained on tick-proof platforms and exposed only to flying hematophagous insects which did not transmit the disease; Dermacentor andersoni as principal vector in eastern Oregon

Anaplasma marginale
Porter, R. L.; and Greve, J. H., 1976, Iowa State Univ. Vet., v. 38 (3), 109-113
Anaplasma marginale, cow, case report, differential diagnosis from leptospirosis, oxytetracycline, poor results, review: Aplington, Iowa

Anaplasma marginale
Purnell, R. E.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 181-185
Theileria gorgonis, exacerbation in 4 splenectomized Connochaetes taurinus, failure to transmit to cattle by blood passage or via Rhipicephalus appendiculatus ticks despite establishment of parasite in ticks; exacerbated Anaplasma marginale infection in C. taurinus was successfully blood-passaged to calves; results indicate that blue wildebeest is probably not a reservoir for T. gorgonis but may be for A. marginale: Tanzania; Kenya

Anaplasma marginale
Reeves, J. D. III; and Swift, B. L., 1977, Vet. Med. and Small Animal Clin., v. 72 (5), 911-914
Anaplasma marginale, transmission to cow via vaccination needle first used to inoculate a carrier cow

Anaplasma marginale, illus.
Renshaw, H. W.; et al., 1977, J. Am. Vet. Med. Ass., v. 170 (3), 334-339
Anaplasma marginale, serologic survey of cattle; inoculation of Odocoileus hemionus hemionus blood into splenectomized calves and subinoculation into non-splenectomized cow, development of parasitemia: Idaho-Utah state boundary, near Stone, Idaho

Anaplasma marginale
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Anaplasma marginale
Richey, E. J.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 35-39
Anaplasma marginale, preservation by lyophilization, preliminary results: protective substances; duration of infectivity; effect of time of collection of infected blood during patent period

Anaplasma marginale
Richey, E. J.; et al., 1977, Am. J. Vet. Research, v. 38 (2), 169-170
Anaplasma marginale, cattle, resistance after chlortetracycline elimination of latent infections is similar to that after killed-antigen vaccination of animals with no record of infection

Anaplasma marginale
Richey, E. J.; et al., 1977, Am. J. Vet. Research, v. 38 (2), 171-172
Anaplasma marginale, latent infections, cows, elimination by low levels of chlortetracycline

Anaplasma marginale, illus.
Ristic, M.; and Carson, C. A., 1977, Advances Exper. Med. and Biol., v. 93, 151-188
bovine anaplasmosis, immunoprophylaxis, review: Anaplasma marginale, biologic properties, antigenic and serologic studies, persistence in immunologically hostile host, various immunogens, immune responses to inactivated A. marginale vaccines, immune response to live attenuated and virulent A. marginale, vaccination studies with attenuated A. marginale, proposed mechanism of protection induced by this vaccine, statistical analysis, application for prevention of anaplasmosis

Anaplasma marginale
Ristic, M.; and Nyindo, M. B. A., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 66-70
Anaplasma marginale, cows inoculated with attenuated vaccine, humoral (complement fixing and agglutinating antibodies) and cell-mediated (macrophage migration inhibition test) immune responses, results demonstrate correlation between cell-mediated immunity and protection

Anaplasma marginale
Roberts, R. H.; and Love, J. N., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 121-122
Anaplasma marginale, Hippelates pusio as possible mechanical vector but no biological cycle found

Anaplasma marginale
Roberts, R. H.; and Love, J. N., 1977, Am. J. Vet. Research, v. 38 (10), 1629-1630
Anaplasma marginale, calves, viability and infectivity after ingestion of infected erythrocytes by potential insect vectors (eye gnat (Hippelates pusio), horse flies (Tabanidae))

Anaplasma marginale
Roby, T. O., 1972, Research Vet. Sc., v. 13 (6), 519-522
Anaplasma marginale, splenectomized calves (exper.), increased survival after subcutaneous treatment with imidocarb, not effective orally

Anaplasma marginale
Roby, T. O., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 100-103
Anaplasma marginale, cattle, pre-acute, acute, and carrier infections, treatment with imidocarb

Anaplasma marginale
Rodriguez, O. N.; Espaine, L.; and Rivas, A., 1975, Rev. Cubana Cien. Vet., v. 5 [on cover], v. 6 [on title page] (1-2), 145-153
Theileria mutans, incidence in cattle with or without Anaplasma marginale, Babesia bigemina and B. argentina natural infections: Cuba

Anaplasma marginale
Sawyer, M.; et al., 1973, Am. J. Vet. Research, v. 34 (10), 1281-1284
quantitative serologic assay (single radial immunodiffusion) for diagnosing congenital infections of cattle (including A. marginale)

Anaplasma marginale
Schreuder, B.E.C.; Uilenberg, G.; and Tondeur, W., 1977, Tropenmed. u. Parasitol., v. 28 (3), 367-371
Theileriidae, reinfection and retransmission experiments in cattle and African buffalo

Anaplasma marginale
Sharma, S. K.; Banerjee, D. P.; and Gautam, O. P., 1977, Ann. Recherches Vet., v. 8 (3), 307-313
Anaplasma marginale, cattle (exper.), dithiosemicarbazone (in combination with oxytetracycline), chloramphenicol and rolitetracycline very efficacious in bringing about clinical recovery and elimination of carrier status, imidocarb could not completely eliminate organisms, gentamicin sulphate ineffective

Anaplasma marginale
Simpson, C. F., 1975, Am. J. Vet. Research, v. 36 (10), 1443-1446
Anaplasma marginale, calves treated with oxytetracycline, adverse effects on morphologic features of anaplasma bodies

Anaplasma marginale
Singh, A.; and Gill, B. S., 1977, Indian J. Animal Sc., v. 47 (4), 224-226
cattle, buffaloes, prevalence: Punjab State

Anaplasma marginale, illus.
Smith, K.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (3), 293-300
Theileria gorgonis, Anaplasma marginale, intra-erythrocytic stages isolated from splenectomized Connochaetes taurinus, fine structure

Anaplasma marginale
Smith, M. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 18 [Demonstration]
cattle: Trinidad

Anaplasma marginale
Smith, M. W., 1977, PANS, v. 23 (1), 27-32
Babesia bigemina, Anaplasma marginale, immunization of susceptible imported cattle using whole blood inoculations from infected donor cattle: Trinidad, imported from Canada

Anaplasma marginale
Swift, B. L.; and Paumer, R. J., 1976, Theriogenology, v. 6 (5), 515-521
Anaplasma marginale, cattle (exper.), vertical (transplacental) transmission, last trimester of gestation

Anaplasma marginale
Thedford, T. R.; and Cook, R. H., 1976, Trop. Animal Health and Prod., v. 8 (4), 202
bovines, field survey, rapid card agglutination test, need for more efficient treatment: Western Kiambu District

Anaplasma marginale
Thompson, K. C., 1976, Vet. Parasitol., v. 2 (2), 223-229
Boophilus microplus, establishment and maintenance of colony free of infection with Anaplasma marginale and Babesia spp., subsequent infection with pure isolate of Babesia bigemina; description of life cycle of Boophilus microplus

Anaplasma marginale
Todorovic, R. A.; Long, R. F.; and McCallon, B. R., 1977, Vet. Microbiol., v. 2 (2), 167-177
Anaplasma marginale, calves, complement fixation and rapid card agglutination tests compared, results indicated that under field conditions rapid card agglutination test was a simpler and more reliable diagnostic test: north coast of Colombia

Anaplasma marginale
Todorovic, R. A.; and Tellez, C. H., 1975, Trop. Animal Health and Prod., v. 7 (3), 125-131
adult cattle (Bos taurus), premunition against Babesia bigemina and B. argentina using Imidocarb or Ganaseg to control post premunition reactions was effective and practical; premunition against Anaplasma marginale using oxytetracycline and Gloxazone was less effective and not practical

Anaplasma marginale
Trees, A. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 287 [Demonstration]
blood from healthy indigenous Nigerian cattle inoculated into splenectomized calf, calf became infected with Babesia bigemina, B. bovis, Anaplasma marginale, Borrelia theileri, Theileria mutans, Eperythrozoon teganodes, E. wenyoni; isolation of pure stabilate of B. bigemina

Anaplasma marginale
Trueblood, M. S.; and Bear, P. D., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 54-58
Anaplasma marginale, cultivation in vitro, use of bovine lymph nodes in explant cultures using raft technique, use of lymph node organ cultures in roller culture bottles with and without antibiotics

Anaplasma marginale
Utterback, W. W.; et al., 1972, Am. J. Vet. Research, v. 33 (1), 257-267
Anaplasma marginale, prevalence in cattle by herd location, survey 1969-1970, relationship of prevalence to variations in ecologic and management practices, predictive value of this data: northern California

Anaplasma marginale
Utterback, W. W.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 3-11
Anaplasma marginale, cattle, prevalence with respect to breed, sex, age, management practices, and ecologic factors: Northern California

Anaplasma marginale
Vaughn, H. W.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 136-137
Anaplasma marginale, cattle, prevalence and distribution: northern Idaho and southeastern Washington

Anaplasma marginale
Vaughn, H. W.; Renshaw, H. W.; and Frank, F. W., 1976, Am. J. Vet. Research, v. 37 (5), 615-617
Anaplasma marginale, detection in elk, serum card test and complement-fixation test gave incomplete and false-positive reactions, plasma card test gave no reactions, results indicate bovine serologic tests for anaplasmosis may be inadequate with elk serum: Clearwater National Forest, Idaho

Anaplasma marginale
Weilgama, D. J.; Jayasekara, M. U.; and Hussain, M., 1975, Ceylon Vet. J., v. 23 (3-4), 49-53
Babesia bigemina, B. argentina, Anaplasma centrale, A. marginale, three breeds of cattle (Ayrshire, Friesian and Shorthorn), haematological changes including parasitaemia and temperature reactions following premunisation with blood from infected cattle: Sri Lanka, imported from New Zealand

Anaplasma ovis, illus.
Gallo, C.; and Balbo, S. M., 1976, Riv. Zootecn. e Vet. (3), 229-245
classification, morphology, epizootiology, transmission, vectors, pathogenicity, immunity, diagnosis, prophylaxis, review

Anaplasma ovis
Gupta, S. L.; Gautam, O. P.; and Chauhan, H. V. S., 1976, Haryana Agric. Univ. J. Research, v. 6 (1), 87-88
Anaplasma ovis, sheep, diagnosis, fluorescent antibody technique, specificity tested, detects low grade infections

Anaplasma ovis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Anaplasma ovis
Uilenberg, G.; and Schreuder, B. E. C., 1976, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 29 (2), 119-126
presence in splenectomized sheep used in transmission and pathogenesis studies of Haematoxenus separatus

Anaplasmosis
Alley, J. L.; and Christenberry, C. C., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 137-140
anaplasmosis, cattle, comparison of rapid card agglutination test with complement fixation test showed overall agreement of 93.9% for unvaccinated animals and 77.9% for vaccinated herds; demonstration that anaplasmosis can be eliminated from a herd of cattle by isolation and aureomycin treatment of infected animals detected exclusively on basis of card agglutination test results

Anaplasmosis
Bedell, D. M.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 91-92
neonatal isoerythrolysis not observed in calves from 12 Angus cows vaccinated with Anaplaz

Anaplasmosis
Collins, W. W., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 97-99
anaplasmosis, cattle, development of feeding programs to provide chlortetracycline at required level in palatable and stable combinations of feed ingredients, correct level of consumption can be achieved by combining suitable formulations and management practices

Anaplasmosis
Galhotra, A. P.; et al., 1977, Indian Vet. J., v. 54 (8), 599-601
anaplasmosis, cattle, pathological changes, post-mortem examination

Anaplasmosis
Hines, H. C., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 77-80
anaplasmosis, cattle, Anaplaz vaccine as a possible cause of isohemolytic disease of calves

Anaplasmosis
Kerr, K. M.; et al., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 80 [Abstract]
neonatal isoerythrolysis produced experimentally in calves from dams hyperimmunized against their sire (an Angus bull that had had a high incidence of neonatal isoerythrolysis in his calves when the dams were inoculated with a bovine blood-based anaplasmosis vaccine)

Anaplasmosis
Kerr, K. M.; McKnelly, S.; and Bridges, C. H., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 86-90
incidence of neonatal isoerythrolysis in calves in a herd of registered Brangus cows previously vaccinated with Anaplaz; lesions, clinical signs, and treatment of affected calves; limitations of predicting which cows will give birth to affected calves; possibilities of selecting bulls for future mating with the cows to prevent recurrence of disease in subsequent calves

Anaplasmosis
McCallon, B. R., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 1-3
anaplasmosis, cattle, preliminary prevalence rates based on 6 month survey, estimated annual loss from anaplasmosis $100 million: United States

Anaplasmosis
Smithies, L. K.; and Erdmann, A. A., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 127-128
anaplasmosis control program: Wisconsin

Anaplasmosis
Stormont, C. J., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 81 [Abstract]
prognosis of neonatal isoerythrolysis based on blood types of cows and bulls taking into account specificity and titers of isoantibodies engendered in cows as result of vaccination with Anaplaz

Anaplasmosis
Tiwari, S. P.; Muley, A. R.; and Mehta, M. L., 1975, JNKVV Research J., v. 9 (3-4), 179-180
anaplasmosis, buffaloes and goats (blood of both), oxytetracycline, good results: Jabalpur

Anaplasmosis
Uilenberg, G., 1976, World Animal Rev. (17), 8-15
babesiasis, theileriasis, anaplasmosis, epizootiology of tick-borne diseases, review

Anc(h)orina Mingazini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ancora Labbe, 1899

Anchorina criodrilii Sciacchitano, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ancora criodrilii (Sciacchitano, 1931) comb. nov.

Anc(h)orina sagittata (Leuckart, 1861) Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ancora sagittata (Leuckart, 1861) Labbe, 1899

Ancora Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis
Syn.: Anc(h)orina Mingazzini, 1891

Ancora criodrilii (Sciacchitano, 1931) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Anchorina criodrilii Sciacchitano, 1931

Ancora lutzi Hasselmann, 1918
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Ancora sagittata (Leuckart, 1861) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Ancyrophora balazyi sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Carabus coriaceus
Carabidae, larvae
(intestine of all): all from Poland

Ancyrophora cornuta n. sp., illus.
Baudoin, J.; and Mouthon, J., 1976, Compt. Rend. Acad. Sc., Paris, v. 282, s. D, (13), 1285-1287
correlation between presence of gregarine parasite and carnivorous diet of host
Dinocras megacephala (intestin)

Ancyrophora gracilis (Schneider): Wellmer 1911
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Ancyrophora stelliformis (Schneider)

Ancyrophora gracilis, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Carabus intricatus
C. violaceus
C. nemoralis

Ancyrophora gracilis (Stein) Leger
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Carabus insulicola: Coree du sud

Ancyrophora philonthi sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Philonthus laevicollis (intestine): Bialowieza, Poland

Ancyrophora stelliformis (Schneider), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Carabus violaceus
Pterostichus vulgaris
(intestine of all): all from Poland

Ancyrophora umbelliformis n. sp., illus.
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
Hydrochara caraboides: Loire-Atlantique (Ouest de la France)

Anisonema Dujardin
Michajłow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (1), 59-62
key to species parasitizing copepods, includes: Anisonema parasiticum; A. naupliorum

Anoplophrya aelosomatis Anderson, 1888
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophryoides aelosomatis (Anderson, 1888) Lom, 1956

Anoplophrya brasili p. p. Leger et Duboscq, 1904
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Durchoniella brasili (Leger et Duboscq, part. 1894) de Puytorac, 1954

Anoplophrya clavata Kent, 1881-82
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella clavata (Leidy, 1855) Cepede, 1910

Anoplophrya debaisieuxi, Delphy, 1922
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mrazekiella debaisieuxi (Delphy, 1922) Cheissin, 1930

Anoplophrya elongata Delphy, 1922
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya elongata (Delphy, 1922) de Puytorac, 1957

Anoplophrya enchytraei (part.) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis commune Lom, 1959

Anoplophrya enchytraei (part.) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis enchytraei de Puytorac, 1954

Anoplophrya enchytraei (part.) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis janovicensis Lom, 1959

Anoplophrya filum Claparede, 1860
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Delphyella filum (Claparede, 1860) de Puytorac, 1969

Anoplophrya filum pp. Schewiakoff, 1896 [et auct.]
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Delphyella filum (Claparede, 1860) de Puytorac, 1969

Anoplophrya fusiformis (part.) Delphy, 1922
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya elongata (Delphy, 1922) de Puytorac, 1957

Anoplophrya herpeni Delphy, 1922
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya pachydrili (Claparede, 1861) Heidenreich, 1935

Anoplophrya inermis Kent, 1881-82
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Acanthophrya inermis (Stein, 1859) Heidenreich, 1935

Anoplophrya maupasi Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophryoides aelosomatis (Anderson, 1888) Lom, 1956

Anoplophrya minuta
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis enchytraei de Puytorac, 1954

Anoplophrya minuta
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis commune Lom, 1959

Anoplophrya minuta
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis janovicensis Lom, 1959

Anoplophrya naidos proparte Ghosh, 1918
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Acanthophrya inermis (Stein, 1859) Heidenreich, 1935

Anoplophrya naidos p. parte Schweier, 1898
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Acanthophrya inermis (Stein, 1859) Heidenreich, 1935

Anoplophrya naidos, p. parte Schewiakoff, 1896
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Acanthophrya inermis (Stein, 1859) Heidenreich, 1935

Anoplophrya nodulata p. p. Ghosh, 1918
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Durchoniella brasili (Leger et Duboscq, part. 1894) de Puytorac, 1954

Anoplophrya nodulata p. p. in Schweier, 1898
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya prolifera (Clap. et Lachm., 1858),Rossolimo, 1926

Anoplophrya nodulata Butschli, 1887, p. p.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya prolifera (Clap. et Lachm., 1858) Rossolimo, 1926

Anoplophrya pachydrili (Clap., 1861) Kent, 1882
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya pachydrili (Claparede, 1861) Heidenreich, 1935

Anoplophrya paradoxa Ross. (part.) Meier, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis commune Lom, 1959

Anoplophrya paradoxa Ross (part.) Meier, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis janovicensis Lom, 1959

Anoplophrya paranaidos Pierantoni, 1909
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella paranaidos (Pierantoni 1909) Heidenrich, 1935

Anoplophrya prolifera (Clap. et Lachm., 1858) Kent, 1882
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya prolifera (Clap. et Lachm., 1858) Rossolimo, 1926

Anoplophrya spiraliformis
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis commune Lom, 1959

Anoplophrya spiraliformis
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis enchytraei de Puytorac, 1954

Anoplophrya spiraliformis
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Buetschliellopsis janovicensis Lom, 1959

Anoplophrya tubificis Kij., 1925 [pro parte?]
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella taupodrili (Kijenskij, 1925) de Puytorac

Anoplophrya tubificis Kijensky, 1925 [pro parte?]
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya pungens (Stein, 1861) Heidenreich, 1935

Anoplophryoidea n. superfam. [nomen nudum]
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Astomatida

Anthemosoma garnhami
  Momen, H., 1976, Parasitology, v. 73 (2), xvi [Abstract]
    Plasmodium spp., Babesia spp., Anthemosoma garnhami, carbohydrate metabolism: enzyme activity, glucose catabolism

Anthemosoma garnhami
  Momen, H.; and Chance, M. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 13-14 [Demonstration]
    Babesia spp., Anthemosoma garnhami, differences in DNA buoyant densities as possible aid to taxonomic studies

Anthemosoma garnhami Landau et al., 1969
  Momen, H.; Homewood, C. A.; and Neame, K. D., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 519-520
    Anthemosoma garnhami-infected mice, red cells show marked increase in permeability as measured by entry of L-glucose

Anthemosoma garnhami
  Vivier, E., 1974, J. Protozool., v. 21 (3), 476 [Abstract]
    Anthemosoma garnhami, organitogenesis of merizoites

Anthonyella Delphy, 1936
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Radiophryidae n. fam., Anthonyellinae n. s. fam.

Anthonyella hoplites (Rossolimo, 1926) Delphy, 1936, illus.
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    synonymy
    Styloscolex sp.
    S. baikalensis
    Teleuscolex korotneffi
    Lamprodrilus sp.
    Lumbriculidae sp.

Anthonyella lumbriculi (Cheissin, 1930) Delphy, 1936, illus.
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Syn.: Radiophrya lumbriculi, Cheissin, 1930
    Styloscolex sp.

Anthonyellinae n. s. fam.
  de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
    Radiophryidae n. fam.
    includes: Anthonyella; Ochridanus; Radiophryopsis

Aphanipteromonas sensu Nicoli [sp.]
  Beaucournu, J. C.; and Deunff, J., [1976], Ann. Parasitol., v. 50 (6), 1975, 831-835
    siphonapteres (proctodeum): France

Ascocystis Grasse, 1953
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Lecudinidae, revision and checklist of species
    diagnosis

Ascocystis barretti (Vavra, 1969) nov. comb.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Lankesteria barretti Vavra, 1969

Ascocystis chagasi (Adler & Mayrink, 1961) Tuzet & Rioux, 1966
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Monocystis chagasi Adler & Mayrink, 1961

Ascocystis clarki (Sanders & Poinar, 1973) comb. nov.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Lankesteria clarki Sanders & Poinar, 1973

Ascocystis culicis (Ross, 1898) Grasse, 1953
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    synonymy

Ascocystis galliardi (Garnham, 1973) comb. nov.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Lankesteria galliardi Garnham, 1973

Ascocystis legeri (Blanchard, 1902) comb. nov.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Monocystis legeri Blanchard, 1902

Ascocystis mackiei (Shortt and Swaminath, 1927) Ormieres, 1965(?)
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    Syn.: Diplocystis sp. Lisova, 1962

Ascocystis mackiei (Shortt & Swaminath, 1927) Ormieres, 1965
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    synonymy

Ascocystis mackiei (Shortt & Swaminath, 1927) Tuzet & Rioux, 1965
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Ascocystis mackiei (Shortt & Swaminath, 1927) Ormieres, 1965

Ascocystis tripteroidesi (Bhatia, 1938) Ormieres, 1965
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Lankesteria tripteroidesi Bhatia, 1938

Aseptorina
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    checklist of species of 6 aseptate gregarine families

Astasia Dujardin
  Michajłow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (1), 59-62
    key to species parasitizing copepods, includes: Astasia mobilis; A. ovorum

Astasia fennica, illus.
  Fize, A.; and Michel, R., 1972, Protistologica, v. 8 (2), 143-153
    Astasia fennica, life cycle, ultrastructure
    Eucyclops serrulatus
    E. macrurus
    Macrocyclops fuscus
    M. albidus
    (tube digestif of all): all from region de Montpellier

Astomatida Schewiakoff, 1896
  de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
    includes: Hoplitophryoidea nv. super-fam.: Anoplophryoidea n. superfam.; Haptophryoidea n. superfam.

Atoxoplasma Garnham, 1950
  Box, E. D., 1975, J. Protozool., v. 22 (2), 165-169
    Isospora serini, description of exogenous stages, asexual stages in macrophages are indistinguishable from parasites previously called avian Toxoplasma, Atoxoplasma, and Lankesterella

Atoxoplasma, illus.
  Box, E. D., 1977, J. Protozool., v. 24 (1), 57-67
    forms resembling parasites called Atoxoplasma or Lankesterella were seen in Serinus canarius infected with Isospora serini

Atoxoplasma Garnham, 1950 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Isospora lacazei (Labbe, 1893)

Atoxoplasma Garnham, 1950 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Isospora serini (Aragao, 1933) Box, 1975

Atoxoplasma [sp.]
  Box, E. M., 1973, J. Protozool., v. 20 (4), 510
    comparative development of Atoxoplasma and Isospora species in canaries

Atoxoplasma [sp.]
  Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
    seasonal occurrence
    Passer domesticus (blood): Sussex, England

Atoxoplasma spp.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    "Toxoplasma sp. auctores from various birds. Mostly Atoxoplasma spp."

Atoxoplasma adiei (Aragao, 1911) Baker, Bennett, Clark & Laird, 1972
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    Syn.: Toxoplasma passeris Rousselot, 1953

Atoxoplasma liothricis (Laveran & Marullaz, 1914) Baker, Bennett, Clark & Laird, 1972
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    Syn.: Toxoplasma liothricis Laveran & Marullaz, 1914

Atoxoplasma paddae (Aragao, 1911) Laird, 1959
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    Syn.: Toxoplasma avium Marullaz, 1913

Auerbachia
  Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
    Myxidium coryphaenoidium, presence of 2 spore forms interpreted as result of polymorphism and not as mixed infection of 2 species; abnormal spores superficially resemble other Myxidium spp. and other genera (Myxobolus, Zschokkella, Auerbachia)

Auerbachia monstrosa Meglitsch, 1960
  Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
    Coelorinchus australis
    C. innotabilis
    (gall bladder of all): all from off New Zealand

Auerbachia pulchra Lom, Noble and Laird, 1975
  Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
    Coelorinchus coelorhincus carminatus (gall bladder): off Key West, Florida
    C. occa (gall bladder): off Lesser Antilles
    Coryphaenoides acrolepis (gall bladder/bile ducts): off northern California
    C. filifer (gall bladder): off Oregon, Washington, B. C., Canada
    C. pectoralis (gall bladder): off northern California
    Macrourus berglax (gall bladder): off Iceland, Newfoundland
    Malacocephalus occidentalis (gall bladder): off Colombia (Caribbean Sea)
    Nezumia bairdii (gall bladder): off Nova Scotia

Babesia
Adam, K. M. G.; Beasley, S. J.; and Blewett, D. A., 1977, Research Vet. Sc., v. 23 (2), 133-138
babesial antibody detected in sera of wild red deer by indirect fluorescent antibody technique, incidence, age and sex of host: Scotland

Babesia
Allison, A. C.; and Clark, I. A., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, Pt. 2), 216-222
specific and non-specific immunity to haemoprotozoa with emphasis on malaria and Babesia infections, workshop report: inherited differences in erythrocytes which influence susceptibility; specific responses of B- and T-lymphocytes to parasite antigens; macrophage activation

Babesia
Aragon, R. S., 1976, Vet. Bull., v. 46 (12), 903-917
Babesia, review of life cycle, pathology, diagnosis, immunity, prophylaxis, and chemotherapy

Babesia
Callow, L. L., 1977, Advances Exper. Med. and Biol., v. 93, 121-149
bovine babesiosis, vaccination, review: biology of Babesia of cattle (taxonomy and distribution, life cycle and transmission by ticks, pathological effects, immunology); need for vaccination (epizootic spread, cattle imported to enzootic area, enzootic instability); approaches to vaccination (premunition, surveillance and treatment, chemoprophylaxis, stimulation of nonspecific immunity, experimental approaches to antigen production); vaccination against B. argentina (= bovis) in Australia; vaccination against B. bigemina; future requirements and developments

Babesia
Doyle, J. J., 1977, Advances Exper. Med. and Biol., v. 93, 27-29
Babesia, antigenic variation, brief review

Babesia
Jadin, J. B.; et al., 1977, J. Protozool., v. 24 (4), 60A [Abstract]
relations between rickettsia and piroplasms

Babesia
Jeon, Y.; Kim, D. S.; and Jeong, S. J., 1975, Research Rep. Office Rural Develop., Min. Agric. and Forest., Korea, v. 17, 45-49
Babesia, cattle, experimental infection for immunization, recovery with persistence of antibodies as shown by indirect fluorescent antibody test

Babesia
Jeon, Y.; and Lee, T. J., 1975, Research Rep. Office Rural Develop., Min. Agric. and Forest., Korea, v. 17, 35-43
Babesia, cattle, experimental infection with Korean strain, detection of antibodies with indirect fluorescent antibody test, chronology of presence of antibodies

Babesia
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Babesia
Kuttler, K. L.; Graham, O. H.; and Trevino, J. L., 1975, Research Vet. Sc., v. 18 (2), 198-200
imidocarb treatment of calves at varying intervals before or after exposure to Babesia-infected Boophilus microplus, at certain times prevented development of clinical infection or rendered next generation of tick larvae incapable of transmitting Babesia infection

Babesia
Leeflang, P.; et al., 1976, Internat. J. Parasitol., v. 6 (2), 159-161
survey of 173 males using indirect fluorescent antibody technique with Babesia bovis, B. bigemina, and B. ratti as antigens, babesial antibodies detected in 54%, no correlation found with mode of living, contact with livestock, incidence of malaria parasitaemias or malarial antibodies, no Babesia organisms found in blood smears or by animal subinoculation, source of antibodies unknown but subclinical infection not likely: Nigeria

Babesia, illus.
Ristic, M., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 831-870
babesiosis, theileriosis, immunology, review

Babesia
Ristic, M., 1976, Vet. Parasitol., v. 2 (1), 31-47
intracellular blood protista: intraerythrocytic behavior, transfer, and circulatory clearance; survival and development within macrophages; persistence of organism in immunologically hostile host; immune responses and protection; serodiagnosis; vaccination

Babesia
Ristic, M.; and Smith, R. D., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 41-63
zoonoses caused by hemoprotozoa, review: trypanosomiasis, malaria, babesiosis

Babesia
  Shortt, H. E., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 519-521
    review of known cases of human infections and their diversity of infection sources

Babesia [sp.], illus.
  Adam, K. M. G.; et al., 1976, Parasitology, v. 73 (1), 1-11
    antibody-positive blood from wild Cervus elaphus produced Babesia infections in splenectomized C. elaphus (also Eperythrozoon infection resembling wenyoni), possible transient deer Babesia infection produced in 1 of 6 bovine calves, indirect fluorescent antibody tests, "Despite their similarities, specific status for B. divergens and the red deer Babesia is probably justified; at present there is insufficient evidence to justify separation of the red deer Babesia from B. capreoli."

Babesia sp., illus.
  Aeschlimann, A.; Brossard, M.; and Quenet, G., 1975, Acta Trop., v. 32 (4), 281-289
    Apodemus sylvaticus
    A. flavicollis
    Clethrionomys glareolus
    Microtus agrestis
    (sang of all): all from Suisse

Babesia sp., illus.
  Bigalke, R. D.; Keep, M. E.; and Schoeman, J. H., 1972, Onderstepoort J. Vet. Research, v. 39 (4), 225-227
    brief description
    Tragelaphus scriptus (blood): Hluhluwe Game Reserve, Natal, Zululand

Babesia spp.
  Bindseil, E., 1976, Medlemsbl. Danske Dyrlaegeforen., v. 59 (1), 11-14
    clinical aspects, diagnosis, pathology, brief review

Babesia sp.
  Blewett, D. A.; Adam, K. M. G.; and Sharman, G. A. M., 1974, J. Protozool., v. 21 (3), 448 [Abstract]
    Cervus elaphus (blood) (nat. and exper.): Scotland

Babesia sp., large, indistinguishable from B. bigemina (Smith and Kilborne, 1893)
  Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
    blood parasites of wild Bovidae, incidence, epizootiological importance
    Connochaetes taurinus (blood): Ngamiland, Botswana

Babesia sp., small, indistinguishable from B. bovis (Babes, 1888)
  Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
    blood parasites of wild Bovidae, incidence, epizootiological importance
    Damaliscus lunatus (blood): Ngamiland, Botswana

Babesia sp.
  Churchill, R. C.; and Best, D. R., 1976, Austral. Vet. J., v. 52 (10), 487 [Letter]
    horse, first recorded case in country: Australia

Babesia [sp.], illus.
  Gorenflot, A.; Piette, M.; and Marchand, A., 1976, Rec. Med. Vet., v. 152 (5), 289-297
    Babesia [sp.], human (blood), optical and scanning electron microscopy, review of human and animal babesiasis: France

Babesia sp., illus.
  Healy, G. R.; Walzer, P. D.; and Sulzer, A. J., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 376-378
    human, asymptomatic infection diagnosed during epidemiologic investigation of transfusion-acquired malaria: Georgia

Babesia sp.
  Irvin, A. D.; and Young, E. R., 1977, Nature, London (5627), v. 269, 407-409
    Babesia spp., in vitro test for screening drug activity which exploits finding that Babesia parasites readily incorporate hypoxanthine in an in vitro culture system, may also be used to detect drug resistance patterns and have possible application in primary screening of antimalarial drugs

Babesia [sp.]
  Mohammed, A. N.; and Leeflang, P., 1975, Samaru Agric. Newsletter, v. 17 (2), 75-77
    cattle (blood): Shika, Nigeria

Babesia spp.
  Osorno, B. M.; et al., 1976, Vet. Parasitol., v. 2 (1), 111-120
    Babesia spp., human, serological survey, 38 reactors from 101 individuals examined, isolation of parasite in hamsters from blood of 3 of these 38 reactors, first documented finding of latent form of human babesiosis: Mexico

Babesia [sp.], illus.
  Reid, J. F.; et al., 1976, Vet. Rec., v. 99 (21), 419
    sheep (blood), first British report, possibly derived from red deer: Argyllshire, Scotland

Babesia [sp.], illus.
  Scharfman, W. B.; and Taft, E. G., 1977, J. Am. Med. Ass., v. 238 (12), 1281-1282
    human babesiosis (Nantucket fever), case report of infection in adult woman after summer visit to Nantucket Island, diagnosis, therapy with chloroquine: Albany, New York

Babesia [sp.], resembling B. microtia
Van Peenen, P. F. D., 1975, Bull. Soc. Path. Exot., v. 68 (3), 303-306
Rattus exulans: Indonesia

Babesia sp.
Wilson, D. E.; and Hirst, S. M., 1977, Wildlife Monogr. (54), Suppl., 3-111
Hippotragus niger: Transvaal; Rhodesia

Babesia [sp.]
Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
Peromyscus truei
Neotoma fuscipes
all from Juniper Hills near Pearblossom, Los Angeles County, California

Babesia n. spec.
Yoeli, M.; and Wisseman, C. L., jr., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 6-7 [Demonstration]
new species recovered from Rattus sp. and Rattus rattoides in Pakistan, histology and pathology in white mice (exper.)

Babesia ardeae
Peirce, M. A., 1973, J. Protozool., v. 20 (5), 543-546
"On the available evidence. . .regarded as valid avian piroplasm. . ."

Babesia argentina
Bachmann, A. W.; et al., 1976, Tropenmed. u. Parasitol., v. 27 (3), 372-376
Babesia argentina, cattle, pre-infection levels of erythrocyte adenosine triphosphate, no significant correlation with susceptibility to infection

Babesia argentina
Bachmann, A. W.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (3), 361-366
Babesia argentina, experimental infection in Droughtmaster cattle which are somewhat resistant to babesiosis, no apparent correlation between hemoglobin types and resistance to infection

Babesia argentina, illus.
Brossard, M., 1976, Rev. Suisse Zool., v. 83 (2), 443-462
morphology, Ixodes ricinus as exper. vector, transmission among exper. cattle; "B. berbera, B. argentina and B. bovis . . . should probably be considered as races of the same species, B. bovis"

Babesia argentina
Callow, L. L., 1976, World Animal Rev. (18), 9-15
Anaplasma centrale, Babesia argentina, B. bigemina, cattle, factors in transmission by Boophilus microplus, immunization with strains from splenectomized calves, review: Australia

Babesia argentina (= bovis)
Callow, L. L., 1977, Advances Exper. Med. and Biol., v. 93, 121-149
bovine babesiosis, vaccination, review: biology of Babesia of cattle (taxonomy and distribution, life cycle and transmission by ticks, pathological effects, immunology); need for vaccination (epizootic spread, cattle imported to enzootic area, enzootic instability); approaches to vaccination (premunition, surveillance and treatment, chemoprophylaxis, stimulation of nonspecific immunity, experimental approaches to antigen production); vaccination against B. argentina (= bovis) in Australia; vaccination against Babesia bigemina; future requirements and developments

Babesia argentina
Callow, L. L.; et al., 1974, Austral. Vet. J., v. 50 (1), 6-11
Babesia argentina, appreciable degree of immunity retained by cattle after sterilization of infections of varying duration by imidocarb, indirect fluorescent antibody test; duration of prior exposure to the parasite influenced the degree of immunity to subsequent challenge with heterologous strains of B. argentina

Babesia argentina
Callow, L. L.; et al., 1976, Austral. Vet. J., v. 52 (10), 446-450
Babesia argentina, unvaccinated cattle, serological testing by indirect fluorescent antibody, strong correlation between infection rate and incidence of Boophilus microplus: southeastern Queensland

Babesia argentina
Callow, L. L.; and Pepper, P. M., 1974, Austral. Vet. J., v. 50 (1), 1-5
Babesia argentina, cattle, measurement of and correlations between fever, changes in the packed cell volume and parasitaemia in evaluation of susceptibility to infection; results suggest that most or all of anemia of B. argentina infections is related to growth and multiplication of parasite, and not to an autoimmune mechanism

Babesia argentina
Callow, L. L.; Quiroga, Q. C.; and McCosker, P. J., 1976, Internat. J. Parasitol., v. 6 (4), 307-310
Babesia argentina, Anaplasma marginale, comparison of strains of each from Australia vs. Bolivia with indirect fluorescent antibody test showed serological identity of the two strains of each parasite, implications for vaccination; since earlier study showed serological identity between B. bovis and B. argentina, the small Babesia of Australia and South America should by priority be called B. bovis

Babesia argentina
Dalgliesh, R. J.; et al., 1976, Exper. Parasitol., v. 40 (1), 124-131
Babesia argentina, calves (exper.) given betamethasone, fatal pulmonary edema, hematological changes, histopathology, findings established occurrence of disseminated intravascular coagulation

Babesia argentina
Dalgliesh, R. J.; and Stewart, N. P., 1977, Austral. Vet. J., v. 53 (4), 176-180
Babesia argentina, increased tolerance to imidocarb after repeated exposure, parasite virulence unaffected, exper.-infected nonsplenectomized calves

Babesia argentina
Emmerson, F. R.; Knott, S. G.; and Callow, L. L., 1976, Austral. Vet. J., v. 52 (10), 451-454
Babesia argentina, beef cattle, field vaccination trials: southeastern Queensland

Babesia argentina
Farlow, G. E., 1976, Internat. J. Parasitol., v. 6 (6), 513-516
Babesia rodhaini, B. argentina, differences in infectivity when incubated in plasma vs. serum, role of glucose in prolonging viability, relevance of findings to living babesial vaccines in which plasma- and serum-based diluents may be used

Babesia argentina
Gill, B. S.; and Khan, M. H., 1972, Indian J. Animal Sc., v. 42 (12), 1071-1073
Babesia bigemina and B. argentina strains injected intravenously into cow calves and buffalo calves, imidocarb dihydrochloride, therapeutic efficacy in calves after splenectomy, subinoculation test

Babesia argentina
Gonzalez, E. F.; Todorovic, R. A.; and Thompson, K. C., 1976, Tropenmed. u. Parasitol., v. 27 (4), 427-437
Anaplasma marginale, Babesia argentina, and B. bigemina in calves (exper.), immunization, economical method using minimum infective doses of parasite stabilates produced mild post-immunization reactions but conferred complete protection against bloodborne challenge, evaluation under laboratory conditions shows promise

Babesia argentina
Goodger, B. V., 1973, Austral. Vet. J., v. 49 (2), 81-84
improved sensitivity of haemagglutination test for Babesia bigemina antibody using antigen obtained from lysate of infected erythrocytes; cross-reaction with B. argentina antisera, comparing cross-reactions of antigens from both species enables differentiation without inhibition techniques

Babesia argentina
Goodger, B. V., 1975, Ztschr. Parasitenk., v. 48 (1), 1-7
Babesia argentina, experimentally infected cattle dying from heavy infection, cold precipitable fibrinogen complex in plasma, possibly formed by proteolytic enzymes from parasite

Babesia argentina
Goodger, B. V., 1976, Internat. J. Parasitol., v. 6 (3), 213-216
Babesia argentina, crude soluble haemagglutination antigen contained fibrinogen, removal of fibrinogen removed most if not all antigenic activity, concluded that antigen was either babesial moiety complexed with fibrinogen or a fibrinogen molecule altered by parasite metabolic activity

Babesia argentina (Lingineres, 1910)
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
as syn. of Babesia bovis (Babes, 1888) Starcovici, 1893

Babesia argentina
Johnston, L. A. Y.; Trueman, K. F.; and Pearson, R. D., 1977, Austral. Vet. J., v. 53 (5), 222-226
Babesia argentina, B. bigemina, cattle (nat. and exper.), comparison of Giemsa and direct fluorescent antibody staining for diagnosis, determination of length of time Babesia may be recognized in tissues after host death; field cases of B. argentina were due to primary infections rather than relapses

Babesia argentina
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Babesia argentina
Kuttler, K. L.; and Johnson, L. W., 1977, Vet. Med. and Small Animal Clin., v. 72 (8), 1354, 1356-1359
Anaplasma marginale, Babesia bigemina, B. argentina, Holstein heifers destined for shipment to Nicaragua, premunition by inoculation with organisms, oxytetracycline given to heifers administered attenuated or non-attenuated A. marginale; apparently resistant to field challenge in Nicaragua

Babesia argentina
Mahoney, D. F.; and Mirre, G. B., 1974, Research Vet. Sc., v. 16 (1), 112-114
extraction of infective material of Babesia argentina from larval stages of Boophilus microplus, use in infecting calves, potential source of erythrocyte-free parasites for vaccination and study of antigens

Babesia argentina
Mahoney, D. F.; and Mirre, G. B., 1977, Research Vet. Sc., v. 23 (1), 126-127
as syn. of Babesia bovis

Babesia argentina
Mahoney, D. F.; and Wright, I. G., 1976, Vet. Parasitol., v. 2 (3), 273-282
Babesia argentina, cattle, immunization with killed antigen against infection with heterologous strain, effect of vaccination on certain pathological parameters

Babesia argentina
Mahoney, D. F.; Wright, I. G.; and Mirre, G. B., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 197-203
Babesia argentina, B. bigemina, calves, persistence of immunity after natural infection in early life and subsequent tick-free maintenance, concluded that animals naturally infected during calfhood are unlikely to require vaccination for protection against babesiosis in later life

Babesia argentina
Parker, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 387-390
Babesia argentina, direct counting technique for estimating high parasitaemias, also suitable for B. bigemina and Plasmodium berghei but not for B. rodhaini or Anaplasma centrale

Babesia argentina
Rajamanickam, C., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (1), 132 [Demonstration]
Babesia argentina infection in 10-day-old calf (blood, brain, spleen) born to cow recently imported from Australia: Malaysia

Babesia argentina
Rodriguez, O. N.; et al., 1975, Rev. Cubana Cien. Vet., v. 5 [on cover], v. 6 [on title page] (1-2), 1-8
Babesia bigemina, B. argentina, splenectomized cattle, correlation between duration of infection, symptomatology and laboratory data, comparison of species

Babesia argentina
Rodriguez, O. N.; Espaine, L.; and Rivas, A., 1975, Rev. Cubana Cien. Vet., v. 5 [on cover], v. 6 [on title page] (1-2), 145-153
Theileria mutans, incidence in cattle with or without Anaplasma marginale, Babesia bigemina and B. argentina natural infections: Cuba

Babesia argentina
Ross, D. R.; and Mahoney, D. F., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 385-392
Babesia argentina in Bos taurus, epizootiological model, computer simulation of parasite rates

Babesia argentina
Roychoudhury, G. K.; Kalita, C. C.; and Roychoudhury, R. K., 1976, Indian Vet. J., v. 53 (5), 364-367
Babesia argentina, exotic calves, clinical symptoms, Berenil, good results, Boophilus microplus recovered from herds: Instructional Livestock Farm, Assam Agricultural University, Khanapara

Babesia argentina, illus.
Todorovic, R., 1976, Vet. Parasitol., v. 2 (1), 97-109
Babesia spp., cattle, review: serodiagnosis, immunization (sterile immunity; premunition), chemoprophylaxis with imidocarb, vectors: Colombia

Babesia argentina
Todorovic, R. A.; and Gonzalez, E. F., 1975, Rev. Inst. Colomb. Agropec., v. 10 (1), 87-99
Babesia bigemina, B. argentina, cattle, killed vaccine produced from infected erythrocytes and plasma, high degree of sterile immunity produced in calves: Colombia

Babesia argentina
Todorovic, R. A.; and Gonzalez, E. F., 1975, Rev. Inst. Colomb. Agropec., v. 10 (2), 243-254
Babesia bigemina, B. argentina, Holstein-Friesian calves immunized with blood from carrier animals or from inoculated, splenectomized calves, challenge by Babesia-infected Boophilus microplus, weight gain records; better results with carrier blood

Babesia argentina
Todorovic, R. A.; Gonzalez, E. F.; and Adams, L. G., 1973, Trop. Animal Health and Prod., v. 5 (4), 234-245
Babesia bigemina, B. argentina, calves, sterile immunity using killed-Babesia vaccine, field-borne challenge with Boophilus microplus infected ticks; possible important role in mechanism of acquired immunity

Babesia argentina
Todorovic, R. A.; and Long, R. F., 1976, Tropenmed. u. Parasitol., v. 27 (2), 169-181
indirect fluorescent antibody test found superior to complement fixation test as epidemiologic tool to detect Babesia bigemina and B. argentina infections in cattle, some cross reactivity in differentiating between spp.: Colombia

Babesia argentina
Todorovic, R. A.; and Tellez, C. H., 1975, Trop. Animal Health and Prod., v. 7 (3), 125-131
adult cattle (Bos taurus), premunition against Babesia bigemina and B. argentina using Imidocarb or Ganaseg to control post premunition reactions was effective and practical; premunition against Anaplasma marginale using oxytetracycline and Gloxazone was less effective and not practical

Babesia argentina
Vizcaino G., O.; and Todorovic, R. A., 1975, Rev. Inst. Colomb. Agropec., v. 10 (1), 77-85
Babesia bigemina, B. argentina, characterization of antigens, complement fixation, immunodiffusion, cross-immunity tests

Babesia argentina
Weilgama, D. J.; Jayasekara, M. U.; and Hussain, M., 1975, Ceylon Vet. J., v. 23 (3-4), 49-53
Babesia bigemina, B. argentina, Anaplasma centrale, A. marginale, three breeds of cattle (Ayrshire, Friesian and Shorthorn), haematological changes including parasitaemia and temperature reactions following premunisation with blood from infected cattle: Sri Lanka, imported from New Zealand

Babesia argentina
Wright, I. G., 1973, Research Vet. Sc., v. 14 (1), 29-34
Babesia argentina, B. babesia, splenectomised calves (exper.), haematology

Babesia argentina
Wright, I. G., 1975, Vet. Parasitol., v. 1 (1), 91-96
purification of esterase from Babesia argentina, probable role in in vitro activation of plasma prekallikrein, implications for mechanism of vasodilatory shock and disseminated intravascular coagulation in acute infections

Babesia bandicootia
Van Peenen, P. F. D.; et al., 1977, J. Protozool., v. 24 (2), 310-312
piroplasms, probably geographic strain of Babesia microti [but those from bandicoot rats are also referred to as B. bandicootia]; occurrence and appearance; transtadial infection of Ixodes granulatus; infectivity for laboratory hosts; effect of syringe passage, splenectomy, and irradiation; pathology; cross-infection experiments; lack of susceptibility of bandicoot rats and spiny rats to Gray strain of B. microti
Bandicota indica nemorivaga (nat. and ? exper.): Taiwan
Rattus coxinga (nat. and exper.): Taiwan
R. norvegicus (exper.)
Mesocricetus auratus (exper.)
Meriones unguiculatus (exper.)
Bandicota indica setifera (exper.)
Ixodes granulatus: Taiwan

Babesia berbera, illus.
Brossard, M., 1976, Rev. Suisse Zool., v. 83 (2), 443-462
morphology, Ixodes ricinus as exper. vector, transmission among exper. cattle; "B. berbera, B. argentina and B. bovis . . . should probably be considered as races of the same species, B. bovis"

Babesia berbera
Weisman, J.; Goldman, M.; and Pipano, E., 1974, J. Protozool., v. 21 (3), 466 [Abstract]
Babesia bigemina, B. berbera, passive transfer of antibodies from cows to calves via colostrum

Babesia bigemina
Bishop, J. P.; et al., 1973, Trop. Animal Health and Prod., v.5 (3), 141-145
Babesia bigemina, isolation, separation and preservation by serially passing through splenectomized calves, stabilate established

Babesia bigemina
Burridge, M. J.; Kimber, C. D.; and McHardy, N., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 191-195
Babesia bigemina, value of dried blood samples as source of antibody for indirect fluorescent antibody test

Babesia bigemina
Callow, L. L., 1976, World Animal Rev. (18), 9-15
Anaplasma centrale, Babesia argentina, B. bigemina, cattle, factors in transmission by Boophilus microplus, immunization with strains from splenectomized calves, review: Australia

Babesia bigemina
Callow, L. L., 1977, Advances Exper. Med. and Biol., v. 93, 121-149
bovine babesiosis, vaccination, review: biology of Babesia of cattle (taxonomy and distribution, life cycle and transmission by ticks, pathological effects, immunology); need for vaccination (epizootic spread, cattle imported to enzootic area, enzootic instability); approaches to vaccination (premunition, surveillance and treatment, chemoprophylaxis, stimulation of nonspecific immunity, experimental approaches to antigen production); vaccination against B. argentina (= bovis) in Australia; vaccination against Babesia bigemia; future requirements and developments

Babesia bigemina
Callow, L. L.; et al., 1974, Austral. Vet. J., v. 50 (1), 12-15
Babesia bigemina, cattle self-cured, drug-cured with imidocarb or with persistent infections showed an appreciable or strong degree of immunity to challenge (the finding is contrary to the concept of premunition); indirect fluorescent antibody test (antibody titre did not reflect degree of resistance to challenge)

Babesia bigemina
Castro, E. R.; and Canabez, F., 1968, Bol. Chileno Parasitol., v. 23 (1-2), 30-33
Babesia bigemina in splenectomized and normal bovines, effects of various levels of radiation on infectivity of whole blood

Babesia bigemina
Corrier, D. E.; and Adams, L. G., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 60-65
Anaplasma marginale, Babesia bigemina, concurrent infections in calves (exper.), clinical course, serological response, pathological manifestations

Babesia bigemina
Dalgliesh, R. J., 1972, Research Vet. Sc., v. 13 (6), 540-545
Babesia bigemina, evaluation of glycerol as a freezing protectant, infectivity of parasitised blood containing glycerol for cattle inoculated intravenously vs. subcutaneously

Babesia bigemina
Decia, C.; and Castro, E. R., 1968, Bol. Chileno Parasitol., v. 23 (1-2), 26-29
Babesia bigemina in calves (exper.), serum proteins and hematologic variations before infection, during prepatent, patent, and convalescent periods

Babesia bigemina
Dwivedi, S. K.; and Gautam, O. P., 1977, Indian J. Animal Sc., v. 46 (12), 1976, 627-629
Babesia bigemina, splenectomized calves, berenil treatment during acute stage, no immunity to challenge with heterologous strain; treatment during carrier stage, survival after heterologous challenge

Babesia bigemina
Dwivedi, S. K.; and Gautam, O. P., 1977, Indian J. Animal Sc., v. 47 (8), 455-457
Babesia bigemina, calves (exper.), glutamic oxalacetic transaminase and glutamic pyruvic transaminase serum levels

Babesia bigemina
Dwivedi, S. K.; Gautam, O. P.; and Banerjee, D. P., 1977, Indian Vet. J., v. 54 (9), 697-702
Babesia bigemina, splenectomized calves, imidocarb dipropionate, therapeutic and prophylactic trials, excellent results, development of acquired immunity following treatment

Babesia bigemina
Dwivedi, S. K.; Sharma, S. P.; and Gautam, O. P., 1976, Indian Vet. J., v. 53 (6), 469-472
Babesia bigemina, exotic and cross-bred cattle, 3 clinical cases, berenil with belamyl as supportive therapy; Boophilus sp. tick recovered from one cow: India

Babesia bigemina, illus.
Friedhoff, K. T.; and Buescher, G., 1976, Ztschr. Parasitenk., v. 50 (3), 345-347
Babesia bigemina, rediscovery of varied stages described by Koch (1905, 1906) as "Strahlenkorper", description, possible evidence for sexual reproduction

Babesia bigemina
Friedhoff, K. T.; and Weber, G., 1973, Isotopes and Radiation Parasitol. III, 165-168
Babesia spp., identification and characterization of certain developmental stages, review

Babesia bigemina
Gill, B. S.; and Khan, M. H., 1972, Indian J. Animal Sc., v. 42 (12), 1071-1073
Babesia bigemina and B. argentina strains injected intravenously into cow calves and buffalo calves, imidocarb dihydrochloride, therapeutic efficacy in calves after splenectomy, subinoculation test

Babesia bigemina
Goldman, M.; and Bukovsky, E., 1973, J. Protozool., v. 20 (4), 531
Babesia bigemina, soluble antigen capable of reacting with antiserum in gel diffusion plates has been extracted from infected plasma, hemolysates of infected red blood cells, and from sonicated parasites

Babesia bigemina
Goldman, M.; and Bukovsky, E., 1975, J. Protozool., v. 22 (2), 262-264
Babesia bigemina-infected bovine blood, extraction of soluble precipitating antigen, preliminary use in gel diffusion test with laboratory and field cattle

Babesia bigemina
Gonzalez, E. F.; Todorovic, R. A.; and Thompson, K. C., 1976, Tropenmed. u. Parasitol., v. 27 (4), 427-437
Anaplasma marginale, Babesia argentina, and B. bigemina in calves (exper.), immunization, economical method using minimum infective doses of parasite stabilates produced mild post-immunization reactions but conferred complete protection against bloodborne challenge, evaluation under laboratory conditions shows promise

Babesia bigemina
Goodger, B. V., 1973, Austral. Vet. J., v. 49 (2), 81-84
improved sensitivity of haemagglutination test for Babesia bigemina antibody using antigen obtained from lysate of infected erythrocytes; cross-reaction with B. argentina antisera, comparing cross-reactions of antigens from both species enables differentiation without inhibition techniques

Babesia bigemina
Graves, I. L.; Adams, W. H.; and Pyakural, S., 1975, Am. J. Vet. Research, v. 36 (6), 843-846
Babesia bigemina, Bos grunniens moved from high to low altitude and challenged with influenza A viruses, hemolytic anemia, possible explanations, death due to Fasciola hepatica and F. gigantica, incidental finding of Bunostomum sp., Trichuris sp., Neoascaris vitulorum, Dictyocaulus sp., coccidia, some reasons for poor survival of yaks at low altitude: Nepal

Babesia bigemina (Smith and Kilborne, 1893)
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
Babesia, validity of different species causing tick-fever in cattle, review

Babesia bigemina
Johnston, L. A. Y.; Trueman, K. F.; and Pearson, R. D., 1977, Austral. Vet. J., v. 53 (5), 222-226
Babesia argentina, B. bigemina, cattle (nat. and exper.), comparison of Giemsa and direct fluorescent antibody staining for diagnosis, determination of length of time Babesia may be recognized in tissues after host death; field cases of B. argentina were due to primary infections rather than relapses

Babesia bigemina
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Babesia bigemina
Kuttler, K. L.; Adams, L. G.; and Todorovic, R. A., 1977, Am. J. Vet. Research, v. 38 (2), 153-156
Babesia bigemina, infected mature cattle, comparison of complement fixation and indirect fluorescent antibody reactions, CF more sensitive than IFA in longer infection, both successful in early infections

Babesia bigemina
Kuttler, K. L.; and Johnson, L. W., 1977, Vet. Med. and Small Animal Clin., v. 72 (8), 1354, 1356-1359
Anaplasma marginale, Babesia bigemina, B. argentina, Holstein heifers destined for shipment to Nicaragua, premunition by inoculation with organisms, oxytetracycline given to heifers administered attenuated or non-attenuated A. marginale; apparently resistant to field challenge in Nicaragua

Babesia bigemina
Leeflang, P.; et al., 1976, Internat. J. Parasitol., v. 6 (2), 159-161
survey of 173 males using indirect fluorescent antibody technique with Babesia bovis, B. bigemina, and B. ratti as antigens, babesial antibodies detected in 54%, no correlation found with mode of living, contact with livestock, incidence of malaria parasitaemias or malarial antibodies, no Babesia organisms found in blood smears or by animal subinoculation, source of antibodies unknown but subclinical infection not likely: Nigeria

Babesia bigemina
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
Boophilus decoloratus confirmed as vector of Babesia bigemina: northern Nigeria

Babesia bigemina
Loehr, K.F., 1972, J. Protozool., v. 19 (4), 658-660
Babesia bigemina, immunity in cattle (exper.), concluded that premunity is followed by sterile immunity which lasts for at least 6 months and thereafter fades gradually with time, also concluded that minimum period of contact between host and parasite is required for acquisition of immunity, capillary tube agglutination test sensitive but unsuitable for detection of carrier animals

Babesia bigemina
Loehr, K. F.; Ross, J. P. J.; and Meyer, H., 1974, Tropenmed. u. Parasitol., v. 25 (2), 217-226
sera of 1505 game animals of 19 different species screened for antibodies to Anaplasma marginale, Babesia bigemina, and Theileria parva, capillary tube agglutination and indirect fluorescent antibody tests, antibodies more prevalent in sera of antelopes grazing in vicinity of non-dipped cattle than in areas where cattle are either dipped regularly or are not present at all, need for studies on transmission of these organisms from game to cattle and vice versa: Kenya; Tanzania; Uganda; Zambia

Babesia bigemina
Mahoney, D. F.; Wright, I. G.; and Mirre, G. B., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 197-203
Babesia argentina, B. bigemina, calves, persistence of immunity after natural infection in early life and subsequent tick-free maintenance, concluded that animals naturally infected during calfhood are unlikely to require vaccination for protection against babesiosis in later life

Babesia bigemina
Mehlitz, D.; and Ehret, R., 1974, Tropenmed. u. Parasitol., v. 25 (1), 3-10
Anaplasma, Babesia, Theileria, cattle, serological survey, capillary-tube agglutination test, indirect fluorescent antibody test, and complement fixation test: Botswana

Babesia bigemina, illus.
Morzaria, S. P.; and Young, A. S., 1977, Research Vet. Sc., v. 23 (1), 55-58
Babesia bigemina, identification of developmental stages in the haemolymph of Boophilus decoloratus using indirect fluorescent antibody technique

Babesia bigemina
Morzaria, S. P.; Young, A. S.; and Hudson, E. B., 1977, Parasitology, v. 74 (3), 291-298
Babesia bigemina, successful experimental transmission of a Kenyan strain by Boophilus decoloratus, production of tick-derived stabilates

Babesia bigemina
Pandey, N. N.; and Mishra, S. S., 1977, Zentralbl. Vet.-Med., Reihe B, v. 24 (7), 522-528
Babesia bigemina in indigenous calves (exper.), macroscopical and histopathological changes: India

Babesia bigemina
Pandey, N. N.; and Mishra, S. S., 1977, Indian Vet. J., v. 54 (11), 880-883
Babesia bigemina, indigenous cattle (exper.), haematological changes and blood glucose level: India

Babesia bigemina
Parker, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 387-390
Babesia argentina, direct counting technique for estimating high parasitaemias, also suitable for B. bigemina and Plasmodium berghei but not for B. rodhaini or Anaplasma centrale

Babesia bigemina
Ranatunga, P.; and Wanduragala, L., 1972, Brit. Vet. J., v. 128 (1), 9-18
Babesia bigemina, Anaplasma centrale, premunization of Jersey cattle imported to Ceylon from New Zealand, reactions and haematology

Babesia bigemina
Rodriguez, O. N.; et al., 1975, Rev. Cubana Cien. Vet., v. 5 [on cover], v. 6 [on title page] (1-2), 1-8
Babesia bigemina, B. argentina, splenectomized cattle, correlation between duration of infection, symptomatology and laboratory data, comparison of species

Babesia bigemina
Rodriguez, O. N.; Espaine, L.; and Rivas, A., 1975, Rev. Cubana Cien. Vet., v. 5 [on cover], v. 6 [on title page] (1-2), 145-153
Theileria mutans, incidence in cattle with or without Anaplasma marginale, Babesia bigemina and B. argentina natural infections: Cuba

Babesia bigemina
Schreuder, B.E.C.; Uilenberg, G.; and Tondeur, W., 1977, Tropenmed. u. Parasitol., v. 28 (3), 367-371
Theileriidae, reinfection and retransmission experiments in cattle and African buffalo

Babesia bigemina
Smith, M. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 18 [Demonstration]
cattle: Trinidad

Babesia bigemina
Smith, M. W., 1977, PANS, v. 23 (1), 27-32
Babesia bigemina, Anaplasma marginale, immunization of susceptible imported cattle using whole blood inoculations from infected donor cattle: Trinidad, imported from Canada

Babesia bigemina
Thompson, K. C., 1976, Vet. Parasitol., v. 2 (2), 223-229
Boophilus microplus, establishment and maintenance of colony free of infection with Anaplasma marginale and Babesia spp., subsequent infection with pure isolate of Babesia bigemina; description of life cycle of Boophilus microplus

Babesia bigemina
Thompson, K. C.; Todorovic, R. A.; and Hidalgo, R. J., 1977, Research Vet. Sc., v. 23 (1), 51-54
Babesia bigemina, antigenic variation in 4 stabilates propagated as acute and chronic blood-borne and tick-borne infections of Colombian cattle, characterization by means of complement fixation, gel diffusion, agar gel electrophoresis, and indirect haemagglutination tests

Babesia bigemina, illus.
Todorovic, R., 1976, Vet. Parasitol., v. 2 (1), 97-109
Babesia spp., cattle, review: serodiagnosis, immunization (sterile immunity; premunition), chemoprophylaxis with imidocarb, vectors: Colombia

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Todorovic, R. A.; and Gonzalez, E. F., 1975, Rev. Inst. Colomb. Agropec., v. 10 (1), 87-99
Babesia bigemina, B. argentina, cattle, killed vaccine produced from infected erythrocytes and plasma, high degree of sterile immunity produced in calves: Colombia

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Todorovic, R. A.; and Gonzalez, E. F., 1975, Rev. Inst. Colomb. Agropec., v. 10 (2), 243-254
Babesia bigemina, B. argentina, Holstein-Friesian calves immunized with blood from carrier animals or from inoculated, splenectomized calves, challenge by Babesia-infected Boophilus microplus, weight gain records; better results with carrier blood

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Babesia bigemina, B. argentina, calves, sterile immunity using killed-Babesia vaccine, field-borne challenge with Boophilus microplus infected ticks; possible important role in mechanism of acquired immunity

Babesia bigemina
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indirect fluorescent antibody test found superior to complement fixation test as epidemiologic tool to detect Babesia bigemina and B. argentina infections in cattle, some cross reactivity in differentiating between spp.: Colombia

Babesia bigemina
Todorovic, R. A.; and Tellez, C. H., 1975, Trop. Animal Health and Prod., v. 7 (3), 125-131
adult cattle (Bos taurus), premunition against Babesia bigemina and B. argentina using Imidocarb or Ganaseg to control post premunition reactions was effective and practical; premunition against Anaplasma marginale using oxytetracycline and Gloxazone was less effective and not practical

Babesia bigemina
Trees, A. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 287 [Demonstration]
blood from healthy indigenous Nigerian cattle inoculated into splenectomized calf, calf became infected with Babesia bigemina, B. bovis, Anaplasma marginale, Borrelia theileri, Theileria mutans, Eperythrozoon teganodes, E. wenyoni; isolation of pure stabilate of B. bigemina

Babesia bigemina
Uilenberg, G.; et al., 1976, Tropenmed. u. Parasitol., v. 27 (3), 329-336
Theileria parva, cattle immunized by infection and treatment method using 3 strains of T. parva survived exposure to natural tick infestation; all controls died from T. parva infections, 3 immunized cattle died from T. mutans or Babesia bigemina infection but none from T. parva

Babesia bigemina
Vizcaino G., O.; and Todorovic, R. A., 1975, Rev. Inst. Colomb. Agropec., v. 10 (1), 77-85
Babesia bigemina, B. argentina, characterization of antigens, complement fixation, immunodiffusion, cross-immunity tests

Babesia bigemina
Weber, G., 1976, Ztschr. Parasitenk., v. 50 (2), 185
Babesia ovis and B. bigemina in blood films, immunocytochemical detection of catabolic enzymes, possible implications

Babesia bigemina
Weilgama, D. J.; Jayasekara, M. U.; and Hussain, M., 1975, Ceylon Vet. J., v. 23 (3-4), 49-53
Babesia bigemina, B. argentina, Anaplasma centrale, A. marginale, three breeds of cattle (Ayrshire, Friesian and Shorthorn), haematological changes including parasitaemia and temperature reactions following premunisation with blood from infected cattle: Sri Lanka, imported from New Zealand

Babesia bigemina
Weisman, J.; Goldman, M.; and Pipano, E., 1974, J. Protozool., v. 21 (3), 466 [Abstract]
Babesia bigemina, B. berbera, passive transfer of antibodies from cows to calves via colostrum

Babesia bigemina
Wright, I. G., 1973, Research Vet. Sc., v. 14 (1), 29-34
Babesia argentina, B. babesia, splenectomised calves (exper.), haematology

Babesia bovis, illus.
Aeschlimann, A.; Brossard, M.; and Quenet, G., 1975, Acta Trop., v. 32 (4), 281-289
bovins (sang): Suisse

Babesia bovis, illus.
Brossard, M., 1976, Rev. Suisse Zool., v. 83 (2), 443-462
morphology, clinical signs in cattle, diagnosis by examination of blood, successful treatment with Berenil or Imizol by intramuscular injection, role of Ixodes ricinus as vector, distribution; "B. berbera, B. argentina and B. bovis . . . should probably be considered as races of the same species, B. bovis"
Ixodes ricinus (nat. and exper.)
cattle
all from Low Plain of the Rhone, Switzerland

Babesia bovis
Callow, L. L.; Quiroga, Q. C.; and McCosker, P. J., 1976, Internat. J. Parasitol., v. 6 (4), 307-310
Babesia argentina, Anaplasma marginale, comparison of strains of each from Australia vs. Bolivia with indirect fluorescent antibody test showed serological identity of the two strains of each parasite, implications for vaccination; since earlier study showed serological identity between B. bovis and B. argentina, the small Babesia of Australia and South America should by priority be called B. bovis

Babesia bovis
Coombs, W. G., 1977, Canad. Vet. J., v. 18 (7), 193-195
cattle (blood), clinical findings, treatment with aminocarbalide (Diampron): Devon, England

Babesia bovis
Dalgliesh, R. J.; et al., 1977, Research Vet. Sc., v. 23 (1), 105-108
Babesia bovis, Bos taurus (exper.), protamine sulphate test as a screening test for disseminated intravascular coagulation

Babesia bovis (=B. argentina)
Dalgliesh, R. J.; and Stewart, N. P., 1976, Austral. Vet. J., v. 52 (11), 543 [Letter]
Babesia bovis, development of infective forms in unfed Boophilus microplus larvae, various temperature conditions, possible use for vaccine

Babesia bovis
Dalgliesh, R. J.; and Stewart, N. P., 1977, Austral. Vet. J., v. 53 (9), 429-431
Babesia bovis, failure of vaccine strains to regain infectivity for Boophilus microplus during long-standing infections in cattle, concluded that the number and frequency of blood passages performed with a strain may determine its infectivity for ticks

Babesia bovis
De Jonghe, R.; and Vermeiren, G., 1977, Vlaams Diergeneesk. Tijdschr., v. 46 (1), 61-62
Babesia bovis outbreak, cattle; Ixodes ricinus found on sick cattle: Wallonie

Babesia bovis (Babes, 1888) Starcovici, 1893
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
Babesia, validity of different species causing tick-fever in cattle, review
synonymy

Babesia bovis
Leeflang, P.; et al., 1976, Internat. J. Parasitol., v. 6 (2), 159-161
survey of 173 males using indirect fluorescent antibody technique with Babesia bovis, B. bigemina, and B. ratti as antigens, babesial antibodies detected in 54%, no correlation found with mode of living, contact with livestock, incidence of malaria parasitaemias or malarial antibodies, no Babesia organisms found in blood smears or by animal subinoculation, source of antibodies unknown but subclinical infection not likely: Nigeria

Babesia bovis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Babesia bovis
Mahoney, D. F.; and Mirre, G. B., 1977, Research Vet. Sc., v. 23 (1), 126-127
Syn.: Babesia argentina
Babesia bovis, observations on dynamics of transovarial transmission by Boophilus microplus allowed selection of larval tick cultures containing a high proportion of infected individuals

Babesia bovis, illus.
Potgieter, F. T.; and Els, H. J., 1976, Onderstepoort J. Vet. Research, v. 43 (3), 123-128
Babesia bovis, merozoite development in salivary glands of Boophilus microplus larvae, ultrastructure

Babesia bovis (Babes, 1888), illus.
Potgieter, F. T.; Els, H. J.; and van Vuuren, A. S., 1976, Onderstepoort J. Vet. Research, v. 43 (1), 1-9
Babesia bovis, ultrastructure of merozoites in gut epithelial cells of Boophilus microplus, electron microscopy

Babesia bovis
Trees, A. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 287 [Demonstration]
blood from healthy indigenous Nigerian cattle inoculated into splenectomized calf, calf became infected with Babesia bigemina, B. bovis, Anaplasma marginale, Borrelia theileri, Theileria mutans, Eperythrozoon teganodes, E. wenyoni; isolation of pure stabilate of B. bigemina

Babesia bovis, illus.
de Vos, A. J.; Imes, G. D.; and Cullen, J. S. C., 1976, Onderstepoort J. Vet. Research, v. 43 (2), 75-77
Babesia bovis, new-born Brown Swiss calf (blood, brain capillaries), neuropathological lesions, fatal infection: Cwaka College, Kwazulu

Babesia bovis (=B. argentina)
Wright, I. G.; and Kerr, J. D., 1977, J. Comp. Path., v. 87 (4), 531-537
Babesia bovis, splenectomized calves, decreases in blood pressure and packed cell volume, plasma kallikrein activation started, possibly by parasite secretions

Babesia caballi
Correa, J.; et al., 1977, Bol. Chileno Parasitol., v. 32 (1-2), 21-22
Babesia equi, B. caballi, serological survey using complement fixation test for evidence of equine piroplasmosis in race horses at Santiago, Chile racetracks

Babesia caballi
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Babesia caballi
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Babesia caballi (Nuttall and Strickland, 1910)
Lima, J. D.; Biondini, J.; and Reis, R., 1976, Arq. Escola Vet. Univ. Fed. Minas Gerais, v. 28 (1), 87-92
Babesia caballi, horse, treated with Ganaseg: Minas Gerais, Brasil

Babesia caballi
Rudolph, W.; Rosende, S.; and Correa, J., 1973, Bol. Chileno Parasitol., v. 28 (1-2), 3-6
Babesia equi, B. caballi, complement fixation micromethod effective diagnostic test for equine piroplasmosis: Chile

Babesia caballi
Smith, M. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 18 [Demonstration]
horses: Trinidad

Babesia caballi
Urcelay, S.; Correa, J.; and Rudolph, W., 1973, Bol. Chileno Parasitol., v. 28 (1-2), 6-9
Babesia equi, B. caballi, serological survey of incidence of equine piroplasmosis in race horses: breeding farms, Santiago Province, Chile

Babesia canis, illus.
Aeschlimann, A.; Brossard, M.; and Quenet, G., 1975, Acta Trop., v. 32 (4), 281-289
chien (sang): Suisse

Babesia canis
Bigalke, R. D., 1976, J. South African Vet. Ass., v. 47 (4), 281-282
Babesia canis, canine, relapses due to failure of immune mechanism, not to inadequate therapy, brief theoretical review

Babesia canis
Button, C., 1976, J. South African Vet. Ass., v. 47 (4), 284-287
dogs, fluid therapy, patient selection, fluid replacement, route of administration and composition

Babesia canis
Bwangamoi, O.; and Isyagi, A. O., 1973, Bull. Epizoot. Dis. Africa, v. 21 (1), 33-37
Babesia canis, dogs, incidence survey: Uganda

Babesia canis
Chiodini, P. L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 27-28 [Demonstration]
Babesia canis, in vitro culture; B. rodhaini and B. divergens in leucocyte-free dilution cultures, effect of imidocarb on leucine and adenosine incorporation, possible use of system for in vitro screening of potential babesicides and also for mode of action studies

Babesia canis
Colgrove, D. J., 1976, J. Am. Vet. Med. Ass., v. 168 (7), 591-592
Babesia canis, dogs, clinical signs, diagnosis, pyrimethamine therapy, possible tick vectors: Oahu Island, Hawaii

Babesia canis
Cowper, S. G., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 415-425
continuous flow apparatus for in vitro maintenance: Schistosoma mansoni, S. haematobium, survival time, carbohydrate metabolism; Plasmodium knowlesi, morphology and carbohydrate metabolism; preliminary attempts to cultivate Trypanosoma vivax and Babesia canis

Babesia canis, illus.
Geyer, S.; and Rathelbeck, H. G., 1976, Kleintier-Praxis, v. 21 (1), 3-11
Babesia canis, 2 case histories, dogs exposed to ticks in Mediterranean area, Acaprin successful in one case

Babesia canis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Babesia canis
Leeflang, P.; Pimentel, W. J.; and Blotkamp, J., 1974, Bull. Epizoot. Dis. Africa, v. 22 (1), 51-53
Babesia canis, Hepatozoon canis, Haemobartonella canis, dog, case report, also infested with Rhipicephalus sanguineus: Nigeria

Babesia canis
Malherbe, W. D.; et al., 1976, J. South African Vet. Med. Ass., v. 47 (1), 29-33
Babesia canis, dogs, acid-base values for blood of poor-prognosis cases, supportive treatment with sodium bicarbonate reduced acidosis: Onderstepoort

Babesia canis
Michael, S. A.; and El Refaii, A. H., 1976, Agric. Research Rev., v. 54 (7), 31-34
Babesia canis, maintenance in laboratory for antigen production, infected canine blood at 6°C in ACD solution, method more advantageous than deep freezing technique

Babesia canis, illus.
Niak, A.; Anwar, M.; and Khatibi, S., 1973, Trop. Animal Health and Prod., v. 5 (3), 200-201
dogs (blood) (nat. and exper.): northern Iran

Babesia canis
Oduye, O. O.; and Dipeolu, O. O., 1976, J. Small Animal Practice, v. 17 (5), 331-337
blood parasites of dogs, single and mixed infections, correlation between incidence and rainfall, degree of parasitaemia, infectivity rate within age groups, no significant difference in host susceptibility of local and exotic breeds to infection: Ibadan, Nigeria

Babesia canis
Smith, M. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 18 [Demonstration]
dogs: Trinidad

Babesia capreoli
Adam, K. M. G.; et al., 1976, Parasitology, v. 73 (1), 1-11
antibody-positive blood from wild Cervus elaphus produced Babesia infections in splenectomized C. elaphus (also Eperythrozoon infection resembling wenyoni), possible transient deer Babesia infection produced in 1 of 6 bovine calves, indirect fluorescent antibody tests, "Despite their similarities, specific status for B. divergens and the red deer Babesia is probably justified; at present there is insufficient evidence to justify separation of the red deer Babesia from B. capreoli."

Babesia caucasica (Yakimoff and Belawine, 1926)
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
as syn. of Babesia divergens (M'Fadyean and Stockman, 1911)

Babesia colchica (Yakimoff, 1928)
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
as syn. of Babesia bovis (Babes, 1888) Starcovici, 1893

Babesia divergens, illus.
Adam, K. M. G.; et al., 1976, Parasitology, v. 73 (1), 1-11
antibody-positive blood from wild Cervus elaphus produced Babesia infections in splenectomized C. elaphus (also Eperythrozoon infection resembling wenyoni), possible transient deer Babesia infection produced in 1 of 6 bovine calves, indirect fluorescent antibody tests, "Despite their similarities, specific status for B. divergens and the red deer Babesia is probably justified; at present there is insufficient evidence to justify separation of the red deer Babesia from B. capreoli."

Babesia divergens
Adam, K. M. G.; and Blewett, D. A., 1974, J. Protozool., v. 21 (3), 448 [Abstract]
Babesia divergens, successful exper. infection of mice, possible practical applications

Babesia divergens, illus.
Aeschlimann, A.; Brossard, M.; and Quenet, G., 1975, Acta Trop., v. 32 (4), 281-289
bovins (sang): Suisse

Babesia divergens
Blewett, D. A.; and Adam, K. M. G., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 284 [Demonstration]
Babesia divergens, serological screening of 20,000 bovine sera, antibody incidence: Scotland

Babesia divergens
Brocklesby, D. W.; Harradine, D. L.; and Young, E. R., 1976, Research Vet. Sc., v. 21 (3), 300-302
Babesia major infected calves not protected against B. divergens; B. divergens infections provide good protection against B. major; B. divergens might be dominant species where both occur

Babesia divergens
Brocklesby, D. W.; and Purnell, R. E., 1977, Nature, London (5592), v. 265, 343
Babesia divergens, calves, failure of BCG to protect against infection

Babesia divergens
Chiodini, P. L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 27-28 [Demonstration]
Babesia canis, in vitro culture; B. rodhaini and B. divergens in leucocyte-free dilution cultures, effect of imidocarb on leucine and adenosine incorporation, possible use of system for in vitro screening of potential babesicides and also for mode of action studies

Babesia divergens
Dolan, T. T., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 274 [Demonstration]
Babesia divergens in calves (exper.), B. rodhaini in rats (exper.), thrombocytopenia with marked hemolytic anemia

Babesia divergens
Donnelly, J.; Joyner, L. P.; and Crossman, P. J., 1972, Research Vet. Sc., v. 13 (6), 511-514
Babesia divergens, cattle, natural outbreak in dairy herd monitored by indirect immunofluorescent antibody test, quantitative estimate of incidence rate of infection, level of infection correlated with period at risk, high incidence, low morbidity, no evidence that amicarbalide protected prophylactically: Sussex

Babesia divergens
Friedhoff, K. T.; and Weber, G., 1973, Isotopes and Radiation Parasitol. III, 165-168
Babesia spp., identification and characterization of certain developmental stages, review

Babesia divergens (M'Fadyean and Stockman, 1911)
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
Babesia, validity of different species causing tick-fever in cattle, review
synonymy

Babesia divergens
Hughes, D. L.; Purnell, R. E.; and Brocklesby, D. W., 1977, Vet. Rec., v. 100 (15), 320-321
Fasciola hepatica in intact and splenectomised calves, effect of subsequent Babesia divergens infections, results show tendency for F. hepatica to exacerbate B. divergens infections of intact calves, but not splenectomised calves

Babesia divergens
Irvin, A. D.; and Young, E. R., 1977, Nature, London (5627), v. 269, 407-409
Babesia spp., in vitro test for screening drug activity which exploits finding that Babesia parasites readily incorporate hypoxanthine in an in vitro culture system, may also be used to detect drug resistance patterns and have possible application in primary screening of antimalarial drugs

Babesia divergens
Irvin, A. D.; and Young, E. R.; and Purnell, R. E., 1977, Parasitology, v. 75 (2), xxviii [Abstract]
Babesia spp. of mice and cattle, uptake of tritiated purines and pyrimidines, uptake of hypoxanthine inhibited in vitro by antibabesial drugs in a degree roughly proportional to drugs' efficacy in vivo, may be useful as primary screening method for active drugs

Babesia divergens
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Babesia divergens
Joyner, L. P.; et al., 1972, Research Vet. Sc., v. 13 (6), 515-518
species-specificity of indirect fluorescent antibody test for differentiating infections with Babesia divergens vs. Babesia major, cattle (nat. and exper.), suitable for use in surveys

Babesia divergens, illus.
Mohammed, A. N., 1976, Tijdschr. Diergeneesk., v. 101 (8), 413-420
experimental transovarial transmission to cattle by Ixodes ricinus

Babesia divergens
Purnell, R. E.; et al., 1976, Vet. Rec., v. 99 (6), 102
Babesia divergens, cattle, diagnosis, microplate enzyme-linked immunosorbent assay

Babesia divergens
Purnell, R. E.; et al., 1976, J. Comp. Path., v. 86 (4), 609-614
Babesia divergens, statistical comparison of isolates from England, Eire, Scotland and Wales with Weybridge strain, pathogenicity for splenectomized calves

Babesia divergens
Purnell, R. E.; et al., 1976, Vet. Rec., v. 99 (21), 415-417
Babesia divergens and Ehrlichia phagocytophila, natural mixed infection, agents isolated, splenectomised calves, reactions after inoculation singly, simultaneously or sequentially, B. divergens infection delayed by E. phagocytophila: Eire

Babesia divergens
Purnell, R. E.; et al., 1977, Parasitology, v. 75 (2), xxxi [Abstract]
Babesia divergens, effects of inoculating different numbers of piroplasms into splenectomized calves, comparison with reaction to irradiated parasites

Babesia divergens
Purnell, R. E.; et al., 1977, Research Vet. Sc., v. 23 (1), 124-125
titration of infective dose in splenectomized calves

Babesia divergens
Purnell, R. E.; et al., 1977, Vet. Rec., v. 100 (1), 4-6
Babesia divergens, splenectomized calves (exper.), mixed infection with Ehrlichia phagocytophila resulted in less marked changes in haematology, apparent suppression of Babesia by Ehrlichia

Babesia divergens
Purnell, R. E.; and Brocklesby, D. W., 1977, Research Vet. Sc., v. 23 (2), 255-256
Babesia divergens, limited protection against isologous and homologous challenge observed in splenectomized calves inoculated with lyophilised plasma from infected animal

Babesia divergens
Purnell, R. E.; Brocklesby, D. W.; and Harradine, D. L., 1975, Vet. Parasitol., v. 1 (2), 199-202
Babesia divergens, B. major, Theileria mutans, successful cryopreservation using either dimethyl sulphoxide or glycerol as cryoprotectants

Babesia divergens
Trees, A. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 277 [Demonstration]
Babesia divergens, use of acridine orange staining to quantitate low levels of parasitemia

Babesia divergens
Voller, A.; Bartlett, A.; and Bidwell, D. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 98-106
application of enzyme immunoassay to diagnosis of human parasitic diseases, technique for microplate enzyme-linked immunosorbent assay

Babesia equi, illus.
Banerjee, D. P.; et al., 1977, Trop. Animal Health and Prod., v. 9 (3), 153-158
Babesia equi, donkeys (vaccinated, infected and carrier intact, and splenectomised), intradermal skin test developed to demonstrate hypersensitivity reaction, leucocyte migration inhibition test developed

Babesia equi, illus.
Carpio, J., 1972, Rev. Invest. Pecuarias, v. 1 (2), 177-183
Babesia equi, isolation from race horses, course of experimental infection in splenectomized and normal horses, first finding in Peru: Lima

Babesia equi
Cordero del Campillo, M.; et al., 1974, An. Fac. Vet. Leon, v. 20, 23-34
Babesia equi, pathology, review of Spanish and Portuguese literature, list of ticks considered present and responsible as vectors in Iberian Peninsula
horse: Leon (NW Spain)

Babesia equi
Cordero del Campillo, M.; et al., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 305-315
case history, horse infected with E[rysipelothrix] insidiosa to produce hyperimmune serum, rapid death from Babesia, considered to be activation of carrier state; possible tick vectors reviewed: Leon (N.W. Spain)

Babesia equi
Cordero del Campillo, M.; et al., 1974, Trab. Estac. Agric. Exper. Leon, v. 11, 11-22
Babesia equi, horse, pathology: Leon (NW Spain)

Babesia equi
Correa, J.; et al., 1977, Bol. Chileno Parasitol., v. 32 (1-2), 21-22
Babesia equi, B. caballi, serological survey using complement fixation test for evidence of equine piroplasmosis in race horses at Santiago, Chile racetracks

Babesia equi, illus.
Frerichs, W. M.; and Holbrook, A. A., 1974, J. Protozool., v. 21 (5), 707-709
Babesia equi, feeding mechanism, organelles involved with ingestion of nutrients (cytostome for taking in hemoglobin and previously undescribed tubule which appears to ingest plasma)

Babesia equi, illus.
Gautam, O. P.; and Dwivedi, S. K., 1976, Indian Vet. J., v. 53 (7), 546-551
Babesia equi, outbreak in mares, clinical pathology, diagnosis, treatment with Berenil, vectors, Hyalomma spp.
mares (nat. and exper.): Hissar, imported from Poland
donkey (exper.)

Babesia equi
Ibanez, E. A.; et al., 1976, Gac. Vet., Buenos Aires (307), v. 38, 7-13
Babesia equi, equine, determination of carriers, application of glucocorticoids to produce hosts for treatment studies

Babesia equi
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Babesia equi, illus.
Mahoney, D. F.; et al., 1977, Austral. Vet. J., v. 53 (10), 461-464
Babesia equi, isolated from horse (nat.) (blood), identification, histopathology, mild disease in intact horse (exper.), acute babesiosis in splenectomised horse (exper.): Bowral, New South Wales, Australia

Babesia equi
Rudolph, W.; Rosende, S.; and Correa, J., 1973, Bol. Chileno Parasitol., v. 28 (1-2), 3-6
Babesia equi, B. caballi, complement fixation micromethod effective diagnostic test for equine piroplasmosis: Chile

Babesia equi
Urcelay, S.; Correa, J.; and Rudolph, W., 1973, Bol. Chileno Parasitol., v. 28 (1-2), 6-9
Babesia equi, B. caballi, serological survey of incidence of equine piroplasmosis in race horses: breeding farms, Santiago Province, Chile

Babesia equi
Yap, L. F.; and Groves, M. G., 1968, Med. J. Malaya, v. 22 (3), 229
Tapirus indicus (blood): National Zoo in Ulu Klang

Babesia equi, illus.
Young, A. S.; and Purnell, R. E., 1973, Bull. Epizoot. Dis. Africa, v. 21 (4), 377-383
development of Babesia equi in the salivary glands of Rhipicephalus evertsi
Equus burchelli (blood): north-western region of Loliondo controlled area, Tanzania

Babesia equi
Young, E.; et al., 1973, Research J. National Parks Republic South Africa (16), 77-81
Equus zebra zebra (blood): Mountain Zebra National Park

Babesia galagolata
Dennig, H. K.; and Hoesl, G., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 79-84
morphology, course of infection, host specificity, clinical signs
Galago crassicaudatus (nat. and exper.): caught in Tanzania and brought to Germany
Macaca mulatta (exper.)
Mus musculus var. alb. (exper.)
rats (exper.)
Cavia porcellcus (exper.)
hamsters (exper.)
gerbils (exper.)

Babesia gibsoni
Botros, B. A. M.; Moch, R. W.; and Barsoum, I. S., 1975, Am. J. Vet. Research, v. 36 (3), 293-296
Babesia gibsoni, splenectomized and nonsplenectomized dogs (exper.), inoculation with fresh and preserved blood, prepatent period, clinical signs, duration of parasitemia, gross pathologic changes, clinicopathologic changes, histopathologic features, relationship of age of dogs to pathogenicity of infection

Babesia gibsoni (blood)
Groves, M. G.; and Yap, L. F., 1968, Med. J. Malaya, v. 22 (3), 229
Canis familiaris: Kuala Lumpur

Babesia gibsoni, illus.
Niak, A.; Anwar, M.; and Khatibi, S., 1973, Trop. Animal Health and Prod., v. 5 (3), 200-201
Vulpes vulpes (blood): northern Iran

Babesia gibsoni
Oduye, O. O.; and Dipeolu, O. O., 1976, J. Small Animal Practice, v. 17 (5), 331-337
blood parasites of dogs, single and mixed infections, correlation between incidence and rainfall, degree of parasitaemia, infectivity rate within age groups, no significant difference in host susceptibility of local and exotic breeds to infection: Ibadan, Nigeria

Babesia gibsoni
Otsuka, H.; and Yoshimura, S., 1976, Bull. Fac. Agric. Univ. Miyazaki, v. 23 (2), 511-515
Babesia gibsoni, dog, transmission experiment using Haemaphysalis flava showed that this tick did not participate in either transovarial or stage-to-stage transmission

Babesia gibsoni
Ruff, M. D.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 297-307
Babesia gibsoni, serum levels of various enzymes in infected dogs with respect to possible liver damage

B[abesia] herpailuri
Goebel, E., 1976, Ztschr. Parasitenk., v. 50 (2), 207
B[abesia] musculi, B. herpailuri, morphology and topography after freeze etching

Babesia herpailuri, illus.
Goebel, E.; and Dennig, H. K., 1976, Ztschr. Parasitenk., v. 49 (2), 97-112
Babesia herpailuri from cats before and after treatment with Imidocarb, ultrastructure of intraerythrocytic trophozoites and merozoites; prophylactic effect of drug may be due to low feeding on hemoglobin

Babesia hylomysci
Hussein, H. S., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 5 [Demonstration]
Babesia microti and B. hylomysci infections in mice on a milk diet, experiments showed dependence of both species on dietary supply of p-aminobenzoic acid (PABA) to synthesize folic and folinic acids

Babesia hylomysci
Hussein, H. S., 1976, Ztschr. Parasitenk., v. 50 (2), 103-108
Babesia hylomysci, mice, ^{59}Fe labelling of hemoglobin demonstrating parasite's preference for mature erythrocytes, direct destruction of erythrocytes as main cause of anemia and autoimmune reaction as probable factor

Babesia hylomysci
Hussein, H. S., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 249-253
Babesia hylomysci, B. microti, mice (exper.), recovery from primary infection of B. microti results in sterile immunity possibly for lifetime, recovery from B. hylomysci results in premunition with infection at subpatent levels with spontaneous relapses, cross-protection occurs between the two species

Babesia hylomysci
Hussein, H. S., 1977, J. Comp. Path., v. 87 (2), 161-167
Babesia hylomysci, mice (exper.), clinical signs, histopathology of liver

Babesia hylomysci
Hussein, H. S., 1977, J. Comp. Path., v. 87 (2), 169-175
Babesia hylomysci, mice (exper.), histopathological changes in kidney

Babesia hylomysci
Momen, H., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 438-439 [Demonstration]
Babesia spp., enzyme variations as significant aid to taxonomic study

Babesia hylomysci
Momen, H.; and Chance, M. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 13-14 [Demonstration]
Babesia spp., Anthemosoma garnhami, differences in DNA buoyant densities as possible aid to taxonomic studies

Babesia hylomysci
Phillips, R. S.; and Wakelin, D., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 276 [Demonstration]
Trichuris muris-infected mice, suppression of immunity by concurrent infection with Babesia spp., severely depressed agglutinating antibody response to sheep red blood cells

Babesia hylomysci
Phillips, R. S.; and Wakelin, D., 1976, Exper. Parasitol., v. 39 (1), 95-100
mice concurrently infected with Babesia and Trichuris muris, marked immunodepression, normal immune expulsion of nematode delayed; Babesia infections had little effect on expulsion of challenge infections of T. muris from previously immunized mice; Babesia infections exerted profound immunodepressive effect on agglutinating antibody response to sheep red blood cells

Babesia karelica (Yakimoff, 1928)
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
as syn. of Babesia divergens (M'Fadyean and Stockman, 1911)

Babesia major, illus.
Aeschlimann, A.; Brossard, M.; and Quenet, G., 1975, Acta Trop., v. 32 (4), 281-289
bovins (sang): Suisse

Babesia major
Brocklesby, D. W.; Harradine, D. L.; and Young, E. R., 1976, Research Vet. Sc., v. 21 (3), 300-302
Babesia major infected calves not protected against B. divergens; B. divergens infections provide good protection against B. major; B. divergens might be dominant species where both occur

Babesia major
Brocklesby, D. W.; and Sellwood, S. A., 1973, Research Vet. Sc., v. 14 (1), 47-52
Haemaphysalis punctata collected from vegetation in Kent and Essex transmitted Babesia major to splenectomised calves, infections produced were relatively mild, Babesia major is not a likely cause of disease in British cattle

Babesia major
Findlay, C. R.; and Begg, T. B., 1977, Vet. Rec., v. 100 (19), 406
Babesia major in Bison bison, case histories, amicarbalide, good results: Kent, Britain

Babesia major (Sergent et al., 1926)
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
Babesia, validity of different species causing tick-fever in cattle, review

Babesia major
Irvin, A. D.; and Young, E. R., 1977, Nature, London (5627), v. 269, 407-409
Babesia spp., in vitro test for screening drug activity which exploits finding that Babesia parasites readily incorporate hypoxanthine in an in vitro culture system, may also be used to detect drug resistance patterns and have possible application in primary screening of antimalarial drugs

Babesia major
Irvin, A. D.; and Young, E. R.; and Purnell, R. E., 1977, Parasitology, v. 75 (2), xxviii [Abstract]
Babesia spp. of mice and cattle, uptake of tritiated purines and pyrimidines, uptake of hypoxanthine inhibited in vitro by antibabesial drugs in a degree roughly proportional to drugs' efficacy in vivo, may be useful as primary screening method for active drugs

Babesia major
Joyner, L. P.; et al., 1972, Research Vet. Sc., v. 13 (6), 515-518
species-specificity of indirect fluorescent antibody test for differentiating infections with Babesia divergens vs. Babesia major, cattle (nat. and exper.), suitable for use in surveys

Babesia major, illus.
Liebisch, A.; Melfsen, J.; and Rahman, M. S., 1976, Berl. u. München. Tierärztl. Wchnschr., v. 89 (24), 477-480
Rind: Nordfriesischen Insel Amrum; Ostfriesischen Insel Norderney und Juist

Babesia major
Mohammed, A. N., 1976, Tijdschr. Diergeneesk., v. 101 (8), 413-420
prevalence in Friesian cattle (blood), failure of experimental transmission by Ixodes ricinus or H[aemaphysalis] punctata: Netherlands

Babesia major
Morzaria, S. P.; et al., 1977, Research Vet. Sc., v. 22 (2), 190-193
Babesia major derived from Haemaphysalis punctata nymphs, cryopreservation, subsequent infectivity for cattle

Babesia major, illus.
Morzaria, S. P.; Bland, P.; and Brocklesby, D. W., 1976, Research Vet. Sc., v. 21 (1), 1-11
Babesia major, ultrastructure in infected Haemaphysalis punctata, characteristic organelles in B. major merozoites confirm this parasite to subphylum Apicomplexa

Babesia major, illus.
Morzaria, S. P.; Bland, P.; and Brocklesby, D. W., 1977, Parasitology, v. 75 (1), 125-130
Babesia major, ultrastructure of penetrating stages of merozoites responsible for causing trans-ovarian infection of Haemaphysalis punctata

Babesia major
Morzaria, S. P.; Brocklesby, D. W.; and Harradine, D. L., 1977, Vet. Rec., v. 100 (23), 484-487
Babesia major, Theileria mutans, cattle (exper.), indirect fluorescent antibody test found to be species specific, possible use in field survey of cattle in Britain

Babesia major
Morzaria, S. P.; Brocklesby, D. W.; and Harradine, D. L., 1977, Research Vet. Sc., v. 23 (2), 261-262
Babesia major, adult female Haemaphysalis punctata could be infected by feeding on infected calves but larvae and nymphs could not, all stages could transmit the parasite to calves, two syringe passages resulted in complete loss of infectivity to ticks

Babesia major
Purnell, R. E.; Brocklesby, D. W.; and Harradine, D. L., 1975, Vet. Parasitol., v. 1 (2), 199-202
Babesia divergens, B. major, Theileria mutans, successful cryopreservation using either dimethyl sulphoxide or glycerol as cryoprotectants

Babesia major
Purnell, R. E.; Brocklesby, D. W.; and Sellwood, S. A., 1973, Isotopes and Radiation Parasitol. III, 169-171
Babesia major-infected bovine erythrocytes, ^{60}Co-irradiation and subsequent injection into splenectomized calves, irradiation at 30 or 50 kR appeared to destroy virulence without altering immunogenicity but results must be interpreted with caution because of the presence of Eperythrozoon tuomii in blood of calves after challenge

Babesia microti
Brandt, F.; Healy, G. R.; and Welch, M., 1977, J. Parasitol., v. 63 (5), 934-937
establishment in golden hamsters of Babesia microti from 7 human infections

Babesia microti, illus.
Clark, I. A.; et al., 1977, Infect. and Immun., v. 17 (2), 430-438
mice, infection with BCG provided good protection against Babesia species, intensity and duration of protection similar to that after natural recovery from babesiosis, parasites degenerated within circulating erythrocytes, neither increased specific immune response nor phagocytosis but a nonspecific soluble effector substance is best explanation for protection

Babesia microti, illus.
Clark, I. A.; et al., 1977, Parasitology, v. 75 (2), 189-196
Babesia microti, abnormal forms in red cells of mice recovering from infection are noninfective, indistinguishable from those present after amicarbalide treatment, and persist in splenectomized hosts, electron microscopy confirms forms as degenerating intra-erythrocytic parasites, probably products of immune response

Babesia microti
Clark, I. A.; Allison, A. C.; and Cox, F. E., 1976, Nature, London (5541), v. 259, 309-311
Babesia, Plasmodium, protection of mice by previous infection with BCG, possible mechanism

Babesia microti, illus.
Clark, I. A.; Cox, F. E. G.; and Allison, A. C., 1977, Parasitology, v. 74 (1), 9-18
mice pre-treated with killed Corynebacterium parvum, completely resistant to infection with Babesia microti or B. rodhaini, protected from death caused by Plasmodium vinckei or P. chabaudi infection, no antibody detected, suggested that non-specific soluble mediator may play important role in protection observed

Babesia microti
Clark, I. A.; Wills, E. J.; and Richmond, J. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 12 [Demonstration]
intra-erythrocytic death of Babesia and Plasmodium as host acquires immunity, possibly caused by soluble mediator liberated by host cells

Babesia microti
Genga, U. E.; and Kreier, J. P., 1976, Ohio J. Sc., v. 76 (4), 188-189
Dermacentor andersoni, transstadial transmission of Babesia microti (Gray strain), hamsters

Babesia microti, illus.
Gravely, S. M.; and Kreier, J. P., 1974, Tropenmed. u. Parasitol., v. 25 (2), 198-206
Babesia microti, removal from infected hamster erythrocytes by continuous-flow ultrasonication, freed parasites collected by differential centrifugation, effectiveness of this system of lysis evaluated by electron microscopy and parasites evaluated serologically by complement-fixation testing, cross-reactions with sera from Plasmodium berghei-infected hamsters may be due to erythrocyte contamination

Babesia microti
Healy, G. R.; and Spielman, A., 1977, Science (4277), v. 195, 507
human babesiosis, answer to comment of McEnroe, W. D., 1977, Science (4277), v. 195, 506-507

Babesia microti
Healy, G. R.; Spielman, A.; and Gleason, N., 1976, Science (4238), v. 192, 479-480
rodents as reservoirs of infection
humans
Microtus pennsylvanicus
Peromyscus leucopus
all from Nantucket Island, Massachusetts

Babesia microti
Hussein, H. S., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 5 [Demonstration]
Babesia microti and B. hylomysci infections in mice on a milk diet, experiments showed dependence of both species on dietary supply of p-aminobenzoic acid (PABA) to synthesize folic and folinic acids

Babesia microti
Hussein, H. S., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 249-253
Babesia hylomysci, B. microti, mice (exper.), recovery from primary infection of B. microti results in sterile immunity possibly for lifetime, recovery from B. hylomysci results in premunition with infection at subpatent levels with spontaneous relapses, cross-protection occurs between the two species

Babesia microti
Irvin, A. D.; and Young, E. R.; and Purnell, R. E., 1977, Parasitology, v. 75 (2), xxviii [Abstract]
Babesia spp. of mice and cattle, uptake of tritiated purines and pyrimidines, uptake of hypoxanthine inhibited in vitro by antibabesial drugs in a degree roughly proportional to drugs' efficacy in vivo, may be useful as primary screening method for active drugs

Babesia microti, illus.
Kreier, J. P.; et al., 1975, Tropenmed. u. Parasitol., v. 26 (1), 9-18
Babesia rodhaini, B. microti, electron microscopy, relationship of stage of development to structure of intra- and extracellular parasites

Babesia microti, illus.
Langreth, S. G., 1976, J. Protozool., v. 23 (2), 215-223
Babesia microti vs. Plasmodium lophurae, feeding mechanisms of parasites freed of host erythrocytes, ferritin uptake of in vitro culture system

Babesia microti
Langreth, S. G.; and Trager, W., 1975, J. Protozool., v. 22 (3), 23A [Abstract]
Babesia microti, Plasmodium lophurae, comparison of feeding mechanisms

Babesia microti
McEnroe, W. D., 1977, Science (4277), v. 195, 506-507
human babesiosis, comment on Healy, G. R.; Spielman, A.; and Gleason, N., 1976, Science (4238), v. 192, 479-480

Babesia microti
Momen, H., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 438-439 [Demonstration]
Babesia spp., enzyme variations as significant aid to taxonomic study

Babesia microti
Momen, H., 1976, Parasitology, v. 73 (2), xvi [Abstract]
Plasmodium spp., Babesia spp., Anthemosoma garnhami, carbohydrate metabolism: enzyme activity, glucose catabolism

Babesia microti
Momen, H.; and Chance, M. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 13-14 [Demonstration]
Babesia spp., Anthemosoma garnhami, differences in DNA buoyant densities as possible aid to taxonomic studies; variations in enzymes and in DNA buoyant densities indicate that B. rodhaini and B. microti are distinct species and not synonyms

Babesia microti, illus.
Parry, M. F.; et al., 1977, J. Am. Med. Ass., v. 238 (12), 1282-1283
Babesia microti, clinical case report in man who had visited Montauk, New York areas, chloroquine therapy successful: Stamford, Connecticut

Babesia microti
Perez, M.; Carson, C. A.; and Ristic, M., 1977, Vet. Parasitol., v. 3 (2), 161-167
Babesia microti, hamsters, cell-mediated immune response measured by leukocyte migration inhibition test, comparison with humoral antibody measurements using indirect fluorescent antibody test

Babesia microti
Phillips, R. S.; and Wakelin, D., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 276 [Demonstration]
Trichuris muris-infected mice, suppression of immunity by concurrent infection with Babesia spp., severely depressed agglutinating antibody response to sheep red blood cells

Babesia microti
Phillips, R. S.; and Wakelin, D., 1976, Exper. Parasitol., v. 39 (1), 95-100
mice concurrently infected with Babesia and Trichuris muris, marked immunodepression, normal immune expulsion of nematode delayed; Babesia infections had little effect on expulsion of challenge infections of T. muris from previously immunized mice; Babesia infections exerted profound immunodepressive effect on agglutinating antibody response to sheep red blood cells

Babesia microti
Playfair, J. H. L.; de Souza, J. B.; and Cottrell, B. J., 1977, Immunology, v. 32 (5), 681-687
Plasmodium spp., Babesia microti, mouse helper T-cell response to parasites measured by using them as carriers for a standard hapten, results show extensive cross-reaction between the 4 parasites as carriers apparently unrelated to known serological cross-reactions but are against the idea that helper T-cells are exclusively responsible for resistance

Babesia microti
Purvis, A., 1976, Parasitology, v. 73 (2), ix [Abstract]
Babesia microti, producing immunodepression in mice

Babesia microti
Purvis, A. C., 1977, Parasitology, v. 75 (2), 197-205
Babesia microti, mice, temporary immunodepression of humoral immune response to sheep red blood cells, cell-mediated responses apparently unaffected, phagocytic activity is increased

Babesia microti
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Babesia microti, illus.
Rudzinska, M. A., 1976, J. Protozool., v. 23 (2), 224-233
intraerythrocytic Babesia microti, ultrastructure with emphasis on plasma membrane, feeding mechanism, and other areas of controversy

Babesia microti
Rudzinska, M. A.; et al., 1975, J. Protozool., v. 22 (3), 28A-29A [Abstract]
Babesia microti, invasion into erythrocytes

Babesia microti, illus.
Rudzinska, M. A.; et al., 1976, Cell and Tissue Research, v. 169 (3), 323-334
Babesia microti invades host erythrocyte through invagination of host plasma membrane with formation of parasitophorous vacuole, vacuolar space disappears and membrane of parasite and vacuole become closely adjacent, outer membrane then disintegrates

Babesia microti
Rudzinska, M. A.; and Trager, W., 1974, J. Protozool., v. 21 (3), 445-446 [Abstract]
Babesia microti, intraerythrocytic merozoites, fine structure

Babesia microti
Rudzinska, M. A.; and Trager, W., 1974, J. Protozool., v. 21 (3), 446 [Abstract]
Babesia microti, intraerythrocytic reproduction, electron microscope study

Babesia microti, illus.
Rudzinska, M. A.; and Trager, W., 1977, Canad. J. Zool., v. 55 (6), 928-938
Babesia microti, intraerythrocytic stage, fine structure and formation of merozoites, schizogony stage lacking

Babesia microti
Ruebush, T. K. II.; et al., 1977, Ann. Int. Med., v. 86 (1), 6-9
Babesia microti infections in humans, 5 case reports, successful treatment with chloroquine phosphate; Ixodes scapularis probably major vector in area: Nantucket Island

Babesia microti
Ruebush, T. K. II; et al., 1977, N. England J. Med., v. 297 (15), 825-827
Babesia microti, results of epidemiologic survey of possible cases of human babesiosis suggest that infections are self-limiting and at times sub-clinical: Nantucket Island, Massachusetts

Babesia microti
Spielman, A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (6), 784-787
seasonal distribution of Ixodes scapularis, the most abundant tick on Peromyscus leucopus on Nantucket Island, demonstration of competence as vector of Babesia microti derived from human infection recently acquired near study site (exper. hamster to hamster transmission)

Babesia microti (Franca, 1910)
Van Peenen, P. F. D.; et al., 1977, J. Protozool., v. 24 (2), 310-312
piroplasms, probably geographic strain of Babesia microti [but those from bandicoot rats are also referred to as B. bandicootia]; occurrence and appearance; transtadial infection of Ixodes granulatus; infectivity for laboratory hosts; effect of syringe passage, splenectomy, and irradiation; pathology; cross-infection experiments; lack of susceptibility of bandicoot rats and spiny rats to Gray strain of B. microti
Bandicota indica nemorivaga (nat. and ? exper.): Taiwan
Rattus coxinga (nat. and exper.): Taiwan
R. norvegicus (exper.)
Mesocricetus auratus (exper.)
Meriones unguiculatus (exper.)
Bandicota indica setifera (exper.)
Ixodes granulatus: Taiwan

Babesia microti
Wolf, R. E., 1977, Clin. Immunol. and Immunopathol., v. 2 (3), 381-394
Babesia microti, effects of antilymphocyte serum and splenectomy on resistance to infection in hamsters, results suggest that although cellular immunity is major factor in host resistance humoral antibody may modify parasitemia and thus give some protection

Babesia moshkovskii (Schurenkova, 1938) Laird and Lari, 1957, illus.
Croft, R. E.; and Kingston, N., 1975, J. Wildlife Dis., v. 11 (2), 229-233
B. moshkovskii, morphology
Falco mexicanus (blood): Wyoming

Babesia moshkovskii
Peirce, M. A., 1973, J. Protozool., v. 20 (5), 543-546
"On the available evidence. . .regarded as valid avian piroplasm. . ."

Babesia motasi
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

B[abesia] musculi
Goebel, E., 1976, Ztschr. Parasitenk., v. 50 (2), 207
B[abesia] musculi, B. herpailuri, morphology and topography after freeze etching

Babesia occidentalis (Yakimoff and Bourzeff, 1927), in part
Hoyte, H. M. D., 1976, Proc. Roy. Soc. Queensland, v. 87, v-xiii
as syn. of Babesia divergens (M'Fadyean and Stockman, 1911)

Babesia ovis
Friedhoff, K. T.; and Weber, G., 1973, Isotopes and Radiation Parasitol. III, 165-168
Babesia spp., identification and characterization of certain developmental stages, review

Babesia ovis
Hashemi-Fesharki, R., 1977, Brit. Vet. J., v. 133 (6), 609-614
Babesia ovis, splenectomized lambs, imidocarb dihydrochloride effective, quinuronium sulphate ineffective

Babesia ovis
Irvin, A. D.; and Young, E. R., 1977, Nature, London (5627), v. 269, 407-409
Babesia spp., in vitro test for screening drug activity which exploits finding that Babesia parasites readily incorporate hypoxanthine in an in vitro culture system, may also be used to detect drug resistance patterns and have possible application in primary screening of antimalarial drugs

Babesia ovis
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Babesia ovis
Khalacheva, M.; and Mechenova, E., 1976, Vet. Med. Nauki, v. 13 (8), 48-55
Babesia ovis, sheep (exper.), erythrocyte count, hemoglobin value and hematocrit increased and urea and bilirubin levels decreased in relation to severity of disease

Babesia ovis
Kiurtov, N., 1973, Vet. Med. Nauki, v. 10 (4), 49-53
Babesia ovis, intact and splenectomized lambs, premunition, role of spleen in antibody production

Babesia ovis
Kiurtov, N., 1976, Vet. Med. Nauki, v. 13 (7), 61-66
Babesia ovis, sheep, direct and indirect immunofluorescence compared with complement fixation; indirect method more readily applicable in diagnostic laboratory, direct method more species specific

Babesia ovis
Weber, G., 1976, Ztschr. Parasitenk., v. 50 (2), 185
Babesia ovis and B. bigemina in blood films, immunocytochemical detection of catabolic enzymes, possible implications

Babesia ratti
Leeflang, P.; et al., 1976, Internat. J. Parasitol., v. 6 (2), 159-161
survey of 173 males using indirect fluorescent antibody technique with Babesia bovis, B. bigemina, and B. ratti as antigens, babesial antibodies detected in 54%, no correlation found with mode of living, contact with livestock, incidence of malaria parasitaemias or malarial antibodies, no Babesia organisms found in blood smears or by animal subinoculation, source of antibodies unknown but subclinical infection not likely: Nigeria

Babesia rodhaini
Annable, C. R.; and Ward, P. A., 1974, J. Immunol., v. 112 (1), 1-8
Babesia rodhaini, rats, presence of proliferative glomerulitis, this renal complication is associated with glomerular deposits of IgG and third component of complement in pattern diagnostic for soluble immune complex-induced nephritis

Babesia rodhaini
Bishop, J. P.; and Kuttler, K. L., 1974, J. Protozool., v. 21 (5), 758-760
Babesia rodhaini, effect of irradiation on infectivity dependent upon irradiation dose, development of acquired resistance after inoculation of irradiated parasitized blood, mice

Babesia rodhaini
Brocklesby, D. W.; and Harradine, D. L., 1973, Research Vet. Sc., v. 14 (3), 397-398
Babesia rodhaini, mice, death delayed by approximately 1 day by prolonged administration of an interferon inducer

Babesia rodhaini
Chapman, W. E.; and Ward, P. A., 1976, J. Immunol., v. 116 (5), 1284-1288
Babesia rodhaini-infected rats, metabolism of third component of complement and IgG, development of hypocomplementemia and immune complex nephritis

Babesia rodhaini
Chapman, W. E.; and Ward, P. A., 1976, J. Immunol., v. 117 (3), 935-938
Babesia rodhaini, rats, depletion of C2, C3, C4, C5, and whole complement in course of infection, no evidence of depletion in alternative (properdin) pathway, findings indicate complement is activated via classical pathway in babesiosis

Babesia rodhaini
Chapman, W. E.; and Ward, P. A., 1977, Science (4285), v. 196, 67-70
Babesia rodhaini, ability to penetrate human erythrocytes depends on factors of the alternative complement pathway (properdin and factor B) as well as ionic magnesium and the third (C3) and fifth (C5) components of complement

Babesia rodhaini
Chiodini, P. L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 27-28 [Demonstration]
Babesia canis, in vitro culture; B. rodhaini and B. divergens in leucocyte-free dilution cultures, effect of imidocarb on leucine and adenosine incorporation, possible use of system for in vitro screening of potential babesicides and also for mode of action studies

Babesia rodhaini
Clark, I. A.; et al., 1977, Infect. and Immun., v. 17 (2), 430-438
mice, infection with BCG provided good protection against Babesia species, intensity and duration of protection similar to that after natural recovery from babesiosis, parasites degenerated within circulating erythrocytes, neither increased specific immune response nor phagocytosis but a nonspecific soluble effector substance is best explanation for protection

Babesia rodhaini
Clark, I. A.; Allison, A. C.; and Cox, F. E., 1976, Nature, London (5541), v. 259, 309-311
Babesia, Plasmodium, protection of mice by previous infection with BCG, possible mechanism

Babesia rodhaini
Clark, I. A.; Cox, F. E. G.; and Allison, A. C., 1977, Parasitology, v. 74 (1), 9-18
mice pre-treated with killed Corynebacterium parvum, completely resistant to infection with Babesia microti or B. rodhaini, protected from death caused by Plasmodium vinckei or P. chabaudi infection, no antibody detected, suggested that non-specific soluble mediator may play important role in protection observed

Babesia rodhaini
Cox, K. O.; Howard, R. J.; and Mitchell, G. F., 1977, Cellular Immunol., v. 32 (1), 223-227
Babesia rodhaini, increased numbers of plaque-forming cells secreting antibodies to modified mouse erythrocytes in spleens of heavily infected BALB/c·nu/+ mice but not in spleens of infected hypothymic BALB/c·nu/nu mice, role of antierythrocyte autoantibodies in pathogenesis controversial

Babesia rodhaini
Dalgliesh, R. J.; and Mellors, L. T., 1977, Cryobiology, v. 14 (4), 492-499
Babesia rodhaini, duration of freezing plateau did not affect parasite survival when cooling was slow and warming fast

Babesia rodhaini
Dalgliesh, R. J.; Swain, A. J.; and Mellors, L. T., 1976, Cryobiology, v. 13 (6), 631-637
frozen Babesia rodhaini, survival, effects of cooling and warming rates and protection by dimethyl sulphoxide

Babesia rodhaini
Dolan, T. T., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 274 [Demonstration]
Babesia divergens in calves (exper.), B. rodhaini in rats (exper.), thrombocytopenia with marked hemolytic anemia

Babesia rodhaini
Dolan, T. T.; and Carr, K. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 274-275 [Demonstration]
Babesia rodhaini-infected rat erythrocytes, light and electron microscopic scanning of cells for damage, good correspondence between damage and percentage of parasitemia

Babesia rodhaini
Farlow, G. E., 1976, Internat. J. Parasitol., v. 6 (6), 513-516
Babesia rodhaini, B. argentina, differences in infectivity when incubated in plasma vs. serum, role of glucose in prolonging viability, relevance of findings to living babesial vaccines in which plasma- and serum-based diluents may be used

Babesia rodhaini
Homewood, C. A.; Neame, K. D.; and Momen, H., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 429-434
Babesia rodhaini-infected mice, red cells show marked increase in permeability as measured by entry of L-glucose

Babesia rodhaini
Irvin, A. D.; and Young, E. R., 1977, Nature, London (5627), v. 269, 407-409
Babesia spp., in vitro test for screening drug activity which exploits finding that Babesia parasites readily incorporate hypoxanthine in an in vitro culture system, may also be used to detect drug resistance patterns and have possible application in primary screening of antimalarial drugs

Babesia rodhaini
Irvin, A. D.; and Young, E. R.; and Purnell, R. E., 1977, Parasitology, v. 75 (2), xxviii [Abstract]
Babesia spp. of mice and cattle, uptake of tritiated purines and pyrimidines, uptake of hypoxanthine inhibited in vitro by antibabesial drugs in a degree roughly proportional to drugs' efficacy in vivo, may be useful as primary screening method for active drugs

Babesia rodhaini, illus.
Kreier, J. P.; et al., 1975, Tropenmed. u. Parasitol., v. 26 (1), 9-18
Babesia rodhaini, B. microti, electron microscopy, relationship of stage of development to structure of intra- and extracellular parasites

Babesia rodhaini
McHardy, N., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 301-306
Babesia rodhaini, mice, stimulation of erythropoiesis can have beneficial effect

Babesia rodhaini
Mitchell, G. F., 1977, Internat. Arch. Allergy and Applied Immunol., v. 53 (4), 385-388
Babesia rodhaini, hypothymic BALB/c.nu/nu mice are more resistant than intact BALB/c. nu/+ mice, proportion of nu/nu survivors is not affected by administration of amicarbalide whereas majority of nu/+ mice develop long-lasting resistance to infection, data suggest that T cell dependent activities are involved in acceleration of parasitaemia and in development of drug-assisted resistance; nu/nu mice are not more resistant than nu/+ mice to Plasmodium berghei

Babesia rodhaini
Momen, H., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 438-439 [Demonstration]
Babesia spp., enzyme variations as significant aid to taxonomic study

Babesia rodhaini
Momen, H., 1976, Parasitology, v. 73 (2), xvi [Abstract]
Plasmodium spp., Babesia spp., Anthemosoma garnhami, carbohydrate metabolism: enzyme activity, glucose catabolism

Babesia rodhaini
Momen, H.; and Chance, M. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 13-14 [Demonstration]
Babesia spp., Anthemosoma garnhami, differences in DNA buoyant densities as possible aid to taxonomic studies; variations in enzymes and in DNA buoyant densities indicate that B. rodhaini and B. microti are distinct species and not synonyms

Babesia rodhaini
Musoke, A. J.; and Cox, H. W., 1977, J. Parasitol., v. 63 (3), 464-470
Plasmodium chabaudi, adaptation to rat host, immune responses in rats and in mice to rat strain, resistance to challenge with homologous and heterologous strains and to Babesia rodhaini, elaboration of soluble serum antigen in mice infected with rat strain

Babesia rodhaini
Neame, K. D.; Homewood, C. A.; and Momen, H., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 8 [Demonstration]
Plasmodium berghei, Babesia rodhaini, effect of intraerythrocytic parasites on the permeability of red-cell membrane

Babesia rodhaini
Nowell, F., 1976, Parasitology, v. 73 (2), xxxi [Abstract]
Babesia rodhaini, parasitaemia regression in mice held at high ambient temperatures, later recrudescence in some mice on return to normal temperatures

Babesia rodhaini
Parker, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 387-390
Babesia argentina, direct counting technique for estimating high parasitaemias, also suitable for B. bigemina and Plasmodium berghei but not for B. rodhaini or Anaplasma centrale

Babesia rodhaini
Phillips. R. S., 1973, Isotopes and Radiation Parasitol. III, 173-177
Babesia rodhaini, protective immunization of mice and rats with irradiated infected red cells, use in demonstrating antigenic variation, review

Babesia rodhaini
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Babesia rodhaini
Soni, J. L.; and Cox, H. W., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 423-430
Plasmodium gallinaceum, chickens, soluble complexes of serum antigen and its antibody may be mediator of acute anemia, serologic identity of serum antigen from malarious chickens and from Babesia rodhaini-infected rats and its distinction from parasite antigen suggest that it might be an autoantigenic macroglobulin

Babesia rodhaini
Thoongsuwan, S.; and Cox, H. W., 1973, Ann. Trop. Med. and Parasitol. v. 67 (4), 373-385
Babesia rodhaini, 3 antigenic variants selected by in vitro treatment with immune globulin, each of 3 strains caused nonspecific serum antigen associated with acute plasmodial and babesial infections to be elaborated without variation or diminution, implications for mechanism of relapse of latent infection

B[abesia] trautmanni
Okon, E. D., 1976, Trop. Animal Health and Prod., v. 8 (2), 96
B[abesia] trautmanni, Eperythrozoon, Haematopinus suis, pigs, incidence of infection according to host age groups: Ibadan, Nigeria

Babesiasis
Amerault, T. E.; and Roby, T. O., 1977, World Animal Rev. (22), 34-38
card test as an accurate and simple procedure for detecting Anaplasma marginale in cattle, possible use for determining wildlife reservoirs of anaplasmosis and detecting babesiosis, review

Babesiasis
Aragon, R. S., 1976, Vet. Bull., v. 46 (12), 903-917
Babesia, review of life cycle, pathology, diagnosis, immunity, prophylaxis, and chemotherapy

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Fameree, L.; Cotteleer, C.; and Antoine, H., 1976, Ann. Med. Vet., v. 120 (6), 421-423
diagnosis, epidemiology, symptoms, sanitation; known cases of bovine babesiosis from 1972-1976: Belgique (province du Luxembourg)

Babesiasis
Guilhon, J.; and Morel, P., 1976, Bull. Acad. Vet. France, v. 49 (1), 157-159
piroplasmosis or babesiosis, human: l'ouest de la France (Sarthe; Calvados)

Babesiasis
Hinaidy, H., 1976, Ztschr. Parasitenk., v. 50 (2), 212-213
Capreolus capreolus
Cervus elaphus
Dama dama
Elaphurus davidianus
all from Austria

Babesiasis
Hinaidy, H. K.; and Supperer, R., 1976, Wien. Tierarztl. Monatsschr., v. 63 (4), 124-125
babesiasis, cattle, differential diagnosis, procedure for preparing and staining blood smears: Austria

Babesiasis
Khalacheva, M.; and Vrubcheva, V., 1976, Vet. Med. Nauki, v. 13 (9), 87-92
babesiasis, sheep (exper.), serum chemistry changes (proteins, enzymes, potassium, sodium)

Babesiasis
Nagar, S. K.; Raizada, R. N.; and Saxena, V. K., 1977, Indian J. Animal Sc., v. 47 (10), 654-663
babesiosis, bovine, ovine, equine, canine, feline, prevalence during 1972-74, composition and ecology of ixodid ticks: Union Territory of Delhi

Babesiasis
Pienaar, J. G., 1977, J. South African Vet. Ass., v. 48 (1), 13-18
cerebral babesiasis in a dog

Babesiasis
Pryce, R. N., 1976, J. South African Vet. Ass., v. 47 (4), 289-290
canine biliary fever, method of determining plasma bicarbonate levels as measure of pH of body fluids, aid in determining metabolic normality and success of treatment

Babesiasis
Scharfman, W. B.; and Taft, E. G., 1977, J. Am. Med. Ass., v. 238 (12), 1281-1282
human babesiosis (Nantucket fever), case report of infection in adult woman after summer visit to Nantucket Island, diagnosis, therapy with chloroquine: Albany, New York

Babesiasis
Soltys, M. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 309-322
immunization against protozoan diseases of animals, review

Babesiasis
Uilenberg, G., 1976, World Animal Rev. (17), 8-15
babesiasis, theileriasis, anaplasmosis, epizootiology of tick-borne diseases, review

Babesioid
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Halcyon albiventris
Dicrurius adsimilis
(blood of all): all from Tanzania

Babesiosis. See Babesiasis.

Babesiosoma-like parasite
Svahn, K., 1976, Norwegian J. Zool., v. 24 (4), 460-461 [Abstract]
Lacerta agilis (erythrocytes): Bornholm, Denmark

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Peirce, M. A., 1973, J. Protozool., v. 20 (5), 543-546
as syn. of Aegyptianella anseris

Bacillidium sp., illus.
Raizada, U., 1976, Current Sc., Bangalore, v. 45 (17), 627-268 [Letter]
Bacillidium sp., possible biological control of cotton pest thrips
Thrips flavus
Scirtothrips oligochaetus

Baculea n. g.
Loubes, C.; and Akbarieh, M., 1977, J. Protozool., v. 24 (4), 61A [Abstract]
microsporidie; mt: Baculea daphniae n. sp.

Baculea daphniae n.g., n.sp. (mt)
Loubes, C.; and Akbarieh, M., 1977, J. Protozool., v. 24 (4), 61A [Abstract]
Daphnia pulex (epithelium intestinal)

Balantidium, illus.
Kan, S. P., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 1-8
Balantidium isolated from human feces, fine structure study of functional morphology and mode of life

Balantidium spp., illus.
McConnell, E. E.; et al., 1974, Onderstepoort J. Vet. Research, v. 41 (3), 97-168
pathological and parasitological survey of 100 free-ranging chacma baboons
Papio ursinus (large intestine): Kruger National Park, Transvaal

Balantidium [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Siganus (digestive tract): Elat, Gulf of Aqaba, Red Sea

Balantidium ? sp.
Peters, W.; Garnham, P.C.C.; and Killick-Kendrick, R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 3 [Demonstration]
histolytica-like amoeba and Balantidium ? sp. in stools of orang-utans suffering from dysentery, treatment with emetine dihydrochloride

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Balamuth, W.; and Siddiqui, W. A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 439-468
Entamoeba histolytica, Balantidium coli, human, immunology, review

Balantidium coli
Biagi, F., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 143 [Letter]
Balantidium coli, children with dysentery, successful nimorazol treatment

Balantidium coli
Botero R., D., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 145 [Letter]
Balantidium coli, children, successful treatment with nitrimidazine: Colombia

Balantidium coli
Contiu, I.; Sahleanu, C. M.; and Purcea, T., 1975, Rev. Crest. Animalelor, v. 25 (8), 51-55
swine dysentery, laboratory diagnosis

Balantidium coli, illus.
Fernandez-Cid, A.; Dexeus, S.; and Temprano, H., 1975, Progr. Obst. y Ginec., v. 18 (5), 505-512
Balantidium coli discovered in cervicovaginal smears of several women during routine cytologic examination, etiology unknown, may be laboratory contamination: Spain

Balantidium coli
Gulanoswka, H.; and Soroczan, W., 1971, Polski Tygod. Lekar., v. 26 (2), 64-65
Balantidium coli vaginal infections in women of rural populations, clinical case reports, treatment with metronidazole: Poland

Balantidium coli (Malmsten, 1857) Stein, 1862
Jain, P. C.; and Kamalapur, S. K., 1974, Indian Vet. J., v. 51 (1), 73
goats (feces): Madhya Pradesh

Balantidium coli, illus.
King, N. W., jr., 1976, Scient. Publication (317). Pan Am. Health Organ., 169-198

Balantidium coli, illus.
Lahiri, V. L.; Elhence, B. R.; and Agarwal, B. M., 1977, Acta Cytol., v. 21 (1), 123-124
Balantidium coli causing fatal case of human peritonitis, morphologic features as seen in routine cytologic smear (Papanicolaou) of ascitic fluid, differential diagnosis from malignancy

Balantidium coli
Lara Aguilera, R.; and Bernal, R. M., 1974, Bol. Med. Hosp. Inf., v. 31 (4), 779-784
Balantidium coli, dysentery in young children, frequent association with Trichuris trichiura infections in endemic areas, case reports, metronidazole: Mexico

Balantidium coli
Layrisse, M.; and Vargas, A., 1975, Progr. Food and Nutrition Sc., v. 1 (10), 645-667
human intestinal parasites, mechanisms by which parasites interfere with host nutrition (competition for nutrients, malabsorption, blood loss, excess nutrient utilization), extensive review

Balantidium coli (Malmsten, 1857) Stein, 1862
Letonja, T.; et al., 1975, Bol. Chileno Parasitol., v. 30 (3-4), 88-89
cerdos (muestras fecales): Santiago, Chile

Balantidium coli
Mwamba, T.; and Pandey, V. S., 1977, Ann. Recherches Vet., v. 8 (2), 167-169
Balantidium coli, pigs, epizootic form of disease causing high losses, terramycin useful treatment: Lubumbashi, Zaire

Balantidium coli
Naquira, F.; et al., Bol. Chileno Parasitol., v. 21 (4), 134-135
Balantidium coli, action in vitro of paromomycin

Balantidium coli, illus.
Norman, J. G.; and Jessop, P., 1973, Med. J. Australia, v. 1 (14), 694-696
Balantidium coli trophozoites discovered in cervo-vaginal smear and fecal specimen of 19-year old girl, probable secondary vaginal contamination, intermittent diarrhea, case report: Perth, Australia

Balantidium coli
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Balantidium coli
Pampiglione, S.; and Ricciardi, M. L., 1975, Riv. Parassitol., Roma, v. 36 (2-3), 89-108
Balantidium coli, discovered during survey of Pygmies (feces) for presence of intestinal parasites, probable contamination from pigs of neighboring Bantu since Pygmies do not raise pigs: Cameroun

Balantidium coli
Pandey, V. S.; and Mwamba, T., 1977, Vet. Parasitol., v. 3 (2), 189-193
Balantidium coli, piglets, treatment with acetarsol alone and in combination with oxytetracycline, combination 100% effective

Balantidium coli, illus.
Radford, A. J., 1973, Med. J. Australia, v. 1 (5), 238-241
Balantidium coli infections in West Irian highland natives who maintain intimate living association with domestic pigs, case reports, medical management: Papua New Guinea

Balantidium coli
Sreemannarayana, O.; Azeez, S. A.; and Zikriah, S., 1976, Indian Vet. J., v. 53 (1), 76
Balantidium coli, cattle, buffaloes, neftin, good results

Balantidium coli
Varma, S.; Malik, P. D.; and Lal, S. S., 1976, Vet. Rec., v. 99 (13), 256
Balantidium coli, consumption of Ascarops strongylina eggs observed during routine fecal examination, pig; possible role in epidemiology of A. strongylina

Balantidium coli
Verhulst, A.; and Cornet, R., 1975, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 28 (3), 301-304
Balantidium coli, pigs, niridazole (Ambilhar) tested in various dosages, effective treatment, no secondary effects in pigs: Republique du Zaire

Balantidium coli
del Villar Ponce, J. P.; et al., 1974, Bol. Med. Hosp. Inf., v. 31 (6), 1195-1200
Balantidium coli, children, differential diagnosis, case reports, distribution associated with close human-swine contact: Mexico

Balantidium entozoon
Jeserich, G., 1973, Mikrokosmos, v. 62 (4), 107-108
Opalina ranarum, Balantidium entozoon, Nyctotherus cordiformis, comparative fine structure morphology

Balbiania
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis

Balbiania gigantea Railliet, 1886
Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
"Balbiania gigantea by Railliet, is now generally recognized as being a Sarcocystis, and the name Sarcocystis gigantea (Railliet, 1886) should be applied to it."

Balbiania gigantea, Railliet, 1886
Heydorn, A.-O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis ovifelis n. spec.

Balbiania gigantea Railliet 1886
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis tenella Railliet 1886

Balbiana gigantia [sic] Railliet, 1886
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora tenella (Railliet, 1886) comb. nov.

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Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Bartonella
Kreier, J. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 24-28
Eubacteriales, Bartonellaceae

Bertarellia Carini, 1930
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses

Besnoitia
Beier, T. V.; Siim, I. K.; and Hutchison, W. M., 1977, Tsitologiia, v. 19 (12), 1362-1368
Eimeriidae, Isosporinae

Besnoitia
D'Haese, J.; Mehlhorn, H.; and Peters, W., 1976, Ztschr. Parasitenk., v. 50 (2), 198-199
coccidia, negative staining to show characteristic structures

Besnoitia Henry 1913
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Sarcocystidae, Toxoplasmatinae
diagnosis, life cycle pattern

Besnoitia Henry 1913
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Eimeriidae
diagnosis

Besnoitia
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
Sarcocystis and related coccidian parasites, general review with discussion of biological aspects of life cycles and new proposal for classification

Besnoitia sp., illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Besnoitia sp.
Guterbock, W. M.; and Levine, N. D., 1977, J. Am. Vet. Med. Ass., v. 170 (12), 1411-1413
cats (feces): east central Illinois

Besnoitia-like organism, illus.
Simpson, C. F.; et al., 1976, Proc. 34. Ann. Meet. Electron Microsc. Soc. America, 144-145
Besnoitia-like organism, aortitis in Calidrus canutus, electron microscopy: Gulf coast of Florida

Besnoitia-like organism, illus.
Simpson, C. F.; Woodard, J. C.; and Forrester, D. J., 1977, Vet. Path., v. 14 (4), 351-360
Besnoitia in Calidris canutus (luminal surface of thoracic aorta), ultrastructure: gulf coast of Florida

Besnoitia sp., Wallace & Frenkel, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora wallacei nom. nov. [i.e., n. sp.]

Besnoitia bennetti, illus.
Bwangamoi, O., 1972, Bull. Epizoot. Dis. Africa, v. 20 (3), 211-220
ringworm (Trichophyton equinum) of horses, concurrent heavy infestation with Boophilus decoloratus and one case of Besnoitia bennetti, case reports; Uasin Gishu Skin Disease is a misleading name and should be abandoned because it refers to clinical syndrome produced by different causal organisms: Kahawa, Kenya

Besnoitia besnoiti (Marotel, 1912), illus.
Basson, P. A.; McCully, R. M.; and Bigalke, R. D., 1970, Onderstepoort J. Vet. Research, v. 37 (2), 105-126
Besnoitia besnoiti, development and pathology of bovine strains in cattle (exper.) and of bovine and antelope strains in rabbits (exper.), macroscopic and microscopic lesions, chronic and acute infections

Besnoitia besnoiti (Marotel, 1912), illus.
Bigalke, R. D.; et al., 1967, Onderstepoort J. Vet. Research, v. 34 (1), 7-28
Besnoitia, susceptibility of rabbits, cattle, and sheep to experimental infection with blue wildebeest and impala strains and subsequent immunity to challenge with bovine strains, suggested all be regarded as distinct strains or biological races of B. besnoiti
Connochaetes taurinus (peripheral veins of limbs and neck)
Aepyceros melampus (peripheral veins of limbs and neck)
Tragelaphus strepsiceros (peripheral veins of limbs and neck)
cattle (blood) (exper.)
rabbits (wide range of internal organs) (exper.)
sheep (peripheral veins, nasal mucosa, endocardium, tendon sheaths) (exper.)

Besnoitia besnoiti, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Besnoitia besnoiti (Marotel, 1912) Henry, 1913
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Besnoitia besnoiti (Marotel 1912) Henry 1913
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Sarcocystis besnoiti Marotel 1912; Gastrocystis besnoiti (Marotel 1912) Brumpt 1913; Globidium besnoiti (Marotel 1912) Wenyon 1926
Bos taurus
Capra hircus
Connochaetes taurina
Aepyceros melangus
Tragelaphus strepsiceros
Oryctolagus cuniculus (exper.)
Ovis aries (exper.)
Meriones tristrami shawi (exper.)
Mesocricetus auratus (exper.)
Spermophilus fulvus (exper.)
Felis catus

Besnoitia besnoiti Marotel, 1912, illus.
Neuman, M., 1974, Tropenmed. u. Parasitol., v. 25 (2), 243-249
Besnoitia besnoiti, cultivation in cell line cultures of baby hamster kidney and HeLa cells and in sheep thyroid monolayer cultures, last gives most favorable conditions for development and growth

Besnoitia besnoiti Henry, 1913
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora besnoiti (Marotel, 1912) comb. nov.

Besnoitia besnoiti
Weiland, G.; and Kaggwa, E., 1976, Ztschr. Parasitenk., v. 50 (2), 177
Besnoitia besnoiti, B. jellisoni, rabbits, indirect immunofluorescence tests, enzyme labelling tests

Besnoitia darlingi, illus.
Smith, D. D.; and Frenkel, J. K., 1977, J. Parasitol., v. 63 (6), 1066-1071
Besnoitia darlingi, isolation from Didelphis marsupialis, production of acute lethal infections in mice and hamsters by inoculation of cysts, chronic infections with formation of tissue cysts obtained in mice by sulfadiazine prophylaxis, cysts fed to cats resulted in shedding of isosporoid oocysts, mouse-to-mouse transmission achieved by injecting triturated tissue containing cysts, serologic testing
Didelphis marsupialis: Kansas City area
mice (exper.)
golden hamsters (exper.)
cats (exper.)

Besnoitia jellisoni, illus.
D'Haese, J.; Mehlhorn, H.; and Peters, W., 1977, Internat. J. Parasitol., v. 7 (6), 505-518
Sarcocystis ovifelis, Besnoitia jellisoni, Eimeria falciformis, pellicular structures, fine structure, implications for mechanism of motility

Besnoitia jellisoni
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Besnoitia wallacei, description, life cycle, serologic tests (cross reactions with B. jellisoni, Toxoplasma gondii, and Sarcocystis muris), infectivity tests, immunity to challenge (slight cross-immunity with B. jellisoni)

Besnoitia jellisoni, illus.
Frenkel, J. K., 1977, Am. J. Path. (418), v. 86 (3), 749-752
Besnoitia jellisoni in Mesocricetus auratus (exper.), chronic infection of hamsters provides a sensitive example of immunosuppression by corticosteroids and cytostatic agents, permits study of corticosteroid effects on chronic infection-immunity in the adrenals or the body as a whole

Besnoitia jellisoni
Frenkel, J. K.; Dubey, J. P.; and Hoff, R. L., 1976, J. Protozool., v. 23 (3), 421-424
Toxoplasma gondii, Besnoitia jellisoni, loss of stages after prolonged passage in tachyzoite stage (T. gondii developed cysts which when fed to cats failed to produce oocysts; B. jellisoni lost capacity to form cysts), phenomena explained by loss of genomes or gene products

Besnoitia jellisoni
Frenkel, J. K.; and Reddy, J. K., 1977, J. Reticuloendothel. Soc., v. 21 (1), 61-68
chemical induction of liver neoplasms is delayed but not prevented in rats chronically infected with Toxoplasma gondii or Besnoitia jellisoni, possible role of activated macrophages in mediating this protection

Besnoitia jellisoni
Krahenbuhl, J. L; Lambert, L. H., jr.; and Remington, J. S., 1976, Cellular Immunol., v. 25 (2), 279-293
Toxoplasma gondii and Besnoitia jellisoni used as form of stimulation for obtaining activated macrophages, effects of activated macrophages on tumor target cells, escape from cytostasis

Besnoitia jellisoni
Krahenbuhl, J. L.; and Remington, J. S., 1974, J. Immunol., v. 113 (2), 507-516
peritoneal macrophages from mice chronically infected with Toxoplama gondii and Besnoitia jellisoni had marked cytostatic effect on variety of tumor target cells

Besnoitia jellisoni
Krick, J. A.; and Remington, J. S., 1975, J. Infect. Dis., v. 131 (6), 665-672
mice infected with Toxoplasma gondii or Besnoitia jellisoni, significant resistance to mixed infections with nocardiosis, possible cell-mediated immunity

Besnoitia jellisoni
Lindberg, R. E.; and Frenkel, J. K., 1977, Infect. and Immun., v. 15 (3), 855-862
Toxoplasma gondii and Besnoitia jellisoni in vitro in hamster peritoneal exudate cells, antigenic stimulation of peritoneal cells and expression of immunity, inhibition of parasite growth by peritoneal macrophages armed either specifically or nonspecifically, expression of immunity by cells derived from hamsters treated with cortisol, effects of cortisol on (1) immune and non-immune lymphocytes, (2) arming of macrophages by lymphocytes, (3) the ability of peritoneal macrophages to destroy antibody-treated parasites

Besnoitia jellisoni, illus.
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Besnoitia jellisoni
McLeod, R.; and Remington, J. S., 1977, Cellular Immunol., v. 34 (1), 156-174
Toxoplasma gondii, Besnoitia jellisoni, mice, studies on specificity of killing of intracellular pathogens by macrophages

Besnoitia jellisoni
Mueller, B. E. G.; Mehlhorn, H.; and Scholtyseck, E., 1973, J. Protozool., v. 20 (4), 505
ultrastructure and origin of cyst wall in Toxoplasmatea

Besnoitia jellisoni
Remington, J. S.; Krahenbuhl, J. L.; and Mendenhall, J. W., 1972, Infect. and Immun., v. 6 (5), 829-834
activated macrophages from mice which were chronically infected with Toxoplasma gondii or Besnoitia jellisoni or which had received Freund complete adjuvant had enhanced capacity to kill intracellular Toxoplasma

Besnoitia jellisoni, illus.
Scholtyseck, E.; Mehlhorn, H.; and Mueller, B.E.G., 1974, J. Protozool., v. 21 (2), 284-294
Sarcocystis tenella, Besnoitia jellisoni, Frenkelia sp., Toxoplasma gondii, nature of cyst, fine structure of cyst wall

Besnoitia jellisoni, illus.
Senaud, J.; Chobotar, B.; and Scholtyseck, E., 1976, Tropenmed. u. Parasitol., v. 27 (2), 145-159
comparative fine structure morphology of Plasmodium cathemerium, Eimeria ferrisi, E. stiedai, Besnoitia jellisoni, and Frenkelia sp. micropores, discussion of micropore function in parasite nutrition

Besnoitia jellisoni
Senaud, J.; and Mehlhorn, H., 1977, J. Protozool., v. 24 (4), 64A [Abstract]
Besnoitia jellisoni, ultrastructural aspects of development in tissue culture

Besnoitia jellisoni (Frenkel, 1953), illus.
Senaud, J.; Mehlhorn, H.; and Scholtyseck, E., J. Protozool., v. 21 (5), 715-720
Besnoitia jellisoni, development in macrophages and cysts from experimentally infected Mus musculus, results concerning origin and formation of inner complex of pellicle during endodyogeny indicate origin of inner membranes from endoplasmic reticulum

Besnoitia jellisoni
Weiland, G.; and Kaggwa, E., 1976, Ztschr. Parasitenk., v. 50 (2), 177
Besnoitia besnoiti, B. jellisoni, rabbits, indirect immunofluorescence tests, enzyme labelling tests

Besnoitia jellisoni
Williams, D. M.; Sawyer, S.; and Remington, J. S., 1976, J. Infect. Dis., v. 134 (6), Dec., 610-623
mice (exper.) whose macrophages were activated by Trypanosoma cruzi or Besnoitia jellisoni were significantly more resistant to intraperitoneal challenge with T. cruzi than were controls; activated macrophages were able to inhibit completely multiplication of T. cruzi, this suggests that the macrophage may play a major role in resistance to infection

Besnoitia jellisoni
Wilson, H. R.; and Fairchild, D. G., 1977, Stain Tech., v. 52 (2), 105-111
modified Whipf's polychrome staining to demonstrate protozoans and tissue components in cutaneous lesions of infected animals

Besnoitia wallacei (Tadros and Laarman, 1976) Dubey, 1977 [n. comb.]
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Besnoitia wallacei, illus.
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Besnoitia wallacei, description, life cycle, serologic tests (cross reactions with B. jellisoni, Toxoplasma gondii, and Sarcocystis muris), infectivity tests, immunity to challenge (slight cross-immunity with B. jellisoni)
Felis catus (feces, intestine) (nat. and exper.): Oahu, Hawaii
Mus musculus (intestine, peritoneum) (exper.)
Rattus norvegicus (exper.)
R. exulans (exper.)

Besnoitia wallacei Tadros and Laarman, 1976
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Mus musculus
Rattus norvegicus
Rattus exulans
Felis catus

Bhatiella Setna, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis, synonymy

Bhatiella iwamusi (Hoshide, 1935) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Cotyloepimeritus iwamusi Hoshide, 1935; Ferraria cornucephala iwamusi Hoshide, 1944

Bhatiella marphysae Setna, 1931 emend.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Bhatiella morphysae Setna, 1931 lapsus calami; Ferraria cornucephali Setna, 1931

Bhatiella morphysae Setna, 1931 lapsus calami
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Bhatiella marphysae Setna, 1931 emend.

Blastocrithidia
Daggett, P. M.; and Decker, J. E., 1974, J. Protozool., v. 21 (3), 429 [Abstract]
comparison of excreted factors of insect trypanosomatids at 25 C vs. 37 C, results indicate secretion products are primarily under parasite not host control

Blastocrithidia sp., similar to B. triatomae Cerisola, del Prado, Rohwedder and Bozzini, 1971
Carvalho, A. L. M.; and Deane, M. P., 1974, J. Protozool., v. 21 (1), 5-8
Trypanosomatidae isolated from Zelus leucogrammus, only Blastocrithidia thought to be a parasite proper of this host spread by cannibalism and ingestion of cysts, other trypanosomatids acquired by feeding on insect prey
Zelus leucogrammus: Brazil
Triatoma infestans (exper.)

Blastocrithidia culicis
Chang, K.-P., 1974, J. Protozool., v. 21 (5), 699-707
ultrastructural evidence suggests that bipolar bodies of Crithidia oncopelti and diplosomes of Blastocrithidia culicis are endosymbiotic bacteria with defective cell walls and that they are subject to destruction by treatment with chloramphenicol but not penicillin

Blastocrithidia culicis, illus.
Chang, K.-P., 1975, J. Protozool., v. 22 (2), 271-276
Blastocrithidia culicis, Crithidia oncopelti, intracellular symbiotes can be eliminated by single chloramphenicol treatment with subsequent reduced growth of flagellates

Blastocrithidia culicis
Chang, K.-P., 1976, J. Protozool., v. 23 (2), 241-244
Blastocrithidia culicis, Crithidia oncopelti, symbiote-free strains: liver extract as essential growth factor in defined medium; cross-reactivity in reciprocal agglutination test with symbiote-containing strains indicates loss of symbiotes does not affect antigenic identity

Blastocrithidia culicis
Hill, G. C.; and Bonilla, C. A., 1974, J. Protozool., v. 21 (5), 632-638
Crithidia fasciculata, isolation of DNA-dependent RNA polymerases, purified nuclear and kinetoplast DNA can serve as templates for these RNA polymerases, this RNA synthesis is inhibited by several trypanocidal drugs, hybridization experiments revealed no homologies between Blastocrithidia culicis and Crithidia fasciculata K-DNA

Blastocrithidia culicis
Paulin, J. J., 1974, J. Protozool., v. 21 (3), 435 [Abstract]
Crithidia fasciculata, Blastocrithidia culicis, Trypanosoma cruzi, three-dimensional reconstructions of chondriome using high voltage electron microscopy

Blastocrithidia culicis, illus.
Paulin, J. J., 1974, Proc. 32. Ann. Meet. Electron Microsc. Soc. America (St. Louis, Missouri, Aug. 13-15), 62-63
Trypanosoma equiperdum, Blastocrithidia culicis, Crithidia fasciculata, 3-dimensional cell structure, high voltage electron microscopy

Blastocrithidia culicis
Paulin, J. J., 1975, J. Protozool., v. 22 (3), 16A [Abstract]
unitary structure of chondriome of 3 spp. of Trypanosomatidae confirmed by application of high voltage electron microscopy

Blastocrithidia culicis
Risby, E. L., 1976, J. Protozool., v. 23 (2), 20A [Abstract]
Crithidia fasciculata, Blastocrithidia culicis, Trypanosoma, phosphohexose isomerase, comparison of characteristics

Blastocrithidia triatomae Cerisola et al. 1971
Haberkorn, A., 1976, Ztschr. Parasitenk., v. 50 (2), 194-195
Blastocrithidia triatomae, course of infection in Triatoma infestans, transmission by coprophagy and cannibalism

Blastocystea new class [nom. nud.]
Zierdt, C. H.; and Tan, H., 1976, Exper. Parasitol., v. 39 (3), 422-430
Protozoa, Sporozoa; includes Blastocystida new order [nom. nud.]

Blastocystida new order [nom. nud.]
Zierdt, C. H.; and Tan, H., 1976, Exper. Parasitol., v. 39 (3), 422-430
Protozoa, Sporozoa, Blastocystea new class [nom. nud.]; includes Blastocystis

Blastocystis
Zierdt, C. H.; and Tan, H., 1976, Exper. Parasitol., v. 39 (3), 422-430
Protozoa, Sporozoa, Blastocystea new class [nom. nud.], Blastocystida new order [nom. nud.]

Blastocystis sp.
Burden, D. J., 1976, Parasitology, v. 73 (2), iv-v [Abstract]
Blastocystis sp., pigs, frequently found in feces of cases of diarrhea and dysentery, causal relationship not established

Blastocystis hominis (Brumpt 1912), illus.
Phillips, B. P.; and Zierdt, C. H., 1976, Exper. Parasitol., v. 39 (3), 358-364
Blastocystis hominis, human, possibly cause of persistent diarrhea, production of similar symptoms in germfree guinea pigs which were inoculated with B. hominis and the enteric flora from symptomatic human patients

Blastocystis hominis, illus.
Zierdt, C. H.; and Tan, H., 1976, Exper. Parasitol., v. 39 (3), 422-430
Blastocystis hominis, 8 axenically grown strains, presence of intracellular bacteria-like spheres and rods, direct relationship between increasing endosymbiont numbers and increasing B. hominis cell size, effect of 3 antibiotics on B. hominis and its endosymbiont

Blastocystis hominis, illus.
Zierdt, C. H.; and Tan, H. K., 1976, Ztschr. Parasitenk., v. 50 (3), 277-283
Blastocystis hominis, in vivo forms from fatal human case, ultrastructure and light microscopy, comparison with in vitro forms

Bodo urinarius
Dyner, E.; and Liwski, R., 1974, Polskie Arch. Med. Wewn., v. 51 (2), 147-154
Trichomonas vaginalis, Bodo urinarius, effects of immunosuppressants in vitro and in vivo (mice)

Bothriopsides acilii Baudoin, 1961
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Bothriopsides histrio (Schneider) Foerster, 1938

Bothriopsides graphoderi Baudoin, 1961 pro parte
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Iorella wegoreki g. n., sp. n.

Bothriopsides histrio (Schneider) Foerster, 1938, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Dytiscus marginalis
Hydrophilus sp.
all from Poland

Bothriopsides histrio (Schneider) pro parte sensu Baudoin, 1961
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Iorella wegoreki g. n., sp. n.

Bothriopsides terpsichorella (Ellis) sensu Foerster, 1938
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Bothriopsides histrio (Schneider) Foerster, 1938

Bothriopsis histrio Schneider, 1875
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Bothriopsides histrio (Schneider) Foerster, 1938

Buchneriella Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Maupasellidae, Maupasellinae

Buchneriella criodrili Heidenreich, 1935, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Criodrilus lacuum
C. ochridanum

Buchneriella pheretimae de Puytorac et Rakotoarivelo, 1965
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42

Buetschliella clignyi, Cepede, 1922
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Spirobuetschliella clignyi (Cepede, 1922) Hovasse, 1950

Buetschliella inermis Tamogradsky, 1914
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Acanthophrya inermis (Stein, 1859) Heidenreich, 1935

Buetschliella naidos Dujardin 1841, in Mackinnon et Adam, 1924
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya naidos (Schmidt, 1846) Heidenreich, 1935

Buetschliellopsis de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Juxtaradiophryinae n. s. fam.

Buetschliellopsis commune Lom, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Fridericia hegemon
F. perrieri
F. lobifera
F. bisetosa
F. bulbosa
Henlea nasuta

Buetschliellopsis enchytraei de Puytorac, 1954, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Fridericia ratzeli
F. lobifera
F. perrieri
F. hegemon
F. dura

Buetschliellopsis janovicensis Lom, 1959
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    synonymy
    Fridericia ratzeli
    F. lobifera
    F. dura
    F. bulbosa
    F. galba
    F. perrieri
    Henlea nasuta

Buetschliellopsis parumciliata Lom, 1954
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Fridericia lobifera
    F. bisetosa
    Buchholzia appendiculata

Buetschliellopsis torpediniformis (Cheissin, 1930)
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Mesenchytraeus bungei

Buxtonella sulcata, illus.
  Henriksen, S. A., 1977, Nord. Vet.-Med., v. 29 (10), 452-457
    Fasciola hepatica, cattle (faeces), mixed infection with Buxtonella sulcata poses no problems in differential diagnosis relative to helminth eggs or coccidia: Kolding, Denmark

Callyntrochlamys
Theodorides, J.; and Desportes, I., 1975, J. Protozool., v. 22 (3), 85A-86A [Abstract]
gregarines of marine invertebrates, organization of epicyte

Caryospora
Cerna, Z., 1974, J. Protozool., v. 21 (4), 481-482
Isospora lacazei oocysts, anomalous sporulation, resemblance to Caryospora

Caryospora
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Caryospora sp.
Cerna, Z., 1973, J. Protozool., v. 20 (4), 535-536
Erithacus rubecula rubecula (feces)

Caryospora brasiliensis Carini, 1932, illus.
Lainson, R.; and Shaw, J. J., 1973, J. Protozool., v. 20 (3), 358-362
redescription
Philodryas olfersi (epithelial cells of small intestine): Barcarena, Para State, north Brazil

Caudospora sp. Frost & Nolan, 1972
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Caudospora sp. Steinhaus, 1951
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Caudospora alaskensis Jamnback, 1970, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Caudospora brevicauda Jamnback
Ebsary, B. A.; and Bennett, G. F., 1975, Canad. J. Zool., v. 53 (8), 1058-1062
Cnephia mutata: insular Newfoundland

Caudospora nasiae Jamnback, 1970, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size and respiration; cytopathology); ecology

Caudospora pennsylvanica Beaudoin and Wills, 1965, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Caudospora polymorpha (Strickland, 1911) n. comb., illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Caudospora simulii Weiser
Ebsary, B. A.; and Bennett, G. F., 1975, Canad. J. Zool., v. 53 (8), 1058-1062
Prosimulium fuscum/mixtum: insular Newfoundland

Caudospora simulii Weiser, 1946, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Cepedea
Mignot, J.-P.; and Brugerolle, G., 1974, J. Protozool., v. 21 (5), 649-658
Opalina, Cepedea, ultrastructure of 2 different types of cysts, cortical morphogenesis

Cephaloidophora
Theodorides, J.; and Desportes, I., 1975, J. Protozool., v. 22 (3), 85A-86A [Abstract]
gregarines of marine invertebrates, organization of epicyte

Cephaloidophora conformis (Diesing, 1851), illus.
Vivares, C. P.; and Van Wormhoudt, A., 1976, Compt. Rend. Soc. Biol., Paris, v. 170 (1), 71-77
Cephaloidophora conformis in Pachygrapsus marmoratus (caecum digestif anterieur), carbohydrate metabolism of host and parasite: region de Marseille

Cephalolobus
Theodorides, J.; and Desportes, I., 1975, J. Protozool., v. 22 (3), 85A-86A [Abstract]
gregarines of marine invertebrates, organization of epicyte

Ceratomyxa [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Siganus (gall bladder)
Mugil cephalus
all from Elat, Gulf of Aqaba, Red Sea

Ceratomyxa californica Jameson 1929
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes paucispinis (gallbladder): off southern California

Ceratomyxa elegans Jameson 1929
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes pinniger
S. rosaceus
(gallbladder of all): all from off southern California

Ceratomyxa hokarari Meglitsch, 1960
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides acrolepis: eastern Pacific

Ceratomyxa hopkinsi Jameson 1929
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes goodei (gallbladder): off northern California

Ceratomyxa laxa Meglitsch 1960
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes serranoides (gallbladder): off southern California

Ceratomyxa sebasta sp. n., illus.
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes carnatus: off central California
S. ensifer: off southern California
S. paucispinis: off northern California
S. pinniger: off northern California
S. serranoides: off central California
(gallbladder of all)

Ceratomyxa vepallida Meglitsch 1960
Moser; M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes carnatus: off northern California
S. ensifer: off southern California
S. paucispinis: off southern California
S. serranoides: off southern California
(gallbladder of all)

Ceratospora Leger, 1892
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Urosporidae
checklist of species

Chagas disease
Amuchastegui, L. M.; Alday, L. E.; and Moreyra, E., 1976, Medicina, Buenos Aires, v. 36 (2), 121-126
cardiac sinus node disfunction and its clinical management in patients affected with chronic Chagasic cardiopathology

Chagas disease
Arendt de Ramos, F. F., 1976, Acta Bioquim. Clin. Latinoam., v. 10 (2), 121-126
Chagas disease, pregnant patients, positive immunological reactions in maternal serum, amniotic fluid, and newborn

Chagas disease
Beer, N.; et al., 1977, Postgrad. Med. J., London (623), v. 53, 537
human Chagas disease, evaluation of effect of digoxin on ventricular automaticity in Chagas cardiomyopathy

Chagas disease
Castagnino, H. E.; et al., 1975, Medicina, Buenos Aires, v. 35 (2), 166-179
case report of man with chronic myocardiopathy of Chagas disease resulting in ventricular aneurysm with severe ventricular tachycardia, successful surgical repair

Chagas disease
Ceneviva, R.; Modena, J. L. P.; and Castelfranchi, P. L., 1971, Medicina, Sao Paulo, v. 4 (2), 37-43
Chagas disease as a predisposing factor in the development of gastroduodenal ulcers in human infections

Chagas disease
Franco, M. F.; and Chamma, L. G., 1973, Internat. Arch. Allergy and Applied Immunol., v. 44 (5), 692-696
schistosomiasis mansoni, Chagas disease, diagnosis, technical modification of indirect immunofluorescent antibody test using filter paper blood eluates, useful as rapid qualitative screening test when dealing with large number of samples

Chagas disease
de Freitas, G.; et al., 1975, J. Protozool., v. 22 (3), 35A [Abstract]
no significant difference in immunoglobulin levels in chronic Chagas disease vs. uninfected controls: Bambui, Minas Gerais, Brazil

Chagas disease
Gerling, D.; Conde, J. E.; and Rabinovich, J. E., 1976, Canad. Entom., v. 108 (4), 427-432
Chagas disease, development of Ooencyrtus trinidadensis and Telenomus costa-limai, egg parasites of vector, Rhodnius prolixus, possible biological control

Chagas disease
Hernandez-Pieretti, O., 1977, Postgrad. Med. J., London (623), v. 53, 533-536
human Chagas disease, echocardiographic findings in persons with chronic Chagas cardiomyopathy compared with other types of cardiomyopathy, use in diagnosis and evaluation of pathology

Chagas disease
Kunzle, J. E.; de Britto-Costa, R.; and Ziliotto, A., jr., 1976, Bol. Chileno Parasitol., v. 31 (1-2), 2-5
Chagas disease in humans, duodenal arteriomesenteric occlusion, case reports, need for consideration in differential diagnosis and etiology of duodenal ileus: Sao Paulo, Brazil

Chagas disease
Miles, M. A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 521-522
Chagas disease, man's role in propagation of infection, habits of temporary residence of rural population in endemic area, possible impact of such temporary movements on dissemination of triatomine bugs and Chagas disease: Brazil

Chagas disease
Sanabria, A.; Croquer, F. J.; and Monsalve, P., 1976, Nouv. Presse. Med., v. 5 (7), 431-432
cardiac arrhythmias resulting from human Chagas disease, clinical trials with carbamazepine to study its anti-arrhythmic action

Chagas disease
dos Santos, R. R.; de Oliveira, J. C. R.; and Rossi, M. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 167
human chronic Chagas disease, presence of IgG and IgM antibodies to neurons suggests immunological mechanism involved in qualitative and quantitative alterations of autonomic nervous system

Chagas disease
Schofield, C. J.; and Moreman, K., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 165-166
Triatoma infestans, absence of sex attractant in adult, significance in detection and sampling of domestic infestations of vectors of human Chagas disease

Chagas disease
Tr. Roy. Soc. Trop. Med. and Hyg., 1976, v. 70 (2), 114-129
African and South American trypanosomiasis, 16th seminar on current status (epidemiology, chemotherapy, immunological research and problems of immunization, infection in domestic animals, vector control)

Cheissinophrya n. gen.
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Hoplitophryidae, Radiophryinae; tod: C. knoeppfleri n.g., n.sp.

Cheissinophrya de Puytorac et Dragesco, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Metaradiophryinae n. s. fam.

Cheissinophrya knoeppfleri n.g., n.sp., illus. (tod)
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Alma emini (tube digestif): Gabon

Cheissinophrya knoeppfleri de Puytorac et Dragesco, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini: Gabon

Chilomastix [sp.]
Brugerolle, G., 1976, J. Protozool., v. 23 (4), 5A [Abstract]
Box salpa (intestin posterieur)

Chilomastix sp., illus.
Koesters, J.; et al., 1976, Bol. Chileno Parasitol., v. 31 (3-4), 84-85
Anser anser domesticus (intestino posterior, ciegos, contenido cloacal)
Anas platyrhynchos (contenido cloacal)
Cairina moschata (contenido cloacal)
Gallus gallus domesticus (nat. and exper.) (contenido cloacal)
hibridos Pekin x mudo (contenido cloacal)
all from Fundo Nalcahue, Mafil, Chile

Chilomastix sp.
Leibovitz, L., 1973, Am. J. Vet. Research, v. 34 (8), 1053-1061
observed in breeder ducks, affected or not affected with necrotic enteritis

Chilomastix sp.
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Rattus norvegicus: Carshalton

Chilomastix aulastomi (Belar), illus.
Brugerolle, G., 1973, J. Protozool., v. 20 (5), 574-585
Chilomastix aulastomi, trophozoite, cyst, ultrastructure
Haemopis sanguisuga (intestin posterieur)

Chilomastix aulastomi Belar, 1921
Nohynkova, E., 1974, J. Protozool., v. 21 (3), 457-458 [Abstract]
Haemopis sanguisuga (intestine): Czechoslovakia

Chilomastix bettencourti, illus.
Grewal, M. S., 1975, Research Bull. Panjab Univ., v. 23 (3-4), 1972, 119-124
Rattus rattus (caecum): Chandigarh, India

Chilomastix mesnili
Barnham, M., 1977, Lancet, London (8047), v. 2, 1077-1078 [Letter]
Chilomastix mesnili, possible pathogenicity in humans, resolution of diarrhea and parasite trophozoites in feces with metronidazole therapy

Chilomastix mesnili
Markell, E. K.; and Quinn, P. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1139-1142
comparison of immediate polyvinyl alcohol fixation with delayed Schaudinn's fixation for demonstrating protozoa in feces, concluded that immediate fixation results in preservation of larger number of organisms and recommends routine use prior to transportation to laboratory

Chilomastix mesnili
Reyes, H.; Olea, M.; and Hernandez, R., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 115-116
survey of food handlers for intestinal parasites shows 50.5% infection rate; Santiago, Chile

Chilomastix mesnili
Sagua, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 58-60
comparison of phenol-alcohol-formalin sedimentation and polyvinyl alcohol fixative tests in diagnosis of human intestinal helminths and protozoa

Chilomastix mesnili
Schenone, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 104-105
metronidazole used for mass therapy of human intestinal protozoa: Chile

Chilomastix mesnili
Vinayak, V. K.; and Sehgal, S. C., 1976, Indian J. Med. Research, v. 64 (9), 1347-1350
human helminthic and protozoan parasites, comparison of nigrosin-methylene blue diagnostic test with formol-ether method and direct examination

Chilomitinae Honigberg, 1963
Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
Trichomonadida Kirby, emend.; Monocercomonadidae Kirby, emend.

Chlamydocystis Dogiel, 1910
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Chlamydocystis captiva Dogiel, 1910
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Chloromyxum esocinum Dogiel, 1934
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Esox lucius: Loch Leven, Scotland

Chloromyxum kabatai sp. n., illus.
Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
Coryphaenoides acrolepis (gall bladder): off Oregon

Chloromyxum subsphaericum Shulman, 1966
Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
Coryphaenoides armatus (gall bladder): off Costa Rica

Chloromyxum truttae Leger, 1906
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Salmo trutta: Loch Leven, Scotland

Ciliate
File, S. K.; McGrew, W. C.; and Tutin, C. E. G., 1976, J. Parasitol., v. 62 (2), 259-261
Pan troglodytes schweinfurthii (feces): Gombe National Park, Tanzania

Clepsidrina cuneata: Pfeiffer, 1910
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina cuneata Stein

Clepsidrina munieri Schneider, 1875
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina munieri (Schneider)

Clepsidrina polymorpha Hammerschmidt, 1838
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina cuneata Stein

Clepsidrina polymorpha var. cuneata: Schneider, 1875
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina cuneata Stein

Cnemidospora lutea Schneider, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Glomeris connexa (intestine): Poland

Coccidia
Bejsovec, J., 1977, J. Protozool., v. 24 (4), 48A [Abstract]
persistence of Coccidia despite changing land use and decreasing numbers of host animals: Bohemia

Coccidia
Birch-Andersen, A.; Ferguson, D. J. P.; and Pontefract, R. D., 1976, Acta Path. et Microbiol. Scand., v. 84B (4), 235-239
technique for obtaining thin sections of coccidian oocysts demonstrated with Eimeria brunetti

Coccidia
Graves, I. L.; Adams, W. H.; and Pyakural, S., 1975, Am. J. Vet. Research, v. 36 (6), 843-846
Babesia bigemina, Bos grunniens moved from high to low altitude and challenged with influenza A viruses, hemolytic anemia, possible explanations, death due to Fasciola hepatica and F. gigantica, incidental finding of Bunostomum sp., Trichuris sp., Neoascaris vitulorum, Dictyocaulus sp., coccidia, some reasons for poor survival of yaks at low altitude: Nepal

Coccidia
Helfer, D. H.; and Koller, L. D., 1976, Cornell Vet., v. 66 (3), 369-371
coccidial forms in intestinal polyps, lamb, probably only coincidental

Coccidia
Jacobs, D.; and Schulze, H. W., 1977, Prakt. Tierarzt, v. 58 (1), 46-48
pig parasites, natural infections, vermitin and dichlorvos effective in field testing

Coccidia
Landau, I., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 403-407
analysis of analogy between Toxoplasma and Hepatozoon life cycles; importance and role of cyst formation in the Coccidia

Coccidia
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Coccidia
Mechow, A., 1975, Avian Path., v. 4 (1), 83-85
improved technique for separating oocysts from large amounts of feces

[Coccidia] koktsidii
Panchin, O. G.; et al., 1975, Veterinariia, Kiev (40), 100-104
helminths and coccidia, sheep, seasonal incidence on pastures, measures for control: Kalanchats'k region, Kherson oblast

Coccidia [sp.]
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Sceloporus occidentalis
S. graciosus
(blood of all): all from central California

Coccidia [sp.]
Carter, J. B., 1976, J. Applied Ecol., v. 13 (1), 103-122
Tipula paludosa
T. oleracea
T. luna
all from north-east England

Coccidia [sp.]?
Carter, J. B., 1976, J. Applied Ecol., v. 13 (1), 103-122
Tipula unca: north-east England

Coccidiae [sp.]
Guildal, J. A.; and Clausen, B., 1973, Norwegian J. Zool., v. 21 (4), 329-330 [Abstract]
Vulpes vulpes (feces): Denmark

Coccidia [sp.], dizoic oocyst
MacLean, S. A.; and Sawyer, T. K., 1976, Tr. Am. Micr. Soc., v. 95 (2), 269 [Abstract]
Scomber scombrus (liver): Chincoteague, Virginia

Coccidia [sp.], polyzoic oocyst
MacLean, S. A.; and Sawyer, T. K., 1976, Tr. Am. Micr. Soc., v. 95 (2), 269 [Abstract]
Scomber scombrus (liver, kidney): Chincoteague, Virginia

Coccidia D
Peters, W.; Tennant, L.; and McDermott, S. N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 8-10 [Demonstration]
lions: Knowsley Safari Park, Prescot, Lancashire

Coccidia [sp.], illus.
Shastri, U. V.; Ghafoor, M. A.; and Krishnamurthy, R., 1976, Indian J. Animal Sc., v. 46 (10), 556-561
Eimeria bareillyi, E. ellipsoidalis, Coccidia [sp.], endogenous development and associated lesions in buffalo calves: Parbhani

Coccidiosis
Chakrabarti, A.; et al., 1977, Indian J. Animal Health, v. 16 (1), 57-58
coccidiosis outbreak in Black Bengal kids, pathology: Baramunda village, Bhubaneswar (Orissa)

Coccidiosis
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Coccidiosis
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

Coccidiosis
Crestian, J., 1977, Clin. Vet., Milano, v. 100 (5), 395-397
carnivores, laboratory diagnosis, review

Coccidiosis
Duerr, U. M., 1976, Tierarztl. Umschau, v. 31 (4), 177-178
coccidiosis of dogs and cats, clinical trials of Trimethoprim-Sulfadiazin, good results

Coccidiosis
Eberhardt, U.; and Oettel, M., 1975, Pharmazie, v. 30 (4), 241-243
coccidiosis, structure-activity relationships of di-thiocarbamoyl hydrazine metal chelating agents, possible use as coccidiostats

Coccidiosis
Elefterescu, A.; Popescu, E.; and Dinu, M., 1976, Rev. Crest. Animalelor, v. 26 (1), 66-69
coccidiosis, chickens, testing of various coccidiostats, formulas for determining "coccidiostatic index" and "performance percentage", Clopydol gave best results

Coccidiosis
Euzeby, J. A., 1976, Vet. Parasitol., v. 1 (4), 289-298
coccidiosis, review and comments on the French literature

Coccidiosis
Finnie, E. P., 1976, Austral. Vet. Practitioner, v. 6 (1), 19-28
brief review of coccidiosis, Macropus giganteus, M. fuliginosus

Coccidiosis
Gabrisch, K., 1976, Prakt. Tierarzt, v. 57, Sondernummer, 37-40
parasites of reptiles, diagnosis, treatment, brief review

Coccidiosis
Isenbuegel, E., 1976, Prakt. Tierarzt, v. 57, Sondernummer, 21-27
Codrinol
Igel

Coccidiosis
Loeliger, H. C., 1974, Prakt. Tierarzt, v. 55, Sondernummer, 6-9
coccidiosis, rabbits, clinical signs, diagnosis, sulfonamide, review

Coccidiosis
Manuel, M. F.; and Alcarde, J. A., 1975, Philippine J. Vet. Med., v. 13 (1-2), 156-162
coccidiosis, broilers raised on deep litter, Amprol Plus, Zoamix, NF-180, comparative drug efficacies, growth and mortality rates

Coccidiosis
Nemeseri, L., 1972, Parasitol. Hungar., v. 5, 217-224
coccidiosis and ascaridiasis in domestic animals and poultry, furidin and rodanin assayed for possible prophylactic use in conditions of large scale animal husbandry

Coccidiosis
Onawunmi, O. A.; and Todd, A. C., 1976, Am. J. Vet. Research, v. 37 (6), 657-660
porcine coccidiosis (exper.), suppression and control with chlortetracycline combination, buquinolate, and lincomycin hydrochloride

Coccidiosis
Panda, N. C.; et al., 1973, Indian Vet. J., v. 50 (1), 22-26
coccidiosis, chicks, Bifuran, sulphaquinoxaline, development of resistance to single coccidiostat over 3 year period; good results against coccidia by alternating drugs yearly

Coccidiosis
Pout, D. D., 1973, Brit. Vet. J., v. 129 (6), 555-567
coccidiosis, lambs, faecal oocyst production, relationship to host age, variation in pattern of initial infection and levels of subsequent burden

Coccidiosis
Pout, D. D., 1976, Vet. Rec., v. 98 (17), 340-341
coccidiosis, sheep, brief review

Coccidiosis
Reichel, F.; Sevcikova, E.; and Firmanova, A., 1977, Veterinarstvi, v. 27 (5), 217-218
coccidiosis, chickens, rabbits, sulfakombin, clinical evaluation

Coccidiosis
Rose, M. E., 1976, Vet. Rec., v. 98 (24), 481-484
poultry coccidiosis, immunity and prospects for immunoprophylaxis, review

Coccidiosis
Spiess, H., 1976, Tierarztl. Umschau, v. 31 (7), 316-319
coccidiosis, poultry, Monensin in feed as prophylaxis, compared with Nicarbazin and Amprolmix

Coccidiosis
Vassiliades, G.; and Toure, S. M., 1975, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 28 (4), 481-489
digestive strongylosis, sheep, morantel tartrate, with or without anticoccidian drug (Cozurone), good control of all except Strongyloides

Coccidiosis
Vindevogel, H.; and Duchatel, J. P., 1976, Ann. Med. Vet., v. 120 (5), 333
incidence of intestinal coccidiosis in wild hares

Coccidiosis
Yvore, P., 1976, Avian Path., v. 5 (4), 237-252
methods for prevention of coccidiosis in poultry, review

Coccidium bigeminum Stiles 1891
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis bigemina (Stiles 1891) comb. n.

Coccidium bigeminum Stiles, 1891 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora miescheriana (Kuhn, 1865) comb. nov.

Coccidium bigeminum Stiles, 1891 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora rivolta (Grassi, 1879) Wenyon, 1923

Coccidium bigeminum var. canis Railliet and Lucet 1891 [pro parte?]
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis bigemina (Stiles 1891) comb. n.

Coccidium bigemina var. canis Railliet and Lucet 1891 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis miescheriana (Kuhn 1865) Labbe 1899

Coccidium bigeminum var. canis (Stiles, 1891) Railliet & Lucet, 1891 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora miescheriana (Kuhn, 1865) comb. nov.

Coccidium bigeminum var. cati Railliet and Lucet 1891
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis muris (Blanchard 1885) Labbe 1899

Coccidium bigeminum var. hominis Railliet and Lucet 1891 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hominis (Railliet and Lucet 1891) comb. n.

Coccidium bigeminum var. hominis Railliet & Lucet, 1891 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora suihominis nom. nov. [i.e., n. sp.]

Coccidium bigeminum var. hominis Railliet & Lucet, 1891 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hominis (Railliet & Lucet, 1891) comb. nov.

Coccidium bigeminum var. putorii Railliet and Lucet 1891
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
nomen nudum

Coccidium bigeminum var. putorii Railliet & Lucet, 1891
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora putorii (Railliet & Lucet, 1891) comb. nov.

Coccidium mitrarium Laveran & Mesnil, 1902
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
as syn. of Eimeria mitraria (Laveran & Mesnil, 1902)

Coccidium rivolta Grassi 1879
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora rivolta (Grassi 1879) Wenyon 1923

Coccidium rivolta Grassi, 1879 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora rivolta (Grassi, 1879) Wenyon, 1923

Coccidium rivoltae Leukart, 1886 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora rivolta (Grassi, 1879) Wenyon, 1923

Cochleomeritus Hoshide, 1944 emend.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Cochleomeritus emersoni sp. n., illus.
Levine, N. D., 1973, J. Protozool., v. 20 (5), 546-548
Diopatra ornata (lumen of intestine): Pacific Ocean off Point Hueneme, near Oxnard, California

Cochleomeritus emersoni Levine, 1973
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Cochleomeritus lysidici Hoshide, 1944
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Cochlosoma sp.
Leibovitz, L., 1973, Am. J. Vet. Research, v. 34 (8), 1053-1061
observed in breeder ducks, affected or not affected with necrotic enteritis

Coelogregarina ephestiae Ghelelovitch, 1948 sensu Weiser, 1954 non Ghelelovitch, 1948
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Mattesia dispora Naville

Coelomophaga n. gen.
Manier, J. F.; Akbarieh, M.; and Bouix, G., 1977, Ztschr. Parasitenk., v. 51 (2), 147-158
Nephridiophagidae
mt: C. chydori n. gen. n. sp.

Coelomophaga chydori n. gen. n. sp. (mt), illus.
Manier, J. F.; Akbarieh, M.; and Bouix, G., 1977, Ztschr. Parasitenk., v. 51 (2), 147-158
Chydorus sphaericus (lumiere intestinale, cellules epitheliales, cavite hemocoelienne): Nord de Montpellier (Herault, France)

Coelophrya n. gen.
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Hoplitophryidae, Radiophryinae; mt: C. roquei n.g., n.sp.

Coelophrya de Puytorac et Dragesco, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Metaracoelophryinae n. sous-fm.

Coelophrya roquei n.g., n. sp., illus. (mt)
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Alma emini (tube digestif): Gabon

Coelophrya roquei de Puytorac et Dragesco, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Coelosporidium
Narasimhamurti, C. C.; and Kalavati, C., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (1), 20-25
genus should be retained

Coelosporidium oithonae n. sp., illus.
Narasimhamurti, C. C.; and Kalavati, C., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (1), 20-25
Oithona rigida (body cavity)

Coelotropha Hennere, 1963
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Grelliidae fam. nov.; diagnosis
includes: C. durchoni (Vivier, 1963) Vivier & Hennere, 1964; C. vivieri Hennere, 1963 (type species)

Coleorhynchus heros (Schneider), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Nepa cinerea (intestine): Poland

Colpoda sp.
Grassmick, R. A.; and Rowley, W. A., 1973, J. Protozool., v. 20 (4), 509
Culex tarsalis larvae: reared in Medical Entomology Laboratory, Iowa State University

Colpoda steini Maupas, 1883
Mattis, D.; and Kvasz, L., 1975, J. Protozool., v. 22 (3), 67A [Abstract]
specific adaptation to body temperature of man
humans (lungs): Indochina

Colponema [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Sparus aurata
Mugil cephalus
Liza ramada
all from Elat, Gulf of Aqaba, Red Sea

Colponema sp.
Paperna, I.; et al., 1977, Aquaculture, v. 10 (3), 195-213
ectoparasites of cultured Sparus aurata, formalin, good results: Elat, Israel

Cometoides
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
new diagnosis

Cometoides capitatus (Leger, 1892), illus.
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
description
Hydrochara caraboides (region posterieure de l'intestin moyen): Loire-Atlantique (Ouest de la France)

Cometoides pileatus n. sp., illus.
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
Hydrophilus piceus: Loire-Atlantique (Ouest de la France)

Conorhynchus Greeff, 1880
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Zygosoma Labbe, 1899

Conorhynchus echiuri Greeff, 1880, pro parte
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Zygosoma gibbosum (Greeff, 1880) Labbe, 1899

Conorhynchus gibbosus Greeff, 1880, pro parte
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Zygosoma gibbosum (Greeff, 1880) Labbe, 1899

Contophrya n. gen.
  de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
    Hoplitophryidae, Contophryinae n.subfam.; tod: C. cachoni n.g., n.sp.

Contophrya de Puytorac et Dragesco, 1969
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Contophryidae n. fam.

Contophrya cachoni n.g., n. sp., illus. (tod)
  de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
    Alma (tube digestif): Gabon

Contophrya cachoni de Puytorac et Dragesco, 1969 illus.
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Alma emini

Contophryidae n. fam.
  de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
    Hoplitophryoidea nv. super-fam.
    includes: Contophrya; Dicontophrya

Contophryinae n.subfam.
  de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
    Hoplitophryidae
    includes: Contophrya n.gen.; Dicontophrya n.gen.

Contortiocorpa Bhatia & Setna, 1938
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Lecudinidae, revision and checklist of species
    diagnosis

Contortiocorpa prashadi Bhatia & Setna, 1938
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Cotyloepimeritus Hoshide, 1935
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Bhatiella Setna, 1931

Cotyloepimeritus iwamusi Hoshide, 1935
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Bhatiella iwamusi (Hoshide, 1935) comb. nov.

Crithidia
  Bos, H. J., 1975, Ztschr. Parasitenk., v. 47 (2), 119-129
    Entamoeba histolytica, human, strains isolated from all 5 symptomatic cases but only 3 out of 10 asymptomatic carrier cases, axenic culture following monoxenic culture with Crithidia, media used; discussion of possible relationship between virulence of strain and ease of culture

Crithidia
  Bos, H. J.; and Hage, A. J., 1975, Ztschr. Parasitenk., v. 47 (2), 79-89
    Entamoeba histolytica, virulent and carrier strains from humans, cultured axenically or with bacteria or Crithidia or passaged through hamster liver, varying virulence in experimental hamster liver infections

Crithidia
  Daggett, P. M.; and Decker, J. E., 1974, J. Protozool., v. 21 (3), 429 [Abstract]
    comparison of excreted factors of insect trypanosomatids at 25 C vs. 37 C, results indicate secretion products are primarily under parasite not host control

Crithidia
  Kallinikova, V. D., 1977, Molec. and Cell. Biochem., v. 14 (1-3), Feb. 4, 55-58
    Kinetoplastida, kinetoplast genetic system, implications for concept of cell differentiation and phylogeny, relationship to phenotype of these flagellates

Crithidia
  Schnur, L. F., 1977, J. Protozool., v. 24 (2), Suppl., 5A-6A [Abstract]
    Leptomonas, Crithidia, Herpetomonas, L[eishmania] tropica, L[eishmania] tropica major, L[eishmania] tropica minor, L[eishmania] donovani, excreted metabolic factors (EF) and homologous antisera, cross-reactions, no EF apparently produced by Trypanosoma cruzi, suggests closer relationship between insect flagellates and leishmanias than between these and trypanosomes like Trypanosoma cruzi

Crithidia
  Simpson, L., 1973, J. Protozool., v. 20 (1), 2-8
    salivarian and stercorarian trypanosomes, kinetoplast DNA, structure and function, review

Crithidia
  Wolstenholme, D. R.; et al., 1974, J. Protozool., v. 21 (5), 622-631
    Crithidia, kinetoplast DNA, structure, replication, informational capacity, physicochemical and electron microscope studies, review with some unpublished results

Crithidia sp.
  Carvalho, A. L. M.; and Deane, M. P., 1974, J. Protozool., v. 21 (1), 5-8
    Trypanosomatidae isolated from Zelus leucogrammus, only Blastocrithidia thought to be a parasite proper of this host spread by cannibalism and ingestion of cysts, other trypanosomatids acquired by feeding on insect prey: Brazil

Crithidia sp.
Injeyan, H. S.; McLaughlin, J.; and Meerovitch, E., 1975, J. Protozool., v. 22 (3), 18A-19A [Abstract]
Crithidia sp., whole and subcellular fractions, cytochemical localization of acid phosphatase

Crithidia sp.
Injeyan, H. S.; and Meerovitch, E., 1974, J. Protozool., v. 21 (5), 738-742
Crithidia sp., inhibition of overall growth and macromolecular synthesis by juvenile hormone, inhibition of RNA synthesis of particular interest, implications for mode of action in insect systems

Crithidia sp.
McLaughlin, J.; Injeyan, H. S.; and Meerovitch, E., 1975, J. Protozool., v. 22 (3), 21A [Abstract]
Crithidia sp., subcellular heterogeneity of acid phosphohydrolase activities

Crithidia sp.
Oelerich, S., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 296-304
Leishmania spp., Trypanosoma cruzi, Crithidia sp., Mycobacterium smegmatis, antigenic analyses and cross reactions, double gel-diffusion, complement-fixation, indirect hemagglutination

Crithidia sp.
Raether, W.; and Uphoff, M., 1976, Internat. J. Parasitol., v. 6 (2), 121-125
Entamoeba histolytica axenic and monoxenic (in association with Crithidia sp.) cultures, survival after storage in liquid nitrogen, semiautomatic vs. automatic freezing, influence of specimen concentration and different plastic storage vessels

Crithidia acanthocephali
Manning, J. E.; and Wolstenholme, D. R., 1976, J. Cell Biol., v. 70 (2, pt. 1), 406-418
Crithidia acanthocephali, mode of replication of nuclear and kinetoplast DNA elucidated by buoyant density shift experiments using deuterium oxide

Crithidia acanthocephali, illus.
Wolstenholme, D. R.; et al., 1974, J. Protozool., v. 21 (5), 622-631
Crithidia, kinetoplast DNA, structure, replication, informational capacity, physicochemical and electron microscope studies, review with some unpublished results

Crithidia deanei, illus.
Camargo, E. P.; and Freymuller, E., 1977, Nature, London (5632), v. 270, 52-53
Crithidia, presence of ornithine carbamoyltransferase in species harboring bacteria-like endosymbionts (C. deanei, C. oncopelti) enables these species to synthesize arginine from ornithine thereby conferring a nutritional advantage on the protozoan host

Crithidia deanei (Carvalho), illus.
Mundim, M. H.; et al., 1974, J. Protozool., v. 21 (4), 518-521
Crithidia deanei, cultivation in defined medium, unexpectedly simple nutritional requirements, bacterial endosymbiont revealed by electron microscopy may provide other essential nutrients

Crithidia deanei
Mundim, M. H.; and Roitman, I., 1977, J. Protozool., v. 24 (2), 329-331
Crithidia deanei, strain cured of its bacteria-like symbiote by chloramphenicol had additional nutritional requirements beyond those of the parent strain

Crithidia euryophthalmi
Daggett, P. M.; and Decker, J. E., 1973, J. Protozool., v. 20 (4), 512
6 insect trypanosomatids, increased survival in mice with repeated passage associated with alteration in secretion

Crithidia fasciculata
Al Chalabi, K.; and Gutteridge, W. E., 1974, J. Protozool., v. 21 (3), 449
Crithidia fasciculata, thymine excretion

Crithidia fasciculata
Al Chalabi, K.; and Gutteridge, W. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146 [Abstract]
Trypanosoma spp., Crithidia fasciculata, C. oncopelti, thymidine phosphorylase activity, salvage pathways, inhibition of activity by suramin and ethidium

Crithidia fasciculata
Al Chalabi, K.; and Gutteridge, W. E., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 268 [Abstract]
Crithidia fasciculata, activity of thymidylate phosphatase

Crithidia fasciculata
Al Chalabi, K.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), xv-xvi [Abstract]
6 spp. of Trypanosomatidae, thymidylate synthetase activity, ranks as potential target for chemotherapeutic attack since it is markedly more sensitive to suramin inhibition than isofunctional mammalian and bacterial

Crithidia fasciculata
Al Chalabi, K.; and Gutteridge, W. E., 1977, Parasitology, v. 74 (3), 299-312
deoxythymidylate catabolism in some trypanosomatids

Crithidia fasciculata
Bacchi, C. J.; et al., 1974, J. Protozool., v. 21 (3), 430 [Abstract]
Crithidia fasciculata, purification and properties of soluble, NAD-linked α-glycerophosphate dehydrogenase, similar preliminary studies with Leptomonas

C[rithidia] fasciculata
Barker, D. C.; and Brack, C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 269 [Abstract]
Trypanosoma brucei, T. mega, Crithidia fasciculata, dark-field electron microscopy of kinetoplast DNA

Crithidia fasciculata
Berens, R. L.; and Marr, J. J., 1977, Exper. Parasitol., v. 43 (2), 407-413
Leishmania donovani, L. brasiliensis, pyruvate kinase, regulatory properties in relation to regulation of glycolysis, comparison with Crithidia fasciculata

Crithidia fasciculata, illus.
Brooker, B. E., 1976, Parasitology, v. 72 (3), 259-267
Crithidia fasciculata, comparison of haptomonads attached to cuticular lining of Anopheles gambiae hindgut with those from mosquito foregut and from rosettes in culture, prominent cell coat only in former suggests cell coat formation is in response to appropriate environmental conditions, cytochemical staining indicates presence of carbohydrates

Crithidia fasciculata, illus.
Cosgrove, W. B., 1973, J. Protozool., v. 20 (2), 191-194
Trypanosomatidae, history and present status of knowledge of kinetoplasts, presidential address

Crithidia fasciculata
Daniels, C. J.; and Palmer, F. B. St. C., 1977, Canad. Fed. Biol. Soc., Programme and Proc. 20. Ann. Meet., v. 20, 74 [Abstract]
Crithidia fasciculata, microsomal preparations, synthesis of phosphatidylinositol

Crithidia fasiculata
Dave, D.; Gutteridge, W. E.; and Richards, W. H. G., 1975, J. Protozool., v. 22 (3), 50A-51A [Abstract]
Crithidia fasiculata vs. rat liver, difference in mechanism for conversion of dihydroorotate to orotate

Crithidia fasciculata
Dewey, V. C.; and Kidder, G. W., 1973, J. Protozool., v. 20 (5), 678-682
Trypanosoma cruzi, growth inhibition by allopurinol, both purines and pyrimidines required for reversal, actual inhibitor is allopurinol ribotide (not free base) acting on orotidine 5'-phosphate decarboxylase

Crithidia fasciculata
Dewey, V. C.; and Kidder, G. W., 1977, Canad. J. Biochem., v. 55 (1), 110-112
Crithidia fasciculata, inhibited growth with adenine analog, 4-aminopyrazolo(3,4-d) pyrimidine, adenine plus a pyrimidine (or a pyrimidine derivative) necessary for reversal of inhibition, adenine analog not inhibitory to enzymes of pyrimidine biosynthetic pathway; if not for its untoward effects in mammals, 4-APP might be suggested as possible therapeutic agent in trypanosomal infections

Crithidia fasciculata
Dixon, H., 1974, J. Protozool., v. 21 (3), 450 [Abstract]
Crithidia fasciculata, effect of inhibitors of bacterial protein synthesis

Crithidia fasciculata
Dooris, P. M.; and McGhee, R. B., 1976, J. Protozool., v. 23 (3), 433-437
Crithidia harmosa, C. fasciculata, differentiation by immunological methods (agglutination, indirect fluorescent antibody) and by polyacrylamide gel slab electrophoresis (number and relative mobilities of component protein bands)

Crithidia fasciculata
Edwards, C.; and Lloyd, D., 1977, J. Gen. Microbiol., v. 100 (2), 339-346
Crithidia fasciculata, subcellular fractionation by differential and zonal centrifugation of digitonin-treated suspension; distribution of enzymes after fractionation; lysosomes and mitochondria separated

Crithidia fasciculata
Goldberg, B.; et al., 1974, J. Protozool., v. 21 (2), 322-326
Leptomonas sp., inhibition by several standard antiprotozoal drugs of growth and oxygen uptake of cells and particulate preparations, possible use as model organism for screening antitrypanosomatid agents as compared to Crithidia fasciculata

Crithidia fasciculata
Grymberg, N.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 35-36
mice, immunization with Crithidia fasciculata (live suspension, ribosomal fraction, and purified RNA) induced certain degree of protection (decrease of parasitaemia) against infection with Trypanosoma cruzi

Crithidia fasciculata
Hajduk, S. L., 1976, J. Protozool., v. 23 (2), 17A [Abstract]
Trypanosoma equiperdum, Crithidia fasciculata, extranuclear DNA, distribution in normal and dyskinetoplastic cells

Crithidia fasciculata
Hill, G. C.; and Bonilla, C. A., 1974, J. Protozool., v. 21 (5), 632-638
Crithidia fasciculata, isolation of DNA-dependent RNA polymerases, purified nuclear and kinetoplast DNA can serve as templates for these RNA polymerases, this RNA synthesis is inhibited by several trypanocidal drugs, hybridization experiments revealed no homologies between Blastocrithidia culicis and Crithidia fasciculata K-DNA

Crithidia fasciculata
Hill, G. C.; and Pettigrew, G. W., 1975, J. Protozool., v. 22 (3), 16A [Abstract]
Crithidia fasciculata, amino acid sequence of cytochrome $\underline{c}_{555}$, differences from C. oncopelti cytochrome $\underline{c}_{557}$

Crithidia fasciculata
Kidder, G. W.; Dewey, V. C.; and Nolan, L. L., 1976, Canad. J. Biochem., v. 54 (1), 32-41
Crithidia fasciculata, enzymes of the orotate biosynthetic pathway

Crithidia fasciculata
Kidder, G. W.; Dewey, V. C.; and Nolan, L. L., 1977, Arch. Biochem. and Biophys., v. 183 (1), 7-12
Crithidia fasciculata, adenine deaminase, properties and localization

Crithidia fasciculata
Leon, W.; Cruz, F. S.; and Vasconcellos, M. E., 1975, J. Protozool., v. 22 (2), 277-280
Crithidia fasciculata, growth inhibition by 4-nitroquinoline 1-oxide, dicoumarol and bovine albumin reversed inhibition

C[rithidia] fasciculata
Linstead, D. J.; Klein, R. A.; and Eldridge, M. V., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 267-268 [Abstract]
Trypanosoma mega, T. brucei, Crithidia fasciculata, presence of 3 enzymes capable of carbon dioxide fixation in culture as source of carbon for metabolism

Crithidia fasciculata
Lipschik, G.; et al., 1976, J. Protozool., v. 23 (2), 30A [Abstract]
trypanosomatids, polyamines and Mg++ content

Crithidia fasciculata
Manaia, A. da C.; and Roitman, I., 1971, Rev. Microbiol., S. Paulo, v. 2 (4), 177-179
Crithidia fasciculata, inhibition of growth by ethidium bromide; more effective at 32°C than 28°C; adenine neutralized the action of ethidium

Crithidia fasciculata
Manaia, A. da C.; and Roitman, I., 1977, J. Protozool., v. 24 (1), 192-195
Crithidia fasciculata grown in presence of ethidium bromide, dyskinetoplasty and growth inhibition, changes in oxidative metabolism and enzyme profiles

Crithidia fasciculata
Marr, J. J.; Birenbaum, M. E.; and Ladenson, J. H., 1977, Exper. Parasitol., v. 42 (2), 322-330
Crithidia fasciculata, aerobic fermentation, in vivo studies, appearance kinetics of intermediates, regulation

Crithidia fasciculata
Miller, P. G. G.; Linstead, D. J.; and Klein, R. A., 1976, Parasitology, v. 73 (2), xvi [Abstract]
5 spp. of trypanosomatids, enzymes of acetyl CoA metabolism, possible routes for synthesis and utilization of acetyl CoA

Crithidia fasciculata
Nichols, J. M.; and Cross, G. A. M., 1976, Biochem. and Biophys. Research Commun., v. 71 (3), 796-802
Crithidia fasciculata, kinetoplast DNA preparations, contamination by extra components probably nuclear DNA

Crithidia fasciculata, illus.
Nichols, J. M.; and Cross, G. A. M., 1977, J. Gen. Microbiol., v. 99 (2), 291-300
Crithidia fasciculata, isolation of mitochondria, mitochondrial RNA, and mitochondrial (kinetoplast) DNA

Crithidia fasciculata
Ozanich, K. M.; and Marr, J. J., 1976, Abst. Ann. Meet. Am. Soc. Microbiol., 141
Crithidia fasciculata, regulation of glycolysis, absence of feedback inhibition of phosphofructokinase

Crithidia fasciculata
Palmer, F. B. St. C., 1974, J. Protozool., v. 21 (1), 160-163
Crithidia fasciculata, biosynthesis of choline and ethanolamine phospholipids

Crithidia fasciculata
Palmer, F. B. St. C., 1976, Biochim. et Biophys. Acta, v. 441 (3), 477-487
Crithidia fasciculata homogenates contain a phosphatase and a phosphodiesterase which hydrolyse triphosphoinositides

Crithidia fasciculata
Palmer, F. B. St. C., 1976, Canad. Fed. Biol. Soc., Programme and Proc. 19. Ann. Meet., v. 19, 32 [Abstract]
Crithidia fasciculata, hydrolysis of triphosphoinositide (TPI) by TPI phosphatase and TPI phosphodiesterase

Crithidia fasciculata
Paulin, J. J., 1974, J. Protozool., v. 21 (3), 435 [Abstract]
Crithidia fasciculata, Blastocrithidia culicis, Trypanosoma cruzi, three-dimensional reconstructions of chondriome using high voltage electron microscopy

Crithidia fasciculata, illus.
Paulin, J. J., 1974, Proc. 32. Ann. Meet. Electron Microsc. Soc. America (St. Louis, Missouri, Aug. 13-15), 62-63
Trypanosoma equiperdum, Blastocrithidia culicis, Crithidia fasciculata, 3-dimensional cell structure, high voltage electron microscopy

Crithidia fasciculata
Paulin, J. J., 1975, J. Protozool., v. 22 (3), 16A [Abstract]
unitary structure of chondriome of 3 spp. of Trypanosomatidae confirmed by application of high voltage electron microscopy

Crithidia fasciculata, illus.
Paulin, J. J., 1977, Exper. Parasitol., v. 41 (2), 283-289
Crithidia fasciculata mitochondrion, reconstruction based on serial thick sections and high-voltage electron microscopy

Crithidia fasciculata, illus.
Pereira, N. M.; et al., 1977, J. Protozool., v. 24 (4), 511-514
Crithidia fasciculata, Herpetomonas samuelpessoai, Leishmania tarentolae, isolation of flagella, 3 types of flagella give similar electrophoretic pattern of proteins, H. samuelpessoai and to a lesser extent C. fasciculata flagella confer protection against Trypanosoma cruzi infections in mice

Crithidia fasciculata
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Crithidia fasciculata
Risby, E. L., 1976, J. Protozool., v. 23 (2), 20A [Abstract]
Crithidia fasciculata, Blastocrithidia culicis, Trypanosoma, phosphohexose isomerase, comparison of characteristics

Crithidia fasciculata
Roitman, I., 1970, Rev. Microbiol., Sao Paulo, v. 1 (2), 93-94
Crithidia fasciculata, growth factors in culture medium above 33°C, lipid and osmotic requirements

Crithidia fasciculata
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Crithidia fasciculata
Simpson, A. M.; and Simpson, L., 1974, J. Protozool., v. 21 (2), 379-382
continuous labeling of Crithidia fasciculata DNA with [^{3}H]thymidine is hindered by apparent presence of a thymidine phosphorylase-like enzyme activity

Crithidia fasciculata, illus.
Simpson, A. M.; and Simpson, L., 1974, J. Protozool., v. 21 (5), 774-781
Crithidia fasciculata, kinetoplast DNA, isolation technique, physical properties of networks and component minicircles

Crithidia fasciculata
Simpson, A. M.; and Simpson, L., 1975, J. Protozool., v. 22 (3), 22A [Abstract]
Crithidia fasciculata, peripheral localization of DNA synthesis within kinetoplast DNA networks

Crithidia fasciculata, illus.
Simpson, A. M.; and Simpson, L., 1976, J. Protozool., v. 23 (4), 583-587
Crithidia fasciculata, Leishmania tarentolae, pulse-labeling of kinetoplast DNA, localization of 2 sites of synthesis within networks and kinetics of labeling of closed minicircles

Crithidia fasciculata, illus.
Simpson, L., 1973, J. Protozool., v. 20 (1), 2-8
salivarian and stercorarian trypanosomes, kinetoplast DNA, structure and function, review

Crithidia fasciculata
Steinert, M.; and Van Assel, S., 1975, J. Protozool., v. 22 (3), 85A [Abstract]
Crithidia, association of circular DNA molecules with kinetoplast and absence of such molecules in nuclear DNA suggests mitochondrial origin

Crithidia harmosa
Daggett, P.-M.; Norman, M.; and Simione, F. P., jr., 1976, J. Protozool., v. 23 (2), 9A-10A [Abstract]
Crithidia harmosa, C. luciliae, successful lyophilization

Crithidia harmosa
Dooris, P. M.; and McGhee, R. B., 1976, J. Protozool., v. 23 (3), 433-437
Crithidia harmosa, C. fasciculata, differentiation by immunological methods (agglutination, indirect fluorescent antibody) and by polyacrylamide gel slab electrophoresis (number and relative mobilities of component protein bands)

Crithidia harmosa McGhee, Hanson, and Schmittner 1969, illus.
Keppel, A. D.; and Janovy, J., jr., 1977, J. Parasitol., v. 63 (5), 879-882
Herpetomonas megaseliae and Crithidia harmosa, growth on blood-agar plates as discrete clone colonies, colony morphology differs in the two genera and species, possible applications

Crithidia hutneri sp. n.
Roitman, I.; et al., 1977, J. Protozool., v. 24 (4), 553-556
growth at high temperature in defined medium, nutritional requirements
Cosmoclopius sp. (final gut): Mambai, State of Goias, Brazil

Crithidia luciliae
Carvalho, A. L. M.; and Deane, M. P., 1974, J. Protozool., v. 21 (1), 5-8
Trypanosomatidae isolated from Zelus leucogrammus, only Blastocrithidia thought to be a parasite proper of this host spread by cannibalism and ingestion of cysts, other trypanosomatids acquired by feeding on insect prey: Brazil

Crithidia luciliae, illus.
Crowe, W.; and Kushner, I., 1977, Arthritis and Rheum., v. 20 (3), 811-814
Crithidia luciliae, immunofluorescent method using Crithidia to detect antibodies to double-stranded DNA, possible use in study of human collagen and nephritic diseases

Crithidia luciliae
Daggett, P. M.; and Decker, J. E., 1973, J. Protozool., v. 20 (4), 512
6 insect trypanosomatids, increased survival in mice with repeated passage associated with alteration in secretion

Crithidia luciliae
Daggett, P. M.; Norman, M.; and Simione, F. P., jr., 1976, J. Protozool., v. 23 (2), 9A-10A [Abstract]
Crithidia harmosa, C. luciliae, successful lyophilization

Crithidia luciliae
Deegan, M. J.; Walker, S. E.; and Lovell, S. E., 1977, Am. J. Clin. Path., v. 68 (1), 103 [Abstract]
Crithidia luciliae which contains kinetoplast composed of non-histone associated DNA used to screen sera for antibodies to double-stranded DNA in evaluation of arthritic conditions

Crithidia luciliae
Gueugnot, J.; et al., 1976, Compt. Rend. Soc. Biol., Paris, v. 170 (4), 782-786
Crithidia luciliae, suspensions treated with 30 lectins of plant and fungus, agglutination, fixation upon membrane and flagella in relation to lectin structure; more precise knowledge of glucidic receptors on membranes; possible use of technique in identification, taxonomy and study of intracellular structures

Crithidia luciliae
Jarzabek-Chorzelska, M.; et al., 1976, Przegl. Dermat., v. 63 (5), 579-584
Crithidia luciliae used as antigen substrate to demonstrate antibodies against native DNA in persons with collagen diseases

Crithidia luciliae
Kleisen, C. M.; et al., 1976, European J. Biochem., v. 64 (1), 153-160
Crithidia luciliae, kinetoplast DNA networks, characterization of novel component of high complexity

Crithidia luciliae
Kleisen, C. M.; Borst, P.; and Weijers, P. J., 1976, European J. Biochem., v. 64 (1), 141-151
Crithidia luciliae, kinetoplast DNA, minicircles are heterogeneous in base sequence

Crithidia luciliae
Newton, B. A.; Steinert, M.; and Borst, P., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 259-260 [Abstract]
Trypanosoma mega, Crithidia luciliae, differentiation by hybridization of complementary RNA with kinetoplast DNA

Crithidia luciliae
Opperdoes, F. R.; et al., 1977, European J. Biochem., v. 76 (1), 29-39
Trypanosoma brucei bloodstream form, subcellular fractionation, localization of glycerol-3-phosphate oxidase in mitochondrion and particulate NAD$^+$-linked glycerol-3-phosphate dehydrogenase in microbodies, supplementary studies with Crithidia luciliae

Crithidia luciliae
Opperdoes, F. R.; de Rijke, D.; and Borst, P., 1976, Comp. Biochem. and Physiol., v. 54 (1B), 7-12
Crithidia luciliae, characterization of mitochondrial adenine nucleotide translocator and ATPase, significant differences between ATPase of trypanosomes and their hosts that could potentially be exploited in chemotherapy

Crithidia luciliae of Carvalho, 1973,1974
Roitman, I.; et al., 1977, J. Protozool., v. 24 (4), 553-556
as syn. of Crithidia luciliae thermophila s. sp. n.

Crithidia luciliae
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Crithidia luciliae
Steinert, M.; and Van Assel, S., 1975, J. Protozool., v. 22 (3), 85A [Abstract]
Crithidia, association of circular DNA molecules with kinetoplast and absence of such molecules in nuclear DNA suggests mitochondrial origin

Crithidia luciliae
Weislogel, P. O.; et al., 1977, Biochim. et Biophys. Acta, v. 478 (2), 167-179
Crithidia luciliae, characterization of kinetoplast DNA networks

Crithidia luciliae
Wolstenholme, D. R.; et al., 1974, J. Protozool., v. 21 (5), 622-631
Crithidia, kinetoplast DNA, structure, replication, informational capacity, physicochemical and electron microscope studies, review with some unpublished results

Crithidia luciliae thermophila s. sp. n.
Roitman, I.; et al., 1977, J. Protozool., v. 24 (4), 553-556
growth at high temperature in defined medium, nutritional requirements
Syn.: Crithidia luciliae of Carvalho, 1973, 1974
Zelus leucogrammus

Crithidia oncopelti
Al Chalabi, K.; and Gutteridge, W. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146 [Abstract]
Trypanosoma spp., Crithidia fasciculata, C. oncopelti, thymidine phosphorylase activity, salvage pathways, inhibition of activity by suramin and ethidium

Crithidia oncopelti
Al Chalabi, K.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), xv-xvi [Abstract]
6 spp. of Trypanosomatidae, thymidylate synthetase activity, ranks as potential target for chemotherapeutic attack since it is markedly more sensitive to suramin inhibition than isofunctional mammalian and bacterial enzymes

Crithidia oncopelti
Al Chalabi, K.; and Gutteridge, W. E., 1977, Parasitology, v. 74 (3), 299-312
deoxythymidylate catabolism in some trypanosomatids

Crithidia oncopelti
Camargo, E. P.; and Freymuller, E., 1977, Nature, London (5632), v. 270, 52-53
Crithidia, presence of ornithine carbamoyltransferase in species harboring bacteria-like endosymbionts (C. deanei, C. oncopelti) enables these species to synthesize arginine from ornithine thereby conferring a nutritional advantage on the protozoan host

Crithidia oncopelti
Chang, K.-P., 1974, J. Protozool., v. 21 (5), 699-707
ultrastructural evidence suggests that bipolar bodies of Crithidia oncopelti and diplosomes of Blastocrithidia culicis are endosymbiotic bacteria with defective cell walls and that they are subject to destruction by treatment with chloramphenicol but not penicillin

Crithidia oncopelti, illus.
Chang, K.-P., 1975, J. Protozool., v. 22 (2), 271-276
Blastocrithidia culicis, Crithidia oncopelti, intracellular symbiotes can be eliminated by single chloramphenicol treatment with subsequent reduced growth of flagellates

Crithidia oncopelti
Chang, K.-P., 1976, J. Protozool., v. 23 (2), 241-244
Blastocrithidia culicis, Crithidia oncopelti, symbiote-free strains: liver extract as essential growth factor in defined medium; cross-reactivity in reciprocal agglutination test with symbiote-containing strains indicates loss of symbiotes does not affect antigenic identity

Crithidia oncopelti
Dwyer, D. M.; and Chang, K.-P., 1976, Proc. Nat. Acad. Sc., U.S.A., v. 73 (3), 852-856
Crithidia oncopelti, bacterial endosymbiote-containing vs. symbiote-free strain, comparison of surface-membrane carbohydrate ligands by lectin-mediated agglutination and lectin-ultrastructure localization

Crithidia oncopelti
Hill, G. C.; and Pettigrew, G. W., 1975, J. Protozool., v. 22 (3), 16A [Abstract]
Crithidia fasciculata, amino acid sequence of cytochrome $\underline{c}_{555}$, differences from C. oncopelti cytochrome $\underline{c}_{557}$

Crithidia oncopelti, illus.
Holwill, M. E. J.; and McGregor, J. L., 1976, J. Exper. Biol., London, v. 65 (1), 229-242
effects of varying concentrations of calcium on motility of different preparations of flagella from Crithidia oncopelti (chemically extracted cells, mechanically detached flagella, and cells treated with ionophores known to transport divalent cations across membranes), changes occurred in bend shape, wave direction and frequency, results suggest that wave reversal in living Crithidia is induced by release of calcium ions within the flagellum following stimulation of the membrane, stroboscopic and high-speed cine photographic techniques

Crithidia oncopelti
Kozlov, I. P.; et al., 1976, Izvest. Akad. Nauk SSSR, s. Biol. (2), 238-244
Crithidia oncopelti, dynamics of accumulation of products of peroxidation of lipids, minimal quantity during exponential growth, increase during stationary phase

Crithidia oncopelti
Pettigrew, G. W.; and Smith, G. M., 1977, Nature, London (5595), v. 265, 661-662
Crithidia oncopelti, cytochrome c557, novel N-terminal protein blocking group identified as dimethylproline

Crithidia oncopelti
Titova, T. S.; et al., 1976, Izvest. Akad. Nauk SSSR, s. Biol. (5), 754-759
Crithidia oncopelti, influence of olivomycine on changes in lipid composition and content of sensitive and resistant C. oncopelti cultures

Crithidia oncopelti
Zaitseva, G. N.; Kolesnikov, A. A.; and Shirshov, A. T., 1977, Molec. and Cell. Biochem., v. 14 (1-3), 47-54
Crithidia oncopelti, kinetoplast DNA, structure, transcription, protein-synthesizing apparatus, protein synthesis controlled by kinetoplast genome

Crithidia oncopelti
Zaitseva, G. N.; Mett, I. L.; and Kolesnikov, A. A., 1976, Biokhimiia, v. 41 (8), 1406-1411
Crithidia oncopelti, DNA-dependent RNA-polymerase activity of isolated kinetoplasts, effect of antibiotics and intercalating agents

Cryptobia catostomi Bower and Woo
Bower, S. M.; and Woo, P. T. K., 1977, Exper. Parasitol., v. 43 (1), 63-68
Cryptobia catostomi, use of in vitro plasma incubation test in study of host specificity (plasma of 6 refractive fishes had cryptobiacidal titers) and of host resistance mechanism (suggested that alternate pathway of complement activation is one mechanism of 'natural immunity' by vertebrates that are related to the susceptible host)

Cryptobia intestinalis (Leger 1905)
Brugerolle, G., 1976, J. Protozool., v. 23 (4), 5A [Abstract]
ultrastructure
Box boops (oesophage)

Cryptobia vaginalis (Hesse, 1910), illus.
Vickerman, K., 1977, J. Protozool., v. 24 (2), 221-233
Cryptobia vaginalis, light and electron microscopic morphology, kinetoplast DNA dispersed throughout single mitochondrion, feeding by pinocytosis through cytopharynx, bacterial symbiotes
Haemopis sanguisuga (vagina)
Hirudo medicinalis (vagina)

Cryptocaryon irritans, illus.
Herkner, H., 1975, Fisch u. Umwelt (1), 139-146
parasites as cause of disease in aquarium fish, review

Cryptosporidium [sp.]
Brownstein, D. G.; et al., 1977, Vet. Path., v. 14 (6), 606-617
Cryptosporidium in snakes, severe hypertrophic gastritis, pathological changes
Elaphe guttata
E. subocularis
Sansinia madagascarensis
Crotalus horridus
all from Baltimore Zoo, Baltimore, Maryland [and/or] National Zoological Park, Washington, D.C.

Cryptosporidium [sp.]
Kennedy, G. A.; Kreitner, G. L.; and Strafuss, A. C., 1977, J. Am. Vet. Med. Ass., v. 170 (3), 348-350
pigs (large intestine)

'?Cryptosporidium' of Bearup (1954) pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

'?Cryptosporidium' of Bearup (1954) pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis ovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975

Cryptosporidium sp. Tyzzer, 1929
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Cryptosporidium [sp.]
Nime, F. A.; et al., 1976, Gastroenterology, v. 70 (4), 592-598
Cryptosporidium as probable cause of severe acute self-limited enterocolitis in 3-year-old child, first report of human infection, light and electron microscopic findings in rectal biopsy, source of infection not established, value of sigmoidoscopy and biopsies for determining etiology of gastrointestinal infections: Vanderbilt University Hospital, Nashville, Tennessee

Cryptosporidium anserinum sp. n., illus.
Proctor, S. J.; and Kemp, R. L., 1974, J. Protozool., v. 21 (5), 664-666
[lapsus p. 666 as C. anserium]
Anser anser (large intestine): Ames, Iowa, USA

Cryptosporidium anserium [lapsus for C. anserinum sp. n.]
Proctor, S. J.; and Kemp, R. L., 1974, J. Protozool., v. 21 (5), 664-666

Cryptosporidium vulpis Wetzel 1938
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
probably as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Cyathodinium indiae n. sp., illus.
Bhaskar Rao, T. S.; and Bhaskar Rao, T., 1976, Current Sc., Bangalore, v. 45 (8), 299-300 [Letter]
Cavia cutleri: Hyderabad region, India

Cyclospora
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Cystobia Mingazzini, 1891 in part
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora Schneider, 1875

Cystobia Mingazzini, 1891 in part
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora Schneider, 1875

Cystobia arenicolae (Cunningham, 1907) Dogiel, 1909
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora arenicolae (Cunningham, 1907) Tregouboff, 1918

Cystobia chiridotae Dogiel, 1906
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora chiridotae (Dogiel, 1906) Goodrich, 1925

Cystobia eupyrgi Dogiel, 1909
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
nomen nudum

Cystobia grassei Changeux, 1961
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora grassei (Changeux, 1961) new combination

Cystobia holothuriae (Schneider, 1858) Mingazzini, 1891
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora holothuriae (Schneider, 1858) Tregouboff, 1918

Cystobia irregularis (Minchin, 1893) Labbe, 1899
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora irregularis (Minchin, 1893) Tregouboff, 1918

Cystobia minchinii Woodcock, 1904
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora minchini (Woodcock, 1904) Tregouboff, 1918

Cystobia schneideri Mingazzini, 1891
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora schneideri (Mingazzini, 1891) new combination

Cystobia stichopi Lutzen, 1967
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora stichopi (Lutzen, 1967) new combination

Cystobia testiculi Tregouboff, 1916
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora testiculi (Tregouboff, 1916) Tregouboff, 1918

Cystoisospora gen. n.
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Sarcocystidae, Toxoplasmatinae
tod: C. felis Wenyon 1923 [n. comb.]
diagnosis, life cycle pattern

Cystoisospora canis (Nemeseri 1959) [n. comb.]
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628

Cystoisospora felis Wenyon 1923 [n. comb.] (tod)
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628

Cystoisospora ohioensis (Dubey 1975) [n. comb.]
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628

Cystoisospora rivolta (Grassi 1879) [n. comb.]
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628

Cystoisospora vulpina (Nieschulz and Bos 1933) [n. comb.]
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628

Cytamoeba [sp.]
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Batrachoseps attenuatus
Aneides lugubrus
Ambystoma tigrinum
(blood of all): all from central California

Cytamoeba [sp.]
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses
Atelopus various: Panama
Eurycea lucifuga: U.S.A.
Leptodactylus fitzingeri: Panama
Leptodactylus spp.: Venezuela
Phrynohyas venulosa: Venezuela
Rana cancrivora: Philippines
R. japonica: Japan
R. rugosa: Japan
Takydromus tachydromoides: Japan

Cytamoeba sp., illus.
Sullivan, J. S.; and Sullivan, J. J., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (1), 126-128
Bufo melanostictus
Rana cancrivora
R. erythraea
R. limnocharis
R. macrodon
R. hosei
Leptobrachium hasselti
(blood of all): all from Peninsular Malaysia

Cytamoeba bacterifera Labbe, 1894
de Sousa, M. A.; and Freire, E. G., 1973, J. Protozool., v. 20 (4), 530
Cytamoeba bacterifera, cytochemistry, failure to transmit from Leptodactylus ocellatus to Bufo crucifer, development

Cytamoeba bacterifera Labbe, 1894
de Sousa, M. A.; and Weigl, D. R., 1975, J. Protozool., v. 22 (3), 36A-37A [Abstract]
Cytamoeba bacterifera, ultrastructure establishes no relationship with Protozoa, may be related to bacteria

Cytauxzoon
Irvin, A. D.; et al., 1972, Vet. Rec., v. 91 (21), 513, 514-516, 517
theilerial piroplasms acquired by Rhipicephalus appendiculatus and R. simus (salivary glands) (exper.) fed on naturally infected Taurotragus oryx pattersonianus were not transmitted to cattle, role of eland in epidemiolgy of cattle theilerioses is questionable: Kenya

Cytauxzoon sp. or Theileria tragelaphi, illus.
Bigalke, R. D.; Keep, M. E.; and Schoeman, J. H., 1972, Onderstepoort J. Vet. Research, v. 39 (4), 225-227
Tragelaphus scriptus (blood): Hluhluwe Game Reserve, Natal, Zululand

Cytauxzoon and/or Theileria [spp.]
Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
blood parasites of wild Bovidae, incidence, epizootiological importance
Syncerus caffer
Connochaetes taurinus
Damaliscus lunatus
Kobus leche
Aepyceros melampus
Hippotragus niger
Tragelaphus strepsiceros
Taurotragus oryx
(blood of all): all from Ngamiland, Botswana

Cytauxzoon spp.
Loehr, K.-F.; and Meyer, H., 1974, Tropenmed. u. Parasitol., v. 25 (3), 288-292
Theileria parva, T. mutans, capillary-tube agglutination and indirect fluorescent antibody tests on sera from Masai cattle (reacted with both species) and 5 game animals (reacted only with T. parva), intra erythrocytic "theileria organisms" found in all game animals but could not be needle-transmitted to splenectomized calves, "one theilerial species which was clearly of T. parva and not of T. mutans antigenicity", "cannot, however, be excluded that these piroplasms may not belong to members of the genus Cytauxzoon"
Connochaetes taurinus
Alcelaphus buselaphus
Aepyceros melampus
Gazella grantii
G. thomsonii
all from Kapiti Plains near Konza, Kenya

Cytauxzoon sp., illus.
McCully, R. M.; Keep, M. E.; and Basson, P. A., 1970, Onderstepoort J. Vet. Research, v. 37 (1), 7-9
Cytauxzoon sp. in Giraffa camelopardalis, both erythrocytic and histiotropic parasites found, pathology, case report: Ubizane game ranch, Zululand (imported from South West Africa)

Cytauxzoon sp.
Stagg, D. A.; et al., 1976, Research Vet. Sc., v. 20 (2), 122-126
theilerial parasite or parasites (Theileria and/or Cytauxzoon), isolation from Taurotragus oryx in Kenya, establishment of five infected cell lines

Cytauxzoon-like organisms, illus.
Wagner, J. E., 1976, J. Am. Vet. Med. Ass., v. 168 (7), 585-588
Cytauxzoon-like organisms, cats, case histories, diagnosis, pathology: southwestern Missouri

Cytauxzoon [sp.]
Wightman, S. R.; Kier, A. B.; and Wagner, J. E., 1977, Feline Pract., v. 7 (3), 23-26
Cytauxzoon, cats (blood), clinical features, pathology, diagnosis, morphology compared to Hemobartonella felis

Cytauxzoon sp., illus.
Wilson, D. E.; and Hirst, S. M., 1977, Wildlife Monogr. (54), Suppl., 3-111
Cytauxzoon sp., intrauterine transfer most probable in Hippotragus niger
Hippotragus niger
H. equinus
all from Transvaal; Rhodesia

Cytauxzoon sp.
Young, E.; et al., 1973, Research J. National Parks Republic South Africa (16), 195-198
Antidorcas marsupialis (blood): Mountain Zebra National Park near Cradock, Cape Province

Cytauxzoonosis-like disease, illus.
Bendele, R. A.; Schwartz, W. L.; and Jones, L. P., 1976, Southwest. Vet., v. 29 (3), 244-246
cytauxzoonosis-like disease, cats (lungs, brain, liver, lymph nodes), case reports, histopathology: southeastern and central Texas

Cytomorpha diazonae Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria diazonae (Mingazzini, 1891) Labbe, 1899

Dactylophorus
Ormieres, R.; and Marques, A., 1976, J. Protozool., v. 23 (4), 15A-16A [Abstract]
Echinomera, Grebnickiella, Dactylophorus, mode of fixation to host

Dactylosoma sp.
Landau, I.; Colley, F. C.; and Yap, L. F., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 144 [Demonstration]
Rana limnocharis (red blood cells): Bukit Lanjan, Selangor

Dactylosoma sp.
Le Bail, O.; and Landau, I., [1975], Ann. Parasitol., v. 49 (6), 1974, 663-668
Bufo marinus (blood): Maripassoula (Guyane francaise)

Dactylosoma spp.
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Rana erythraea
R. hosei
R. limnocharis
R. cancrivora
R. macrodon
all from Malayan Peninsula

Dactylosoma ranarum (Kruse, 1890), illus.
Miyata, A., 1976, Nettai Igaku (Trop. Med.), v. 18 (3), 135-141
Dactylosoma ranarum, survey, incidence in tadpoles and frogs, parasite development and morphology
Rana rugosa (blood): Nagasaki City vicinity

Dactylosomes
Saoud, M. F. A., 1976, Rev. Zool. Africaine, v. 90 (2), 313-322
Chrysichthys auratus (blood): Sudan

Davisia
Moser, M.; and Noble, E. R., 1975, J. Parasitol., v. 61 (4), 691-694
species containing cavity in lateral appendages distinguished from those with "solid" appendages

Davisia anoplopoma sp. n., illus.
Moser, M.; and Noble, E. R., 1975, J. Parasitol., v. 61 (4), 691-694
Anoplopoma fimbria (urinary bladder and kidney tubules): off southern and central California

Davisia anoplopoma Moser and Noble, 1975
Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
Anoplopoma fimbria (urinary bladder): off central California

Davisia coryphaenoidia Yoshino & Noble, 1973
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides acrolepis: eastern Pacific

Davisia coryphaenoidia Yoshino and Noble, 1973
Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
Coryphaenoides acrolepis (urinary bladder/kidney): off northern Mexico, southern and northern California, Oregon, B. C.
C. cinereus (urinary bladder): Bering Sea
C. pectoralis (urinary bladder/kidney): off northern California, Washington

Davisia newfoundlandia Yoshino and Noble, 1973
Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
Macrourus berglax (urinary bladder/kidney): off Newfoundland

Davisia pectoralis sp. n., illus.
Moser, M.; and Noble, E. R., 1975, J. Parasitol., v. 61 (4), 691-694
Coryphaenoides pectoralis (urinary bladder and kidney tubules): off northern California

Davisia pectoralis Moser and Noble, 1975
Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
Coryphaenoides acrolepis (urinary bladder/kidney)
C. pectoralis (urinary bladder)
all from off northern California

Davisia reginae sp. n., illus.
Love, M. S.; and Moser, M., 1976, J. Parasitol., v. 62 (6), 982-983
Sebastes serranoides (urinary bladders): off central California
S. maliger (urinary bladders): northern California
Embiotoca lateralis (urinary bladders): off central California
Damalichthys vacca (urinary bladders): off central California

Defretinella Hennere, 1966
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Grelliidae fam. nov.
diagnosis
includes: D. eulaliae Hennere, 1966 (type species)

Dehornia n. gen.
Porchet Hennere, E., 1972, Protistologica, v. 8 (2), 245-255
probably a Coccidia
mt: Dehornia sthenelais n. sp.

Dehornia sthenelais n. gen., sp. n. (mt), illus.
Porchet Hennere, E., 1972, Protistologica, v. 8 (2), 245-255
sporogenesis, light and electron microscopy
Sthenelais boa (coelomique): region de Roscoff

Dehornia sthenelais
Vivier, E.; and Porchet, E., 1976, J. Protozool., v. 23 (4), 22A
Aggregata eberthi, Hepatozoon domerguei, Dehornia sthenelais, crystalline inclusions

Delphyella de Puytorac, 1969
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Jirovecellinae n. s. fam.

Delphyella filum (Claparede, 1860) de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Clitellio arenarius

Dermocystidium spp.
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Salmo trutta: Loch Leven, Scotland

Dermocystidium marinum, illus.
Perkins, F. O., 1976, J. Parasitol., v. 62 (6), 959-974
Dermocystidium marinum zoospores, conoid and other sporozoan-like organelles, fine structure, results indicate D. marinum is protozoan in subphylum Apicomplexa and is most closely related to coccidian Sporozoasida

Dermocystidium marinum Mackin, Owen and Collier
Perkins, F. O., 1976, J. Protozool., v. 23 (2), 10A [Abstract]
Dermocystidium marinum, ultrastructure, considered to be member of Apicomplexa, closely related to coccidian Sporozoa

Desmophrya Raabe, 1933
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Radiophryinae nov. comb.

Desmophrya contorta Raabe, 1933, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Pisidium casertanum

Dicoelophrya n. gen.
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Hoplitophryidae, Radiophryinae; no type designated

Dicoelophrya de Puytorac et Dragesco, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Metaracoelophryinae n. sous-fm.

Dicoelophrya almae sp. nov., illus.
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Alma emini (tube digestif): Gabon

Dicoelophrya almae de Puytorac et Dragesco, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Dicoelophrya calliste n.g., n.sp., illus.
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Alma emini (tube digestif): Gabon

Dicoelophrya calliste de Puytorac et Dragesco, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Dicontophrya n. gen.
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Hoplitophryidae, Contophryinae n.subfam.; tod: D. grassei n.g., n. sp.

Dicontophrya de Puytorac et Dragesco, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Contophryidae n. fam.

Dicontophrya grassei n.g., n. sp., illus. (tod)
de Puytorac, P.; and Dragesco, J., 1969, Biol. Gabon., v. 5 (1), 5-27
Alma emini: Gabon

Dicontophrya grassei de Puytorac et Dragesco, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Dicontophrya rara (Katashima, 1952 [sic]) de Puytorac, 1970
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Radiophrya rara, Katashima, 1951[sic]
Pheretima yamada

Didymophyes gigantea, illus.
Hildebrand, H. F., 1976, Ztschr. Parasitenk., v. 49 (3), 193-215
Didymophyes gigantea, electron microscopy of epimerite and protomerite, relationship between parasite surface and host cell, periparasitic space, modified structure of host cell
Oryctes nasicornis (Darm): Prafrance (Cevennen) and Lesquin (Departement Nord), Frankreich

Didymophyes gigantea Stein, illus.
Hildebrand, H. F.; and Vinckier, D., 1975, J. Protozool., v. 22 (2), 200-213
Didymophyes gigantea, morphology during syzygy and growth and development of trophozoite, locomotion
Oryctes nasicornis (intestin moyen): Prafrance, Cevennes, France and Lesquin, Nord, France

Didymophyes onthophagi Foerster, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Onthophagus fracticornis (gastric caeca): Poland

Didymophyes paradoxa Stein, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Geotrupes stercorarius
G. stercorosus
G. vernalis
(intestine of all): all from Poland

Didymophyes rara Leger, 1892
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Didymophyes paradoxa Stein

Dientamoeba Jepps & Dobell
Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
Monocercomonadidae Kirby, emend.; Dientamoebinae Grasse, emend.
diagnosis

Dientamoeba fragilis Jepps & Dobell, illus.
Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
Dientamoeba fragilis, diagnosis and redescription; light and electron microscopic observations of binucleate stages

Dientamoeba fragilis
de Carneri, I.; Grassi, L.; and Trane, F., 1972, Parassitologia, v. 14 (1), 113
Entamoeba histolytica, E. coli, Dientamoeba fragilis, prevalence in an unselected hospital population: Milano

Dientamoeba fragilis
Chwirot, E.; and Kasprzak, W., 1975, Polski Tygod. Lekar., v. 30 (6), 261-263
Entamoeba histolytica, E. hartmanni, Dientamoeba fragilis, humans, clinical trials to evaluate effectiveness of metronidazole, good results with minimal side effects

Dientamoeba fragilis
Desser, S. S.; and Yang, Y. J., 1976, Canad. Med. Ass. J., v. 114 (4), 290-291
Dientamoeba fragilis infection in child resulting in severe intestinal symptoms, diagnosis only after multiple fecal examinations, diphetarsone treatment; possible role of Enterobius vermicularis as vector of Dientamoeba fragilis in human infections

Dientamoeba fragilis
Dwyer, D. M., 1974, J. Protozool., v. 21 (1), 139-145
Trichomonas gallinae, Histomonas meleagridis, Dientamoeba fragilis, Entamoeba invadens, E. histolytica, antigenic relationships analyzed by immunoelectrophoretic techniques

Dientamoeba fragilis
Markell, E. K.; and Quinn, P. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1139-1142
comparison of immediate polyvinyl alcohol fixation with delayed Schaudinn's fixation for demonstrating protozoa in feces, concluded that immediate fixation results in preservation of larger number of organisms and recommends routine use prior to transportation to laboratory

Dientamoeba fragilis
Ockert, G.; and Schmidt, T., 1976, J. Hyg., Epidemiol., Microbiol. and Immunol., v. 20 (1), 76-81
Enterobius vermicularis, confirmation as vector of Dientamoeba fragilis

Dientamoeba fragilis
Subiabre, V., 1969, Bol. Chileno Parasitol., v. 24 (3-4), 166-167
Dientamoeba fragilis, demonstration in human feces using polyvinilic alcohol or Teleman's concentration method: Chile

Dientamoeba fragilis
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Dientamoeba fragilis, illus.
Yang, J.; and Scholten, T. H., 1976, Am. J. Clin. Path., v. 65 (5), 715-718
celestine blue B stain given as alternative to hematoxylin in routine staining of fecal smears for Protozoa, comparative laboratory trials

Dientamoeba fragilis
Yang, J.; and Scholten, T., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 16-22
Dientamoeba fragilis in humans, extensive epidemiologic survey, pathology, fecal examination diagnostic methods, periodicity, frequent occurrence in presence of Enterobius vermicularis suggests possibility of helminths as vectors: Toronto, Canada

Dientamoebinae Grasse, emend. [n. rank]
Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
Trichomonadida Kirby, emend.; Monocercomonadidae Kirby, emend.
diagnosis; includes: Dientamoeba Jepps & Dobell

Diplauxis Vivier, Ormieres & Tuzet, 1964
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Diplauxis hatti Vivier, Ormieres & Tuzet, 1964
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Diplauxis schreveli Porchet-Hennere & Fischer, 1973
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Diplocystidae Bhatia, 1930
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
diagnosis; checklist of genera and species of 6 aseptate gregarine families

Diplocystis Kunstler, 1887
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Diplocystidae
diagnosis; checklist of species

Diplocystis sp. Lisova, 1962
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Ascocystis mackiei (Shortt and Swaminath, 1927) Ormieres, 1965(?)

Diplocystis sp. of Lisova (1962)
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
? as syn. of Ascocystis mackiei (Shortt & Swaminath, 1927) Ormieres, 1965

Diplocystis phryganeae Berg von Emme, 1912
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Faucispora phryganeae (Berg von Emme, 1912) Baudoin, 1967

Diplodina Woodcock, 1906
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora Schneider, 1875

Diplodina gonadipertha Djakonov, 1923
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora gonadipertha (Djakonov, 1923) new combination

Diplodina irregularis (Minchin, 1893) Woodcock, 1904
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora irregularis (Minchin, 1893) Tregouboff, 1918

Diplodina minchinii (Woodcock, 1904) Woodcock, 1906
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora minchini (Woodcock, 1904) Tregouboff, 1918

Diplomonadida Wenyon, 1926, emend.
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
diagnosis; includes: Enteromonadina subord. nov.; Diplomonadina Wenyon, emend.

Diplomonadina Wenyon, emend.
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Diplomonadida
diagnosis; includes: Hexamita (type genus); Trepomonas; Trigonomonas; Spironucleus; Octomitus; Giardia

Diplospora bigemina von Wasielewski 1904 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora felis Wenyon 1923

Diplospora bigemina von Wasielewski 1904 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora rivolta (Grassi 1879) Wenyon 1923

Diplospora bigemina Wasielewski, 1904 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora felis (Wasielewski, 1904) Wenyon, 1923

Diplospora bigemina Wasielewski, 1904 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora rivolta (Grassi, 1879) Wenyon, 1923

Diplospora lacazii Labbe, 1893 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora lacazei (Labbe, 1893)

Diplospora lacazii Labbe, 1893 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Diplospora rivoltae Labbe, 1893 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora lacazei (Labbe, 1893)

Diplospora rivoltae Labbe, 1893 [pro parte?]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Doliocystis rhyncoboli Crawley, 1903
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ulivina rhyncoboli (Crawley, 1903) Kamm, 1922

Dorisiella
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Drepanidium lacertarum Danilewsky, 1886
Svahn, K., 1975, Norwegian J. Zool., v. 23 (4), 277-295
as syn. of Karyolysus lacertae Danilewsky, 1886

Duboscqia coptotermi sp. n., illus.
Kalavati, C.; and Narasimhamurti, C. C., 1976, J. Parasitol., v. 62 (2), 323-324
Coptotermes heimi (epithelial cells of midgut): Andhra University Campus, Waltair, India

Duboscqia penetrans Thorne, 1940, illus.
Mankau, R., 1975, J. Protozool., v. 22 (1), 31-34
incorrectly classified as microsporidan protozoan, possible affinities to Bacillus

Dubosquia penetrans, illus.
Morone De Lucia, M. R.; and Grimaldi De Zio, S., 1973, Nematol. Mediterran., v. 1 (1), 66-68
Xiphinema mediterraneum (gonads)

Dubosquia. See Duboscqia.

Durchoniella de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Durchoniellinae n. s. fam.

Durchoniella brasili (Leger et Duboscq, part. 1894) de Puytorac, 1954, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Audouinia tentaculata

Durchoniella cirratuli de Puytorac, 1954, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Cirratulus cirratus

Durchoniella dodecaceriae de Puytorac et Schrevel, 1965
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Dodecaceria caulleryi
D. concharum

Durchoniella legeri duboscqui (Leger et Duboscq part., 1894) de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Audouinia tentaculata

Durchoniellinae n. s. fam.
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam.
includes: Durchoniella; Hovasseiella

East Coast Fever
Irvin, A. D.; et al., 1972, Vet. Rec., v. 91 (21), 513, 514-516, 517
theilerial piroplasms acquired by Rhipicephalus appendiculatus and R. simus (salivary glands) (exper.) fed on naturally infected Taurotragus oryx pattersonianus were not transmitted to cattle, role of eland in epidemiolgy of cattle theilerioses is questionable: Kenya

Echinomera
Ormieres, R.; and Marques, A., 1976, J. Protozool., v. 23 (4), 15A-16A [Abstract]
Echinomera, Grebnickiella, Dactylophorus, mode of fixation to host

Echinomera hispida (Schneider), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Lithobius forficatus
L. calcaratus
(intestine of all): all from Poland

Echinomera hispida, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Lithobius forficatus

Echinomera horrida
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Lithobius mutabilis

Echinomera leptoiuli sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Leptoiulus proximus (intestine): Siemianice, Poland

Echiurocystis bullis (Noble, 1938) Levine, 1977
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Enterocystis bullis Noble, 1938

Echiurocystis greeffi (Noble, 1938) Levine, 1977
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Enterocystis greeffi Noble, 1938

Echiurocystis yumushii (Iitsuka, 1933) Levine, 1977
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
synonymy

Eimeria
Cuckler, A. C., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 371-397
coccidiosis and histomoniasis in avian hosts, host-parasite relationships, immunity, review

Eimeria
D'Haese, J.; Mehlhorn, H.; and Peters, W., 1976, Ztschr. Parasitenk., v. 50 (2), 198-199
coccidia, negative staining to show characteristic structures

Eimeria
Duszynski, D. W.; and Speer, C. A., 1976, Ztschr. Parasitenk., v. 48 (3-4), 191-197
Isospora arctopitheci, I. bigemina, sporozoite excystation, lack of Stieda body, comparison with other Isospora, Eimeria and Sarcocystis species

Eimeria
Huchzermeyer, F. W., 1976, J. South African Vet. Ass., v. 47 (4), 253-254
Eimeria, chickens, immunization by vaccination with low dose of Eimeria and controlled exposure, method of choice in coccidiosis control, brief review: Rhodesia

Eimeria
Joyner, L. P., 1963, State Vet. J. (54), v. 18, 214-219
domestic fowl, life history, pathology, species incidence, preventive chemotherapy, brief review

Eimeria
Kozar, Z., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 871-912
toxoplasmosis and coccidiosis in mammalian hosts, immunology, review

Eimeria
Long, P. L., 1971, Symposia Brit. Soc. Parasitol., v. 9, 65-75
maintenance of Eimeria and Histomonas in vivo, extensive review: maintenance of Eimeria (purification of species, exper. infections in chickens, factors affecting susceptibility of chickens and chicken embryos to infection; freeze-preservation); maintenance of Histomonas meleagridis (infection with ground-up infected tissues; use of in vitro cultures to obtain in vivo infections; use of embryonated Heterakis gallinarum ova to induce histomoniasis in vivo)

Eimeria
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Eimeria
Mechow, A., 1975, Avian Path., v. 4 (1), 83-85
improved technique for separating oocysts from large amounts of feces

Eimeria
Ryley, J. F., 1975, Symposia Brit. Soc. Parasitol., v. 13, 43-58
coccidia of domestic fowl, pathology, extensive review (effects on host cell, on structure and function of adjacent cells and tissues, and on extraintestinal tissues and systems)

Eimeria
Soltys, M. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 309-322
immunization against protozoan diseases of animals, review

Eimeria
Speer, C. A.; and Duszynski, D. W., 1974, J. Protozool., v. 21 (3), 424 [Abstract]
relationship between sporocyst structure and process of excystation in coccidia

Eimeria Schneider, 1875
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
Eimeriidae fam. emend., Eimeriinae subfam. emend.
definition, key

Eimeria
Trigg, P. I.; and Gutteridge, W. E., 1977, Symposia Brit. Soc. Parasitol., v. 15, 57-81
parasitic Protozoa, morphological, biochemical, and physiological changes occurring during life cycles, symposium presentation

Eimeria
Williams, J. C.; and Knox, J. W., 1976, Am. J. Vet. Research, v. 37 (4), 453-464
failure of stocker cattle to achieve projected weight gains at high stocking rates on Coastal bermudagrass pastures even with supplemental feeding and anthelmintic control of parasitism

Eimeria [sp.]
Bali, H. S.; and Kalra, I. S., 1975, J. Research, Punjab Agric. Univ., v. 12 (3), 313-316
white leghorn birds: Punjab State, India

Eimeria sp.
Bejsovec, J., 1977, J. Protozool., v. 24 (4), 48A [Abstract]
persistence of Coccidia despite changing land use and decreasing numbers of host animals
Perdix perdix perdix: Bohemia

Eimerian species
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Lophura leucomelana
Francolinus francolinus
Pycnonotus jocosus
(feces of all): all from Delhi Zoo

Eimeria sp.
Bisseru, B.; and Lim, K. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 412 [Demonstration]
Corvus splendens protegatus (intestine): Klang, Selangor, Malaysia

Eimeria sp.
Brittain, P. C.; and Voth, D. R., 1975, J. Wildlife Dis., v. 11 (2), 269-271
Lepus californicus melanotis (intestinal areas): Rocky Mountain Arsenal near Denver, Colorado

Eimeria spp.
Christensson, D.; and Rehbinder, C., 1975, Nord. Vet.-Med., v. 27 (10), 496-498
gastrointestinal parasites of reindeer calves, none found in first month of life, increasing infection with age: Norrbotten

Eimeria spp.
Colley, F. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 85 [Demonstration]
Callosciurus prevostii
Sundasciurus hippurus
Petaurista petaurista
Aeromys tephromeles
Rattus fulvescens
R. sabanus
R. edwardsi
R. tiomanicus
Trichys lipura
Tupaia glis ferruginea
(feces of all): all from Malaysia

Eimeria spp.
Crompton, D. W. T., 1976, Proc. 11. Poultry Sc. Symp., 193-245
gastrointestinal parasites, Gallus gallus, review; Eimeria spp., extensive study of pathologic changes in host alimentary tract

Eimeria spp., "thin-walled coccidia"
Davidson, W. R., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 211-217
epizootiologic and pathologic study of endoparasites of selected populations of gray squirrels
Sciurus carolinensis (small intestine): southeastern United States

Eimeria sp.
Davronov, O., 1974, Uzbek. Biol. Zhurnal (3), 51-52
Eimeria sp., potential for biological control of Mus musculus (exper.)

Eimeria sp.
Fameree, L.; and Cotteleer, C., 1976, J. Protozool., v. 23 (4), 10A [Abstract]
coccidiosis, cats, prevalence: Belgium

Eimeria sp.
Faust, B. S.; and Pappas, P. W., 1977, J. Zoo Animal Med., v. 8 (1), 18-23
Anser anser domesticus
Columba livia
Phoenicopterus chiliensis
P. ruber
Sturnus vulgaris
(feces of all): all from Columbus (Ohio) Zoo

Eimeria sp.
Foreyt, W. J.; and Todd, A. C., 1976, J. Parasitol., v. 62 (3), 496
prevalence survey in domestic mink: mink ranches, Wisconsin

Eimeria sp.
Foreyt, W. J.; Todd, A. C.; and Hartsough, G. R., 1977, Am. J. Vet. Research, v. 38 (3), 391-394
Isospora laidlawi, Eimeria vison and Eimeria sp. in Mustela vison (exper.), anticoccidial activity of eight compounds

Eimeria spp.
Forrester, D. J.; et al., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 55-59
Grus canadensis tabida (feces): Florida

Eimeria sp.
Forrester, D. J.; Bush, A. O.; and Williams, L. E., jr., 1975, J. Parasitol., v. 61 (3), 547-548
Grus canadensis pratensis (feces): Florida

Eimeria [spp.]
Kutzer, E.; and Frey, H., 1976, Ztschr. Parasitenk., v. 50 (2), 213-214
Lepus europaeus

Eimeria sp.
Landau, I.; et al., 1975, Compt. Rend. Acad. Sc., Paris, v. 281, s. D, Sc. Nat. (22), 1721-1723
Eimeria sp., crustacean infected with oocysts from fish feces, freed sporozoites remain as latent stage in intestinal wall; possible alimentary pyramid concentrating infective forms in aquatic environment
Gymnothorax moringa "murene" (foie, intestin): Petit Bourg, Guadeloupe
Mysidacea (intestinal wall) (exper.)

Eimeria spp.
Long, P. L.; and Millard, B. J., 1977, Avian Path., v. 6 (3), 227-233
coccidiosis, turkeys treated with sulphaquinoxaline and monensin, evaluation of infection by number of oocysts in litter, good control and improved weight gain with monensin

Eimeria sp.
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Rattus sabanus
R. whiteheadi
R. rajah pellax
R. muelleri
Echinosorex gymnurus
Nycticebus coucang
Sundasciurus tenuis
all from Bukit Lanjan Forest Reserve, West Malaysia

Eimeria sp., illus.
Munson, D. A., 1974, J. Wildlife Dis., v. 10 (3), 256-262
Liparis atlanticus (intestine): Rye, New Hampshire

Eimeria spp.
Norton, C. C., 1976, Folia Vet. Latina, v. 6 (3), 218-238
pheasant coccidia, description, review of literature

Eimeria sp., illus.
Ovezmukhammedov, A.; and Berdyev, A. S., 1975, Izvest. Akad. Nauk Turkmensk. SSR, s. Biol. Nauk (6), 55-59
brief description
Felis margarita: Turkmenistan

Eimeria C
Peters, W.; Tennant, L.; and McDermott, S. N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 8-10 [Demonstration]
lions: Knowsley Safari Park, Prescot, Lancashire

Eimeria sp.
Prestwood, A. K.; Pursglove, S. R.; and Hayes, F. A., 1976, J. Wildlife Dis., v. 12 (3), 380-385
survey of parasites of Odocoileus virginianus and Ovis aries on common range, deer unlikely reservoir host for sheep parasites
Ovis aries: Hardy County, West Virginia

Eimeria sp.
Schulte, J. W.; Klimstra, W. D.; and Dyer, W. G., 1976, J. Wildlife Management, v. 40 (3), 579-581
Odocoileus virginianus clavium (feces): Big Pine Key, Florida

Eimeria sp.
Volf, K.; and Volfova, M., 1974, Veterinarstvi, v. 24 (3), 125-126
srnci zvere
jeleni zvere
all from Trebic District

Eimeria sp. similar to E. iowaensis
Wacha, R. S.; and Christiansen, J. L., 1975, J. Protozool., v. 22 (4), 453-454
Natrix sipedon sipedon: Louisa Co., Iowa

Eimeria sp., illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
description, prevalence
Graptemys geographica (feces): Iowa

Eimeria sp.
Wheat, B. E., 1975, J. Alabama Acad. Sc., v. 46 (3-4), 97 [Abstract]
Pipistrellus subflavus (cecum): Clarke County, Alabama

Eimeria [sp.]
Wobeser, G., 1974, J. Wildlife Dis., v. 10 (3), 249-255
Eimeria [sp.] similar to E. truncata causing renal coccidiosis in Anas platyrhynchos and A. acuta, pathologic changes, ureteral trematodes and cestodes also present in A. platyrhynchos: Saskatoon, Saskatchewan

Eimeria acerinae sp. n., illus.
Pellerdy, L.; and Molnar, K., 1971, Parasitol. Hungar., v. 4, 121-124
Acerina cernua (intestinal scrapings): Hungarian reaches of the Danube River

Eimeria acervulina
Augustin, H.; Senaud, J.; and Doens-Juteau, O., 1976, J. Protozool., v. 23 (4), 3A [Abstract]
Eimeria acervulina, microgametogenesis

Eimeria acervulina, illus.
Babu, K. S. J.; Seshadri, S. J.; and Mohiyuddeen, S., 1976, Indian Vet. J., v. 53 (1), 47-54
coccidiosis, poultry, field cases, diagnosis, pathology, relationship of infection to age and sex of host, mixed infection with E. necatrix

Eimeria acervulina
Balauca, N., 1976, Arch. Exper. Vet.-Med., v. 30 (6), 903-912
Eimeria acervulina, E. necatrix, and E. mitis in cage-kept hens (exper.), mixed infections with Clostridium perfringens causing necrotic enteritis, little pathologic changes from mono-infections

E[imeria] acervulina
Balauca, N.; et al., 1976, Arch. Exper. Vet.-Med., v. 30 (6), 913-923
E[imeria] acervulina, E[imeria] necatrix, and E[imeria] mitis in ground-kept hens (exper.), mixed infections with Clostridium perfringens causing necrotic enteritis, higher mortality in ground-kept hens than cage-kept hens

Eimeria acervulina
Carbo'Baptista, N.; Larbier, M.; and Yvore, P., 1976, Avian Path., v. 5 (3), 187-194
Eimeria acervulina, chicks, decreased intestinal absorption of ^{14}C L-lysine and water accompanied by an increase in tissue water content and in secretion rate of mineral ions (Na$^+$ and K$^+$)

Eimeria acervulina
Chapman, H. D., 1976, Avian Path., v. 5 (4), 283-290
Eimeria tenella, E. acervulina, E. maxima, incidence in broiler house used for commercial drug trials, resistance to various anticoccidials

Eimeria acervulina
Denek, J.; and Strossova, Z., 1977, J. Protozool., v. 24 (2), Suppl., 14A [Abstract]
field strains of Coccidia, development of resistance to clopidol: CSSR

E[imeria] acervulina
Douglas, A. W.; et al., 1977, J. Med. Chem., v. 20 (7), 939-943
4-chlorobenzaldehyde 1-(4-chlorophenyl)-4(1H)-pyridinylidene hydrazone fluorosulfonate, broad-spectrum anticoccidial activity in chickens, action similar to that of robenidine in laboratory trials with E[imeria] tenella and E[imeria] acervulina; robenidine-resistant E[imeria] tenella strain also resistant to new compound

Eimeria acervulina
El-Moukdad, A. R., 1976, Wien. Tierarztl. Monatsschr., v. 63 (12), 399-405
Eimeria necatrix, E. acervulina, E. tenella, chickens, effect of various disinfectants on oocyst development

Eimeria acervulina
El-Moukdad, A. R., 1977, Wien. Tierarztl. Monatsschr., v. 64 (3), 85-91
effect of disinfectants on eggs of Ascaris suum, Toxascaris leonina, small horse strongyles and coccidia oocysts

Eimeria acervulina
Foure, N.; et al., 1974, Bull. Inform. Station Exper. Avicult. Ploufragan (Cotes-du-Nord), v. 14 (3), 51-56
Eimeria tenella, E. necatrix, E. acervulina, Clopidol + methyl-benzoquate, presence of fully sensitive or double resistant strains

Eimeria acervulina
Foure, N.; and Bennejean, G., 1974, Bull. Inform. Station Exper. Avicult. Ploufragan (Cotes-du-Nord), v. 14 (3), 57-60
Eimeria, chickens, Halofuginone, comparison with methyl-benzoquate and Clopidol, good results

Eimeria acervulina
Giordani, G.; Govoni, S.; and Bergami, W., 1977, Avicoltura, Bologna, v. 46 (1), 41-46
Eimeria spp., breeder hens, continuously fed sulphachloropyrazine + diaveridine + ethopabate, no undesirable effects on egg production, values statistically equal for control and coccidiostat fed birds

Eimeria acervulina
Graham, D. W.; et al., 1977, Experientia, v. 33 (10), 1274-1276
Eimeria tenella, E. acervulina, anticoccidial activity of riboflavine antagonists

Eimeria acervulina
Hein, H. E., 1976, Exper. Parasitol., v. 39 (3), 415-421
Eimeria acervulina, E. brunetti, E. maxima, pathogenic effects of single vs. mixed infections with low doses of oocysts in chickens, changes in mean weight and oocyst output

Eimeria acervulina
Hein, H. E., 1976, Exper. Parasitol., v. 40 (2), 250-260
Eimeria acervulina, E. brunetti, E. maxima, E. necatrix, immunogenic potential under precisely defined conditions which provide standards enabling comparisons between species and strains to be made, found that high level of resistance to reinfection was conferred by multiple infections with relatively low doses of oocysts

Eimeria acervulina
Jeffers, T. K., 1977, Poultry Science, v. 56 (5), 1725 [Abstract]
Eimeria tenella, E. maxima, E. acervulina, broiler flocks, no development of monensin resistance following long-term drug exposure

Eimeria acervulina
Joyner, L. P.; et al., 1975, Avian Path., v. 4 (1), 17-33
Eimeria acervulina-infected chickens, amino acid malabsorption and intestinal leakage of plasma proteins, food intake and growth, results suggest that anorexia and protein leakage from gut are major factors in pathogenesis

Eimeria acervulina
Joyner, L. P.; and Norton, C. C., 1976, Parasitology, v. 72 (1), 115-125
Eimeria maxima, E. acervulina, chicks, immunity arising from continuous low-level infections was stronger and/or more enduring than that produced by single inoculations of comparable numbers of oocysts

Eimeria acervulina
Joyner, L. P.; and Norton, C. C., 1977, Parasitology, v. 75 (2), 155-164
Eimeria spp., anticoccidial effects of amprolium alone or with ethopabate, dinitolmide, and monensin with particular reference to oocyst sporulation, significance of effects on gametogony and sporogony

Eimeria acervulina
Karlsson, T.; and Reid, W. M., 1977, Brit. Poultry Sc., v. 18 (4), 497-501
Eimeria acervulina, E. necatrix, E. brunetti, E. tenella, chickens (exper.), immunity challenge techniques used to determine prevalence in Georgia (USA), lesion scores as main criterion to indicate exposure

Eimeria acervulina
Kohls, R. E.; Engle, A. T.; and Butters, H. E., 1972, Avian Dis., v. 16 (4), 907-914
coccidiosis in laying chickens (exper.), continuous buquinolate medication, good protection with increased egg production and egg quality, immune status at termination

Eimeria acervulina
Kolev, V.; et al., 1976, Vet. Med. Nauk., v. 13 (4), 3-12
coccidiosis, incidence in poultry at large poultry dressing combine, birds imported from France and England as possible source of infection, prophylactic measures for better disease control: Ruse, Bulgaria

Eimeria acervulina
Kuil, H.; et al., 1977, Vet. Parasitol., v. 3 (1), 33-40
Eimeria maxima, Eimeria acervulina, chickens, effect of inoculation dose on indirect fluorescent antibody response, difference in immunogenicity between two species substanttiated by difference in IFA response after challenge, reinoculation with Eimeria maxima indicated that birds were immune but single infection with Eimeria acervulina did not result in solid immunity

Eimeria acervulina
Kumar, A.; Pandey, M. D.; and Rawat, J. S., 1976, Indian J. Poultry Sc., v. 11 (1), 11-13
Eimeria acervulina, E. necatrix, mixed infection, chickens (exper.), serum electrolyte changes, decreased percentage of total body water, dehydration

Eimeria acervulina
Kumar, A.; and Rawat, J. S., 1976, Indian J. Animal Sc., v. 45 (3), 1975, 154-156
Eimeria necatrix, E. acervulina, chickens (exper.), effect on serum enzymes, blood glucose and cholesterol

Eimeria acervulina, illus.
Long, P. L.; et al., 1976, Folia Vet. Latina, v. 6 (3), 201-217
Eimeria spp., avian, procedure, key and laboratory methods for identification, pathology, experimental infection procedures, factors affecting development, instructive review

Eimeria acervulina
Long, P. L.; and Millard, B. J., 1976, Parasitology, v. 73 (3), 327-336
Eimeria praecox, E. maxima, E. acervulina, site finding and site specificity in chickens

Eimeria acervulina
Long, P. L.; and Millard, B. J., 1976, Ztschr. Parasitenk., v. 48 (3-4), 287-290
Eimeria spp., immunized chickens, oocysts produced after corticosteroid treatment in some birds, possibly endogenous cycle continues or remains dormant; possible means of diagnosis of occult infection in apparently immune hosts

Eimeria acervulina
Martin Vazquez, A., 1975, An. Inst. Invest. Vet., Madrid, v. 23, 1974-1975, 169-178
Eimeria acervulina, chickens, changes of level of plasmatic ureo-stable alkaline phosphatase

Eimeria acervulina, illus.
Michael, E., 1976, Ztschr. Parasitenk., v. 49 (1), 33-40
Eimeria acervulina, sporozoites within chicken intestinal epithelium macrophages 5-6 days post infection, ultrastructure, macrophage structural changes, possible mechanisms of sporozoites survival and transport to development site

Eimeria acervulina
Miller, B. M.; et al., 1977, Poultry Science, v. 56 (6), 2039-2044
Eimeria spp., chickens, efficacy and tolerance of two 6-amino-9-(substituted benzyl) purines, good results

Eimeria acervulina
Mylari, B. L.; et al., 1977, J. Med. Chem., v. 20 (4), 475-483
Eimeria spp. in chickens, 1-benzyl-6-azauracils as potential new anticoccidials, comparison trials with several experimental and commercial anticoccidials

Eimeria acervulina (Tyzzer, 1929), illus.
Naciri-Bontemps, M., 1976, Ann. Recherches Vet., v. 7 (3), 223-230
Eimeria acervulina cycle, grown in primary culture of chicken kidney cells, inoculation with sporozoites for schizogony, merozoites for gamogony

Eimeria acervulina
Oettel, M.; et al., 1975, Pharmazie, v. 30 (5), 321-324
Eimeria tenella; E. acervulina, drug screening in chickens comparing 1.6-di[1-methylallyl]-dithiocarbamoylhydrazine and meticlorpindol

Eimeria acervulina
Oikawa, H.; et al., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 271-279
Eimeria spp., chickens, survey during 1973 neither seasonal nor regional distribution differences were observed; oocyst detection rate significantly low in chicks up to 30 days old: Japan

Eimeria acervulina
Patterson, D. S. P.; et al., 1975, Avian Path., v. 4 (1), 11-16
Eimeria acervulina-infected chickens, intestinal malabsorption of amino acids, in vitro studies, results generally support view of depressed absorption but demonstration was not as technically straightforward as earlier reports suggested

Eimeria acervulina
Pesti, G. M.; and Combs, G. F., jr., 1976, Poultry Science, v. 55 (6), 2265-2274
Eimeria, site selectivity of various species for infection in particular regions of chick gastrointestinal tract used as tool to study loci of selenium absorption, potentiation of selenium deficiency by intestinal parasitism

Eimeria acervulina
Reid, W. M., 1975, Avian Dis., v. 19 (4), 802-811
Eimeria spp., chicks medicated with roxarsone and zoalene produced higher oocyst counts than unmedicated controls confirming the conclusion that oocyst counts are an unsatisfactory index to anticoccidial activity

Eimeria acervulina
Reid, W. M.; et al., 1976, Poultry Science, v. 55 (4), 1436-1441
Eimeria tenella, E. acervulina, chickens, high environmental temperatures, reduction of anticoccidial feed intake, results suggest decreases in feed and anticoccidial intake during heat-stress could seldom cause coccidiosis breaks under field conditions

Eimeria acervulina
Reid, W. M.; et al., 1977, Poultry Science, v. 56 (1), 66-71
coccidiosis, chicks, monensin, development of immunity dependent upon drug level, treatment time, and oocyst exposure

Eimeria acervulina
Rollinson, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 436-437 [Demonstration]
Eimeria spp., attempted differentiation of chicken coccidia by electrophoretic variation of enzymes

Eimeria acervulina
Rollinson, D., 1976, Parasitology, v. 73 (2), iv [Abstract]
Eimeria mivati, E. acervulina, electrophoretic forms of enzymes, high frequency of interspecific variation, crosses produce strains characterized by glucose phosphate isomerase-2 (characteristic of E. acervulina) and ability to passage in eggs (E. mivati), results support Long's view that E. mivati Houghton isolate belongs to species E. acervulina

Eimeria acervulina
Rose, M. E., 1976, Vet. Rec., v. 98 (24), 481-484
poultry coccidiosis, immunity and prospects for immunoprophylaxis, review

Eimeria acervulina
Ruff, M. D.; and Britton, W. M., 1976, Poultry Science, v. 55 (5), 1712-1716
Eimeria acervulina, E. tenella, white Leghorn hens, severe infection, lower egg production, reduced yolk color

Eimeria acervulina
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria acervulina
Ruff, M. D.; Witlock, D. R.; and Smith, R. R., 1976, Exper. Parasitol., v. 39 (2), 244-251
comparison of effects of Eimeria tenella and E. acervulina infection on methionine absorption by avian intestine: importance of gut region infected; specific kinetic parameters affected; effect of intestinal pH; morphological changes in intestinal mucosa which might account for transport changes

Eimeria acervulina
Ryley, J. F.; and Hardman, L., 1977, Parasitology, v. 75 (2), xv [Abstract]
Eimeria spp., effect of vitamin K-deficient diets on infection and in particular on effectiveness of anti-coccidial drugs, concluded that use of deficient diet in screening and evaluating drugs is justified

Eimeria acervulina
Ryley, J. F.; and Hardman, L., 1977, J. Protozool., v. 24 (4), 43A [Abstract]
Eimeria mivati, E. acervulina, speciation studies using drug resistance as genetic marker, immunization experiments indicate little if any cross-immunity

Eimeria acervulina
Shirley, M. W., 1977, J. Protozool., v. 24 (4), 43A-44A [Abstract]
establishment of infections in chickens and/or eggs by single sporozoites or single sporocysts

Eimeria acervulina Tyzzer, 1929
Singh, C. V.; Joshi, H. C.; and Shah, H. L., 1976, Pantnagar J. Research, v. 1 (1), 63-66
Eimeria spp., White Leghorn chicks, changes in blood biochemistry, possible aid in diagnosis

Eimeria acervulina
Sykes, A. H., 1970, Proc. Nutrition Soc., v. 29 (1), 16 A
Eimeria acervulina, infection in chicks (exper.) resulted in reduction of weight gain, some actual weight loss and impaired utilization of absorbed energy

Eimeria acervulina
Tarozzi, F., 1974, Folia Vet. Latina, v. 4 (2), 275-286
strain of Eimeria acervulina resistant to amprolium + ethopabate and to sulphachlorpyrazine, efficacy of sulphachlorpyrazine + diaveridine + ethopabate, chickens

Eimeria acervulina
Turk, D. E., 1977, Poultry Science, v. 56 (5), 1763 [Abstract]
Eimeria spp., chicks, iron absorption, decreased by some species, increased during recovery period

Eimeria acervulina
Weppelman, R. M.; et al., 1977, Poultry Science, v. 56 (5), 1550-1559
Eimeria tenella, E. acervulina, E. maxima, chicks, narasin, monensin, lasalocid, comparison of efficacy, resistance and tolerance

Eimeria acervulina
Witlock, D. R.; and Ruff, M. D., 1977, J. Parasitol., v. 63 (2), 193-199
5 Eimeria spp. compared, intestinal mucosal surface damage in infected chickens observed by scanning electron microscopy

Eimeria acervulina
Wojton, T., 1976, Med. Wet., v. 32 (2), 78
Eimeria acervulina, E. maxima, poultry, efficacy of coccidot and sulfatyf

Eimeria acervulina
Yvore, P., 1974, Folia Vet. Latina, v. 4 (3), 408-425
intestinal coccidiosis (E. acervulina), caecal coccidiosis (E. tenella), comparison of nutritional effect in fowl, E. tenella causing insignificant losses, E. acervulina causing severe disease, interference with absorption

Eimeria acervulina
Yvore, P.; and Hubert, J., 1977, Rec. Med. Vet., v. 153 (4), 287-291
Eimeria tenella, E. acervulina, halofuginone added to drinking water of infected chickens (exper.), curative effects compared with action of darvisul and amprolium, good results

Eimeria acervulina
Zukovic, M.; and Brglez, J., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 11-16
coccidiosis of domestic fowl, survey, first record of Eimeria brunetti, control, review: Croatia and Slovenia

Eimeria acervulina var. diminuta, illus.
Shirley, M. W.; Millard, B. J.; and Long, P. L., 1977, Parasitology, v. 75 (2), 165-176
Eimeria acervulina var. diminuta and Eimeria acervulina var. mivati compared, growth in vitro, response to anticoccidial drugs, electrophoretic mobility profiles of four enzymes

Eimeria acervulina var. mivati, illus.
Long, P. L., 1974, Folia Vet. Latina, v. 4 (2), 289-300
culture in chick embryos, review

Eimeria acervulina var. mivati
Long, P. L.; and Millard, B. J., 1976, Ztschr. Parasitenk., v. 48 (3-4), 287-290
Eimeria spp., immunized chickens, oocysts produced after corticosteroid treatment in some birds, possibly endogenous cycle continues or remains dormant; possible means of diagnosis of occult infection in apparently immune hosts

Eimeria acervulina var. mivati
Long, P. L.; and Millard, B. J., 1977, Avian Path., v. 6 (1), 77-92
chickens immunised against Eimeria using repeated low level infections with attenuated strains of E. tenella and E. acervulina var. mivati alone or together with E. brunetti, E. maxima, E. necatrix

Eimeria acervulina var. mivati
Shirley, M. W., 1975, J. Protozool., v. 22 (3), 55A-56A [Abstract]
Eimeria spp., lactate dehydrogenase, glucose phosphate isomerase, characteristic electrophoretic mobilities for different species, parasite identification and other possible applications

Eimeria acervulina var. mivati
Shirley, M. W., 1977, J. Protozool., v. 24 (4), 43A-44A [Abstract]
establishment of infections in chickens and/or eggs by single sporozoites or single sporocysts

Eimeria acervulina var. mivati
Shirley, M. W.; and Lee, D. L., 1977, J. Parasitol., v. 63 (2), 390-392
Eimeria spp., isoelectric focusing of enzymes, compared with starch gel electrophoresis, inter- and intraspecies differences

Eimeria acervulina var. mivati, illus.
Shirley, M. W.; Millard, B. J.; and Long, P. L., 1977, Parasitology, v. 75 (2), 165-176
Eimeria acervulina var. diminuta and Eimeria acervulina var. mivati compared, growth in vitro, response to anticoccidial drugs, electrophoretic mobility profiles of four enzymes

Eimeria adenoeides
Anderson, W. I.; et al., 1977, Avian Path., v. 6 (2), 125-130
Eimeria meleagrimitis, E. gallopavonis, E. adenoeides, turkey poults, altered intestinal pH values

Eimeria adenoeides
Anderson, W. I.; Reid, W. M.; and McDougald, L. R., 1976, Avian Dis., v. 20 (2), 387-394
single and mixed infections of Eimeria adenoeides, E. meleagrimitis, and E. gallopavonis, turkeys, monensin, drug efficacy in laboratory and floor-pen experiments, good results

Eimeria adenoeides
Joyner, L. P., 1973, Folia Vet. Latina, v. 3 (1), 110-123
Eimeria adenoeides, E. meleagrimitis, coccidiosis in turkeys, pathogenicity, age resistance, control, drug resistance, review

Eimeria adenoeides
Joyner, L. P.; and Norton, C. C., 1973, Research Vet. Sc., v. 14 (3), 279-284
Eimeria meleagrimitis known to be sulphaquinoxaline-resistant, activity of amprolium and ethopabate separately or together or in combination with sulphaquinoxaline in Mixture A.E.S., results indicate that activity of mixture was due to ethopabate component although one strain proved resistant to it also, activity of ethopabate reversed by p-aminobenzoic acid; Eimeria adenoeides, effect of amprolium, sulphaquinoxaline, and ethopabate separately and in combination, ethopabate on its own was completely ineffective

Eimeria adenoeides
McDougald, L. R., 1976, Poultry Science, v. 55 (6), 2442-2447
Eimeria adenoeides, E. meleagrimitis, E. gallopavonis, turkeys, monensin, good efficacy, increased host growth

Eimeria adleri Yakimoff and Gousseff, 1936, illus.
Ovezmukhammedov, A.; and Berdyev, A. S., 1975, Izvest. Akad. Nauk Turkmensk. SSR, s. Biol. Nauk (6), 55-59
brief description
Vulpes vulpes: Turkmenistan

Eimeria ahsata
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria ahsata
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria ahsata
Catchpole, J.; Norton, C. C.; and Joyner, L. P., 1976, Parasitology, v. 72 (2), 137-147
4 Eimeria spp., lambs, monospecific or multispecific infections, pattern and level of oocyst production, clinical symptoms

Eimeria ahsata
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

Eimeria ahsata
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Ovis ammon musimon: School Forest Enterprise, University of Agriculture Brno, Krtiny

E[imeria] ahsata
Glebezdin, V. S., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 76-80
coccidiosis, sheep, infection rates, host age, season: Turkmenistan

Eimeria ahsata
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria ah-sa-ta
Pav, J., 1974, Veterinarstvi, v. 24 (11), 513-515
mufloni: Vlkov preserve

Eimeria ahsata
Pout, D. D.; Norton, C. C.; and Catchpole, J., 1973, Brit. Vet. J., v. 129 (6), 568-582
Eimeria spp., lambs (exper.), patterns of oocyst production in single and mixed infections, results indicate that continuous oocyst production as observed in the field could be achieved through exponential dosing, limitations of laboratory infections

Eimeria ah-sa-ta
Taylor, S. M.; et al., 1973, Vet. Rec., v. 93 (17), 461-464
Eimeria spp., lambs kept on straw bedding vs. expanded metal flooring showed higher fecal oocyst counts, diarrhea, greater weight loss, and some fatalities; causal agents of disease appear to be E. ninaekohlyakimovae and/or E. arloingi with concurrent increase of intestinal Cl[ostridium] welchii type A

Eimeria alabamensis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria alabamensis, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves (duodenum, jejunum): Veliko Turnovo Okrug, Bulgaria

Eimeria alabamensis
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria alabamensis, illus.
Soekardono, S.; Ernst, J. V.; and Benz, G. W., 1975, Vet. Parasitol., v. 1 (1), 19-33
Eimeria alabamensis, calves (exper.), prepatent and patent periods, low pathogenicity, description of oocyst, sporulation of oocysts at various temperatures

Eimeria americana Carvalho, 1943, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria anguillae Leger & Hollande, 1922, illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
description
Anguilla rostrata (epithelium of gut's anterior portion, feces): Matamek River, Quebec

Eimeria anseris (Kotlan, 1932), illus.
Betke, P.; and Wilhelm, A., 1976, Monatsh. Vet.-Med., v. 31 (15), 585-589
Eimeria anseris, geese, peracute outbreak, clinical symptoms, laboratory diagnosis, control with sulfadimidin: Cottbus

Eimeria anseris
Szalay, G.; and Sari, I., 1976, Magy. Allat. Lapja, v. 98, v. 31 (2), 95-96
Eimeria nocens, E. anseris, large scale goose farms, control with Sulfamix

Eimeria arabiana Veisov
Musaev, M. A.; and Ismailov, S. G., 1976, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (6), 71-75
Eimeria arabiana, various infectious doses, variability of measurements and shape of oocysts, variation in length of patent period
Meriones vinogradovi (nat. and exper.): Dzhulfinsk section, Azerbaidzhan Antiplague Institute

Eimeria arloingi (E. ovina)
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria arloingi
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria arloingi
Baker, N. F.; Walters, G. T.; and Fisk, R. A. 1972, Am. J. Vet. Research, v. 33 (1), 83-86
Eimeria arloingi, E. parva, E. intricata, amprolium, control in feedlot lambs

Eimeria arloingi (Marotel, 1905) Martin, 1909
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Capra ibex (feces): Delhi Zoo

E[imeria] arloingi
Chroust, K., 1973, Acta Vet. Brno, v. 42 (1), 45-52
lambs naturally infected with E[imeria] ninakohlyakimovae, E. arloingi, E. parva, E. intricata, E. faurei, and E. crandallis, treated with Clopidol administered in feed, effective control with dose rates of 500 mg/kg feed for 14 days or 250 mg/kg feed for 56 days

Eimeria arloingi
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Ovis ammon musimon: School Forest Enterprise, University of Agriculture Brno, Krtiny

E[imeria] arloingi
Glebezdin, V. S., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 76-80
coccidiosis, sheep, infection rates, host age, season: Turkmenistan

E[imeria] arloingi
Misra, S. C., 1972, Indian J. Animal Research, v. 6 (2), 95-96
parasitic gastro-enteritis, goats, epidemiology, seasonal incidence: Orissa

Eimeria arloingi
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria arloingi
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Ovis aries: Chile

Eimeria arloingi
Osman, H. S.; Restani, R.; and Canestri-Trotti, G., 1972, Parassitologia, v. 14 (1), 167-173
sheep (feces): Somalia

Eimeria arloingi
Pav, J., 1974, Veterinarstvi, v. 24 (11), 513-515
mufloni: Vlkov preserve

E[imeria] arloingi 'B'
Pout, D. D., 1974, Brit. Vet. J., v. 130 (1), 54-60
E[imeria] crandallis, E[imeria] arloingi 'B', lambs (exper.), effects on host growth, food intake, faecal oocyst production and clinical symptoms; results indicate that diet may influence the total body response to infection in experimental animals

Eimeria arloingi A
Pout, D. D.; Norton, C. C.; and Catchpole, J., 1973, Brit. Vet. J., v. 129 (6), 568-582
Eimeria spp., lambs (exper.), patterns of oocyst production in single and mixed infections, results indicate that continuous oocyst production as observed in the field could be achieved through exponential dosing, limitations of laboratory infections

Eimeria arloingi B, illus.
Pout, D. D.; Norton, C. C.; and Catchpole, J., 1973, Brit. Vet. J., v. 129 (6), 568-582
Eimeria spp., lambs (exper.), patterns of oocyst production in single and mixed infections, results indicate that continuous oocyst production as observed in the field could be achieved through exponential dosing, limitations of laboratory infections

Eimeria arloingi
Reed, D. E.; et al., 1976, J. Am. Vet. Med. Ass., v. 169 (9), 975-979
Odocoileus virginianus (rectal contents): South Dakota

Eimeria arloingi
Reid, J. F. S., 1976, Vet. Rec., v. 98 (25), 496-499
gastrointestinal nematodes, coccidiosis, diarrhea of sheep, age and seasonal factors: Britain

Eimeria arloingi
Richter, S.; Karlovic, M.; and Rukavina, A.; 1976, Vet. Glasnik, v. 30 (9), 757-762
incidence, severe infections
goats (intestine, feces): Velebita

Eimeria arloingi
Rivellini, P.; Fenizia, D.; and Rania, U., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 494-498
incidence, sheep: Campania and Lucania

Eimeria arloingi
Taylor, S. M.; et al., 1973, Vet. Rec., v. 93 (17), 461-464
Eimeria spp., lambs kept on straw bedding vs. expanded metal flooring showed higher fecal oocyst counts, diarrhea, greater weight loss, and some fatalities; causal agents of disease appear to be E. ninaekohl-yakimovae and/or E. arloingi with concurrent increase of intestinal Cl[ostridium] welchii type A

Eimeria arloingi
Zielinski, J., 1972, Med. Wet., v. 28 (9), 566-567
parasites, sheep, Nilverm, copper sulfate

Eimeria ascotensis
Joseph, T., 1973, J. Protozool., v. 20 (4), 509
Sciurus niger rufiventer: northern Indiana

Eimeria attenuata sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1974, J. Protozool., v. 21 (4), 483-489
Thamnophis sirtalis parietalis (feces): Iowa
Natrix sipedon sipedon (feces): Windy Hills Shooting Refuge, 5 mi S Muscatine, Louisa Co., Iowa

Eimeria attenuata Wacha & Christiansen
Wacha, R. S.; and Christiansen, J. L., 1975, J. Protozool., v. 22 (4), 453-454
Natrix sipedon sipedon: Louisa Co., Iowa

Eimeria auburnensis Christensen et Porter, 1939
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (8), 68-70
Bos grunniens domesticus (feces): Tadzhikistan

Eimeria auburnensis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria auburnensis, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves (jejunum, ileum): Veliko Turnovo Okrug, Bulgaria

Eimeria auburnensis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria auburnensis
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria audubonii Duszynski and Marquardt, 1969, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria auriti n. sp., illus.
Mirza, M. Y., 1975, Bull. Nat. Hist. Research Cent. Univ. Baghdad, v. 6 (1), 39-52
Hemiechinus auritus (feces): Baghdad

Eimeria baghdadensis n. sp., illus.
Mirza, M. Y., 1975, Bull. Nat. Hist. Research Cent. Univ. Baghdad, v. 6 (1), 39-52
Mus musculus (feces): Baghdad

Eimeria banffensis n. sp., illus.
Lepp, D. L.; Todd, K. S., jr.; and Samuel, W. M., 1973, Tr. Am. Micr. Soc., v. 92 (2), 305-307
Ochotona princeps (feces): Jumpingpound and Sibbald Creeks areas, near Banff, Alberta, Canada

Eimeria bareillyi
Shastri, U. V.; Ghafoor, M. A.; and Krishnamurthy, R., 1976, Indian J. Animal Sc., v. 46 (10), 556-561
Eimeria bareillyi, E. ellipsoidalis, Coccidia [sp.], endogenous development and associated lesions in buffalo calves: Parbhani

Eimeria bareillyi, illus.
Shastri, U. V.; and Krishnamurthi, R., 1976, Indian J. Animal Sc., v. 45 (1), 1975, 46-47
Eimeria bareillyi, buffaloes, pathology, postmortem findings: Parbhani

Eimeria bateri, Bhatia, Pandey and Pande, 1965, illus.
Martialay Valle, F., 1975, An. Inst. Invest. Vet., Madrid, v. 23, 1974-1975, 131-148
Eimeria bateri, pathogenic for one day old quails, little pathogenicity for quails more than 2 weeks old
Coturnix coturnix japonica: near Madrid

Eimeria bateri, illus.
Sanchez Acedo, C.; and Garcia Reguera, V., 1972, Rev. Iber. Parasitol., v. 32 (3-4), 271-276
Eimeria bateri, morphology, incidence in farm quail, inefficacy of sulfacetacine
Coturnix coturnix (intestino delgado, ciegos): farm near Zaragoza

Eimeria bentongi n. sp., illus.
Colley, F. C.; and Mullin, S. W., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 289-291
Hylomys suillus (intestinal contents): Janda Baik, Bentong, Pahang, Malaysia

Eimeria betica n. sp., illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria bonasae
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Tetrastes bonasia: Kazakhstan

Eimeria bovis Fiebiger, 1912
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (8), 68-70
Bos grunniens domesticus (feces): Tadzhikistan

Eimeria bovis
Benz, G. W.; and Ernst, J. V., 1976, Am. J. Vet. Research, v. 37 (8), 895-899
Cooperia punctata and/or Eimeria bovis-infected calves, reduced alkaline phosphatase activities in intestinal mucosa

Eimeria bovis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria bovis
Danielli, Y.; and Neuman, M., 1975, Refuah Vet., v. 32 (4), 94-95, 153-154
mixed parasites, cattle, good results following repeated chemotherapy: Birkat Ata

Eimeria bovis
Fitzgerald, P. R.; and Mansfield, M. E., 1973, J. Protozool., v. 20 (1), 121-126
Eimeria bovis, Holstein-Friesian calves (exper.), monensin incorporated in pelleted feed protected against severe clinical coccidiosis, observations on oocyst discharge in feces, clinical signs, weight gains, food consumption, hemoglobin, packed cell volume, total serum protein, sodium and potassium content of serum, and differential white cell count

Eimeria bovis
Jensen, J. B.; et al., 1976, J. Parasitol., v. 62 (2), 195-198
Eimeria spp., effect of substituting various gases for CO_2 in established excystation procedure, evidence that role of CO_2 is that of allosteric effector enhancing action of reducing agent

Eimeria bovis
Julian, R. J.; Harrison, K. B.; and Richardson, J. A., 1976, Mod. Vet. Pract., v. 57 (9), 711-712, 716, 718
Eimeria zurnii, E. bovis, cattle, nervous signs, pathology, severe enteritis, possibility of toxins from coccidia: Canada

Eimeria bovis, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves (jejunum, ileum, caecum, colon): Veliko Turnovo Okrug, Bulgaria

Eimeria bovis
Klesius, P. H.; et al., 1977, Exper. Parasitol., v. 41 (2), 480-490
Eimeria bovis, cattle, evidence for cell-mediated immune response shown by delayed hypersensitivity skin tests and lymphocyte blastogenesis with antigens extracted from oocysts of E. bovis and E. stiedai, cross-reactivity between two species

Eimeria bovis
Klesius, P. H.; and Fudenberg, H. H., 1977, Clin. Immunol. and Immunopathol., v. 8 (2), 238-246
cattle, transfer of cell-mediated immunity to Eimeria bovis antigen with bovine transfer factor (unfractionated or alcohol precipitates)

Eimeria bovis
Klesius, P. H.; and Kristensen, F., 1977, Clin. Immunol. and Immunopathol., v. 7 (2), 240-252
transfer factor, characterization of biological activity by criteria of passive transfer of delayed hypersensitivity reactivity, lymphocyte stimulation, and protective effects against bovine and rabbit coccidiosis (Eimeria bovis, Eimeria stiedae)

Eimeria bovis
Miner, M. L.; and Jensen, J. B., 1976, Am. J. Vet. Research, v. 37 (9), 1043-1045
Eimeria bovis, E. zuernii, calves (exper.), decoquinate administered orally in grain mix suppressed oocyst discharge and bloody diarrhea, clinical signs did not recur for 23 days after treatment was discontinued

Eimeria bovis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria bovis
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria bovis
Ryley, J. F.; Gordon, R. W.; and Stark, J. R., 1974, J. Protozool., v. 21 (4), 596-598
Eimeria bovis, E. stiedai, storage polysaccharides shown to be amylopectin

Eimeria brasiliensis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria brasiliensis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria brunetti
Allen, W. M.; et al., 1973, J. Comp. Path., v. 83 (3), 369-375
Eimeria brunetti, chickens experimentally infected at 2 different dose rates, physiopathological changes, plasma protein and electrolyte alterations, weight loss

Eimeria brunetti, illus.
Birch-Andersen, A.; Ferguson, D. J. P.; and Pontefract, R. D., 1976, Acta Path. et Microbiol. Scand., v. 84B (4), 235-239
technique for obtaining thin sections of coccidian oocysts demonstrated with Eimeria brunetti

Eimeria brunetti, illus.
Ferguson, D. J. P.; et al., 1977, Acta Path. et Microbiol. Scand., v. 85B (1), 67-77
Eimeria brunetti, ultrastructural changes occurring during microgametogony, ultrastructure of microgamete

Eimeria brunetti, illus.
Ferguson, D. J. P.; et al., 1977, Acta Path. et Microbiol. Scand., v. 85B (1), 78-88
Eimeria brunetti, ultrastructural changes occurring during macrogametogony, ultrastructure of macrogamete

Eimeria brunetti, illus.
Ferguson, D. J. P.; et al., 1977, Acta Path. et Microbiol. Scand., v. 85B (1), 110-112
Eimeria brunetti, ultrastructure of late sporoblast, initiation of sporozoite formation

Eimeria brunetti, illus.
Ferguson, D. J. P.; et al., 1977, Acta Path. et Microbiol. Scand., v. 85B (3), 201-211
Eimeria brunetti, domestic fowls, formation and structure of oocyst wall

Eimeria brunetti, illus.
Ferguson, D. J. P.; et al., 1977, Acta Path. et Microbiol. Scand., v. 85B (6), 363-373
Eimeria brunetti, micropores, ultrastructure, distribution, and apparent functional status in various developmental forms

Eimeria brunetti
Giordani, G.; Govoni, S.; and Bergami, W., 1977, Avicoltura, Bologna, v. 46 (1), 41-46
Eimeria spp., breeder hens, continuously fed sulphachloropyrazine + diaveridine + ethopabate, no undesirable effects on egg production, values statistically equal for control and coccidiostat fed birds

Eimeria brunetti
Hein, H. E., 1976, Exper. Parasitol., v. 39 (3), 415-421
Eimeria acervulina, E. brunetti, E. maxima, pathogenic effects of single vs. mixed infections with low doses of oocysts in chickens, changes in mean weight and oocyst output

Eimeria brunetti
Hein, H. E., 1976, Exper. Parasitol., v. 40 (2), 250-260
Eimeria acervulina, E. brunetti, E. maxima, E. necatrix, immunogenic potential under precisely defined conditions which provide standards enabling comparisons between species and strains to be made, found that high level of resistance to reinfection was conferred by multiple infections with relatively low doses of oocysts

Eimeria brunetti
Helmboldt, C. F.; and Bryant, E. S., 1971, Avian Dis., v. 15 (4), 775-780
Eimeria brunetti, E. maxima, presence of oocysts in chickens with necrotic enteritis, etiologic relationship not clear; differentiation of E. brunetti coccidiosis from necrotic enteritis

Eimeria brunetti
Joyner, L. P.; and Norton, C. C., 1977, Parasitology, v. 74 (2), 205-213
Eimeria maxima, genetic transfer of drug resistance, results support view that drug resistance factors are transferred at zygote formation, phenomenon took place between variants of the same species (Eimeria maxima var. indentata) but not between different species (Eimeria brunetti), transference not affected by treatment with acriflavine

Eimeria brunetti
Joyner, L. P.; and Norton, C. C., 1977, Parasitology, v. 75 (2), 155-164
Eimeria spp., anticoccidial effects of amprolium alone or with ethopabate, dinitolmide, and monensin with particular reference to oocyst sporulation, significance of effects on gametogony and sporogony

Eimeria brunetti
Karlsson, T.; and Reid, W. M., 1977, Brit. Poultry Sc., v. 18 (4), 497-501
Eimeria acervulina, E. necatrix, E. brunetti, E. tenella, chickens (exper.), immunity challenge techniques used to determine prevalence in Georgia (USA), lesion scores as main criterion to indicate exposure

Eimeria brunetti
Kohls, R. E.; Engle, A. T.; and Butters, H. E., 1972, Avian Dis., v. 16 (4), 907-914
coccidiosis in laying chickens (exper.), continuous buquinolate medication, good protection with increased egg production and egg quality, immune status at termination

Eimeria brunetti, illus.
Long, P. L.; et al., 1976, Folia Vet. Latina, v. 6 (3), 201-217
Eimeria spp., avian, procedure, key and laboratory methods for identification, pathology, experimental infection procedures, factors affecting development, instructive review

Eimeria brunetti
Long, P. L.; and Millard, B. J., 1976, Ztschr. Parasitenk., v. 48 (3-4), 287-290
Eimeria spp., immunized chickens, oocysts produced after corticosteroid treatment in some birds, possibly endogenous cycle continues or remains dormant; possible means of diagnosis of occult infection in apparently immune hosts

Eimeria brunetti
Long, P. L.; and Millard, B. J., 1977, Avian Path., v. 6 (1), 77-92
chickens immunised against Eimeria using repeated low level infections with attenuated strains of E. tenella and E. acervulina var. mivati alone or together with E. brunetti, E. maxima, E. necatrix

Eimeria brunetti
Miller, B. M.; et al., 1977, Poultry Science, v. 56 (6), 2039-2044
Eimeria spp., chickens, efficacy and tolerance of two 6-amino-9-(substituted benzyl) purines, good results

Eimeria brunetti
Movsesijan, M.; et al., 1975, Acta Vet. Beograd, v. 25 (5), 255-259
Eimeria spp., chickens administered single oral dose of irradiated oocysts resistant to subsequent challenge dose of non-irradiated oocysts, irradiated and non-irradiated E. tenella antigenically more potent than E. necatrix and E. brunetti and produced highest titres of specific antibodies

Eimeria brunetti
Mylari, B. L.; et al., 1977, J. Med. Chem., v. 20 (4), 475-483
Eimeria spp. in chickens, 1-benzyl-6-azauracils as potential new anticoccidials, comparison trials with several experimental and commercial anticoccidials

Eimeria brunetti
Oikawa, H.; et al., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 271-279
Eimeria spp., chickens, survey during 1973 neither seasonal nor regional distribution differences were observed; oocyst detection rate significantly low in chicks up to 30 days old: Japan

Eimeria brunetti
Pesti, G. M.; and Combs, G. F., jr., 1976, Poultry Science, v. 55 (6), 2265-2274
Eimeria, site selectivity of various species for infection in particular regions of chick gastrointestinal tract used as tool to study loci of selenium absorption, potentiation of selenium deficiency by intestinal parasitism

Eimeria brunetti
Reid, W. M., 1975, Avian Dis., v. 19 (4), 802-811
Eimeria spp., chicks medicated with roxarsone and zoalene produced higher oocyst counts than unmedicated controls confirming the conclusion that oocyst counts are an unsatisfactory index to anticoccidial activity

Eimeria brunetti
Reid, W. M.; et al., 1977, Poultry Science, v. 56 (1), 66-71
coccidiosis, chicks, monensin, development of immunity dependent upon drug level, treatment time, and oocyst exposure

Eimeria brunetti Levine, 1942
Reid, W. M.; and Johnson, J., 1974, Folia Vet. Latina, v. 4 (4), 585-602
Eimeria brunetti in chickens, severity of pathogenicity, lesion scoring and weight gains, economic losses, diagnostic techniques

Eimeria brunetti
Rollinson, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 436-437 [Demonstration]
Eimeria spp., attempted differentiation of chicken coccidia by electrophoretic variation of enzymes

Eimeria brunetti
Rose, M. E., 1976, Vet. Rec., v. 98 (24), 481-484
poultry coccidiosis, immunity and prospects for immunoprophylaxis, review

Eimeria brunetti
Ruff, M. D.; et al., 1975, Avian Path., v. 4 (1), 73-81
Eimeria brunetti, effects on intestinal pH in conventional and gnotobiotic chickens

Eimeria brunetti
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria brunetti
Ryley, J. F.; and Hardman, L., 1977, Parasitology, v. 75 (2), xv [Abstract]
Eimeria spp., effect of vitamin K-deficient diets on infection and in particular on effectiveness of anti-coccidial drugs, concluded that use of deficient diet in screening and evaluating drugs is justified

Eimeria brunetti
Sokolic, A.; et al., 1976, Brit. Vet. J., v. 132 (4), 416-422
chickens, single immunizing oral dose of irradiated oocysts of Eimeria brunetti, E. necatrix, E. tenella protects against severe simultaneous challenge with these three spp.

Eimeria brunetti
Turk, D. E., 1977, Poultry Science, v. 56 (5), 1763 [Abstract]
Eimeria spp., chicks, iron absorption, decreased by some species, increased during recovery period

Eimeria brunetti, illus.
Witlock, D. R.; and Ruff, M. D., 1977, J. Parasitol., v. 63 (2), 193-199
5 Eimeria spp. compared, intestinal mucosal surface damage in infected chickens observed by scanning electron microscopy

Eimeria brunetti
Zukovic, M.; and Brglez, J., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 11-16
coccidiosis of domestic fowl, survey, first record of Eimeria brunetti, control, review
Croatia and Slovenia

Eimeria bukidnonensis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria bukidnonensis Tubangui 1931, illus.
Courtney, C. H.; Ernst, J. V.; and Benz, G. W., 1976, J. Parasitol., v. 62 (3), 372-376
redescription of oocysts, cattle (feces), comparison of measurements, attempt to determine patent and prepatent periods

Eimeria bukidnonensis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria bukidnonensis
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria callosciuri Colley, 1971
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Callosciurus notatus
C. nigrovittatus
all from Bukit Lanjan Forest Reserve, West Malaysia

Eimeria cameli
Gill, H. S., 1976, Indian Vet. J., v. 53 (11), 897-898
Camelus dromedarius (feces): States of Punjab, Haryana, and Rajasthan

Eimeria campania Carvalho, 1943, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria canadensis Bruce, 1921
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (8), 68-70
Bos grunniens domesticus (feces): Tadzhikistan

Eimeria canadensis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria canadensis
Danielli, Y.; and Neuman, M., 1975, Refuah Vet., v. 32 (4), 94-95, 153-154
mixed parasites, cattle, good results following repeated chemotherapy: Birkat Ata

Eimeria canadensis, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves: Veliko Turnovo Okrug, Bulgaria

Eimeria canadensis (Bruce, 1921), illus.
Mueller, B. E. G.; deVos, A. J.; and Hammond, D. M., 1973, J. Protozool., v. 20 (2), 293-297
Eimeria canadensis, development of first generation schizonts and merozoites in various bovine cell cultures

Eimeria canadensis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria capreoli
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (1), 65-77
roe deer: Czechoslovakia

Eimeria capreoli
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Capreolus capreolus: School Forest Enterprise, University of Agriculture Brno, Krtiny

Eimeria capreoli
Volf, K.; and Volfova, M., 1974, Veterinarstvi, v. 24 (3), 125-126
srnci zvere: Trebic District

Eimeria carinii
Mishra, G. S.; and Gonzalez, J. P., 1975, Arch. Inst. Pasteur Tunis, v. 52 (1-2), 71-87
Rattus norvegicus (intestin): Tunis, Tunisie
rat blanc (exper.)

Eimeria carpelli
Kocyłowski, B.; et al., 1976, Bull. Vet. Inst. Pulawy, v. 20 (1-2), 12-17
Eimeria carpelli, carp (intestine), incidence correlated with host age and state of culture and hygiene of ponds, amprolium chloride, good results, higher efficacy than nitrofural: Poland

Eimeria carpelli
Kurashvili, B. E., 1975, Izvest. Akad. Nauk Gruzinsk. SSR, s. Biol., v. 1 (4), 317-320
antagonistic and synergetic interrelationships between intestinal parasites

Eimeria catostomi sp. n., illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
Catostomus commersoni
Hypentelium nigricans
(epithelium of gut's anterior portion and feces of all): all from Grand River and Laurel Creek near Waterloo, Bronte Creek near Milton, and Conestogo River near Conestogo, Ontario

Eimeria caucasica
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Alectoris kakelik: Kazakhstan

Eimeria caviae
Prosl, H., 1976, Ztschr. Parasitenk., v. 50 (2), 214
Meerschweinchen

Eimeria cerdonis, illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria cerdonis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Eimeria cheetali
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Antelope cervicapra (feces): Lucknow Zoo

Eimeria chinchillae De Vos & Van der Westhuizen, 1968
De Vos, A. J., 1970, Onderstepoort J. Vet. Research, v. 37 (1), 29-36
Eimeria chinchillae, experimental studies on host range; pathology, morphology, and development in hosts; findings indicate a lack of host specificity
Praomys natalensis (feces, large intestine, small intestine)
Rhabdomys pumilio (feces, large intestine, small intestine)
white mice (feces, large intestine, small intestine)
white rats (feces)
Otomys irroratus (feces)
Mystromys albicaudatus (feces)
Arvicanthis niloticus (feces, wall of rectum)
chinchillas (feces, large intestine)
(all exper.)

?Eimeria chinchillae De Vos & Van der Westhuizen, 1968
De Vos, A. J., 1970, Onderstepoort J. Vet. Research, v. 37 (1), 29-36
Eimeria chinchillae, experimental studies on host range; pathology, morphology, and development in hosts; findings indicate a lack of host specificity
Saccostomus campestris (feces) (exper.)

Eimeria chinchillae De Vos & Van der Westhuizen, 1968, illus.
De Vos, A. J.; and Dobson, Lynne D., 1970, Onderstepoort J. Vet. Research, v. 37 (4), 185-189
Rhabdomys pumilio (feces, wall of caecum and colon, small intestine) (nat. and exper.): near the Veterinary Research Institute, Onderstepoort, Pretoria district
chinchillas (feces) (exper.)

Eimeria chinkari Pande et al., 1970
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Gazella gazella (feces): Lucknow Zoo

Eimeria choudari Bhatia et al., 1972
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Columba spp. (feces): Lucknow Zoo

Eimeria chrysemydis Deeds & Jahn, 1939, illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
redescription, prevalence
Chrysemys picta belli (feces): Iowa

Eimeria cicaki sp. n., illus.
Else, J. G.; and Colley, F. C., 1975, J. Protozool., v. 22 (4), 455-457
Gehyra mutilata
Hemidactylus frenatus
(feces, epithelial cells of small intestine of all): all from Kuala Lumpur, Malaysia

Eimeria coecicola Kheisin, 1947, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria coecicola Kheissin, 1947, illus.
Norton, C. C.; Catchpole, J.; and Rose, M. E., 1977, Parasitology, v. 75 (1), 1-7
Eimeria stiedai, presence of oocyst residuum confirmed, differential diagnosis from Eimeria coecicola and Eimeria neoleporis

Eimeria coecicola
Rollinson, D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21-22 [Demonstration]
Eimeria spp. of mammals, electrophoretic techniques to differentiate spp. by variations in mobility of enzymes

Eimeria colchici Norton, 1967
Bejsovec, J., 1973, J. Protozool., v. 20 (4), 535
seasonal incidence
Phasianus colchicus: Central Bohemia

Eimeria colchici Norton, 1967
Bejsovec, J., 1974, J. Protozool., v. 21 (3), 454 [Abstract]
Eimeria colchici, E. phasiani, incidence of oocysts in various biotopes of an agricultural landscape over a 6-year period: Central Bohemia

Eimeria colchici, illus.
Norton, C. C., 1976, Folia Vet. Latina, v. 6 (3), 218-238
pheasants, description, review of literature: United Kingdom

Eimeria colini sp. n., illus.
Fisher, J. W.; and Kelley, G. L., 1977, J. Parasitol., v. 63 (2), 200-202
Colinus virginianus (feces): Jasper County, Iowa

Eimeria collanuli sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1974, J. Protozool., v. 21 (4), 483-489
Diadophis punctatus arnyi: Exline City Dump, Pleasant Township, Appanoose Co., Iowa

Eimeria confusa
Davidson, W. R., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 211-217
epizootiologic and pathologic study of endoparasites of selected populations of gray squirrels
Sciurus carolinensis (small intestine): southeastern United States

Eimeria confusa
Forrester, D. J.; et al., 1977, J. Parasitol., v. 63 (6), 1045
Sciurus carolinensis (feces): Jacksonville, Duval County, northern Florida

Eimeria confusa Joseph, 1969
Joseph, T., 1973, J. Protozool., v. 20 (4), 509
Sciurus niger rufiventer (exper.)

Eimeria contorta Haberkorn, 1971
Cerna, Z., 1975, J. Protozool., v. 22 (3), 60A [Abstract]
"Our preliminary experiments suggested that E[imeria] contorta isolated from rats and transmissible to mice may, evidently, be identical in its antigenic reaction to that of the mouse coccidians E. falciformis and E. falciformis var. pragensis"

Eimeria contorta
Heil, R., 1974, Hamatologische Untersuchungen an Saugetieren und Kuken nach experimentellen Kokzidien-Infektionen, 137 pp.
Rattus norvegicus var. albinus, experimental infection, haemoglobin and haematocrit values, differential blood cell counts

Eimeria contorta
Mueller, B. E. G., 1975, J. Protozool., v. 22 (3), 11A [Abstract]
Eimeria contorta, sporozoite ultrastructure and transformation into schizonts

Eimeria contorta Haberkorn, 1971, illus.
Mueller, B. E. G., 1975, Ztschr. Parasitenk., v. 47 (1), 23-34
Eimeria contorta, asexual development from sporozoite to first-generation merozoite in tissue culture, possible feeding function of some structures, electron microscopy

Eimeria contorta Haberkorn, 1971, illus.
Mueller, B. E. G., 1975, Ztschr. Parasitenk., v. 47 (2), 91-101
Eimeria contorta, development, first generation merozoites to second generation schizonts and merozoites in mouse, electron microscopy

Eimeria contorta
Stockdale, P. H. G., 1976, Canad. J. Zool., v. 54 (7), 1064-1068
Eimeria contorta and E. nieschulzi in rats, E. zuernii in calves, production of patent infections using oocysts, sporocysts, or sporozoites

Eimeria coturnicis
Zaprianov, M., 1976, Vet. Med. Nauki, v. 13 (5), 78-83
Eimeria spp. of domestic Alectoris graeca cypriates (feces), highest incidence in birds up to two months of age and in May-June

Eimeria crandallis
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria crandallis
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria crandallis Honess, 1942
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Capra ibex (feces): Delhi Zoo

Eimeria crandallis
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Eimeria crandallis
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

E[imeria] crandallis
Chroust, K., 1973, Acta Vet. Brno, v. 42 (1), 45-52
lambs naturally infected with E[imeria] ninakohlyakimovae, E. arloingi, E. parva, E. intricata, E. faurei, and E. crandallis, treated with Clopidol administered in feed, effective control with dose rates of 500 mg/kg feed for 14 days or 250 mg/kg feed for 56 days

Eimeria crandallis
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Ovis ammon musimon: School Forest Enterprise, University of Agriculture Brno, Krtiny

E[imeria] crandallis
Glebezdin, V. S., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 76-80
coccidiosis, sheep, infection rates, host age, season: Turkmenistan

Eimeria crandallis
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria crandallis
Pav, J., 1974, Veterinarstvi, v. 24 (11), 513-515
mufloni: Vlkov preserve

E[imeria] crandallis
Pout, D. D., 1974, Brit. Vet. J., v. 130 (1), 54-60
E[imeria] crandallis, E[imeria] arloingi 'B', lambs (exper.), effects on host growth, food intake, faecal oocyst production and clinical symptoms; results indicate that diet may influence the total body response to infection in experimental animals

Eimeria crandallis, illus.
Pout, D. D.; Norton, C. C.; and Catchpole, J., 1973, Brit. Vet. J., v. 129 (6), 568-582
Eimeria spp., lambs (exper.), patterns of oocyst production in single and mixed infections, results indicate that continuous oocyst production as observed in the field could be achieved through exponential dosing, limitations of laboratory infections

Eimeria crandallis
Taylor, S. M.; et al., 1973, Vet. Rec., v. 93 (17), 461-464
Eimeria spp., lambs kept on straw bedding vs. expanded metal flooring showed higher fecal oocyst counts, diarrhea, greater weight loss, and some fatalities; causal agents of disease appear to be E. ninaekohlyakimovae and/or E. arloingi with concurrent increase of intestinal Cl[ostridium] welchii type A

Eimeria crotalviridis sp. n., illus.
Duszynski, D. W.; et al., 1977, J. Protozool., v. 24 (3), 359-361
excystation of sporozoites; ultrastructure of oocyst wall
Crotalus viridis viridis: 10 mi W of Los Lunas, Valencia County, and 60 mi NW Clovis, Curry County, New Mexico

Eimeria cylindrica Wilson, 1931
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (8), 68-70
Bos grunniens domesticus (feces): Tadzhikistan

Eimeria cylindrica
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria cylindrica, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves (duodenum, jejunum): Veliko Turnovo Okrug, Bulgaria

Eimeria cylindrica
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria cylindrica
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria dalli sp. n., illus.
Clark, G. W.; and Colwell, D. A., 1974, J. Protozool., v. 21 (2), 197-199
Ovis dalli (feces): Crescent mountain, Kenai Peninsula, Alaska

Eimeria dathei nov. sp., illus.
Tscherner, W., 1976, Zool. Garten N. F., v. 46 (1-2), 52-53
Kobus ellipsiprymnus: Tierpark Berlin

Eimeria dawari n. sp., illus.
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Muntiacus muntijaka (feces): Lucknow Zoo

Eimeria debliecki
Levine, N. D.; and Cechner, P. E., 1973, J. Protozool., v. 20 (4), 510-511
Eimeria spp. and Isospora canis oocysts fed to pigs, no evidence that Sarcocystis infections resulted

Eimeria debliecki
Levine, N. D.; Cechner, P. E.; and Meyer, R. C., 1974, Indian Vet. J., v. 51 (1), 57-59
pigs fed oocysts of porcine Eimeria spp. and of Isospora canis, "Neither Sarcocystis nor any other protozoon was found, except for Eimeria spp. in the intestinal epithelium of pigs that had been inoculated with porcine Eimeria spp."

Eimeria debliecki, illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria debliecki
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Eimeria debliecki
Onawunmi, O. A.; and Todd, A. C., 1976, Am. J. Vet. Research, v. 37 (6), 657-660
porcine coccidiosis (exper.), suppression and control with chlortetracycline combination, buquinolate, and lincomycin hydrochloride

E[imeria] debliecki, illus.
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian J. Animal Sc., v. 46 (10), 567-570
coccidia, pigs (exper.), measurements of 4 types of mature schizonts recovered from single pig (small intestine)

E[imeria] debliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1977, Indian J. Animal Sc., v. 47 (9), 597-600
E[imeria] debliecki, E. neodebliecki, E. perminuta, Isospora suis, pigs (feces), sporulation time: Aligarh

Eimeria debliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1977, Indian Vet. J., v. 54 (5), 360-362
Eimeria spp., pigs, amprolsol-MSD fully effective if administered just after second day of infection

Eimeria debliecki (Douwes, 1921), illus.
Yvore, P., 1976, Rev. Med. Vet., v. 152 (1), 25-32
pigs (nat. and exper.): France

Eimeria delagei marginata Deeds & Jahn, 1939, illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
redescription, prevalence
Chrysemys picta belli
Graptemys geographica
G. pseudogeographica
(feces of all): all from Iowa

Eimeria dericksoni, illus.
Wacha, R. S.; and Christiansen, J. L., 1977, J. Protozool., v. 24 (3), 357-359
diagnosis; oocysts redescribed
Trionyx spiniferus: Iowa

Eimeria dhamini sp. nov., illus.
Mandal, R.; and Mukherjee, R. N., 1977, Science and Culture, v. 43 (11), 479-480 [Letter]
Ptyas mucosus (small intestine): Zoological Garden, Lucknow, U.P.

Eimeria diminuta sp.n., illus.
Fernando, M. A.; and Remmler, O., 1973, J. Protozool., v. 20 (3), 357
Gallus lafayettei (feces): north central Ceylon
Gallus domesticus (exper.)

Eimeria dispersa, illus.
Doran, D. J.; and Augustine, P. C., 1977, J. Protozool., v. 24 (1), 172-176
Eimeria dispersa and E. gallopavonis in primary chicken and turkey kidney cell cultures, infectivity, survival, development

Eimeria dissanaikei sp. n., illus.
Fernando, M. A.; and Remmler, O., 1973, J. Protozool., v. 20 (1), 43-45
Gallus lafayettei (feces): north central Ceylon
G. domesticus (exper.)

Eimeria dromedarii
Gill, H. S., 1976, Indian Vet. J., v. 53 (11), 897-898
Camelus dromedarius (feces): States of Punjab, Haryana, and Rajasthan

Eimeria duncani sp. n., illus.
Varghese, T., 1977, J. Protozool., v. 24 (2), 209-210
Halcyon sancta (feces): Wau, New Guinea Highlands, Papua New Guinea

Eimeria duodenalis Norton, 1967, illus.
Fisher, J. W.; and Wacha, R. S., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 226-227
Phasianus colchicus (feces): private game farm, Jasper County, Iowa

Eimeria duodenalis, illus.
Norton, C. C., 1976, Folia Vet. Latina, v. 6 (3), 218-238
pheasants, description, review of literature: United Kingdom

Eimeria duodenalis
Utebaeva, M. K.; and Svanbaev, S. K., 1975, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (6), 27-32
Eimeria spp., pheasants, incidence, seasonal distribution
Phasianus colchicus mongolicus
Chrysolophus pictus
Syrmaticus reevesi
Crossoptilon tibetanum
Euplocomus nycthomerus
Phasianus colchicus lorenzi
all from Kazakhstan

Eimeria ellipsoidalis Becket et Frye, 1929
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (8), 68-70
Bos grunniens domesticus (feces): Tadzhikistan

Eimeria ellipsoidalis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria ellipsoidalis, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves (duodenum, jejunum): Veliko Turnovo Okrug, Bulgaria

Eimeria ellipsoidalis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria ellipsoidalis
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria ellipsoidalis, illus.
Shastri, U. V.; Ghafoor, M. A.; and Krishnamurthy, R., 1976, Indian J. Animal Sc., v. 46 (10), 556-561
Eimeria bareillyi, E. ellipsoidalis, Coccidia [sp.], endogenous development and associated lesions in buffalo calves: Parbhani

Eimeria elliptica sp. n., illus.
Sayin, F.; Dincer, S.; and Meric, I., 1977, J. Protozool., v. 24 (2), 210-212
Spalax leucodon (intestine): Lalahan district, Ankara, Turkey

Eimeria environ Honess, 1939, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria etheostomae sp. n., illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
Etheostoma exile
E. nigrum
(epithelium of gut's anterior portion and feces of all): all from Bronte Creek near Milton and Laurel Creek near Waterloo, Ontario

Eimeria europaea
Haupt, W.; and Hartung, J., 1977, Monatsh. Vet.-Med., v. 32 (9), 339-341
Lepus europaeus: Leipzig area

Eimeria europaea
Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
Lepus europaeus: Austria

Eimeria europaea Pellerdy, 1956, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria exigua Yakimoff, 1934, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria exigua Yakimoff, 1934 *of* Morgan and Waller, 1940
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria minima Carvalho, 1943

Eimeria exigua var. septentrionalis Yakimoff et al., 1936 *of* Madsen, 1938
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria septentrionalis Yakimoff, Matschoulsky and Spartansky, 1936

Eimeria falciformis Eimer, 1870
Cerna, Z., 1975, J. Protozool., v. 22 (3), 60A [Abstract]
"Our preliminary experiments suggested that E[imeria] contorta isolated from rats and transmissible to mice may, evidently, be identical in its antigenic reaction to that of the mouse coccidians E. falciformis and E. falciformis var. pragensis"

Eimeria falciformis
Cox, A. B.; Duncan S.; and Levy, C. K., 1977, J. Parasitol., v. 63 (5), 927-929
Eimeria falciformis, effects of cobalt irradiation on infectivity and immunogenicity of sporulated oocysts, mice

Eimeria falciformis, illus.
D'Haese, J.; Mehlhorn, H.; and Peters, W., 1977, Internat. J. Parasitol., v. 7 (6), 505-518
Sarcocystis ovifelis, Besnoitia jellisoni, Eimeria falciformis, pellicular structures, fine structure, implications for mechanism of motility

Eimeria falciformis
Heil, R., 1974, Hamatologische Untersuchungen an Saugetieren und Kuken nach experimentellen Kokzidien-Infektionen, 137 pp.
Mus musculus var. albinus, experimental infection, haemoglobin and haematocrit values, differential blood cell counts

Eimeria falciformis
Wright, A. C.; and Levy, C. K., 1974, J. Protozool., v. 21 (3), 424 [Abstract]
Eimeria falciformis, experiments to determine the extent to which age of unsporulated oocyst's affects their responses to ^{60}Co gamma irradiation, possible effect of prior cold storage upon results

Eimeria falciformis var. pragensis
Cerna, Z., 1975, J. Protozool., v. 22 (3), 60A [Abstract]
"Our preliminary experiments suggested that E[imeria] contorta isolated from rats and transmissible to mice may, evidently, be identical in its antigenic reaction to that of the mouse coccidians E. falciformis and E. falciformis var. pragensis"

Eimeria faurei
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria faurei
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria faurei
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Eimeria faurei
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

E[imeria] faurei
Chroust, K., 1973, Acta Vet. Brno, v. 42 (1), 45-52
lambs naturally infected with E[imeria] ninakohlyakimovae, E. arloingi, E. parva, E. intricata, E. faurei, and E. crandallis, treated with Clopidol administered in feed, effective control with dose rates of 500 mg/kg feed for 14 days or 250 mg/kg feed for 56 days

Eimeria faurei (Moussu and Marotel, 1902) Martin, 1909, illus.
Duszynski, D. W.; Samuel, W. M.; and Gray, D. R., 1977, Canad. J. Zool., v. 55 (6), 990-999
redescription
Ovibos moschatus: Nunivak Island, Alaska; game farm, Alberta; Bathurst Island, Northwest Territories, and breeding station, Ft. Chimo, Quebec, Canada; Bardu breeding station, Norway
Ovis canadensis: Montana
(feces of all)

Eimeria faurei, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

E[imeria] faurei
Glebezdin, V. S., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 76-80
coccidiosis, sheep, infection rates, host age, season: Turkmenistan

E[imeria] faurei
Misra, S. C., 1972, Indian J. Animal Research, v. 6 (2), 95-96
parasitic gastro-enteritis, goats, epidemiology, seasonal incidence: Orissa

Eimeria faurei
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria faurei
Osman, H. S.; Restani, R.; and Canestri-Trotti, G., 1972, Parassitologia, v. 14 (1), 167-173
sheep
goats
(feces of all): all from Somalia

Eimeria faurei
Pav, J., 1974, Veterinarstvi, v. 24 (11), 513-515
mufloni: Vlkov preserve

Eimeria faurei
Rivellini, P.; Fenizia, D.; and Rania, U., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 494-498
incidence, sheep: Campania and Lucania

Eimeria faurei
Svanbaev, S. K.; and Dzerzhinskii, V. A., 1976, Vestnik Sel'skokhoz. Nauki Kazakhstana (1), 70-72
Eimeria faurei, lambs, furacillin and furazolidone suppress parasite development in first half of prepatent period, treatment not useful in second half of prepatent period

Eimeria faurei
Volf, K.; and Volfova, M., 1974, Veterinarstvi, v. 24 (3), 125-126
srnci zvere
jeleni zvere
all from Trebic District

Eimeria faurei
Zielinski, J., 1972, Med. Wet., v. 28 (9), 566-567
parasites, sheep, Nilverm, copper sulfate

Eimeria felina
Fameree, L.; and Cotteleer, C., 1976, J. Protozool., v. 23 (4), 10A [Abstract]
coccidiosis, cats, prevalence: Belgium

Eimeria felina
Loeffler, K., 1974, Prakt. Tierarzt, v. 55, Sondernummer, 78-80
cat diseases, prophylaxis and therapy, brief review

Eimeria fernandoae sp. n., illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
Catostomus commersoni
Hypentelium nigricans
(epithelium of gut's anterior portion and feces of all): all from Bronte Creek near Milton, Conestogo River near Conestogo, and Grand River near Waterloo, Ontario

Eimeria ferrisi Levine & Ivens, 1965, illus.
Ankrom, S. L.; Chobotar, B.; and Ernst, J. V., 1975, J. Protozool., v. 22 (3), 317-323
Eimeria ferrisi, redescription of sporulated oocysts, life cycle and observations on endogenous stages in mice
Mus musculus (caecum, colon) (exper.)

Eimeria ferrisi Levine and Ivens 1965, illus.
Chobotar, B.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 111-121
Eimeria ferrisi, macrogamonts and macrogametes in epithelial cells of Mus musculus cecum and colon, ultrastructure, development, oocyst wall formation

Eimeria ferrisi Levine and Ivens 1965, illus.
Scholtyseck, E.; et al., 1977, Ztschr. Parasitenk., v. 51 (3), 229-240
Eimeria ferrisi, mice, electron microscopy of microgamogony

Eimeria ferrisi, illus.
Senaud, J.; Chobotar, B.; and Scholtyseck, E., 1976, Tropenmed. u. Parasitol., v. 27 (2), 145-159
comparative fine structure morphology of Plasmodium cathemerium, Eimeria ferrisi, E. stiedai, Besnoitia jellisoni, and Frenkelia sp. micropores, discussion of micropore function in parasite nutrition

Eimeria ferrisi Levine and Ivens, 1965, illus.
Senaud, J.; Chobotar, B.; and Scholtyseck, E., 1976, Ztschr. Parasitenk., v. 49 (3), 217-222
Eimeria ferrisi, asexual development in intestinal lumen of mouse, multinuclear stages apparently identical with endogenesis, ultrastructure

Eimeria ferruginea Colley, 1970
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Tupaia glis: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria francolini n. sp., illus.
Swarup, M.; and Chauhan, M. P. S., 1976, Science and Culture, v. 42 (3), 166-167 [Letter]
Francolinus francolinus (feces): Delhi Zoo

Eimeria furonis
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Mustela]: Kazakhstan fur farms

Eimeria gadi (Fiebiger, 1913), illus.
Odense, P. H.; and Logan, V. H., 1976, J. Protozool., v. 23 (4), 564-571
prevalence, distribution, seasonal variation, morphology, pathologic implications
Melanogrammus aeglefinus
Enchelyopus cimbrius
(swim bladder of all): all from Nova Scotian fishing banks

Eimeria galateai sp. n., illus.
Varghese, T., 1977, J. Protozool., v. 24 (2), 209-210
Tanysiptera galatea (feces): Brown River Area, Port Mosby, Papua New Guinea

Eimeria gallopavonis
Anderson, W. I.; et al., 1977, Avian Path., v. 6 (2), 125-130
Eimeria meleagrimitis, E. gallopavonis, E. adenoeides, turkey poults, altered intestinal pH values

Eimeria gallopavonis
Anderson, W. I.; Reid, W. M.; and McDougald, L. R., 1976, Avian Dis., v. 20 (2), 387-394
single and mixed infections of Eimeria adenoides, E. meleagrimitis, and E. gallopavonis, turkeys, monensin, drug efficacy in laboratory and floor-pen experiments, good results

Eimeria gallopavonis Hawkins, illus.
Doran, D. J.; and Augustine, P. C., 1977, J. Protozool., v. 24 (1), 172-176
Eimeria dispersa and E. gallopavonis in primary chicken and turkey kidney cell cultures, infectivity, survival, development

Eimeria gallopavonis
McDougald, L. R., 1976, Poultry Science, v. 55 (6), 2442-2447
Eimeria adenoeides, E. meleagrimitis, E. gallopavonis, turkeys, monensin, good efficacy, increased host growth

Eimeria gonzalezcastroi nom. nov.
Lizcano Herrera, J.; and Romero-Rodriguez, J., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 391
Syn.: E. gonzalezi Lizcano-Herrera y Romero-Rodriguez, 1972

Eimeria gonzalezi Lizcano-Herrera y Romero-Rodriguez, 1972
Lizcano Herrera, J.; and Romero-Rodriguez, J., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 391
as syn. of E. gonzalezcastroi nom. nov.

Eimeria gorakhpuri Bhatia & Pande, 1967
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Numida meleagris (feces): Delhi Zoo; Lucknow Zoo

Eimeria granulosa Christensen, 1938, illus.
Duszynski, D. W.; Samuel, W. M.; and Gray, D. R., 1977, Canad. J. Zool., v. 55 (6), 990-999
redescription
Ovibos moschatus (feces): Nunivak Island, Alaska; Bathurst Island, Northwest Territories, Canada; Bardu breeding station, Norway
Ovis canadensis (feces): Montana

Eimeria granulosa, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

Eimeria granulosa
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria granulosa
Osman, H. S.; Restani, R.; and Canestri-Trotti, G., 1972, Parassitologia, v. 14 (1), 167-173
sheep (feces): Somalia

Eimeria granulosa
Taylor, S. M.; et al., 1973, Vet. Rec., v. 93 (17), 461-464
Eimeria spp., lambs kept on straw bedding vs. expanded metal flooring showed higher fecal oocyst counts, diarrhea, greater weight loss, and some fatalities; causal agents of disease appear to be E. ninaekohlyakimovae and/or E. arloingi with concurrent increase of intestinal Cl[ostridium] welchii type A

Eimeria graptemydos sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
prevalence
Graptemys geographica
G. pseudogeographica
Chrysemys picta belli
(feces of all): all from Iowa

Eimeria groenlandica Madsen, 1938 emend., illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Syn.: Eimeria perforans var. groenlandica Madsen, 1938 pro parte
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria gruis Yakimoff & Matschoulsky 1935, illus.
Courtney, C. H.; et al., 1975, J. Parasitol., v. 61 (4), 695-699
description, morphology compared with E. reichenowi
Grus canadensis pratensis (feces): Florida
G. canadensis tabida (feces): Florida; Arizona
G. canadensis canadensis (feces): Texas

Eimeria haematodi sp. n., illus.
Varghese, T., 1977, J. Parasitol., v. 63 (2), 210-211
Trichoglossus haematodus (feces): Port Moresby and Wau, Papua New Guinea

Eimeria helmisophis sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1974, J. Protozool., v. 21 (4), 483-489
Carphophis amoenus vermis: 2 mi SE Winterset, Madison Co., Iowa

Eimeria hispidi sp. nov., illus.
Bastardo de San Jose, T., 1974, Acta Biol. Venez., v. 8 (3-4), 567-578
endogenous cycle in host intestine
Tropidurus hispidus (epitelio y submucosa del intestino delgado): El Veladero, Estado Guarico, Venezuela

Eimeria hoffmani sp. n., illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
Umbra limi (epithelium of gut's anterior portion, feces): Bronte Creek near Milton, Ontario

Eimeria honessi Carvalho, 1943 emend., illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria honessi Landers, 1952
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
not syn. of Eimeria honessi Carvalho, 1943 emend.

Eimeria hungarica
Haupt, W.; and Hartung, J., 1977, Monatsh. Vet.-Med., v. 32 (9), 339-341
Lepus europaeus: Leipzig area

Eimeria hungarica
Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
Lepus europaeus: Austria

Eimeria hungarica Pellerdy, 1956, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria hungarica Pellerdy, 1956, illus.
Romero Rodriguez, J., 1975, Rev. Iber. Parasitol., v. 35 (1-2), 157-161
Lepus granatensis
Oryctolagus cuniculus ferox
all from Granada, Spain

Eimeria hydrophis sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1974, J. Protozool., v. 21 (4), 483-489
Natrix sipedon sipedon: Windy Hills Shooting Refuge, 5 mi S Muscatine, Louisa Co., Iowa; Van Buren Co., Iowa; Sweet Marsh, Bremer Co., Iowa
N. rhombifera rhombifera: Schoolbus Dike, Windy Hills Shooting Refuge, 4-1/2 mi S Muscatine, Muscatine Co., Iowa
(feces of all)

Eimeria hydrophis Wacha & Christiansen
Wacha, R. S.; and Christiansen, J. L., 1975, J. Protozool., v. 22 (4), 453-454
Natrix sipedon sipedon: Louisa Co., Iowa

Eimeria illinoisensis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria imantauica
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Alopex lagopus]: Kazakhstan fur farms

Eimeria indentata sp.n., illus.
Fernando, M. A.; and Remmler, O., 1973, J. Protozool., v. 20 (1), 43-45
Gallus lafayettei (feces): north central Ceylon
Gallus domesticus (exper.)

Eimeria indianensis sp. n., illus.
Joseph, T., 1974, J. Protozool., v. 21 (1), 12-15
prepatent and patent periods, host immunity
Didelphis virginiana (nat. and exper.) (feces from rectum, intestine): Northern Indiana, U.S.A.

Eimeria indianensis
Joseph, T., 1974, Proc. Indiana Acad. Sc., v. 83, 1973, 467 [Abstract]
Didelphis virginiana: St. Joseph's, Elkhart, La Porte and Marshall Counties, Indiana

Eimeria intestinalis
Catchpole, J.; and Norton, C. C., 1975, J. Protozool., v. 22 (3), 49A [Abstract]
Eimeria intestinalis, rabbits (exper.), pathogenic effects increased with higher dose levels, developmental stages observed, sulphaquinoxaline in food was partially effective as preventive but not clopidol, treatment in drinking water with sulphadimidine was successful but not amprolium + ethopabate

Eimeria intestinalis Kheisin, 1948, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria intestinalis
Rollinson, D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21-22 [Demonstration]
Eimeria spp. of mammals, electrophoretic techniques to differentiate spp. by variations in mobility of enzymes

Eimeria intestinalis Cheissin 1948, illus.
Romero Rodriguez, J.; and Lizcano Herrera, J., 1972, Rev. Iber. Parasitol., v. 32 (3-4), 199-206
Eimeria intestinalis, E. sculpta, morphology, first report from Oryctolagus cuniculus domestica in Spain

Eimeria intestinalis
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria intricata
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria intricata
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria intricata
Baker, N. F.; Walters, G. T.; and Fisk, R. A., 1972, Am. J. Vet. Research, v. 33 (1), 83-86
Eimeria arloingi, E. parva, E. intricata, amprolium, control in feedlot lambs

Eimeria intricata
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Eimeria intricata
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

E[imeria] intricata
Chroust, K., 1973, Acta Vet. Brno, v. 42 (1), 45-52
lambs naturally infected with E[imeria] ninakohlyakimovae, E. arloingi, E. parva, E. intricata, E. faurei, and E. crandallis, treated with Clopidol administered in feed, effective control with dose rates of 500 mg/kg feed for 14 days or 250 mg/kg feed for 56 days

Eimeria intricata, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

E[imeria] intricata
Glebezdin, V. S., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 76-80
coccidiosis, sheep, infection rates, host age, season: Turkmenistan

E[imeria] intricata
Misra, S. C., 1972, Indian J. Animal Research, v. 6 (2), 95-96
parasitic gastro-enteritis, goats, epidemiology, seasonal incidence: Orissa

Eimeria intricata
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria intricata
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Ovis aries: Chile

Eimeria intricata
Pav, J., 1974, Veterinarstvi, v. 24 (11), 513-515
mufloni: Vlkov preserve

Eimeria intricata
Rivellini, P.; Fenizia, D.; and Rania, U., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 494-498
incidence, sheep: Campania and Lucania

Eimeria iowaensis sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1974, J. Protozool., v. 21 (4), 483-489
Thamnophis sirtalis parietalis (feces): Iowa

Eimeria iroquoina Molnar & Fernando, 1974
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
variation in cyprinid hosts
Nocomis biguttatus
Notropis heterolepis
N. rubellus
Pimephales notatus
Rhinichthys atratulus
Semotilus atromaculatus
(epithelium of gut's anterior portion and feces of all): all from Ontario

Eimeria irresidua Kessel and Jankiewicz, 1931, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria irresidua Kessel and Jankiewicz, 1931 *of* Gill and Ray (1960) in Lepus ruficaudatus
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria campania Carvalho, 1943

Eimeria irresidua
Lineburg, A.; and Pastuszko, J., 1972, Med. Wet., v. 28 (4), 219-220
coccidiosis, rabbits, Sulfatyf, Anticoccid

Eimeria irresidua form campanius Carvalho, 1943
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria campania Carvalho, 1943

Eimeria juniataensis sp. n., illus.
Pluto, T. G.; and Rothenbacher, H., 1976, J. Parasitol., v. 62 (2), 207-208
Graptemys geographica (feces): Raystown Branch, Juniata River, Huntingdon County, Pennsylvania

Eimeria kanchili n. sp., illus.
Mullin, S. W.; and Colley, F. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 441-443
Tragulus javanicus (intestinal contents): Subang Forest Preserve, Selangor, West Malaysia

Eimeria karatauica sp. nov., illus.
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Alectoris kakelik (intestine): Karatau, Kazakhstan

Eimeria kofoidi
Zaprianov, M., 1976, Vet. Med. Nauki, v. 13 (5), 78-83
Eimeria spp. of domestic Alectoris graeca cypriates (feces), highest incidence in birds up to two months of age and in May-June

Eimeria kotlani Graubmann et Grafner, 1964
Antukhaev, I. K., 1977, Tsitologiia, v. 19 (3), 288-292
Eimeria kotlani, ultrastructure of macrogamete; development within host cell nucleus; structure of wall forming bodies; nutrition presumably by pinocytosis

Eimeria labbeana Pinto, 1928
Gut, J., 1974, J. Protozool., v. 21 (3), 455 [Abstract]
Eimeria labbeana in Streptopelia decaocto (exper.), life cycle

Eimeria labbeana
Gut, J.; Jurkovic, P.; and Bedrnik, P., 1977, J. Protozool., v. 24 (2), Suppl., 14A [Abstract]
Eimeria labbeana, in vitro cultivation

Eimeria labbeana
Hunt, S.; and O'Grady, J., 1976, Austral. Vet. J., v. 52 (8), 390 [Letter]
pigeons: outer suburb of Sydney

Eimeria labbeana (Labbe), illus.
Mandal, A. K., 1977, Indian J. Animal Health, v. 16 (1), 5-8
Isospora lacazei, Eimeria labbeana, distribution patterns of DNA and RNA in various stages of life cycle

Eimeria labbeana Pinto, 1928, illus.
Varghese, T., 1975, J. Protozool., v. 22 (1), 66-71
Eimeria labbeana first generation merozoites, fine structure

Eimeria labbeana, illus.
Varghese, T., 1976, Ztschr. Parasitenk., v. 49 (1), 25-32
Eimeria labbeana, endogenous stages, fine structure of feeding organelles

Eimeria labbeana, illus.
Varghese, T., 1976, Ztschr. Parasitenk., v. 50 (3), 227-235
Eimeria labbeana from Columba livia, microgametogenesis, fine structure of microgamonts and microgametes

Eimeria labbeana, illus.
Varghese, T., 1977, J. Protozool., v. 24 (3), 376-382
Eimeria labbeana, fine structure of 3 generations of meronts, merogony, and merozoites with special reference to rhoptry-microneme system

Eimeria lafayettei sp.n., illus.
Fernando, M. A.; and Remmler, O., 1973, J. Protozool., v. 20 (1), 43-45
Gallus lafayettei (feces): north central Ceylon

Eimeria lalahanensis sp. n., illus.
Sayin, F.; Dincer, S.; and Meric, I., 1977, J. Protozool., v. 24 (2), 210-212
Spalax leucodon (intestine): Lalahan district, Ankara, Turkey

Eimeria lampropeltis Anderson et al.
Wacha, R. S.; and Christiansen, J. L., 1974, J. Protozool., v. 21 (4), 483-489
as possible syn. of E. zamenis Phisalix, 1921

Eimeria lancasterensis
Forrester, D. J.; et al., 1977, J. Parasitol., v. 63 (6), 1045
Sciurus carolinensis (feces): Jacksonville, Duval County, northern Florida

Eimeria lancasterensis Joseph, 1969, "similar to"
Joseph, T., 1973, J. Protozool., v. 20 (4), 509
Sciurus niger rufiventer: northern Indiana

Eimeria larimerensis, illus.
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Eimeria leimadophi sp. n., illus.
Lainson, R.; and Shaw, J. J., 1973, J. Protozool., v. 20 (3), 358-362
Leimadophis poecilogyrus (small intestine): Serra dos Carajas, Para State, north Brazil

Eimeria leninogorica sp. nov., illus.
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Tetrastes bonasia (intestine): Leninogorska, Kazakhstan

Eimeria leporis
Haupt, W.; and Hartung, J., 1977, Monatsh. Vet.-Med., v. 32 (9), 339-341
Lepus europaeus: Leipzig area

Eimeria leporis
Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
Lepus europaeus: Austria

Eimeria leporis Nieschulz, 1923, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria leporis Nieschulz of Morgan and Waller, 1940 and Waller and Morgan, 1941
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria neoleporis Carvalho, 1942

Eimeria leuckarti (Flesch, 1883) Reichenow, 1940, illus.
Canestri-Trotti, G.; and Restani, R., 1972, Parassitologia, v. 14 (2-3), 245-250
Eimeria leuckarti, incidence, description
horse (feces): Italy
donkey (feces): Abruzzo, Italy

Eimeria leuckarti, illus.
McQueary, C. A.; Worley, D. E.; and Catlin, J. E., 1977, Am. J. Vet. Research, v. 38 (10), 1673-1674
Eimeria leuckarti, horses, prevalence and life cycle: southwestern Montana

Eimeria leuckarti
Sheahan, B. J., 1976, Vet. Rec., v. 99 (11), 213-214
Eimeria leuckarti, horse, acute case, pathology: Ireland

Eimeria leuckarti, illus.
Sutoh, M.; et al., 1976, National Inst. Animal Health Quart., v. 16 (2), 59-64
pathology
foals (small intestine): Japan

Eimeria leucodonica sp. n., illus.
Veisov, A. M., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 82-85
Spalax leucodon: vicinity of Talinsk region, Armenian SSR

Eimeria liophi sp. n., illus.
Lainson, R.; and Shaw, J. J., 1973, J. Protozool., v. 20 (3), 358-362
Liophis cobella (feces): Belem, Para State, north Brazil

Eimeria lophurae n. sp., illus.
Chauhan, M. P. S.; Paliwal, U. C. D.; and Swarup, M., 1976, Current Sc., Bangalore, v. 45 (16), 591-592 [Letter]
Lophura leucomenala (feces): Delhi Zoo

Eimeria lutotestudinis sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
prevalence
Kinosternon flavescens spooneri: Iowa

Eimeria lyruri
Bejsovec, J., 1977, J. Protozool., v. 24 (4), 48A [Abstract]
persistence of Coccidia despite changing land use and decreasing numbers of host animals
Lyrurus tetrix: Bohemia

Eimeria lyruri
Pav, J.; and Zajicek, D., 1974, Veterinarstvi, v. 24 (11), 517-520
Lyrus tetrix
Tetrao urogallus
all from CSSR

Eimeria macyi sp. n., illus.
Wheat, B. E., 1975, J. Parasitol., v. 61 (5), 920-922
Pipistrellus subflavus (cecal contents): Lion's Den Cave, Clarke County, Alabama

Eimeria magna
Burgaz, I., 1973, Norwegian J. Zool., v. 21 (4), 327 [Abstract]
crossinfection of mountain hares (Lepus timidus) with tame rabbit coccidia (Eimeria stiedai, E. magna) and tame rabbits with mountain hare coccidia (E. media)

Eimeria magna
Danforth, H. D., 1975, J. Protozool., v. 22 (3), 11A [Abstract]
Eimeria magna, fine structure of in vivo and in vitro asexual stages

Eimeria magna
Fitzgerald, P. R., 1976, J. Protozool., v. 23 (1), 150-154
Eimeria stiedai, E. perforans, E. magna, rabbits, control by administration of antibiotics A204 sodium, A28695A, and A204 normal propyl in pelleted feed

Eimeria magna
Jensen, J. B.; and Edgar, S. A., 1976, J. Parasitol., v. 62 (2), 203-206
Eimeria magna sporozoites, penetration into cultured cells, effects of antiphagocytic agents and microtubule inhibitors, results suggest that phagocytosis is not mechanism of entry but that sporozoite motility is of primary importance

Eimeria magna, illus.
Jensen, J. B.; and Edgar, S. A., 1976, J. Parasitol., v. 62 (6), 988-992
Eimeria magna, evidence of secretory function of rhoptries during penetration of cultured cells

Eimeria magna Perard, 1925, illus.
Jensen, J. B.; and Hammond, D. M., 1975, J. Protozool., v. 22 (3), 411-415
Eimeria magna, penetration into cultured cells, ultrastructure of invasion process

Eimeria magna Perard, 1925, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria magna Perard, 1925 of Henry, 1932
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria townsendii Carvalho, 1943

Eimeria magna
Lineburg, A.; and Pastuszko, J., 1972, Med. Wet., v. 28 (4), 219-220
coccidiosis, rabbits, Sulfatyf, Anticoccid

Eimeria magna, illus.
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Eimeria magna
Rollinson, D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21-22 [Demonstration]
Eimeria spp. of mammals, electrophoretic techniques to differentiate spp. by variations in mobility of enzymes

Eimeria magna
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria magna, illus.
Ryley, J. F.; and Robinson, T. E., 1976, Ztschr. Parasitenk., v. 50 (3), 257-275
Eimeria magna, rabbits, schizogony, gametogony, excystation of oocysts; mild pathogenic effects

Eimeria magna, illus.
Speer, C. A.; et al., 1973, J. Protozool., v. 20 (2), 274-281
Eimeria magna, fine structure of young, intermediate, and mature macrogamonts

Eimeria magna, illus.
Speer, C. A.; and Danforth, H. D., 1976, J. Protozool., v. 23 (1), 109-115
Eimeria magna, microgametogenesis in rabbits and in kidney cell cultures, fine structure of developmental stages

Eimeria maior Honess, 1939, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria mandali Banik & Ray, 1964
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Pavo cristatus (feces): Lucknow Zoo

Eimeria maralikiensis sp. n., illus.
Veisov, A. M., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 82-85
Spalax leucodon: vicinity of Talinsk and Maralik Aniisk regions, Armenian SSR

Eimeria marsica, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

Eimeria marsica, illus.
Norton, C. C.; and Catchpole, J., 1976, Parasitology, v. 72 (1), 111-114
redescription
Ovis aries (nat. and exper.): England; Wales

Eimeria marsica
Rivellini, P.; Fenizia, D.; and Rania, U., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 494-498
incidence, sheep: Campania and Lucania

Eimeria mascoutini sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
prevalence
Trionyx spiniferus (feces): Muscatine County, Iowa

Eimeria mascoutini, illus.
Wacha, R. S.; and Christiansen, J. L., 1977, J. Protozool., v. 24 (3), 357-359
photographs and additional measurements of oocysts
Trionyx spiniferus: Iowa

Eimeria mastomyis n. sp., illus.
De Vos, A. J.; and Dobson, Lynne D., 1970, Onderstepoort J. Vet. Research, v. 37 (4), 185-189
Praomys natalensis (intestine): near the Veterinary Research Institute, Onderstepoort, Pretoria district

Eimeria matsubayashii Tsunoda, 1952, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria matsubayashii Tsunoda, 1952, illus.
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria maxima, illus.
Babu, K. S. J.; Seshadri, S. J.; and Mohiyuddeen, S., 1976, Indian Vet. J., v. 53 (1), 47-54
coccidiosis, poultry, field cases, diagnosis, pathology, relationship of infection to age and sex of host

Eimeria maxima
Chapman, H. D., 1976, Avian Path., v. 5 (4), 283-290
Eimeria tenella, E. acervulina, E. maxima, incidence in broiler house used for commercial drug trials, resistance to various anticoccidials

E[imeria] maxima
Dvorak, M.; et al., 1977, Veterinaria, Praha, v. 19 (1-2), 31-73
coccidiosis, poultry, preclinical tests using darvisul as standard for comparison, screening trials with sulphonamides and diaminopyrimidines; development of preparation Sulfakombin suitable for administration in water

Eimeria maxima
Foure, N.; and Bennejean, G., 1974, Bull. Inform. Station Exper. Avicult. Ploufragan (Cotes-du-Nord), v. 14 (3), 57-60
Eimeria, chickens, Halofuginone, comparison with methyl-benzoquate and Clopidol, good results

Eimeria maxima
Giordani, G.; Govoni, S.; and Bergami, W., 1977, Avicoltura, Bologna, v. 46 (1), 41-46
Eimeria spp., breeder hens, continuously fed sulphachloropyrazine + diaveridine + ethopabate, no undesirable effects on egg production, values statistically equal for control and coccidiostat fed birds

Eimeria maxima
Hein, H. E., 1976, Exper. Parasitol., v. 39 (3), 415-421
Eimeria acervulina, E. brunetti, E. maxima, pathogenic effects of single vs. mixed infections with low doses of oocysts in chickens, changes in mean weight and oocyst output

Eimeria maxima
Hein, H. E., 1976, Exper. Parasitol., v. 40 (2), 250-260
Eimeria acervulina, E. brunetti, E. maxima, E. necatrix, immunogenic potential under precisely defined conditions which provide standards enabling comparisons between species and strains to be made, found that high level of resistance to reinfection was conferred by multiple infections with relatively low doses of oocysts

Eimeria maxima
Helmboldt, C. F.; and Bryant, E. S., 1971, Avian Dis., v. 15 (4), 775-780
Eimeria brunetti, E. maxima, presence of oocysts in chickens with necrotic enteritis, etiologic relationship not clear; differentiation of E. brunetti coccidiosis from necrotic enteritis

Eimeria maxima
Jeffers, T. K., 1977, Poultry Science, v. 56 (5), 1725 [Abstract]
Eimeria tenella, E. maxima, E. acervulina, broiler flocks, no development of monensin resistance following long-term drug exposure

Eimeria maxima
Joyner, L. P.; and Norton, C. C., 1975, J. Protozool., v. 22 (3), 52A-53A [Abstract]
Eimeria maxima, transfer of drug resistance from one strain to another when passaged together in chicks

Eimeria maxima
Joyner, L. P.; and Norton, C. C., 1976, Parasitology, v. 72 (1), 115-125
Eimeria maxima, E. acervulina, chicks, immunity arising from continuous low-level infections was stronger and/or more enduring than that produced by single inoculations of comparable numbers of oocysts

Eimeria maxima
Joyner, L. P.; and Norton, C. C., 1976, Parasitology, v. 73 (2), iii [Abstract]
Eimeria maxima, transferred drug resistance, balance of evidence supports view that transference phenomenon takes place by genetic recombination

Eimeria maxima
Joyner, L. P.; and Norton, C. C., 1977, Parasitology, v. 74 (2), 205-213
Eimeria maxima, genetic transfer of drug resistance, results support view that drug resistance factors are transferred at zygote formation, phenomenon took place between variants of the same species (Eimeria maxima var. indentata) but not between different species (Eimeria brunetti), transference not affected by treatment with acriflavine

Eimeria maxima
Joyner, L. P.; and Norton, C. C., 1977, Parasitology, v. 75 (2), 155-164
Eimeria spp., anticoccidial effects of amprolium alone or with ethopabate, dinitolmide, and monensin with particular reference to oocyst sporulation, significance of effects on gametogony and sporogony

Eimeria maxima
Kohls, R. E.; Engle, A. T.; and Butters, H. E., 1972, Avian Dis., v. 16 (4), 907-914
coccidiosis in laying chickens (exper.), continuous buquinolate medication, good protection with increased egg production and egg quality, immune status at termination

Eimeria maxima
Kolev, V.; et al., 1976, Vet. Med. Nauk., v. 13 (4), 3-12
coccidiosis, incidence in poultry at large poultry dressing combine, birds imported from France and England as possible source of infection, prophylactic measures for better disease control: Ruse, Bulgaria

Eimeria maxima
Kuil, H.; et al., 1977, Vet. Parasitol., v. 3 (1), 33-40
Eimeria maxima, Eimeria acervulina, chickens, effect of inoculation dose on indirect fluorescent antibody response, difference in immunogenicity between two species substanttiated by difference in IFA response after challenge, reinoculation with Eimeria maxima indicated that birds were immune but single infection with Eimeria acervulina did not result in solid immunity

Eimeria maxima
Kuil, H.; and Dankert-Brands, S., 1976, Vet. Parasitol., v. 2 (3), 293-298
Eimeria tenella, E. maxima, development of indirect fluorescent antibody titers in infected chickens fed rations medicated with metichlorpindol and/or methylbenzoquate, concluded that development of parasite is necessary to stimulate host to produce circulating fluorescent antibodies

Eimeria maxima, illus.
Long, P. L.; et al., 1976, Folia Vet. Latina, v. 6 (3), 201-217
Eimeria spp., avian, procedure, key and laboratory methods for identification, pathology, experimental infection procedures, factors affecting development, instructive review

Eimeria maxima
Long, P. L.; and Millard, B. J., 1976, Parasitology, v. 73 (3), 327-336
Eimeria praecox, E. maxima, E. acervulina, site finding and site specificity in chickens

Eimeria maxima
Long, P. L.; and Millard, B. J., 1976, Ztschr. Parasitenk., v. 48 (3-4), 287-290
Eimeria spp., immunized chickens, oocysts produced after corticosteroid treatment in some birds, possibly endogenous cycle continues or remains dormant; possible means of diagnosis of occult infection in apparently immune hosts

Eimeria maxima
Long, P. L.; and Millard, B. J., 1977, Avian Path., v. 6 (1), 77-92
chickens immunised against Eimeria using repeated low level infections with attenuated strains of E. tenella and E. acervulina var. mivati alone or together with E. brunetti, E. maxima, E. necatrix

Eimeria maxima
Miller, B. M.; et al., 1977, Poultry Science, v. 56 (6), 2039-2044
Eimeria spp., chickens, efficacy and tolerance of two 6-amino-9-(substituted benzyl) purines, good results

Eimeria maxima
Mylari, B. L.; et al., 1977, J. Med. Chem., v. 20 (4), 475-483
Eimeria spp. in chickens, 1-benzyl-6-azauracils as potential new anticoccidials, comparison trials with several experimental and commercial anticoccidials

Eimeria maxima
Norton, C. C.; and Hein, H. E., 1976, Parasitology, v. 72 (3), 345-354
Eimeria maxima, comparison of 3 strains (Weybridge, Houghton, and a fresh field isolate), oocyst measurements, pathogenicity, oocyst production, immunity to homologous not heterologous challenge, chickens

Eimeria maxima
Oikawa, H.; et al., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 271-279
Eimeria spp., chickens, survey during 1973 neither seasonal nor regional distribution differences were observed; oocyst detection rate significantly low in chicks up to 30 days old: Japan

Eimeria maxima
Pesti, G. M.; and Combs, G. F., jr., 1976, Poultry Science, v. 55 (6), 2265-2274
Eimeria, site selectivity of various species for infection in particular regions of chick gastrointestinal tract used as tool to study loci of selenium absorption, potentiation of selenium deficiency by intestinal para-

Eimeria maxima
Reid, W. M., 1975, Avian Dis., v. 19 (4), 802-811
Eimeria spp., chicks medicated with roxarsone and zoalene produced higher oocyst counts than unmedicated controls confirming the conclusion that oocyst counts are an unsatisfactory index to anticoccidial activity

Eimeria maxima
Reid, W. M.; et al., 1977, Poultry Science, v. 56 (1), 66-71
coccidiosis, chicks, monensin, development of immunity dependent upon drug level, treatment time, and oocyst exposure

Eimeria maxima
Rollinson, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 436-437 [Demonstration]
Eimeria spp., attempted differentiation of chicken coccidia by electrophoretic variation of enzymes

Eimeria maxima
Rose, M. E., 1974, Infect. and Immun., v. 10 (4), 862-871
Eimeria tenella, E. maxima, chickens, phagocytosis of sporozoites and sporocysts in vitro by peritoneal exudate cells from immunized animals, role of antibody, species-specificity

Eimeria maxima
Rose, M. E., 1976, Vet. Rec., v. 98 (24), 481-484
poultry coccidiosis, immunity and prospects for immunoprophylaxis, review

Eimeria maxima
Rose, M. E.; and Hesketh, P., 1976, Parasitology, v. 73 (1), 25-37
Eimeria maxima, determination of life-cycle stages which induce protective immunity (second generation schizont probably most concerned), stages affected by immune response (sexual stages most susceptible), chickens

Eimeria maxima
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria maxima
Ryley, J. F.; and Hardman, L., 1977, Parasitology, v. 75 (2), xv [Abstract]
Eimeria spp., effect of vitamin K-deficient diets on infection and in particular on effectiveness of anti-coccidial drugs, concluded that use of deficient diet in screening and evaluating drugs is justified

Eimeria maxima
Shirley, M. W.; and Lee, D. L., 1977, J. Parasitol., v. 63 (2), 390-392
Eimeria spp., isoelectric focusing of enzymes, compared with starch gel electrophoresis, inter- and intraspecies differences

Eimeria maxima
Smith, R. R.; and Ruff, M. D., 1975, Poultry Science, v. 54 (6), 2081-2086
Eimeria spp., rapid technique using sodium bicarbonate, ether and centrifugation, cleaning and concentration of oocysts; storage stability of E. tenella and E. mivati oocysts at 4°C, comparison with sedimentation cleaned oocysts

Eimeria maxima Tyzzer, 1929
Singh, C. V.; Joshi, H. C.; and Shah, H. L., 1976, Pantnagar J. Research, v. 1 (1), 63-66
Eimeria spp., White Leghorn chicks, changes in blood biochemistry, possible aid in diagnosis

Eimeria maxima
Weppelman, R. M.; et al., 1977, Poultry Science, v. 56 (5), 1550-1559
Eimeria tenella, E. acervulina, E. maxima, chicks, narasin, monensin, lasalocid, comparison of efficacy, resistance and tolerance

Eimeria maxima, illus.
Witlock, D. R.; and Ruff, M. D., 1977, J. Parasitol., v. 63 (2), 193-199
5 Eimeria spp. compared, intestinal mucosal surface damage in infected chickens observed by scanning electron microscopy

Eimeria maxima
Wojton, T., 1976, Med. Wet., v. 32 (2), 78
Eimeria acervulina, E. maxima, poultry, efficacy of coccidot and sulfatyf

Eimeria maxima var. indentata
Joyner, L. P.; and Norton, C. C., 1977, Parasitology, v. 74 (2), 205-213
Eimeria maxima, genetic transfer of drug resistance, results support view that drug resistance factors are transferred at zygote formation, phenomenon took place between variants of the same species (Eimeria maxima var. indentata) but not between different species (Eimeria brunetti), transference not affected by treatment with acriflavine

Eimeria mayarii
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Rattus norvegicus: Carshalton

Eimeria mayurai Bhatia & Pande, 1966
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Pavo cristatus (feces): Delhi Zoo; Lucknow Zoo

Eimeria mayurai
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Pavo cristatus: Kazakhstan

Eimeria mecistophori n. sp., illus.
Narasimhamurti, C. C., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (4), 141-147
Eimeria mecistophori n. sp. in Mecistocephalus punctifrons (gut), life cycle: University Campus, Waltair

Eimeria media
Burgaz, I., 1973, Norwegian J. Zool., v. 21 (4), 327 [Abstract]
crossinfection of mountain hares (Lepus timidus) with tame rabbit coccidia (Eimeria stiedai, E. magna) and tame rabbits with mountain hare coccidia (E. media)

Eimeria media Kessel, 1929, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria media Kessel, 1929 of Honess, 1939, variety of
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria honessi Carvalho, 1943 emend.

Eimeria media
Lineburg, A.; and Pastuszko, J., 1972, Med. Wet., v. 28 (4), 219-220
coccidiosis, rabbits, Sulfatyf, Anticoccid

Eimeria media
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria media form honessi Carvalho, 1943
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria honessi Carvalho, 1943 emend.

Eimeria megalostiedai sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 35-37
Clemmys insculpta (intestine): Floyd Township, Floyd County, Iowa

Eimeria megalostomata
Utebaeva, M. K.; and Svanbaev, S. K., 1975, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (6), 27-32
Eimeria spp., pheasants, incidence, seasonal distribution
Phasianus colchicus mongolicus
Chrysolophus pictus
Syrmaticus reevesi
Crossoptilon tibetanum
Euplocomus nycthomerus
Phasianus colchicus lorenzi
all from Kazakhstan

Eimeria meleagrimitis
Anderson, W. I.; et al., 1977, Avian Path., v. 6 (2), 125-130
Eimeria meleagrimitis, E. gallopavonis, E. adenoeides, turkey poults, altered intestinal pH values

Eimeria meleagrimitis
Anderson, W. I.; Reid, W. M.; and McDougald, L. R., 1976, Avian Dis., v. 20 (2), 387-394
single and mixed infections of Eimeria adenoides, E. meleagrimitis, and E. gallopavonis, turkeys, monensin, drug efficacy in laboratory and floor-pen experiments, good results

Eimeria meleagrimitis
Joyner, L. P., 1973, Folia Vet. Latina, v. 3 (1), 110-123
Eimeria adenoeides, E. meleagrimitis, coccidiosis in turkeys, pathogenicity, age resistance, control, drug resistance, review

Eimeria meleagrimitis
Joyner, L. P.; and Norton, C. C., 1973, Research Vet. Sc., v. 14 (3), 279-284
Eimeria meleagrimitis known to be sulphaquinoxaline-resistant, activity of amprolium and ethopabate separately or together or in combination with sulphaquinoxaline in Mixture A.E.S., results indicate that activity of mixture was due to ethopabate component although one strain proved resistant to it also, activity of ethopabate reversed by p-aminobenzoic acid; Eimeria adenoeides, effect of amprolium, sulphaquinoxaline, and ethopabate separately and in combination, ethopabate on its own was completely ineffective

Eimeria meleagrimitis
Long, P. L.; and Millard, B. J., 1977, Avian Path., v. 6 (3), 227-233
coccidiosis, turkeys treated with sulphaquinoxaline and monensin, evaluation of infection by number of oocysts in litter, good control and improved weight gain with monensin

Eimeria meleagrimitis, illus.
Long, P. L.; Millard, B. J.; and Shirley, M. W., 1977, Parasitology, v. 75 (2), 177-182
Eimeria meleagrimitis from turkeys, oocyst measurements, sporulation and prepatent times, pathogenicity, cross-immunity tests, electrophoretic analysis of enzymes, documentation of strain variation shows that extreme caution should be used in identifying species of Eimeria from the turkey by their oocyst characters

Eimeria meleagrimitis
McDougald, L. R., 1976, Poultry Science, v. 55 (6), 2442-2447
Eimeria adenoeides, E. meleagrimitis, E. gallopavonis, turkeys, monensin, good efficacy, increased host growth

Eimeria micropteri sp. n., illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
Micropterus dolomieui
M. salmoides
(epithelium of gut's anterior portion and feces of all): Conestogo River near Conestogo and Laurel Creek near Waterloo, Ontario

Eimeria micruri sp. n., illus.
Lainson, R.; and Shaw, J. J., 1973, J. Protozool., v. 20 (3), 358-362
Micrurus filiformis (feces): Belem, Para State, north Brazil

Eimeria minima Carvalho, 1943, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Syn.: Eimeria exigua Yakimoff, 1934 of Morgan and Waller, 1940
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria minima Yakimoff, 1934 of Gill and Ray, 1960
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria hungarica Pellerdy, 1956

Eimeria mitis
Balauca, N., 1976, Arch. Exper. Vet.-Med., v. 30 (6), 903-912
Eimeria acervulina, E. necatrix, and E. mitis in cage-kept hens (exper.), mixed infections with Clostridium perfringens causing necrotic enteritis, little pathologic changes from mono-infections

E[imeria] mitis
Balauca, N.; et al., 1976, Arch. Exper. Vet.-Med., v. 30 (6), 913-923
E[imeria] acervulina, E[imeria] necatrix, and E[imeria] mitis in ground-kept hens (exper.), mixed infections with Clostridium perfringens causing necrotic enteritis, higher mortality in ground-kept hens than cage-kept hens

Eimeria mitis
Giordani, G.; Govoni, S.; and Bergami, W., 1977, Avicoltura, Bologna, v. 46 (1), 41-46
Eimeria spp., breeder hens, continuously fed sulphachloropyrazine + diaveridine + ethopabate, no undesirable effects on egg production, values statistically equal for control and coccidiostat fed birds

Eimeria mitis
Martinez Gomez, F.; et al., 1976, Arch. Zootecn., Cordoba (99), v. 25, 273-285
Eimeria necatrix, E. mitis, chickens, pronifer administered in animal feed, efficacy similar to commercial coccidiostat, need for further investigations

Eimeria mitis
Razdan, H. N.; Bansal, S. R.; and Gautam, O. P., 1976, Indian Vet. J., v. 53 (4), 255-259
saquadil, mixed infection of Eimeria tenella, E. necatrix, E. mitis, chicks, drug efficacy, good results

Eimeria mitis
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria mitis Tyzzer, 1929
Singh, C. V.; Joshi, H. C.; and Shah, H. L., 1976, Pantnagar J. Research, v. 1 (1), 63-66
Eimeria spp., White Leghorn chicks, changes in blood biochemistry, possible aid in diagnosis

Eimeria mitis
Zukovic, M.; and Brglez, J., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 11-16
coccidiosis of domestic fowl, survey, first record of Eimeria brunetti, control, review Croatia and Slovenia

Eimeria mitraria (Laveran & Mesnil, 1902), illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
redescription, prevalence
Syn.: Coccidium mitrarium Laveran & Mesnil, 1902
Chrysemys picta belli
Graptemys geographica
G. pseudogeographica
Kinosternon flavescens spooneri
(feces of all): all from Iowa

Eimeria mivati
Kolev, V.; et al., 1976, Vet. Med. Nauk., v. 13 (4), 3-12
coccidiosis, incidence in poultry at large poultry dressing combine, birds imported from France and England as possible source of infection, prophylactic measures for better disease control: Ruse, Bulgaria

Eimeria mivati
Long, P. L., 1973, Folia Vet. Latina, v. 3 (1), 89-109
Eimeria tenella, Eimeria mivati, growth in developing chick embryos and cultured cells, incubation temperature and route of inoculation, testing of anticoccidials, serial passage in embryos, review

Eimeria mivati
Reid, W. M., 1975, Avian Dis., v. 19 (4), 802-811
Eimeria spp., chicks medicated with roxarsone and zoalene produced higher oocyst counts than unmedicated controls confirming the conclusion that oocyst counts are an unsatisfactory index to anticoccidial activity

Eimeria mivati
Reid, W. M.; et al., 1977, Poultry Science, v. 56 (1), 66-71
coccidiosis, chicks, monensin, development of immunity dependent upon drug level, treatment time, and oocyst exposure

Eimeria mivati
Rollinson, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 436-437 [Demonstration]
Eimeria spp., attempted differentiation of chicken coccidia by electrophoretic variation of enzymes

Eimeria mivati
Rollinson, D., 1976, Parasitology, v. 73 (2), iv [Abstract]
Eimeria mivati, E. acervulina, electrophoretic forms of enzymes, high frequency of interspecific variation, crosses produce strains characterized by glucose phosphate isomerase-2 (characteristic of E. acervulina) and ability to passage in eggs (E. mivati), results support Long's view that E. mivati Houghton isolate belongs to species E. acervulina

Eimeria mivati
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria mivati
Ryley, J. F.; and Hardman, L., 1977, J. Protozool., v. 24 (4), 43A [Abstract]
Eimeria mivati, E. acervulina, speciation studies using drug resistance as genetic marker, immunization experiments indicate little if any cross-immunity

Eimeria mivati
Smith, R. R.; and Ruff, M. D., 1975, Poultry Science, v. 54 (6), 2081-2086
Eimeria spp., rapid technique using sodium bicarbonate, ether and centrifugation, cleaning and concentration of oocysts; storage stability of E. tenella and E. mivati oocysts at 4°C, comparison with sedimentation cleaned oocysts

Eimeria mivati Edgar & Seibold 1964, illus.
Wheat, B. E.; et al., 1976, Ztschr. Parasitenk., v. 50 (2), 125-136
Eimeria mivati, ultrastructure of macrogametogenesis

Eimeria mivati, illus.
Witlock, D. R.; and Ruff, M. D., 1977, J. Parasitol., v. 63 (2), 193-199
5 Eimeria spp. compared, intestinal mucosal surface damage in infected chickens observed by scanning electron microscopy

Eimeria miyairii Ohira, 1912
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Rattus whiteheadi: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria moschati sp. n., illus.
Duszynski, D. W.; Samuel, W. M.; and Gray, D. R., 1977, Canad. J. Zool., v. 55 (6), 990-999
Ovibos moschatus (feces): Nunivak Island, Alaska; game farm, Alberta; Bathurst Island, Northwest Territories, and breeding station, Ft. Chimo, Quebec, Canada; Bardu breeding station, Norway

Eimeria mundaragi
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria muuli Mullin, Colley and Stevens, 1972
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Chiropodomys gliroides: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria myotis
Wheat, B. E., 1975, J. Parasitol., v. 61 (5), 920-922
"described from unsporulated oocysts, and since the system of classification in the Eimeriidae developed by Hoare (1933) is based on the number of sporocysts and sporozoites present within a sporulated oocyst, these species are herein considered nomina nuda."

Eimeria nadsoni
Bejsovec, J., 1977, J. Protozool., v. 24 (4), 48A [Abstract]
persistence of Coccidia despite changing land use and decreasing numbers of host animals
Lyrurus tetrix: Bohemia

Eimeria nadsoni
Pav, J.; and Zajicek, D., 1974, Veterinarstvi, v. 24 (11), 517-520
Lyrus tetrix
Tetrao urogallus
all from CSSR

Eimeria nagpurensis Gill and Ray, 1960, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria natricis sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1975, J. Protozool., v. 22 (4), 453-454
Natrix sipedon sipedon (feces): Louisa Co., Iowa

Eimeria nebulosa n. sp., illus.
Colley, F. C.; and Else, J. G., [1976], Ann. Parasitol., v. 50 (6), 1975, 669-673
Varanus nebulosus (feces, epithelial cells of small intestine): Malaysia

Eimeria necatrix, illus.
Babu, K. S. J.; Seshadri, S. J.; and Mohiyuddeen, S., 1976, Indian Vet. J., v. 53 (1), 47-54
coccidiosis, poultry, field cases, diagnosis, pathology, relationship of infection to age and sex of host, mixed infection with E. acervulina

Eimeria necatrix
Balauca, N., 1976, Arch. Exper. Vet.-Med., v. 30 (6), 903-912
Eimeria acervulina, E. necatrix, and E. mitis in cage-kept hens (exper.), mixed infections with Clostridium perfringens causing necrotic enteritis, little pathologic changes from mono-infections

E[imeria] necatrix
Balauca, N.; et al., 1976, Arch. Exper. Vet.-Med., v. 30 (6), 913-923
E[imeria] acervulina, E[imeria] necatrix, and E[imeria] mitis in ground-kept hens (exper.), mixed infections with Clostridium perfringens causing necrotic enteritis, higher mortality in ground-kept hens than cage-kept hens

Eimeria necatrix
Chhabra, M. B.; and Ruprah, N. S., 1976, Indian Vet. J., v. 53 (4), 260-262
Eimeria tenella, Eimeria necatrix, single and mixed infections, chickens, comparative efficacy of 4 coccidiostats (Bifuran, Cycostat, Deccox, Embazin)

Eimeria necatrix (Johnson, 1930), illus.
Dubremetz, J. F., 1975, J. Protozool., v. 22 (1), 71-84
Eimeria necatrix, second generation schizonts, differentiation of daughter merozoites

E[imeria] necatrix
Dvorak, M.; et al., 1977, Veterinaria, Praha, v. 19 (1-2), 31-73
coccidiosis, poultry, preclinical tests using darvisul as standard for comparison, screening trials with sulphonamides and diaminopyrimidines; development of preparation Sulfakombin suitable for administration in water

Eimeria necatrix
El-Moukdad, A. R., 1976, Wien. Tierarztl. Monatsschr., v. 63 (12), 399-405
Eimeria necatrix, E. acervulina, E. tenella, chickens, effect of various disinfectants on oocyst development

Eimeria necatrix
El-Moukdad, A. R., 1977, Wien Tierarztl. Monatsschr., v. 64 (3), 85-91
effect of disinfectants on eggs of Ascaris suum, Toxascaris leonina, small horse strongyles and coccidia oocysts

Eimeria necatrix
Enigk, K.; and Dey-Hazra, A., 1976, Vet. Parasitol., v. 2 (2), 177-185
Eimeria necatrix, chickens, maltase and saccharase activity of intestinal mucosa during mild and severe infections

Eimeria necatrix, illus.
Fernando, M. A.; and Pasternak, J., 1977, Parasitology, v. 74 (1), 19-26
Eimeria necatrix, methods for isolating and purifying infected chick intestinal cells containing second-generation schizonts at various times post-infection

Eimeria necatrix
Fernando, M. A.; and Pasternak, J., 1977, Parasitology, v. 74 (1), 27-32
Eimeria necatrix, isolation of host-cell nuclei from chick intestinal cells infected with second-generation schizonts, DNA and RNA composition

Eimeria necatrix
Foure, N.; et al., 1974, Bull. Inform. Station Exper. Avicult. Ploufragan (Cotes-du-Nord), v. 14 (3), 51-56
Eimeria tenella, E. necatrix, E. acervulina, Clopidol + methyl-benzoquate, presence of fully sensitive or double resistant strains

Eimeria necatrix
Foure, N.; and Bennejean, G., 1974, Bull. Inform. Station Exper. Avicult. Ploufragan (Cotes-du-Nord), v. 14 (3), 57-60
Eimeria, chickens, Halofuginone, comparison with methyl-benzoquate and Clopidol, good results

Eimeria necatrix
Giordani, G.; Govoni, S.; and Bergami, W., 1977, Avicoltura, Bologna, v. 46 (1), 41-46
Eimeria spp., breeder hens, continuously fed sulphachloropyrazine + diaveridine + ethopabate, no undesirable effects on egg production, values statistically equal for control and coccidiostat fed birds

Eimeria necatrix
Gross, W. B., 1976, Poultry Science, v. 55 (4), 1508-1512
Eimeria necatrix, chickens with high levels of plasma corticosterone housed in environment of considerable social interaction had more active phagocytic defense and fewer schizonts than chickens with low levels of plasma corticosterone housed in environment with minimized social interaction

Eimeria necatrix
Hein, H. E., 1976, Exper. Parasitol., v. 40 (2), 250-260
Eimeria acervulina, E. brunetti, E. maxima, E. necatrix, immunogenic potential under precisely defined conditions which provide standards enabling comparisons between species and strains to be made, found that high level of resistance to reinfection was conferred by multiple infections with relatively low doses of oocysts

Eimeria necatrix
Karlsson, T.; and Reid, W. M., 1977, Brit. Poultry Sc., v. 18 (4), 497-501
Eimeria acervulina, E. necatrix, E. brunetti, E. tenella, chickens (exper.), immunity challenge techniques used to determine prevalence in Georgia (USA), lesion scores as main criterion to indicate exposure

Eimeria necatrix
Kohls, R. E.; Engle, A. T.; and Butters, H. E., 1972, Avian Dis., v. 16 (4), 907-914
coccidiosis in laying chickens (exper.), continuous buquinolate medication, good protection with increased egg production and egg quality, immune status at termination

Eimeria necatrix
Kolev, V.; et al., 1976, Vet. Med. Nauk., v. 13 (4), 3-12
coccidiosis, incidence in poultry at large poultry dressing combine, birds imported from France and England as possible source of infection, prophylactic measures for better disease control: Ruse, Bulgaria

Eimeria necatrix
Kumar, A.; Pandey, M. D.; and Rawat, J. S., 1976, Indian J. Poultry Sc., v. 11 (1), 11-13
Eimeria acervulina, E. necatrix, mixed infection, chickens (exper.), serum electrolyte changes, decreased percentage of total body water, dehydration

Eimeria necatrix
Kumar, A.; and Rawat, J. S., 1976, Indian J. Animal Sc., v. 45 (3), 1975, 154-156
Eimeria necatrix, E. acervulina, chickens (exper.), effect on serum enzymes, blood glucose and cholesterol

Eimeria necatrix, illus.
Long, P. L.; et al., 1976, Folia Vet. Latina, v. 6 (3), 201-217
Eimeria spp., avian, procedure, key and laboratory methods for identification, pathology, experimental infection procedures, factors affecting development, instructive review

Eimeria necatrix
Long, P. L.; and Millard, B. J., 1976, Ztschr. Parasitenk., v. 48 (3-4), 287-290
Eimeria spp., immunized chickens, oocysts produced after corticosteroid treatment in some birds, possibly endogenous cycle continues or remains dormant; possible means of diagnosis of occult infection in apparently immune hosts

Eimeria necatrix
Long, P. L.; and Millard, B. J., 1977, Avian Path., v. 6 (1), 77-92
chickens immunised against Eimeria using repeated low level infections with attenuated strains of E. tenella and E. acervulina var. mivati alone or together with E. brunetti, E. maxima, E. necatrix

Eimeria necatrix
Martinez Gomez, F.; et al., 1976, Arch. Zootecn., Cordoba (99), v. 25, 273-285
Eimeria necatrix, E. mitis, chickens, pronifer administered in animal feed, efficacy similar to commercial coccidiostat, need for further investigations

Eimeria necatrix
Miller, B. M.; et al., 1977, Poultry Science, v. 56 (6), 2039-2044
Eimeria spp., chickens, efficacy and tolerance of two 6-amino-9-(substituted benzyl) purines, good results

Eimeria necatrix
Movsesijan, M.; et al., 1975, Acta Vet. Beograd, v. 25 (5), 255-259
Eimeria spp., chickens administered single oral dose of irradiated oocysts resistant to subsequent challenge dose of non-irradiated oocysts, irradiated and non-irradiated E. tenella antigenically more potent than E. necatrix and E. brunetti and produced highest titres of specific antibodies

Eimeria necatrix
Mylari, B. L.; et al., 1977, J. Med. Chem., v. 20 (4), 475-483
Eimeria spp. in chickens, 1-benzyl-6-azauracils as potential new anticoccidials, comparison trials with several experimental and commercial anticoccidials

Eimeria necatrix
Pesti, G. M.; and Combs, G. F., jr., 1976, Poultry Science, v. 55 (6), 2265-2274
Eimeria, site selectivity of various species for infection in particular regions of chick gastrointestinal tract used as tool to study loci of selenium absorption, potentiation of selenium deficiency by intestinal parasitism

Eimeria necatrix
Razdan, H. N.; Bansal, S. R.; and Gautam, O. P., 1976, Indian Vet. J., v. 53 (4), 255-259
saquadil, mixed infection of Eimeria tenella, E. necatrix, E. mitis, chicks, drug efficacy, good results

Eimeria necatrix, illus.
Regal, D. S., 1976, Zentralbl. Vet.-Med., Reihe B, v. 23 (9), 744-763
Eimeria necatrix, chickens (exper.), developmental stages in middle section of small intestine, caecum and large intestine, light and electron microscopy

Eimeria necatrix, illus.
Regal, D. S., 1977, Zentralbl. Vet.-Med., Reihe B, v. 24 (4), 297-316
Eimeria necatrix in chickens (exper.), developmental stages of schizogony and gametogony in various sections of the intestine, light and electron microscopy

Eimeria necatrix, illus.
Regal, D. S., 1977, Zentralbl. Vet.-Med., Reihe B, v. 24 (9), 708-721
Eimeria necatrix, formation and histochemistry of inner and outer oocyst shells, light and electron microscopy

Eimeria necatrix
Reid, W. M., 1975, Avian Dis., v. 19 (4), 802-811
Eimeria spp., chicks medicated with roxarsone and zoalene produced higher oocyst counts than unmedicated controls confirming the conclusion that oocyst counts are an unsatisfactory index to anticoccidial activity

Eimeria necatrix
Reid, W. M.; et al., 1977, Poultry Science, v. 56 (1), 66-71
coccidiosis, chicks, monensin, development of immunity dependent upon drug level, treatment time, and oocyst exposure

Eimeria necatrix
Rollinson, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 436-437 [Demonstration]
Eimeria spp., attempted differentiation of chicken coccidia by electrophoretic variation of enzymes

Eimeria necatrix
Rose, M. E., 1976, Vet. Rec., v. 98 (24), 481-484
poultry coccidiosis, immunity and prospects for immunoprophylaxis, review

Eimeria necatrix
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria necatrix
Ryley, J. F.; and Hardman, L., 1977, Parasitology, v. 75 (2), xv [Abstract]
Eimeria spp., effect of vitamin K-deficient diets on infection and in particular on effectiveness of anti-coccidial drugs, concluded that use of deficient diet in screening and evaluating drugs is justified

Eimeria necatrix Johnson, 1930
Singh, C. V.; Joshi, H. C.; and Shah, H. L., 1976, Pantnagar J. Research, v. 1 (1), 63-66
Eimeria spp., White Leghorn chicks, changes in blood biochemistry, possible aid in diagnosis

Eimeria necatrix
Sokolic, A.; et al., 1976, Brit. Vet. J., v. 132 (4), 416-422
chickens, single immunizing oral dose of irradiated oocysts of Eimeria brunetti, E. necatrix, E. tenella protects against severe simultaneous challenge with these three spp.

Eimeria necatrix, illus.
Stockdale, P. H. G.; and Fernando, M. A., 1975, Research Vet. Sc., v. 19 (2), 204-208
Eimeria necatrix, chicks, development of second generation schizonts, type of cell parasitized by this stage and pathogenesis of lesions it causes

Eimeria necatrix
Turk, D. E., 1977, Poultry Science, v. 56 (5), 1763 [Abstract]
Eimeria spp., chicks, iron absorption, decreased by some species, increased during recovery period.

Eimeria necatrix
Witlock, D. R.; and Ruff, M. D., 1977, J. Parasitol., v. 63 (2), 193-199
5 Eimeria spp. compared, intestinal mucosal surface damage in infected chickens observed by scanning electron microscopy

Eimeria necatrix
Wojton, T., 1976, Med. Wet., v. 32 (12), 741-742
Eimeria necatrix, poultry, control, amprol plus and coccidot effective, anticoccid and sulfatyf were less potent

Eimeria necatrix
Zukovic, M.; and Brglez, J., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 11-16
coccidiosis of domestic fowl, survey, first record of Eimeria brunetti, control, review: Croatia and Slovenia

Eimeria neitzi spec. nov., illus.
McCully, R. M.; et al., 1970, Onderstepoort J. Vet. Research, v. 37 (1), 45-58
histopathology; development
Aepyceros melampus (uterus): Kruger National Park

Eimeria neodebliecki
Levine, N. D.; and Cechner, P. E., 1973, J. Protozool., v. 20 (4), 510-511
Eimeria spp. and Isospora canis oocysts fed to pigs, no evidence that Sarcocystis infections resulted

Eimeria neodebliecki
Levine, N. D.; Cechner, P. E.; and Meyer, R. C., 1974, Indian Vet. J., v. 51 (1), 57-59
pigs fed oocysts of porcine Eimeria spp. and of Isospora canis, "Neither Sarcocystis nor any other protozoon was found, except for Eimeria spp. in the intestinal epithelium of pigs that had been inoculated with porcine Eimeria spp."

Eimeria neodebliecki, illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria neodebliecki
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Eimeria neodebliecki
Onawunmi, O. A.; and Todd, A. C., 1976, Am. J. Vet. Research, v. 37 (6), 657-660
porcine coccidiosis (exper.), suppression and control with chlortetracycline combination, buquinolate, and lincomycin hydrochloride

E[imeria] neodebliecki, illus.
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian J. Animal Sc., v. 46 (10), 567-570
coccidia, pigs (exper.), measurements of 4 types of mature schizonts recovered from single pig (small intestine)

E[imeria] neodebliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1977, Indian J. Animal Sc., v. 47 (9), 597-600
E[imeria] debliecki, E. neodebliecki, E. perminuta, Isospora suis, pigs (feces), sporulation time: Aligarh

Eimeria neodebliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1977, Indian Vet. J., v. 54 (5), 360-362
Eimeria spp., pigs, amprolsol-MSD fully effective if administered just after second day of infection

Eimeria neoirresidua Duszynski and Marquardt, 1969, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria neoleporis Carvalho, 1942, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria neoleporis Carvalho of Pellerdy, 1954, 1965
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
not syn. of Eimeria neoleporis Carvalho, 1942

Eimeria neoleporis Carvalho, 1942
Norton, C. C.; Catchpole, J.; and Rose, M. E., 1977, Parasitology, v. 75 (1), 1-7
Eimeria stiedai, presence of oocyst residuum confirmed, differential diagnosis from Eimeria coecicola and Eimeria neoleporis

Eimeria neoleporis
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria nesokiai n. sp., illus.
Mirza, M. Y., 1975, Bull. Nat. Hist. Research Cent. Univ. Baghdad, v. 6 (1), 39-52
Nesokia indica (feces): Al-Jadiria, Baghdad

Eimeria nieschulzi Dieben 1924
Conder, G. A.; and Duszynski, D. W., 1977, J. Parasitol., v. 63 (2), 206-209
Eimeria nieschulzi, oocysts exposed to heat and/or Co-60 gamma-radiation, attenuation, reduced pathogenesis, subsequent immunity to challenge, rats

Eimeria nieschulzi Dieben, 1924, illus.
Conder, G. A.; and Duszynski, D. W., 1977, J. Protozool., v. 24 (1), 177-181
Eimeria nieschulzi, effects of heat and cobalt-60 gamma-radiation on oocyst structure and sporozoite excystation

Eimeria nieschulzi
Duszynski, D. W.; and Conder, G. A., 1977, Internat. J. Parasitol., v. 7 (1), 83-88
Eimeria nieschulzi oocyst sporulation, influence of external factors and self-regulating mechanisms

Eimeria nieschulzi, illus.
Grewal, M. S., 1975, Research Bull. Panjab Univ., v. 23 (3-4), 1972, 119-124
Rattus rattus: Chandigarh, India

Eimeria nieschulzi
Heil, R., 1974, Hamatologische Untersuchungen an Saugetieren und Kuken nach experimentellen Kokzidien-Infektionen, 137 pp.
Rattus norvegicus var. albinus, experimental infection, haemoglobin and haematocrit values, differential blood cell counts

Eimeria nieschulzi, illus.
Liburd, E. M.; Armstrong, W. D.; and Mahrt, J. L., 1973, Cellular Immunol., v. 7 (3), 444-452
Eimeria nieschulzi in rats, investigation of normal immune response and of adoptive immunity with primed thoracic duct lymphocytes

Eimeria nieschulzi
Liburd, E. M.; Pabst, H. F.; and Armstrong, W. D., 1972, Cellular Immunol., v. 5 (3), 487-489
Eimeria nieschulzi in rats as an animal model for evaluation of dialyzable transfer factor, partial but significant immunity induced by injection of transfer factor from immune syngeneic animals

Eimeria nieschulzi
Mayberry, L. F., 1973, J. Protozool., v. 20 (4), 504-505
Eimeria nieschulzi-infected rat intestine, incorporation of nucleic acid precursors

Eimeria nieschulzi, Dieben 1924, illus.
Mayberry, L. F.; and Marquardt, W. C., 1974, J. Protozool., v. 21 (4), 599-603
Eimeria nieschulzi, nucleic acid precursor incorporation by parasite and by control and parasitized rat jejunal villus epithelial cells

Eimeria nieschulzi
Mishra, G. S.; and Gonzalez, J. P., 1975, Arch. Inst. Pasteur Tunis, v. 52 (1-2), 71-87
Rattus norvegicus (intestin grele, gros intestin): Tunis, Tunisia
rat blanc (exper.)

Eimeria nieschulzi Dieben, 1924
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Rattus surifer: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria nieschulzi
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Rattus norvegicus: Carshalton

Eimeria nieschulzi
Stockdale, P. H. G., 1976, Canad. J. Zool., v. 54 (7), 1064-1068
Eimeria contorta and E. nieschulzi in rats, E. zuernii in calves, production of patent infections using oocysts, sporocysts, or sporozoites

Eimeria ninakohlyakimovae
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria ninakohlyakimovae
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria ninakohlyakimovae
Bergstrom, R. C.; and Jolley, W. R., 1977, Vet. Med. and Small Animal Clin., v. 72 (6), 1052-1053
Eimeria spp., sheep, monensin, toxicity at certain dosage levels, appears to be of little value as therapeutic agent but results suggest that it may be effective as a prophylactic coccidiostat or coccidiocide

Eimeria ninakohlyakimovae
Bergstrom, R. C.; Kinnison, J. L.; and Werner, B. A., 1977, Am. J. Vet. Research, v. 38 (6), 887-888
Trichostrongylus colubriformis, Eimeria ninakohlyakimovae, changes in weight gains, feed conversion efficiency, and wool fiber diameter in lambs maintained on 2 different diets (good ration; marginal diet)

Eimeria ninakohlyakimovae
Bergstrom, R. C.; and Maki, L. R., 1977, Livestock Advis., v. 2 (8), 35-39
Eimeria ninakohlyakimovae, lambs (exper.), monensin sodium, supressed oocyst count, good feed conversion and body weight gains following treatment

Eimeria ninakohlyakimovae
Cabaret, J., 1976, Rev. Elevage et Med. Vet. Pays Trop. v. 29 (3), 221-226
sheep, goats, survey, treatment, economic importance: Kaedi area (Mauritania)

Eimeria ninakohlyakimovae
Catchpole, J.; Norton, C. C.; and Joyner, L. P., 1976, Parasitology, v. 72 (2), 137-147
4 Eimeria spp., lambs, monospecific or multispecific infections, pattern and level of oocyst production, clinical symptoms

Eimeria ninakohlyakimovae
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Eimeria ninakohlyakimovae
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

E[imeria] ninakohlyakimovae
Chroust, K., 1973, Acta Vet. Brno, v. 42 (1), 45-52
lambs naturally infected with E[imeria] ninakohlyakimovae, E. arloingi, E. parva, E. intricata, E. faurei, and E. crandallis, treated with Clopidol administered in feed, effective control with dose rates of 500 mg/kg feed for 14 days or 250 mg/kg feed for 56 days

Eimeria ninakohlyakimovae
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Ovis ammon musimon: School Forest Enterprise, University of Agriculture Brno, Krtiny

Eimeria ninaekohljakimovi
Dzerzhinskii, V. A.; and Svanbaev, S. K., 1975, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 34-36
Eimeria ninaekohljakimovi, endogenous developmental cycle in lambs, schizonts and schizogony

Eimeria ninakohlyakimovae, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

E[imeria] ninakohljakimovae
Glebezdin, V. S., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 76-80
coccidiosis, sheep, infection rates, host age, season: Turkmenistan

Eimeria ninakohlyakimovae
Khan, M. Z.; Kazam, A. R.; and Sabri, M. A., 1973, Pakistan J. Scient. Research, Lahore, v. 25 (3-4), 262-264
Eimeria ninakohlyakimovae, pathology, case report
Angora goat: Rakh Khaviewala, Pakistan

Eimeria ninakohlyakimovae
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria ninakohlyakimovae
Osman, H. S.; Restani, R.; and Canestri-Trotti, G., 1972, Parassitologia, v. 14 (1), 167-173
sheep
goats
(feces of all): all from Somalia

Eimeria ninae-kohl-yakimovae
Pav, J., 1974, Veterinarstvi, v. 24 (11), 513-515
mufloni: Vlkov preserve

Eimeria ninakohlyakimovae
Rama, S.; Murtuza, M.; and Singh, C. D. N., 1976, Indian Vet. J., v. 53 (11), 895-896
Eimeria parva, E. ninakohlyakimovae, sheep (exper.), serum levels of calcium, magnesium and blood inorganic phosphorus, no significant changes after infection

Eimeria ninakohlyakimovae, illus.
Rama, S. P.; Singh, C. D. N.; and Sinha, B. K., 1977, Indian J. Animal Sc., v. 47 (11), 735-738
Eimeria parva, sheep (exper.), mixed infections with Eimeria ninakohlyakimovae, pathology

Eimeria ninakohlyakimovae
Rivellini, P.; Fenizia, D.; and Rania, U., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 494-498
incidence, sheep: Campania and Lucania

Eimeria ninakohlyakimovae
Rollinson, D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21-22 [Demonstration]
Eimeria spp. of mammals, electrophoretic techniques to differentiate spp. by variations in mobility of enzymes

Eimeria ninaekohlyakimovae
Taylor, S. M.; et al., 1973, Vet. Rec., v. 93 (17), 461-464
Eimeria spp., lambs kept on straw bedding vs. expanded metal flooring showed higher fecal oocyst counts, diarrhea, greater weight loss, and some fatalities; causal agents of disease appear to be E. ninaekohlyakimovae and/or E. arloingi with concurrent increase of intestinal Cl[ostridium] welchii

Eimeria nocens
Szalay, G.; and Sari, I., 1976, Magy. Allat. Lapja, v. 98, v. 31 (2), 95-96
Eimeria nocens, E. anseris, large scale goose farms, control with Sulfamix

Eimeria noelleri
Gill, H. S., 1976, Indian Vet. J., v. 53 (11), 897-898
Camelus dromedarius (feces): States of Punjab, Haryana, and Rajasthan

Eimeria nonbrumpti Levine, 1953
Cerna, Z., 1973, J. Protozool., v. 20 (4), 535-536
Dendrocopus major pinetorum (feces)

Eimeria notati Colley and Mullin, 1971
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Callosciurus notatus: Bukit Lanjan Forest Reserve. West Malaysia

Eimeria ontarioensis
Davidson, W. R., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 211-217
epizootiologic and pathologic study of endoparasites of selected populations of gray squirrels
Sciurus carolinensis (small intestine): southeastern United States

Eimeria ontarioensis
Forrester, D. J.; et al., 1977, J. Parasitol., v. 63 (6), 1045
Sciurus carolinensis (feces): Jacksonville, Duval County, northern Florida

Eimeria ontarioensis Lee and Dorney 1971
Joseph, T., 1973, J. Protozool., v. 20 (4), 509
Sciurus niger rufiventer: northern Indiana

Eimeria oomingmakensis sp. n., illus.
Duszynski, D. W.; Samuel, W. M.; and Gray, D. R., 1977, Canad. J. Zool., v. 55 (6), 990-999
Ovibos moschatus (feces): Bathurst Island, Northwest Territories; Bardu breeding station, Norway

Eimeria otomyis n. sp., illus.
De Vos, A. J.; and Dobson, Lynne D., 1970, Onderstepoort J. Vet. Research, v. 37 (4), 185-189
Otomys irroratus (intestine): near the Veterinary Research Institute, Onderstepoort, Pretoria district

Eimeria ovibovis sp. n., illus.
Duszynski, D. W.; Samuel, W. M.; and Gray, D. R., 1977, Canad. J. Zool., v. 55 (6), 990-999
Ovibos moschatus (feces): game farm, Alberta; Bathurst Island, Northwest Territories, and breeding station, Ft. Chimo, Quebec, Canada; Bardu breeding station, Norway

Eimeria ovina
Catchpole, J.; Norton, C. C.; and Joyner, L. P., 1976, Parasitology, v. 72 (2), 137-147
4 Eimeria spp., lambs, monospecific or multispecific infections, pattern and level of oocyst production, clinical symptoms

Eimeria ovina
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Eimeria ovina
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

Eimeria ovina (= E. arloingi)
Chapman, H.D.; Lewis, J.A.; and Searle, R.M., 1973, Research Vet. Sc., v. 14 (3), 369-375
growth of lambs during acquisition of natural infection of coccidia acquired at pasture, no difference in growth between untreated and amprolium-sulphadimidine treated groups; absence of effect on growth of identical medication regime in relatively coccidia-free lambs indicates lack of toxicity of drugs; results demonstrate that natural subclinical infections of coccidia are not important under this husbandry system

Eimeria ovina Levine and Ivens, 1970, illus.
Duszynski, D. W.; Samuel, W. M.; and Gray, D. R., 1977, Canad. J. Zool., v. 55 (6), 990-999
redescription
Ovibos moschatus (feces): Nunivak Island, Alaska; game farm, Alberta; Bathurst Island, Northwest Territories, and breeding station, Ft. Chimo, Quebec, Canada; Bardu breeding station, Norway
Ovis canadensis (feces): Montana

Eimeria ovina, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

Eimeria pachylepyron Colley and Mullin, 1972
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Nycticebus coucang: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria pacifica Ormsbee, 1939, illus.
Fisher, J. W.; and Wacha, R. S., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 226-227
Phasianus colchicus (feces): private game farm, Jasper County, Iowa

Eimeria pacifica
Utebaeva, M. K.; and Svanbaev, S. K., 1975, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (6), 27-32
Eimeria spp., pheasants, incidence, seasonal distribution
Phasianus colchicus mongolicus
Chrysolophus pictus
Syrmaticus reevesi
Crossoptilon tibetanum
Euplocomus nycthomerus
Phasianus colchicus lorenzi
all from Kazakhstan

Eimeria padulensis n. sp., illus.
Romero Rodriguez, J.; and Lizcano Herrera, J., 1974, Rev. Iber. Parasitol., v. 34 (1-2), 29-31
Alectoris rufa (intestino delgado)

Eimeria pahangi Colley and Mullin, 1971
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Callosciurus notatus: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria pallida
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria pallida
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria pallida
Bergstrom, R. C.; and Jolley, W. R., 1977, Vet. Med. and Small Animal Clin., v. 72 (6), 1052-1053
Eimeria spp., sheep, monensin, toxicity at certain dosage levels, appears to be of little value as therapeutic agent but results suggest that it may be effective as a prophylactic coccidiostat or coccidiocide

Eimeria pallida
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Eimeria pallida
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

Eimeria pallida, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

Eimeria pallida
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria panda
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (1), 65-77
roe deer: Czechoslovakia

Eimeria panda
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Capreolus capreolus: School Forest Enterprise, University of Agriculture Brno, Krtiny

Eimeria paradisaeai sp. n., illus.
Varghese, T., 1977, J. Parasitol., v. 63 (5), 887-889
Paradisaea raggiana (feces): Wau Highlands, Papua New Guinea

Eimeria parahi Pande et al., 1970
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Axis porcinus (feces): Lucknow Zoo

Eimeria parva
Ajayi, J. A., 1976, Parasitology, v. 72 (3), 335-343
8 ovine Eimeria spp., effects of temperature and Aureo-S-700 on sporulation, viability, and infectivity of oocysts

Eimeria parva
Ajayi, J. A.; and Todd, A. C., 1977, Brit. Vet. J., v. 133 (2), 166-174
Eimeria spp., lambs, two levels of aureomycin-sulphamethazine supplement, smaller dose permitted resistance to be acquired, higher dose suppressed coccidial growth and multiplication but did not permit development of resistance

Eimeria parva
Baker, N. F.; Walters, G. T.; and Fisk, R. A. 1972, Am. J. Vet. Research, v. 33 (1), 83-86
Eimeria arloingi, E. parva, E. intricata, amprolium, control in feedlot lambs

Eimeria parva
Bergstrom, R. C.; and Jolley, W. R., 1977, Vet. Med. and Small Animal Clin., v. 72 (6), 1052-1053
Eimeria spp., sheep, monensin, toxicity at certain dosage levels, appears to be of little value as therapeutic agent but results suggest that it may be effective as a prophylactic coccidiostat or coccidiocide

Eimeria parva
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 1-6
lambs under a husbandry system with crowded indoor housing, performance in 2 separate trials: growth during acquisition of natural mixed coccidial infections; pathogenicity of an artificial infection of primarily Eimeria ninakohlyakimovae, growth, blood changes

Eimeria parva
Chapman, H. D., 1974, Research Vet. Sc., v. 16 (1), 7-11
course of mixed coccidial infection acquired by lambs born at pasture, immunity to challenge following this natural infection and following artificial infection (with primarily Eimeria ninakohlyakimovae), betamethasone administration caused increases in oocyst output

E[imeria] parva
Chroust, K., 1973, Acta Vet. Brno, v. 42 (1), 45-52
lambs naturally infected with E[imeria] ninakohlyakimovae, E. arloingi, E. parva, E. intricata, E. faurei, and E. crandallis, treated with Clopidol administered in feed, effective control with dose rates of 500 mg/kg feed for 14 days or 250 mg/kg feed for 56 days

Eimeria parva
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Ovis ammon musimon: School Forest Enterprise, University of Agriculture Brno, Krtiny

Eimeria parva, illus.
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

E[imeria] parva
Glebezdin, V. S., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 76-80
coccidiosis, sheep, infection rates, host age, season: Turkmenistan

Eimeria parva
Mossalam, I., 1972, Parasitol. Hungar., v. 5, 375-382
incidence survey in sheep (feces) and morphological comparisons with data described previously: Egypt

Eimeria parva
Osman, H. S.; Restani, R.; and Canestri-Trotti, G., 1972, Parassitologia, v. 14 (1), 167-173
sheep
goats
(feces of all): all from Somalia

Eimeria parva
Pav, J., 1974, Veterinarstvi, v. 24 (11), 513-515
mufloni: Vlkov preserve

Eimeria parva
Rama, S.; Murtuza, M.; and Singh, C. D. N., 1976, Indian Vet. J., v. 53 (11), 895-896
Eimeria parva, E. ninakohlyakimovae, sheep (exper.), serum levels of calcium, magnesium and blood inorganic phosphorus, no significant changes after infection

Eimeria parva, illus.
Rama, S. P.; Singh, C. D. N.; and Sinha, B. K., 1977, Indian J. Animal Sc., v. 47 (11), 735-738
Eimeria parva, sheep (exper.), mixed infections with Eimeria ninakohlyakimovae, pathology

Eimeria parva
Rivellini, P.; Fenizia, D.; and Rania, U., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 494-498
incidence, sheep: Campania and Lucania

Eimeria parva
Taylor, S. M.; et al., 1973, Vet. Rec., v. 93 (17), 461-464
Eimeria spp., lambs kept on straw bedding vs. expanded metal flooring showed higher fecal oocyst counts, diarrhea, greater weight loss, and some fatalities; causal agents of disease appear to be E. ninaekohlyakimovae and/or E. arloingi with concurrent increase of intestinal Cl[ostridium] welchii type A

Eimeria parva
Volf, K.; and Volfova, M., 1974, Veterinarstvi, v. 24 (3), 125-126
srnci zvere: Trebic District

Eimeria pattersoni
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Lyruris tetrix: Kazakhstan

Eimeria paulistana da Fonseca, 1933, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria pavonis Mandal, 1965
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Pavo cristatus (feces): Delhi Zoo

Eimeria pelandoki n. sp., illus.
Mullin, S. W.; and Colley, F. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 441-443
Tragulus javanicus (intestinal contents): Subang Forest Preserve, Selangor, West Malaysia

Eimeria pelecani sp. n., illus.
Courtney, C. H.; and Ernst, J. V., 1975, J. Parasitol., v. 61 (6), 1081-1082
Pelecanus occidentalis carolinensis (feces): Bird Keys, Lee County, Florida

Eimeria pellerdyi
Gill, H. S., 1976, Indian Vet. J., v. 53 (11), 897-898
Camelus dromedarius (feces): States of Punjab, Haryana, and Rajasthan

Eimeria pellita
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria perforans
Fitzgerald, P. R., 1976, J. Protozool., v. 23 (1), 150-154
Eimeria stiedai, E. perforans, E. magna, rabbits, control by administration of antibiotics A204 sodium, A28695A, and A204 normal propyl in pelleted feed

Eimeria perforans (Leuckart, 1879) Sluiter and Swellengrebel, 1912, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria perforans
Lineburg, A.; and Pastuszko, J., 1972, Med. Wet., v. 28 (4), 219-220
coccidiosis, rabbits, Sulfatyf, Anticoccid

Eimeria perforans
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria perforans
Wosu, N. J.; et al., 1977, Lab. Animal Sc., v. 27 (2), 210-216
Encephalitozoon cuniculi, rabbits (exper.), diagnosis by immunofluorescence and intradermal test, no cross reactions between E. cuniculi and experimentally induced Toxoplasma gondii, Eimeria perforans, or E. stiedai

Eimeria perminuta, illus.
Martinez-Gomez, F.; Hernandèz-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

E[imeria] perminuta, illus.
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian J. Animal Sc., v. 46 (10), 567-570
coccidia, pigs (exper.), measurements of 4 types of mature schizonts recovered from single pig (small intestine)

E[imeria] perminuta
Upadhyay, A. N.; and Ahluwalia, S. S., 1977, Indian J. Animal Sc., v. 47 (9), 597-600
E[imeria] debliecki, E. neodebliecki, E. perminuta, Isospora suis, pigs (feces), sporulation time: Aligarh

Eimeria perminuta
Upadhyay, A. N.; and Ahluwalia, S. S., 1977, Indian Vet. J., v. 54 (5), 360-362
Eimeria spp., pigs, amprolsol-MSD fully effective if administered just after second day of infection

Eimeria phasiani Tyzzer, 1929
Bejsovec, J., 1973, J. Protozool., v. 20 (4), 535
seasonal incidence
Phasianus colchicus: Central Bohemia

Eimeria phasiani Tyzzer, 1929
Bejsovec, J., 1974, J. Protozool., v. 21 (3), 454 [Abstract]
Eimeria colchici, E. phasiani, incidence of oocysts in various biotopes of an agricultural landscape over a 6-year period: Central Bohemia

Eimeria phasiani
Bejsovec, J., 1974, J. Protozool., v. 21 (3), 454 [Abstract]
Eimeria spp. of wild galliform birds, failure of transmission to domestic galliform birds even in localities with prolonged and intensive contact or by experimental infection: Czechoslovakia

Eimeria phasiani Tyzzer, 1929, illus.
Fisher, J. W.; and Wacha, R. S., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 226-227
Phasianus colchicus (feces): private game farm, Jasper County, Iowa

Eimeria phasiani, illus.
Norton, C. C., 1976, Folia Vet. Latina, v. 6 (3), 218-238
pheasants, description, review of literature: United Kingdom

Eimeria phasiani
Utebaeva, M. K.; and Svanbaev, S. K., 1975, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (6), 27-32
Eimeria spp., pheasants, incidence, seasonal distribution
Phasianus colchicus mongolicus
Chrysolophus pictus
Syrmaticus reevesi
Crossoptilon tibetanum
Euplocomus nycthomerus
Phasianus colchicus lorenzi
all from Kazakhstan

Eimeria phasiani
Zaprianov, M., 1976, Vet. Med. Nauki, v. 13 (5), 78-83
Eimeria spp. of domestic Alectoris graeca cypriates (feces), highest incidence in birds up to two months of age and in May-June

Eimeria pintoensis da Fonseca, 1932, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria piriformis Kotlan and Pospesch, 1934, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Syn.: Eimeria piriformis Marotel and Guilhon, 1941 non E. piriformis Lubimov, 1934
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria piriformis Kheisin, 1948 of Pellerdy, 1953
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria intestinalis Kheisin, 1948

Eimeria piriformis Marotel and Guilhon, 1941 non E. piriformis Lubimov, 1934
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria piriformis Kotlan and Pospesch, 1934

Eimeria piriformis, illus.
Romero Rodriguez, J.; and Lizcano Herrera, J., 1972, Rev. Iber. Parasitol., v. 32 (3-4), 199-206

Eimeria piriformis
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria plecoti
Wheat, B. E., 1975, J. Parasitol., v. 61 (5), 920-922
"described from unsporulated oocysts, and since the system of classification in the Eimeriidae developed by Hoare (1933) is based on the number of sporocysts and sporozoites present within a sporulated oocyst, these species are herein considered nomina nuda."

Eimeria poecilogyri Carini, 1933, illus.
Lainson, R.; and Shaw, J. J., 1973, J. Protozool., v. 20 (3), 358-362
redescription
Leimadophis poecilogyrus (small intestine): Serra dos Carajas, Para State, north Brazil

Eimeria polita
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Eimeria polita (Pellerdy, 1949), illus.
Yvore, P., 1976, Rev. Med. Vet., v. 152 (1), 25-32
pigs: France

Eimeria polydaedali sp. n., illus.
Borst, G. H. A.; Peters, M. L.; and Weijerman, W., 1975, Ztschr. Parasitenk., v. 47 (3), 187-189
Varanus (Polydaedalus) niloticus (intestinal contents, feces): imported from Ghana

Eimeria ponderosa
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (1), 65-77
roe deer: Czechoslovakia

Eimeria ponderosa
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Capreolus capreolus: School Forest Enterprise, University of Agriculture Brno, Krtiny

Eimeria ponderosa
Volf, K.; and Volfova, M., 1974, Veterinarstvi, v. 24 (3), 125-126
srnci zvere: Trebic District

Eimeria porci
Levine, N. D.; and Cechner, P. E., 1973, J. Protozool., v. 20 (4), 510-511
Eimeria spp. and Isospora canis oocysts fed to pigs, no evidence that Sarcocystis infections resulted

Eimeria porci
Levine, N. D.; Cechner, P. E.; and Meyer, R. C., 1974, Indian Vet. J., v. 51 (1), 57-59
pigs fed oocysts of porcine Eimeria spp. and of Isospora canis, "Neither Sarcocystis nor any other protozoon was found, except for Eimeria spp. in the intestinal epithelium of pigs that had been inoculated with porcine Eimeria spp."

Eimeria porci, illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria porci
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Eimeria porci (Vetterling, 1965), illus.
Yvore, P., 1976, Rev. Med. Vet., v. 152 (1), 25-32
pigs: France

Eimeria poudrei Duszynski and Marquardt, 1969, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria praecox, illus.
Long, P. L.; et al., 1976, Folia Vet. Latina, v. 6 (3), 201-217
Eimeria spp., avian, procedure, key and laboratory methods for identification, pathology, experimental infection procedures, factors affecting development, instructive review

Eimeria praecox
Long, P. L.; and Millard, B. J., 1976, Parasitology, v. 73 (3), 327-336
Eimeria praecox, E. maxima, E. acervulina, site finding and site specificity in chickens

Eimeria praecox
Rose, M. E., 1976, Vet. Rec., v. 98 (24), 481-484
poultry coccidiosis, immunity and prospects for immunoprophylaxis, review

Eimeria praecox
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria praecox
Shirley, M. W.; and Lee, D. L., 1977, J. Parasitol., v. 63 (2), 390-392
Eimeria spp., isoelectric focusing of enzymes, compared with starch gel electrophoresis, inter- and intraspecies differences

Eimeria pragensis
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Mus musculus: Carshalton

Eimeria pretoriensis n. sp., illus.
De Vos, A. J.; and Dobson, Lynne D., 1970, Onderstepoort J. Vet. Research, v. 37 (4), 185-189
Rhabdomys pumilio (feces, small intestine): near the Veterinary Research Institute, Onderstepoort, Pretoria district

Eimeria procera
Bejsovec, J., 1974, J. Protozool., v. 21 (3), 454 [Abstract]
Eimeria spp. of wild galliform birds, failure of transmission to domestic galliform birds even in localities with prolonged and intensive contact or by experimental infection: Czechoslovakia

Eimeria procera
Bejsovec, J., 1977, J. Protozool., v. 24 (4), 48A [Abstract]
persistence of Coccidia despite changing land use and decreasing numbers of host animals
Perdix perdix perdix: Bohemia

Eimeria procera, illus.
Romero Rodriguez, J.; and Lizcano Herrera, J., 1974, Rev. Iber. Parasitol., v. 34 (1-2), 29-31

Eimeria procera
Zaprianov, M., 1976, Vet. Med. Nauki, v. 13 (5), 78-83
Eimeria spp. of domestic Alectoris graeca cypriates (feces), highest incidence in birds up to two months of age and in May-June

Eimeria pseudemydis Lainson, 1968, illus.
Bone, L. W., 1975, J. Wildlife Dis., v. 11 (2), 290-291
description
Pseudemys scripta elegans (feces): Lonoke County, Arkansas

Eimeria pseudogeographica sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
prevalence
Graptemys pseudogeographica
Chrysemys picta belli
(feces of all): all from Iowa

Eimeria pumilioi n. sp., illus.
De Vos, A. J.; and Dobson, Lynne D., 1970, Onderstepoort J. Vet. Research, v. 37 (4), 185-189
Rhabdomys pumilio (feces, caecum, colon, small intestine): near the Veterinary Research Institute, Onderstepoort, Pretoria district

Eimeria punctata
Osman, H. S.; Restani, R.; and Canestri-Trotti, G., 1972, Parassitologia, v. 14 (1), 167-173
sheep
goats
(feces of all): all from Somalia

Eimeria pungitii sp. n., illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
Pungitius pungitius (epithelium of gut's anterior portion, feces): Matamek River, Quebec

Eimeria punjabensis Gill and Ray, 1960, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria rajasthani
Gill, H. S., 1976, Indian Vet. J., v. 53 (11), 897-898
Camelus dromedarius (feces): States of Punjab, Haryana, and Rajasthan

Eimeria reichenowi Yakimoff & Matschoulsky 1935, illus.
Courtney, C. H.; et al., 1975, J. Parasitol., v. 61 (4), 695-699
description, morphology compared with E. gruis
Grus canadensis pratensis (feces): Florida
G. canadensis tabida (feces): Florida; Arizona
G. canadensis canadensis (feces): Texas

Eimeria residualis n.sp., illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria rhabdomyis n. sp., illus.
De Vos, A. J.; and Dobson, Lynne D., 1970, Onderstepoort J. Vet. Research, v. 37 (4), 185-189
Rhabdomys pumilio (feces, caecum, colon): near the Veterinary Research Institute, Onderstepoort, Pretoria district

Eimeria robertsoni
Haupt, W.; and Hartung, J., 1977, Monatsh. Vet.-Med., v. 32 (9), 339-341
Lepus europaeus: Leipzig area

Eimeria robertsoni
Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
Lepus europaeus: Austria

Eimeria robertsoni Madsen, 1938, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria robertsoni
Samoil, H. P.; and Samuel, W. M., 1977, J. Parasitol., v. 63 (2), 203-205
Eimeria robertsoni, life cycle in Lepus americanus (nat. and exper.), failure of cross-transmission to Oryctolagus cuniculus: Alberta

Eimeria ruficaudati Gill and Ray, 1960, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria sabani Colley and Mullin, 1971
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Rattus sabanus: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria salvelini sp. n., illus.
Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
Salvelinus fontinalis (epithelium of gut's anterior portion, feces): Matamek River, Quebec

Eimeria sardari n. sp., illus.
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Muntiacus muntijaka (feces): Lucknow Zoo

Eimeria scabra
Levine, N. D.; and Cechner, P. E., 1973, J. Protozool., v. 20 (4), 510-511
Eimeria spp. and Isospora canis oocysts fed to pigs, no evidence that Sarcocystis infections resulted

Eimeria scabra
Levine, N. D.; Cechner, P. E.; and Meyer, R. C., 1974, Indian Vet. J., v. 51 (1), 57-59
pigs fed oocysts of porcine Eimeria spp. and of Isospora canis, "Neither Sarcocystis nor any other protozoon was found, except for Eimeria spp. in the intestinal epithelium of pigs that had been inoculated with porcine Eimeria spp."

Eimeria scabra, illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria scabra
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Eimeria scabra (Henry, 1931), illus.
Yvore, P., 1976, Rev. Med. Vet., v. 152 (1), 25-32
pigs: France

Eimeria sculpta Madsen, 1938, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria sculpta Madsen 1938, illus.
Romero Rodriguez, J.; and Lizcano Herrera, J., 1972, Rev. Iber. Parasitol., v. 32 (3-4), 199-206
Eimeria intestinalis, E. sculpta, morphology, first report from Oryctolagus cuniculus domestica in Spain

Eimeria semisculpta
Haupt, W.; and Hartung, J., 1977, Monatsh. Vet.-Med., v. 32 (9), 339-341
Lepus europaeus: Leipzig area

Eimeria semisculpta
Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
Lepus europaeus: Austria

Eimeria semisculpta Madsen, 1938, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria semisculpta Madsen, 1938 of Pellerdy, 1956 in part
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
as syn. of Eimeria campania Carvalho, 1943

Eimeria separata, illus.
Grewal, M. S., 1975, Research Bull. Panjab Univ., v. 23 (3-4), 1972, 119-124
Rattus rattus: Chandigarh, India

Eimeria separata
Marquardt, W. C., 1976, J. Protozool., v. 23 (2), 287-290
host-parasite interactions in Coccidia, past-president's address with emphasis on: winter coccidiosis caused by Eimeria zuernii; pathogenesis of mixed infections; genetic basis of host specificity of E. separata

Eimeria separata
Mayberry, L. F.; et al., 1975, J. Protozool., v. 22 (3), 28A [Abstract]
Eimeria separata will complete life cycle only in certain strains of mouse

Eimeria separata Becker and Hall, 1931
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Rattus muelleri
R. tiomanicus
all from Bukit Lanjan Forest Reserve, West Malaysia

Eimeria separata
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Rattus norvegicus: Carshalton

Eimeria septentrionalis
Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
Lepus europaeus: Austria

Eimeria septentrionalis Yakimoff, Matschoulsky and Spartansky, 1936, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Syn.: Eimeria exigua var. septentrionalis Yakimoff et al., 1936 of Madsen, 1938
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria sigmodontis, illus.
Ernst, J. V.; Todd, K. S., jr.; and Barnard, W. P., 1977, Internat. J. Parasitol., v. 7 (5), 373-381
Eimeria sigmodontis, description of endogenous stages
Sigmodon hispidus (mucosal epithelial cells of cecum and colon) (exper.)

Eimeria sinensis Chen, 1956, illus.
Molnar, K., 1976, Acta Vet., Budapest, v. 26 (3), 303-312
description, developmental stages, pathology
Hypophthalmichthys mollitrix
Aristichthys nobilis
(gut of all): all from pond farms of Hungary

Eimeria sipedon sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1975, J. Protozool., v. 22 (4), 453-454
Natrix sipedon sipedon (feces): Louisa Co., Iowa

Eimeria spalacis sp. n., illus.
Sayin, F.; Dincer, S.; and Meric, I., 1977, J. Protozool., v. 24 (2), 210-212
Spalax leucodon (intestine): Lalahan district, Ankara, Turkey

Eimeria spinosa Henry, 1931, illus.
Santos, S. M.; do Amaral, V.; and Reboucas, M. M., 1975, Biologico, S. Paulo, v. 41 (12), 361-363
Sus scrofa domesticus (feces): Serra Negra, state of Sao Paulo, Brazil

Eimeria spinosa (Henry, 1931), illus.
Yvore, P., 1976, Rev. Med. Vet., v. 152 (1), 25-32
pigs: France

Eimeria stefanskii
Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
Lepus europaeus: Austria

Eimeria stefanskii Pastuszko, 1961, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria stiedai
Burgaz, I., 1973, Norwegian J. Zool., v. 21 (4), 327 [Abstract]
crossinfection of mountain hares (Lepus timidus) with tame rabbit coccidia (Eimeria stiedai, E. magna) and tame rabbits with mountain hare coccidia (E. media)

Eimeria stiedai
Dennis, M. V.; Klesius, P. H.; and Dixon, C. F., 1976, J. Alabama Acad. Sc., v. 47 (3), 116 [Abstract]
Eimeria stiedai sporozoites, rabbit peritoneal macrophages, cell-mediated immune response, phagocytosis in presence or absence of serum, immune rabbit serum has no significant effect on phagocytosis

Eimeria stiedai
Fitzgerald, Paul R., 1974, J. Protozool., v. 21 (2), 336-338
Eimeria stiedai, transmission from donor to recipient rabbits by blood transfusion, data suggest that sporozoites either free in blood or inside macrophages or monocytes are the route by which E. stiedai reaches the liver

Eimeria stiedai
Fitzgerald, P. R., 1976, J. Protozool., v. 23 (1), 150-154
Eimeria stiedai, E. perforans, E. magna, rabbits, control by administration of antibiotics A204 sodium, A28695A, and A204 normal propyl in pelleted feed

Eimeria stiedai
Frandsen, J. C., 1975, J. Protozool., v. 22 (3), 20A [Abstract]
Eimeria stiedai, glucose 6-phosphate dehydrogenase, partial characterization

Eimeria stiedai (Lindemann, 1865) Kisskalt & Hartmann, 1907
Frandsen, J. C., 1976, Comp. Biochem. and Physiol., v. 54 (4B), 537-541
Eimeria stiedai, glucose 6-phosphate dehydrogenase, partial purification and some properties

Eimeria stiedai
Frandsen, J. C.; and Ennis, T. H., 1974, J. Protozool., v. 21 (3), 433-434 [Abstract]
Eimeria stiedai, properties of glucose 6-phosphate dehydrogenase

Eimeria stiedai
Heil, R., 1974, Hamatologische Untersuchungen an Saugetieren und Kuken nach experimentellen Kokzidien-Infektionen, 137 pp.
Oryctolagus cuniculus, experimental infection, haemoglobin and haematocrit values, differential blood cell counts

Eimeria stiedae, illus.
Heller, G., 1972, Protistologica, v. 8 (1), 43-51
Eimeria stiedae, structure and formation of conoid, rhoptries, and micronemes

Eimeria stiedai
Jensen, J. B.; et al., 1976, J. Parasitol., v. 62 (2), 195-198
Eimeria spp., effect of substituting various gases for CO_2 in established excystation procedure, evidence that role of CO_2 is that of allosteric effector enhancing action of reducing agent

Eimeria stiedai
Jolley, W. R.; et al., 1976, J. Parasitol., v. 62 (2), 199-202
Eimeria stiedai, E. tenella, formation of sulfhydryl groups in walls of oocysts subjected to in vitro excystation, effect of cysteine hydrochloride-CO_2 complex on oocyst wall proteins

Eimeria stiedai
Jolley, W. R.; and Nyberg, P. A., 1974, Proc. Helminth. Soc. Washington, v. 41 (2), 259-260
Eimeria tenella, E. stiedai, in vitro excystation, formation of carbon dioxide-cysteine complex in incubation fluid

Eimeria stiedae
Klesius, P. H.; et al., 1977, Exper. Parasitol., v. 41 (2), 480-490
Eimeria bovis, cattle, evidence for cell-mediated immune response shown by delayed hypersensitivity skin tests and lymphocyte blastogenesis with antigens extracted from oocysts of E. bovis and E. stiedai, cross-reactivity between two species

Eimeria stiedai
Klesius, P. H.; Kramer, T. T.; and Frandsen, J. C., 1976, Exper. Parasitol., v. 39 (1), 59-68
Eimeria stiedai, rabbits, detection of delayed hypersensitivity by skin testing with oocyst antigen extract, skin reactivity passively transferred with lymphocyte suspensions and cell-free transfer factor but not with serum from infected skin-reactive animals

Eimeria stiedae
Klesius, P. H.; and Kristensen, F., 1977, Clin. Immunol. and Immunopathol., v. 7 (2), 240-252
transfer factor, characterization of biological activity by criteria of passive transfer of delayed hypersensitivity reactivity, lymphocyte stimulation, and protective effects against bovine and rabbit coccidiosis (Eimeria bovis, Eimeria stiedae)

Eimeria stiedai (Lindemann, 1865) Kisskalt and Hartmann, 1907, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria stiedai (Lindemann, 1865) Kisskalt & Hartmann, 1907, illus.
Norton, C. C.; Catchpole, J.; and Rose, M. E., 1977, Parasitology, v. 75 (1), 1-7
Eimeria stiedai, presence of oocyst residuum confirmed, differential diagnosis from Eimeria coecicola and Eimeria neoleporis

Eimeria stiedai
Pellerdy, L.; Schrecke, W.; and Duerr, U., 1971, Parasitol. Hungar., v. 4, 115-120
Eimeria stiedai, resistance of newborn laboratory rodents (exper.) to infection

Eimeria stiedae
Rollinson, D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21-22 [Demonstration]
Eimeria spp. of mammals, electrophoretic techniques to differentiate spp. by variations in mobility of enzymes

Eimeria stiedai
Romero Rodriguez, J.; and Sanguinetti de la Torre, M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 289-300
description
Oryctolagus cuniculus domestica: Spain

Eimeria stiedai
Ryley, J. F.; Gordon, R. W.; and Stark, J. R., 1974, J. Protozool., v. 21 (4), 596-598
Eimeria bovis, E. stiedai, storage polysaccharides shown to be amylopectin

Eimeria stiedai, illus.
Senaud, J.; Chobotar, B.; and Scholtyseck, E., 1976, Tropenmed. u. Parasitol., v. 27 (2), 145-159
comparative fine structure morphology of Plasmodium cathemerium, Eimeria ferrisi, E. stiedai, Besnoitia jellisoni, and Frenkelia sp. micropores, discussion of micropore function in parasite nutrition

Eimeria stiedai
Stingl, H., 1974, Das extrahepatische Gallengangsystem des Kaninchens und seine Veranderungen unter dem Einfluss einer experimentellen Eimeria stiedai-Infektion, 76 pp.
Eimeria stiedai, rabbits, pathological changes in bile duct system

Eimeria stiedai (Lindemann, 1865) Kisskalt and Hartmann, 1907, illus.
Varga, I., 1976, Acta Vet., Budapest, v. 26 (1), 105-112
Eimeria stiedai in Oryctolagus cuniculus (exper.), successful transmission to Lepus europaeus, pathology, reisolated oocysts from L. europaeus proved infective to both L. europaeus and O. cuniculus

Eimeria stiedai, illus.
Varga, I., 1976, Magy. Allat. Lapja, v. 98, v. 31 (11), 726-730
Eimeria stiedai, experimental transmission to Lepus europaeus (gall bladder) and Oryctolagus cuniculus, both susceptible to infection, histological alterations in liver

Eimeria stiedai
Wosu, N. J.; et al., 1977, Lab. Animal Sc., v. 27 (2), 210-216
Encephalitozoon cuniculi, rabbits (exper.), diagnosis by immunofluorescence and intradermal test, no cross reactions between E. cuniculi and experimentally induced Toxoplasma gondii, Eimeria perforans, or E. stiedai

Eimeria stiedai
Yvore, P.; and Guillaume, J., 1976, Ann. Recherches Vet., v. 7 (4), 343-348
Eimeria stiedai, rabbits, disturbance in lipid and energy metabolism

Eimeria subepithelialis, illus.
Marincek, M.; and Miljkovic, G., 1972, Acta Parasitol. Iugoslavica, v. 3 (2), 79-88
Eimeria subepithelialis, cytochemical localization of alkaline phosphatase in various life-cycle stages

Eimeria subspherica Christensen, 1941
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (8), 68-70
Bos grunniens domesticus (feces): Tadzhikistan

Eimeria subspherica
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria subspherica Christensen, 1941, illus.
Ernst, J. V.; and Courtney, C. H., 1977, Proc. Helminth. Soc. Washington, v. 44 (1), 97-98
Eimeria subspherica, calves (exper.), prepatent and patent periods, redescription of sporulated oocyst

Eimeria subspherica, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves: Veliko Turnovo Okrug, Bulgaria

Eimeria subspherica
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria subsphaerica
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria suis
Levine, N. D.; and Cechner, P. E., 1973, J. Protozool., v. 20 (4), 510-511
Eimeria spp. and Isospora canis oocysts fed to pigs, no evidence that Sarcocystis infections resulted

Eimeria suis
Levine, N. D.; Cechner, P. E.; and Meyer, R. C., 1974, Indian Vet. J., v. 51 (1), 57-59
pigs fed oocysts of porcine Eimeria spp. and of Isospora canis, "Neither Sarcocystis nor any other protozoon was found, except for Eimeria spp. in the intestinal epithelium of pigs that had been inoculated with porcine Eimeria spp."

Eimeria suis, illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Eimeria suis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Eimeria suis (Noller, 1921), illus.
Yvore, P., 1976, Rev. Med. Vet., v. 152 (1), 25-32
pigs: France

Eimeria superba
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (1), 65-77
roe deer: Czechoslovakia

Eimeria superba
Dyk, V.; and Chroust, K., 1974, Acta Vet. Brno, v. 43 (2), 123-131
helminths and coccidians of Ovis ammon musimon and Capreolus capreolus, intensity variation with age of host, lack of evidence for parasite exchange between mouflons and roe deer
Capreolus capreolus: School Forest Enterprise, University of Agriculture Brno, Krtiny

Eimeria surifer Colley and Mullin, 1971
Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
Rattus surifer: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria sylvilagi Carini, 1940, illus.
Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
Coccidia of Leporidae, checklist with brief descriptions and hosts

Eimeria symmetrica sp. n., illus.
Fernando, M. A.; and Remmler, O., 1973, J. Protozool., v. 20 (1), 43-45
Gallus lafayettei (feces): north central Ceylon
G. domesticus (exper.)

Eimeria talikiensis sp. n., illus.
Veisov, A. M., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 82-85
Spalax leucodon: vicinity of Talinsk and Maralik Aniisk regions, Armenian SSR

Eimeria taterae sp. n., illus.
Mirza, M. Y.; and al-Rawas, A. Y., 1975, J. Protozool., v. 22 (1), 23-24
Tatera indica (feces): Baghdad district, Iraq

Eimeria teetari Bhatia et al., 1966
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Francolinus francolinus (feces): Lucknow Zoo

Eimeria tenella
Abu Ali, N.; et al., 1976, Vet. Parasitol., v. 1 (4), 309-316
Eimeria tenella, chickens inoculated orally vs. subcutaneously, comparison of circulating antibody response using fluorescent antibody titration

Eimeria tenella
Anderson, W. I.; et al., 1976, Avian Dis., v. 20 (4), 752-755
Eimeria tenella, chickens (exper.), lifecycle stages in bursa of Fabricius, histological examinations

Eimeria tenella
Anderson, W. I.; et al., 1977, Avian Dis., v. 21 (4), 637-641
Eimeria tenella, development of immunity in chicks experimentally infected with infectious bursal disease virus at various times before and during coccidial challenge, results indicate that viral infection prior or concurrent with Eimeria tenella immunization significantly reduced immune protection

Eimeria tenella
Anderson, W. I.; Reid, W. M.; and Johnson, J. K., 1976, Poultry Science, v. 55 (4), 1429-1435
Eimeria tenella, chickens, high environmental temperature and humidity elevated body temperatures of host impairing survival and development of some parasites resulting in decreased severity of coccidiosis, impractical as control measure because of problems of timing the heat treatment and raising the temperature in large chicken houses

Eimeria tenella, illus.
Babu, K. S. J.; Seshadri, S. J.; and Mohiyuddeen, S., 1976, Indian Vet. J., v. 53 (1), 47-54
coccidiosis, poultry, field cases, diagnosis, pathology, relationship of infection to age and sex of host

Eimeria tenella
Bajwa, R. S.; and Gill, B. S., 1977, Ann. Recherches Vet., v. 8 (2), 181-186
Eimeria tenella, progeny of irradiated oocysts, sporulation rate, pathogenicity, immunogenicity, reproductive potential, concluded that radiation effects are not transmissible to progeny

Eimeria tenella
Bajwa, R. S.; and Gill, B. S., 1977, Indian J. Animal Sc., v. 47 (3), 126-130
Eimeria tenella, chickens, amprolium in drinking water, development of immunity; treatment more effective with lighter infections, immunization more successful with heavier infections

Eimeria tenella
Bedrnik, P., 1975, J. Protozool., v. 22 (3), 58A [Abstract]
Eimeria tenella, successful propagation in tissue cultures derived from immune chickens suggests predominant role of circulating antibodies in immunity against coccidia

Eimeria tenella
Bedrnik, P., 1977, J. Protozool., v. 24 (2), Suppl., 12A-13A [Abstract]
Eimeria tenella, failure to transfer immunity with immune spleen lymphocytes in vitro or in vivo (chicks)

Eimeria tenella
Bedrnik, P.; and Jurkovic, P., 1977, J. Protozool., v. 24 (4), 48A-49A [Abstract]
Eimeria tenella, chickens, comparison of two different antigens for indirect immunofluorescent test

Eimeria tenella, illus.
Browning, R. F.; Patton, W. H.; and Lytle, C. F., 1976, Exper. Parasitol., v. 39 (2), 195-203
Eimeria tenella-infected kidney cell cultures, increased incorporation of tritium-labeled thymidine in both parasitized and unparasitized cells, apparent parasite-induced stimulation of DNA synthesis

Eimeria tenella
Cerna, Z.; Louckova, M.; and Danek, J., 1974, J. Protozool., v. 21 (3), 455 [Abstract]
Eimeria tenella, chickens, antibody formation blocked by clopidol but not by zoalene

Eimeria tenella
Chapman, H. D., 1976, Avian Path., v. 5 (4), 283-290
Eimeria tenella, E. acervulina, E. maxima, incidence in broiler house used for commercial drug trials, resistance to various anticoccidials

Eimeria tenella
Chapman, H. D., 1976, Vet. Parasitol., v. 2 (2), 187-196
Eimeria tenella Houghton strain, chickens, monensin and lasalocid unable to completely control infection at 125 ppm, control achieved at higher levels but monensin was toxic, resistance could not be developed to either drug after 12 serial passages, however serial passage in presence of monensin resulted in increase in pathogenicity

Eimeria tenella
Chapman, H. D., 1976, Vet. Parasitol., v. 1 (4), 299-308
Eimeria tenella, delayed treatment used to study mode of action of 7 anticoccidial drugs in the chicken and chicken embryo

Eimeria tenella
Chapman, H. D., 1976, Parasitology, v. 73 (3), 265-273
Eimeria tenella, factors influencing development of resistance to robenidine, chickens

Eimeria tenella
Chapman, H. D., 1976, Parasitology, v. 73 (3), 275-282
Eimeria tenella, infections in chicken embryo used to study drug resistance to decoquinate, clopidol, robenidine, and amprolium

Eimeria tenella
Chapman, H. D., 1977, Parasitology, v. 75 (2), xxvi [Abstract]
Eimeria tenella, effect of different enzymes on in vitro excystation

Eimeria tenella
Chhabra, M. B.; and Ruprah, N. S., 1976, Indian Vet. J., v. 53 (4), 260-262
Eimeria tenella, Eimeria necatrix, single and mixed infections, chickens, comparative efficacy of 4 coccidiostats (Bifuran, Cycostat, Deccox, Embazin)

Eimeria tenella
Clarke, P. L., 1977, Parasitology., v. 75 (2), iii [Abstract]
Eimeria tenella, distribution in caecum of domestic chicks

Eimeria tenella
Denek, J.; and Strossova, Z., 1977, J. Protozool., v. 24 (2), Suppl., 14A [Abstract]
field strains of Coccidia, development of resistance to clopidol: CSSR

Eimeria tenella
Doens-Juteau, O., 1974, J. Protozool., v. 21 (3), 470 [Abstract]
Eimeria tenella, macrogamete, oocyst, surface ultrastructure

Eimeria tenella
Doran, D. J., 1974, Proc. Helminth. Soc. Washington, v. 41 (2), 169-173
Eimeria tenella, production of first- and second-generation merozoites in primary cultures of chicken kidney cells, attempts to obtain development in cell culture using merozoites of the first and second generations

Eimeria tenella, illus.
Doran, D. J.; and Augustine, P. C., 1973, J. Protozool., v. 20 (5), 658-661
Eimeria tenella, comparative development from sporozoites to oocysts in primary kidney cell cultures from chickens vs. guinea fowl, partridges, pheasants, quail, and turkeys

Eimeria tenella
Doran, D. J.; and Augustine, P. C., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 126-128
Eimeria tenella, comparative oocyst production, primary cultures of chicken kidney cells, 15 media systems

Eimeria tenella
Doran, D. J.; Vetterling, J. M.; and Augustine, P. C., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 77-80
Eimeria tenella, chickens, in vivo and in vitro comparison of Wisconsin, Weybridge, and Beltsville strains, pathogenicity, oocyst production, infectivity

E[imeria] tenella
Douglas, A. W.; et al., 1977, J. Med. Chem., v. 20 (7), 939-943
4-chlorobenzaldehyde 1-(4-chlorophenyl)-4(1H)-pyridinylidene hydrazone fluorosulfonate, broad-spectrum anticoccidial activity in chickens, action similar to that of robenidine in laboratory trials with E[imeria] tenella and E[imeria] acervulina; robenidine-resistant E[imeria] tenella strain also resistant to new compound

E[imeria] tenella
Dvorak, M.; et al., 1977, Veterinaria, Praha, v. 19 (1-2), 31-73
coccidiosis, poultry, preclinical tests using darvisul as standard for comparison, screening trials with sulphonamides and diaminopyrimidines; development of preparation Sulfakombin suitable for administration in water

Eimeria tenella
El-Moukdad, A. R., 1976, Wien. Tierarztl. Monatsschr., v. 63 (12), 399-405
Eimeria necatrix, E. acervulina, E. tenella, chickens, effect of various disinfectants on oocyst development

Eimeria tenella
El-Moukdad, A. R., 1977, Wien Tierarztl. Monatsschr., v. 64 (3), 85-91
effect of disinfectants on eggs of Ascaris suum, Toxascaris leonina, small horse strongyles and coccidia oocysts

Eimeria tenella
Enigk, K.; et al., 1975, Zentralbl. Vet.-Med., Reihe B, v. 22 (8), 687-702
survival of resistant external stages of parasites during fermentation of liquid cattle manure at high temperatures

Eimeria tenella
Foure, N.; et al., 1974, Bull. Inform. Station Exper. Avicult. Ploufragan (Cotes-du-Nord), v. 14 (3), 51-56
Eimeria tenella, E. necatrix, E. acervulina, Clopidol + methyl-benzoquate, presence of fully sensitive or double resistant strains

Eimeria tenella
Foure, N.; and Bennejean, G., 1974, Bull. Inform. Station Exper. Avicult. Ploufragan (Cotes-du-Nord), v. 14 (3), 57-60
Eimeria, chickens, Halofuginone, comparison with methyl-benzoquate and Clopidol, good results

Eimeria tenella
Fu, H.-M.; and Lee, Y.-C., 1976, J. Chinese Soc. Vet. Sc., v. 2 (2), 51-55
Eimeria tenella, chickens inoculated with chemical attenuated oocysts, safe initial infection, satisfactory protection against reinfection

Eimeria tenella
Giambrone, J. J.; et al., 1977, Poultry Science, v. 56 (1), 243-246
Eimeria tenella, increased susceptibility in chicks previously exposed to infectious bursal disease virus

Eimeria tenella
Giordani, G.; Govoni, S.; and Bergami, W., 1977, Avicoltura, Bologna, v. 46 (1), 41-46
Eimeria spp., breeder hens, continuously fed sulphachloropyrazine + diaveridine + ethopabate, no undesirable effects on egg production, values statistically equal for control and coccidiostat fed birds

Eimeria tenella
Graham, D. W.; et al., 1977, Experientia, v. 33 (10), 1274-1276
Eimeria tenella, E. acervulina, anticoccidial activity of riboflavine antagonists

Eimeria tenella
Heil, R., 1974, Hamatologische Untersuchungen an Saugetieren und Kuken nach experimentellen Kokzidien-Infektionen, 137 pp.
Gallus domesticus, experimental infection, haemoglobin and haematocrit values, differential blood cell counts

Eimeria tenella
Huang, R. J.; and Lee, Y. C., 1975, J. Chinese Soc. Vet. Sc., v. 1 (2), 104-110
Eimeria tenella, chickens, immunization by use of anticoccidials

Eimeria tenella
Hubert, J.; Yvore, P.; and Kerboeuf, D., 1976, Ann. Recherches Vet., v. 7 (1), 83-90
parasite survival in liquid manure, antiparasitic action of xylene

Eimeria tenella, illus.
Jeffers, T. K., 1975, J. Parasitol., v. 61 (6), 1083-1090
Eimeria tenella, attempt to genetically alter developmental rate in order to elucidate mechanisms controlling length of prepatent period, selection for precociousness was accompanied by attenuation, lack of pathogenicity apparently due to defective second-generation schizogony

Eimeria tenella
Jeffers, T. K., 1976, Avian Dis., v. 20 (4), 649-653
Eimeria tenella, chickens, reduction of anticoccidial drug resistance by massive introduction of drug-sensitive attenuated strain in floor pens

Eimeria tenella
Jeffers, T. K., 1976, Ztschr. Parasitenk., v. 50 (3), 251-255
Eimeria tenella, decoquinate-sensitive, precocious strain crossed with decoquinate-resistant, normally developing strain, genetic recombination

Eimeria tenella
Jeffers, T. K., 1977, Poultry Science, v. 56 (5), 1725 [Abstract]
Eimeria tenella, E. maxima, E. acervulina, broiler flocks, no development of monensin resistance following long-term drug exposure

Eimeria tenella
Jensen, J. B.; et al., 1976, J. Parasitol., v. 62 (2), 195-198
Eimeria spp., effect of substituting various gases for CO_2 in established excystation procedure, evidence that role of CO_2 is that of allosteric effector enhancing action of reducing agent

Eimeria tenella
Jolley, W. R.; et al., 1976, J. Parasitol., v. 62 (2), 199-202
Eimeria stiedai, E. tenella, formation of sulfhydryl groups in walls of oocysts subjected to in vitro excystation, effect of cysteine hydrochloride-CO_2 complex on oocyst wall proteins

Eimeria tenella
Jolley, W. R.; and Nyberg, P. A., 1974, Proc. Helminth. Soc. Washington, v. 41 (2), 259-260
Eimeria tenella, E. stiedai, in vitro excystation, formation of carbon dioxide-cysteine complex in incubation fluid

Eimeria tenella
Karlsson, T.; and Reid, W. M., 1977, Brit. Poultry Sc., v. 18 (4), 497-501
Eimeria acervulina, E. necatrix, E. brunetti, E. tenella, chickens (exper.), immunity challenge techniques used to determine prevalence in Georgia (USA), lesion scores as main criterion to indicate exposure

Eimeria tenella
Karlsson, T.; and Reid, W. M., 1977, Poultry Science, v. 56 (5), 1727-1728 [Abstract]
Eimeria tenella, broilers given immunizing doses of E. tenella failed to develop immunity while being medicated with monensin; developed partial immunity if medicated with robenidine or zoalene

Eimeria tenella
Kimura, N.; et al., 1976, Poultry Science, v. 55 (4), 1375-1383
Eimeria tenella, chickens, qualitative and quantitative study on intestinal flora, bacteriological and histopathological changes in ceca, probable that proliferation of clostridia and enterobacteria retards recovery from cecal coccidiosis

Eimeria tenella
Kohls, R. E.; Engle, A. T.; and Butters, H. E., 1972, Avian Dis., v. 16 (4), 907-914
coccidiosis in laying chickens (exper.), continuous buquinolate medication, good protection with increased egg production and egg quality, immune status at termination

Eimeria tenella
Kolev, V.; et al., 1976, Vet. Med. Nauk., v. 13 (4), 3-12
coccidiosis, incidence in poultry at large poultry dressing combine, birds imported from France and England as possible source of infection, prophylactic measures for better disease control: Ruse, Bulgaria

Eimeria tenella, illus.
Kouwenhoven, B.; and Kuil, H., 1976, Vet. Parasitol., v. 2 (3), 283-292
Eimeria tenella, chickens, rabbits, demonstration of circulating antibodies by indirect immunofluorescent antibody test using sporozoites and second-stage schizonts as antigen

Eimeria tenella
Kuil, H., 1977, Trop. and Geogr. Med., v. 29 (3), 316 [Abstract]
Eimeria tenella, in vitro cultivation

Eimeria tenella
Kuil, H.; and Dankert-Brands, S., 1976, Vet. Parasitol., v. 2 (3), 293-298
Eimeria tenella, E. maxima, development of indirect fluorescent antibody titers in infected chickens fed rations medicated with metichlorpindol and/or methylbenzoquate, concluded that development of parasite is necessary to stimulate host to produce circulating fluorescent antibodies

Eimeria tenella
Lee, E. H.; et al., 1976, Vet. Rec., v. 99 (23), 466-467 [Letter]
Eimeria tenella, chickens, resistance to robenidine: Ontario

Eimeria tenella
Lee, E. H.; and Fernando, M. A., 1977, Canad. J. Comp. Med., v. 41 (4), 466-470
Eimeria tenella, robenidine-resistant strain recovered from broiler chickens infected with acute cecal coccidiosis, dose-response, infections established in chickens maintained at 4 times the recommended level of robenidine but no oocysts recovered when drug level increased eightfold; no infections found in chickens infected with sensitive strain of E. tenella and treated with robenidine

Eimeria tenella
Lee, E.-H.; Remmler, O.; and Fernando, M. A., 1977, J. Parasitol., v. 63 (1), 155-156
Eimeria tenella, results of single sporocyst infections suggest that sporozoites are sexually undifferentiated

Eimeria tenella
Lee, Y. C.; Hsieh, F. F.; and Chien, C. Y., 1976, J. Chinese Soc. Vet. Sc., v. 2 (1), 44-47
Eimeria tenella, broilers, Stenorol in feed, superior to Elancoban or Amprol-plus as prophylactic treatment, no remarkable differences among treatments in weight gain and feed consumption

Eimeria tenella, illus.
Long, P. L., 1973, Folia Vet. Latina, v. 3 (1), 89-109
Eimeria tenella, Eimeria mivati, growth in developing chick embryos and cultured cells, incubation temperature and route of inoculation, testing of anticoccidials, serial passage in embryos, review

Eimeria tenella, illus.
Long, P. L., 1974, Folia Vet. Latina, v. 4 (2), 289-300
culture in chick embryos, review

Eimeria tenella, illus.
Long, P. L.; et al., 1976, Folia Vet. Latina, v. 6 (3), 201-217
Eimeria spp., avian, procedure, key and laboratory methods for identification, pathology, experimental infection procedures, factors affecting development, instructive review

Eimeria tenella
Long, P. L.; and Millard, B. J., 1975, J. Protozool., v. 22 (3), 53A [Abstract]
Eimeria tenella embryo-adapted strain, failure to regain pathogenicity for chickens after 62 embryo passages, single immunizing dose confers protection against pathogenic strain, successful infection of duck and quail embryos

Eimeria tenella
Long, P. L.; and Millard, B. J., 1976, Ztshhr. Parasitenk., v. 48 (3-4), 287-290
Eimeria spp., immunized chickens, oocysts produced after corticosteroid treatment in some birds, possibly endogenous cycle continues or remains dormant; possible means of diagnosis of occult infection in apparently immune hosts

Eimeria tenella
Long, P. L.; and Millard, B. J., 1977, Avian Path., v. 6 (1), 77-92
chickens immunised against Eimeria using repeated low level infections with attenuated strains of E. tenella and E. acervulina var. mivati alone or together with E. brunetti, E. maxima, E. necatrix

Eimeria tenella, illus.
Long, P. L.; and Rose, M. E., 1976, Ztschr. Parasitenk., v. 48 (3-4), 291-294
Eimeria tenella, limited in vitro growth in macrophages from chicken peritoneal exudates, trophozoites and first generation schizonts developed; possible means for study of cellular immune responses

Eimeria tenella, illus.
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Eimeria tenella
Ludvik, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Sarcocystis tenella, Toxoplasma gondii, Eimeria tenella, negative-stained preparations, ultrastructure

Eimeria tenella
Luecht, I.; and Scholtyseck, E., 1976, Ztschr. Parasitenk., v. 50 (2), 208
Eimeria tenella, embryo-adapted strain, fine structure

Eimeria tenella, illus.
McDougald, L. R.; and Galloway, R. B., 1976, Exper. Parasitol., v. 40 (3), 314-319
Eimeria tenella, various anticoccidial drugs, in vitro and in vivo techniques used to observe effects on (1) viability of extracellular sporozoites, (2) penetration of sporozoites into host cells, (3) survival of intracellular sporozoites for 24 hr postinoculation

Eimeria tenella, illus.
McDougald, L. R.; and Jeffers, T. K., 1976, J. Protozool., v. 23 (4), 530-534
Eimeria tenella precocious vs. normal strains, comparative development in vitro, pathogenicity attenuation and reduction of prepatent period in precocious strain clearly resulted from omission of portion of life cycle (2nd generation schizogony)

Eimeria tenella, illus.
McDougald, L. R.; and Jeffers, T. K., 1976, Science (4236), v. 192, 258-259
Eimeria tenella, selection for precociousness resulted in strain with only one asexual generation prior to gametogony in vitro

Eimeria tenella
McLoughlin, D. K.; and Chute, M. B., 1973, Avian Dis., v. 17 (4), 717-721
Eimeria tenella, chickens (exper.), nequinate, good results against a sensitive strain and strains resistant to non-quinoline coccidiostats, development of nequinate-resistant strain, cross-resistance study

Eimeria tenella, illus.
Madden, P. A.; and Vetterling, J. M., 1977, J. Parasitol., v. 63 (4), 607-610
Eimeria tenella, development of microgametes from microgamont stage to maturity and fertilization, scanning electron microscopy

Eimeria tenella (Railliet et Lucet, 1895)
Makoova, Z.; and Jurasek, V., 1975, J. Protozool., v. 22 (3), 66A-67A [Abstract]
Eimeria tenella, chickens (exper.), blood picture

E[imeria] tenella
Manuel, M. F.; and Morales, E. G., 1975, Philippine J. Vet. Med., v. 13 (1-2), 1974, 147-155
Akiba caulleryi, chickens, clinical signs, infectivity and mortality rates, prophylactic value of sulfamonomethoxine administered in feed, good results; drug not effective against E[imeria] tenella

Eimeria tenella
Manuel, M. F.; Parawan, O. O.; and Alonso, N. V. II, 1975, Philippine J. Vet. Med., v. 13 (1-2), 1974, 137-146
Eimeria tenella, chicks, clinical signs, cecal pathology, mortality rate, therapeutic value of various water-soluble coccidiostats

Eimeria tenella
Manuel, M. F.; and Rubio, M. E. A., 1975, Philippine J. Vet. Med., v. 14 (2), 65-73
Eimeria tenella, resistance of native, Vantress broiler and white leghorn chicks compared

Eimeria tenella
Markus, M. B., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 247-248
Eimeria tenella, passage through Lumbricus terrestris does not affect infectivity for chicks, implies possibility of cat-earthworm-bird-cat cycle for Toxoplasma gondii

Eimeria tenella, illus.
Meingassner, J. G.; and Schick, R., 1977, Wien. Tierarztl. Monatsschr., v. 64 (3), 76-79
Eimeria tenella, hens, comparison of amprolium + ethopabate and clopidol for protection of chick embryos, only clopidol was effective

Eimeria tenella
Miller, B. M.; et al., 1977, Poultry Science, v. 56 (6), 2039-2044
Eimeria spp., chickens, efficacy and tolerance of two 6-amino-9-(substituted benzyl) purines, good results

Eimeria tenella
Mitrovic, M.; and Schildknecht, E. G., 1975, Poultry Science, v. 54 (3), 750-756
Eimeria tenella, chicks, efficacy of lasalocid against sulfaquinoxaline, nicarbazin, zoalene, amprolium, clopidol and 4-hydroxyquinoline resistant E. tenella strains; no resistance to lasalocid induced

Eimeria tenella
Mohammed, M. A.; Atef, M.; and Shihata, I. M., 1976, Egypt. J. Vet. Sc., v. 11 (1-2), 1974, 1-5
Eimeria tenella, poultry, efficiency of insecticides and disinfectants against oocysts

Eimeria tenella
Morisawa, Y.; Kataoka, M.; and Kitano, N., 1977, J. Med. Chem., v. 20 (4), 483-487
Eimeria tenella, synthesis and anticoccidial activity of nitropyridinecarboxamides and derivatives, chickens

Eimeria tenella
Movsesijan, M.; et al., 1975, Acta Vet. Beograd, v. 25 (5), 255-259
Eimeria spp., chickens administered single oral dose of irradiated oocysts resistant to subsequent challenge dose of non-irradiated oocysts, irradiated and non-irradiated E. tenella antigenically more potent than E. necatrix and E. brunetti and produced highest titres of specific antibodies

E[imeria] tenella
Musaev, M. A.; and Elichev, Ia. Ia., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (3), 84-93
E[imeria] tenella, chickens, infection and immunization with large doses of oocysts, changes in blood serum protein and protein fraction levels

Eimeria tenella
Mylari, B. L.; et al., 1977, J. Med. Chem., v. 20 (4), 475-483
Eimeria spp. in chickens, 1-benzyl-6-azauracils as potential new anticoccidials, comparison trials with several experimental and commercial anticoccidials

Eimeria tenella
Oettel, M.; et al., 1975, Pharmazie, v. 30 (5), 321-324
Eimeria tenella; E. acervulina, drug screening in chickens comparing 1.6-di[1-methylallyl]-dithiocarbamoylhydrazine and meticlorpindol

Eimeria tenella
Oikawa, H.; et al., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 271-279
Eimeria spp., chickens, survey during 1973 neither seasonal nor regional distribution differences were observed; oocyst detection rate significantly low in chicks up to 30 days old: Japan

Eimeria tenella, illus.
Ouellette, C. A.; Strout, R. G.; and McDougald, L. R., 1973, J. Protozool., v. 20 (1), 150-153
Eimeria tenella cultured in embryonic chick kidney cells, incorporation of tritiated cytidine, thymidine, and uridine into DNA and RNA

Eimeria tenella, illus.
Ouellette, C. A.; Strout, R. G.; and McDougald, L. R., 1974, J. Protozool., v. 21 (2), 398-400
Eimeria tenella, asexual forms cultured in vitro, dependence upon thymidylate synthetase reaction as a source of thymidylic acid for DNA synthesis

Eimeria tenella
Panitz, E., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 111-112
Eimeria tenella, chicks, megalomicin complex, anticoccidial activity, effect of structural changes of components

Eimeria tenella
Panitz, E.; and Stahl, C. A., 1977, Proc. Helminth. Soc. Washington, v. 44 (1), 102-104
Eimeria tenella, chickens, prophylactic and therapeutic activity of flutamide

Eimeria tenella
Penev, P.; Nikolova, M.; and Stefanova, M., 1975, Vet. Med. Nauk., v. 12 (9), 46-50
Eimeria tenella, infection of chickens previously infected with Marek's disease, inability to build up stable immunity against subsequent Eimeria tenella infection

Eimeria tenella
Penev, P.; and Stefanova, M., 1975, Vet. Med. Nauki, v. 12 (5), 111-115
Eimeria tenella, immunogenic ability of oocysts following treatment with gamma rays

Eimeria tenella
Pesti, G. M.; and Combs, G. F., jr., 1976, Poultry Science, v. 55 (6), 2265-2274
Eimeria, site selectivity of various species for infection in particular regions of chick gastrointestinal tract used as tool to study loci of selenium absorption, potentiation of selenium deficiency by intestinal parasitism

Eimeria tenella
Razdan, H. N.; Bansal, S. R.; and Gautam, O. P., 1976, Indian Vet. J., v. 53 (4), 255-259
saquadil, mixed infection of Eimeria tenella, E. necatrix, E. mitis, chicks, drug efficacy, good results

Eimeria tenella, illus.
Rehman, B., 1974, Pakistan J. Sc., v. 26 (1-6), 89-91
Eimeria tenella, white leghorn chickens (exper), life cycle studies, prepatent period

Eimeria tenella
Reid, W. M., 1975, Avian Dis., v. 19 (4), 802-811
Eimeria spp., chicks medicated with roxarsone and zoalene produced higher oocyst counts than unmedicated controls confirming the conclusion that oocyst counts are an unsatisfactory index to anticoccidial activity

Eimeria tenella
Reid, W. M.; et al., 1976, Poultry Science, v. 55 (4), 1436-1441
Eimeria tenella, E. acervulina, chickens, high environmental temperatures, reduction of anticoccidial feed intake, results suggest decreases in feed and anticoccidial intake during heat-stress could seldom cause coccidiosis breaks under field conditions

Eimeria tenella
Reid, W. M.; et al., 1977, Poultry Science, v. 56 (1), 66-71
coccidiosis, chicks, monensin, development of immunity dependent upon drug level, treatment time, and oocyst exposure

Eimeria tenella
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Eimeria tenella
Rollinson, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 436-437 [Demonstration]
Eimeria spp., attempted differentiation of chicken coccidia by electrophoretic variation of enzymes

Eimeria tenella
Rollinson, D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21 [Demonstration]
Eimeria tenella, development in Coturnix c. japonica eggs to oocysts capable of sporulation

Eimeria tenella, illus.
Rose, M. E., 1974, Infect. and Immun., v. 10 (4), 862-871
Eimeria tenella, E. maxima, chickens, phagocytosis of sporozoites and sporocysts in vitro by peritoneal exudate cells from immunized animals, role of antibody, species-specificity

Eimeria tenella
Rose, M. E., 1975, J. Protozool., v. 22 (3), 55A [Abstract]
Eimeria tenella, sporozoites, interactions with phagocytic cells from normal vs. infected chickens

Eimeria tenella
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poultry coccidiosis, immunity and prospects for immunoprophylaxis, review

Eimeria tenella
Rose, M. E., 1977, Exper. Parasitol., v. 42 (1), 129-141
Eimeria tenella, chickens injected in wattle with different antigens, skin hypersensitivity measured at intervals throughout immunization by infection and also after injection of antigens in Freund's complete adjuvant, attempted transfer of skin hypersensitivity with serum or cells, correlation of skin hypersensitivity with in vitro tests

Eimeria tenella
Ruff, M. D.; and Britton, W. M., 1976, Poultry Science, v. 55 (5), 1712-1716
Eimeria acervulina, E. tenella, white Leghorn hens, severe infection, lower egg production, reduced yolk color

Eimeria tenella
Ruff, M. D.; Reid, W. M.; and Rahn, A. P., 1976, Am. J. Vet. Research, v. 37 (8), 963-967
coccidiosis in broilers, different levels of monensin, body weight gain, feed conversion, lesion score, mortality, skin and blood pigmentation

Eimeria tenella
Ruff, M. D.; Witlock, D. R.; and Smith, R. R., 1976, Exper. Parasitol., v. 39 (2), 244-251
comparison of effects of Eimeria tenella and E. acervulina infection on methionine absorption by avian intestine: importance of gut region infected; specific kinetic parameters affected; effect of intestinal pH; morphological changes in intestinal mucosa which might account for transport changes

Eimeria tenella
Ryley, J. F.; et al., 1976, Parasitology, v. 73 (3), 311-326
Eimeria tenella, separation of oocysts from feces, development of efficient procedure applicable on a large scale

Eimeria tenella
Ryley, J. F.; and Hardman, L., 1977, Parasitology, v. 75 (2), xv [Abstract]
Eimeria spp., effect of vitamin K-deficient diets on infection and in particular on effectiveness of anti-coccidial drugs, concluded that use of deficient diet in screening and evaluating drugs is justified

Eimeria tenella
Ryley, J. F.; and Wilson, R. G., 1976, Parasitology, v. 73 (2), 137-148
Eimeria tenella, practical considerations involved in screening drugs in cell culture and in the chicken, analysis for incidence of anticoccidial activity and host cell toxicity based on 11,550 compounds screened in vitro, correlation of activity in vitro and in vivo attempted, results show that screening compounds in vitro is not satisfactory or reliable alternative to screening in chickens, but in vitro studies can play a part in evaluation of active drugs

Eimeria tenella
Ryley, J. F.; and Wilson, R. G., 1976, Parasitology, v. 73 (3), 287-309
Eimeria tenella, features of anticoccidial activity of 9 older anticoccidials re-investigated both in vivo and in cell culture, methodology discussed in relation to more active and more recent anticoccidials, further experiments with robenidine reported

Eimeria tenella
Sherkov, S., 1976, Vet. Med. Nauki, v. 13 (9), 93-99
Eimeria tenella, chickens, thiamine and egg white in feed, effects on host pathomorphology and on mortality and oocyst production

Eimeria tenella
Shirley, M. W., 1975, J. Protozool., v. 22 (3), 55A-56A [Abstract]
Eimeria spp., lactate dehydrogenase, glucose phosphate isomerase, characteristic electrophoretic mobilities for different species, parasite identification and other possible applications

Eimeria tenella
Shirley, M. W.; and Lee, D. L., 1977, J. Parasitol., v. 63 (2), 390-392
Eimeria spp., isoelectric focusing of enzymes, compared with starch gel electrophoresis, inter- and intraspecies differences

Eimeria tenella
Shirley, M. W.; and Millard, B. J., 1976, Parasitology, v. 73 (3), 337-341
Eimeria tenella, single sporozoite infections in chicken embryos, results demonstrate bisexual nature of sporozoite, suggest that sexual differentiation is influenced by environmental factors, and show that true clones of Eimeria can be established from individual sporozoites

Eimeria tenella
Singh, J.; and Hussain, O., 1977, Ann. Parasitol., v. 52 (4), 397-402
Eimeria tenella, chicks with different levels of infection, comparison of therapeutic activity of sulphadimethoxine vs. bifuran and of effect of medication on development of immunity

Eimeria tenella
Smith, R. R.; and Ruff, M. D., 1975, Poultry Science, v. 54 (6), 2081-2086
Eimeria spp., rapid technique using sodium bicarbonate, ether and centrifugation, cleaning and concentration of oocysts; storage stability of E. tenella and E. mivati oocysts at 4°C, comparison with sedimentation cleaned oocysts

Eimeria tenella
Sofield, W. L.; and Strout, R. G., 1974, J. Protozool., v. 21 (3), 434 [Abstract]
Eimeria tenella, amino acids essential for asexual development in vitro

Eimeria tenella
Sokolic, A.; et al., 1976, Brit. Vet. J., v. 132 (4), 416-422
chickens, single immunizing oral dose of irradiated oocysts of Eimeria brunetti, E. necatrix, E. tenella protects against severe simultaneous challenge with these three spp.

Eimeria tenella
Stotish, R. L.; et al., 1976, J. Biol. Chem., v. 251 (2), 302-307
Eimeria tenella, glycoprotein unique to cytoplasm of unsporulated oocyst, purification and partial characterization, disappearance from cytoplasm during sporulation, possible incorporation into sporozoite membranes, studies on possible role in immunity inconclusive

Eimeria tenella, illus.
Stotish, R. L.; Simashkevich, P. M.; and Wang, C. C., 1977, J. Parasitol., v. 63 (6), 1124-1126
Eimeria tenella, separation of sporozoites, sporocysts, and oocysts by centrifugal elutriation

Eimeria tenella, illus.
Stotish, R. L.; and Wang, C. C., 1975, J. Parasitol., v. 61 (4), 700-703
Eimeria tenella, merozoites, preparation and purification in sufficient quantity and purity for biochemical study

Eimeria tenella
Svoboda, S., 1973, Acta Vet. Brno, v. 42 (1), 53-59
Eimeria tenella, white leghorn chicks, passive transfer of immunity by leucocytes obtained from immunized donors on 16th day after single application of oocysts, intraperitoneal vs. intravenous administration

Eimeria tenella
Symons, L. E. A.; and Jones, W. O., 1977, Exper. Parasitol., v. 42 (1), 194-202
Eimeria tenella-infected chickens, altered protein synthesis, compared to hosts infected with bacteria or with intestinal nematodes, evidence that loss of protein may increase apparent concentration of DNA and RNA of muscle and liver

Eimeria tenella
Tamas, T.; et al., 1977, Poultry Science, v. 56 (5), 1761 [Abstract]
Eimeria tenella, poultry, battery tests with MK-302, compared with six marketed coccidiostats; floor pen trials with birds in initial trial infected with five-species mixture of oocysts via feed; no cross resistance with other coccidiostats

Eimeria tenella
Turk, D. E., 1977, Poultry Science, v. 56 (5), 1763 [Abstract]
Eimeria spp., chicks, iron absorption, decreased by some species, increased during recovery period

Eimeria tenella
Unni, A. K. K.; and Ramakrishnan, A., 1975, Kerala J. Vet. Sc., v. 6 (1-2), 141-145
Eimeria tenella, efficacy of amprolium, chickens, good results

Eimeria tenella
Vetterling, J. M., 1973, J. Protozool., v. 20 (4), 510
Eimeria tenella will not complete life cycle in close phylogenetic relatives of Gallus gallus even though it will do so in cell culture of these birds; 1 or 2 sporozoites were able to develop to macrogametes in Alectoris graeca but did not produce patent infection; E. tenella does not develop parenterally in its definitive host or close phylogenetic relatives

Eimeria tenella, illus.
Vetterling, J. M., 1976, J. Protozool., v. 23 (1), 155-158
Eimeria tenella, host specificity, among 8 genera of gallinaceous birds only breeds of Gallus gallus developed patent infections

Eimeria tenella
Vetterling, J. M.; and Waldrop, H. R., 1976, J. Protozool., v. 23 (3), 397-402
Eimeria tenella and chick kidney cell cultures, demonstration of phosphatases, effect of fixation (type, concentration, and duration)

Eimeria tenella, illus.
Wang, C. C., 1976, Biochem. Pharmacol., v. 25 (3), 343-349
Eimeria tenella, relationship of respiration to sporulation and excystation, inhibition of respiration during sporulation and excystation by quinolone coccidiostats, amquinate-resistant strain much less subject to inhibition by quinolones, 2-hydroxynaphthoquinone coccidiostats are equally effective inhibitors against wild type and amquinate-resistant mutant

Eimeria tenella
Wang, C. C.; and Stotish, R. L., 1975, J. Parasitol., v. 61 (5), 923-927
Eimeria tenella, in vitro excystation, pancreatic chymotrypsin as essential enzyme

Eimeria tenella, illus.
Wang, C. C.; and Stotish, R. L., 1975, J. Protozool., v. 22 (3), 438-443
Eimeria tenella oocysts during sporulation, changes in nucleic acids and proteins

Eimeria tenella
Wang, C. C.; Stotish, R. L.; and Poe, M., 1975, J. Protozool., v. 22 (4), 564-568
Eimeria tenella oocysts, dihydrofolate reductase, isolation and purification, kinetic parameters and molecular weight estimate; pyrimethamine is potent inhibitor of activity of E. tenella dihydrofolate reductase but less effective inhibitor of dihydrofolate reductase of chicken liver, difference may explain in vivo therapeutic action of pyrimethamine, opposite results with methotrexate

Eimeria tenella, illus.
Wang, C. C.; Weppelman, R. M.; and Lopez-Ramos, B., 1975, J. Protozool., v. 22 (4), 560-564
Eimeria tenella oocysts, amylopectin granules, isolation, purification, composition, identification of amylopectin phosphorylase, preliminary studies on properties and regulation

Eimeria tenella
Washburn, K. W., 1975, Avian Dis., v. 19 (4), 791-801
Eimeria tenella, chickens with mutant and normal hemoglobin types compared under conditions of hematopoietic stress from coccidiosis infection and blood loss from mechanical bleeding

Eimeria tenella
Weppelman, R. M.; et al., 1977, Poultry Science, v. 56 (5), 1550-1559
Eimeria tenella, E. acervulina, E. maxima, chicks, narasin, monensin, lasalocid, comparison of efficacy, resistance and tolerance

Eimeria tenella
Weppelman, R. M.; Battaglia, J. A.; and Wang, C. C., 1977, Exper. Parasitol., v. 42 (1), 56-66
Eimeria tenella, selection and frequency of drug-resistant mutants in chickens medicated with optimal levels of drugs, mutants resistant to amquinate or glycarbylamide were isolated but this procedure failed to yield mutants resistant to amprolium, nicarbazin, robenidine, or monensin

Eimeria tenella
Weppelman, R. M.; Vandenheuvel, W. J. A.; and Wang, C. C., 1976, Lipids, v. 11 (3), 209-215
Eimeria tenella, oocysts, fatty acids and nonsaponifiable lipids, analysis by gas liquid chromatography and mass spectrometry, changes during sporulation, distribution of nonsaponifiable lipids within oocysts before and after sporulation

Eimeria tenella
Wilson, R. G., 1976, Parasitology, v. 73 (3), 283-286
Eimeria tenella, short-term application of 6 drugs in tissue culture, evaluation of coccidiocidal vs. coccidiostatic effect, necessity of monitoring technique to confirm drug withdrawal, importance of realistic level of drug dose

Eimeria tenella
Wyatt, R. D.; Ruff, M. D.; and Page, R. K., 1975, Avian Dis., v. 19 (4), 730-740
chicks, interaction of aflatoxin with Eimeria tenella infection and monensin: E. tenella and aflatoxin in combination significantly increased mortality and weight depression, and caused more severely reduced hemoglobin, packed cell volume, and plasma pigmentation; monensin sodium did not completely prevent mortality and weight depression in a mixed infection; coccidial lesion scores were less for combination of E. tenella and aflatoxicosis than for coccidiosis alone

Eimeria tenella
Yvore, P., 1974, Folia Vet. Latina, v. 4 (3), 408-425
intestinal coccidiosis (E. acervulina), caecal coccidiosis (E. tenella), comparison of nutritional effect in fowl, E. tenella causing insignificant losses, E. acervulina causing severe disease, interference with absorption

Eimeria tenella
Yvore, P.; and Hubert, J., 1977, Rec. Med. Vet., v. 153 (4), 287-291
Eimeria tenella, E. acervulina, halofuginone added to drinking water of infected chickens (exper.), curative effects compared with action of darvisul and amprolium, good results

Eimeria tenella
Zaprianov, M., 1976, Vet. Med. Nauki, v. 13 (5), 78-83
Eimeria spp. of domestic Alectoris graeca cypriates (feces), highest incidence in birds up to two months of age and in May-June

Eimeria tenella
Zukovic, M.; and Brglez, J., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 11-16
coccidiosis of domestic fowl, survey, first record of Eimeria brunetti, control, review: Croatia and Slovenia

Eimeria tenggilingi sp. n., illus.
Else, J. G.; and Colley, F. C., 1976, J. Protozool., v. 23 (4), 487-488
Manis javanica (feces): Primary forest, Ulu Jempol Valley, Negri Sembilan, Malaysia

Eimeria tetartooimia Wacha, 1973, illus.
Fisher, J. W.; and Wacha, R. S., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 226-227
Phasianus colchicus (feces): private game farm, Jasper County, Iowa

Eimeria tetradacrutata sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
prevalence
Chrysemys picta belli (bile): Iowa

Eimeria tetricis
Bejsovec, J., 1974, J. Protozool., v. 21 (3), 454 [Abstract]
Eimeria spp. of wild galliform birds, failure of transmission to domestic galliform birds even in localities with prolonged and intensive contact or by experimental infection: Czechoslovakia

Eimeria tetricis
Bejsovec, J., 1977, J. Protozool., v. 24 (2), Suppl., 13A [Abstract]
failure to experimentally infect Phasianus colchicus, Perdix perdix, or Gallus gallus f. domestica

Eimeria tetricis
  Bejsovec, J., 1977, J. Protozool., v. 24 (2), Suppl., 13A [Abstract]
    Eimeria tetricis, seasonal fluctuation in occurrence in Lyrurus tetrix: West Bohemia

Eimeria tetricis
  Bejsovec, J., 1977, J. Protozool., v. 24 (4), 48A [Abstract]
    persistence of Coccidia despite changing land use and decreasing numbers of host animals
    Lyrurus tetrix: Bohemia

Eimeria tetricis
  Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
    Lyrurus tetrix: Kazakhstan

Eimeria theileri n. sp., illus.
  De Vos, A. J.; and Dobson, Lynne D., 1970, Onderstepoort J. Vet. Research, v. 37 (4), 185-189
    Praomys natalensis (intestine): near the Veterinary Research Institute, Onderstepoort, Pretoria district

Eimeria townsendi
  Haupt, W.; and Hartung, J., 1977, Monatsh. Vet.-Med., v. 32 (9), 339-341
    Lepus europaeus: Leipzig area

Eimeria townsendi
  Kutzer, E.; and Frey, H., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (24), 480-483
    Lepus europaeus: Austria

Eimeria townsendii Carvalho, 1943, illus.
  Levine, N. D.; and Ivens, V., 1972, J. Protozool., v. 19 (4), 572-581
    Coccidia of Leporidae, checklist with brief descriptions, synonymy, and hosts

Eimeria traguli n. sp., illus.
  Mullin, S. W.; and Colley, F. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 441-443
    Tragulus javanicus (intestinal contents): Subang Forest Preserve, Selangor, West Malaysia

Eimeria traguli, illus.
  Colley, F. C.; and Mullin, S. W., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 444-452
    Eimeria traguli, events in sporulation of oocyst population

Eimeria truncata
  Wobeser, G., 1974, J. Wildlife Dis., v. 10 (3), 249-255
    Eimeria [sp.] similar to E. truncata causing renal coccidiosis in Anas platyrhynchos and A. acuta, pathologic changes, ureteral trematodes and cestodes also present in A. platyrhynchos: Saskatoon, Saskatchewan

Eimeria truttae Leger & Hesse, 1919, illus.
  Molnar, K.; and Hanek, G., 1974, J. Protozool., v. 21 (4), 489-493
    description
    Salvelinus fontinalis (epithelium of gut's anterior portion, feces): Matamek River, Quebec

Eimeria tupaiae Mullin, Colley and Stevens, 1972
  Mullin, S. W.; Colley, F. C.; and Welch, Q. B., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 93-98
    Eimeria spp., mark-release-recapture techniques used to study ecology of coccidia in Malaysian rain-forest mammals, seasonal variations, reinfection records, immunity
    Tupaia glis: Bukit Lanjan Forest Reserve, West Malaysia

Eimeria turkmenica sp. n., illus.
  Sayin, F.; Dincer, S.; and Meric, I., 1977, J. Protozool., v. 24 (2), 210-212
    Spalax leucodon (intestine): Lalahan district, Ankara, Turkey

Eimeria tuzdili sp. n., illus.
  Sayin, F.; Dincer, S.; and Meric, I., 1977, J. Protozool., v. 24 (2), 210-212
    Spalax leucodon (intestine): Lalahan district, Ankara, Turkey

Eimeria ustkamenogorica sp. nov., illus.
  Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
    Tetrastes bonasia (intestine): Leninogorska, Kazakhstan

Eimeria ventriosa
  Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
    Tetrao urogallus: Kazakhstan

Eimeria vesicostieda sp. n., illus.
  Wacha, R. S.; and Christiansen, J. L., 1977, J. Protozool., v. 24 (3), 357-359
    Trionyx spiniferus (feces): Chichaqua Wildlife Refuge, Polk Co., Iowa

Eimeria viridis (Labbe, 1893), illus.
  Sanchez-Acedo, C.; Otero, J.; and Albala-Perez, F., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 245-252
    Rhinolophus ferrum equinum: Spain

Eimeria vison
  Foreyt, W. J.; and Todd, A. C., 1976, J. Parasitol., v. 62 (3), 496
    prevalence survey in domestic mink: mink ranches, Wisconsin

Eimeria vison
  Foreyt, W. J.; Todd, A. C.; and Hartsough, G. R., 1977, Am. J. Vet. Research, v. 38 (3), 391-394
    Isospora laidlawi, Eimeria vison and Eimeria sp. in Mustela vison (exper.), anticoccidial activity of eight compounds

Eimeria vison
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Mustela]: Kazakhstan fur farms

Eimeria wenrichi Saxe, Levine, and Ivens 1960
Winchell, E. J., 1977, J. Parasitol., v. 63 (4), 756-757
Microtus breweri (ceca, large intestines, small intestines): Muskeget Island, 5 miles west of Nantucket, Massachusetts

Eimeria weybridgensis
Catchpole, J.; Norton, C. C.; and Joyner, L. P., 1976, Parasitology, v. 72 (2), 137-147
4 Eimeria spp., lambs, monospecific or multispecific infections, pattern and level of oocyst production, clinical symptoms

Eimeria weybridgensis
Euzeby, J., 1977, Rev. Med. Vet., Toulouse, v. 128 (10), 1303-1316
ovine Eimeria, brief descriptions, life cycles, pathology, modes of infection, pathogenicity, recommended treatment for clinical coccidiosis, review

Eimeria weybridgensis
Rollinson, D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21-22 [Demonstration]
Eimeria spp. of mammals, electrophoretic techniques to differentiate spp. by variations in mobility of enzymes

Eimeria wyomingensis
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria wyomingensis Huizinga and Winger 1942, illus.
Courtney, C. H.; Ernst, J. V.; and Benz, G. W., 1976, J. Parasitol., v. 62 (3), 372-376
redescription of oocysts, cattle (nat. and exper.) (feces), comparison of measurements, patent and prepatent periods

Eimeria wyomingensis, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves (jejunum, ileum): Veliko Turnovo Okrug, Bulgaria

Eimeria wyomingensis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria wyomingensis
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria yakisevi
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Tetrao urogallus: Kazakhstan

Eimeria zamenis Phisalix, 1921, illus.
Wacha, R. S.; and Christiansen, J. L., 1974, J. Protozool., v. 21 (4), 483-489
redescription, possible synonomy of E. lampropeltis Anderson et al.
Coluber constrictor flaviventris: Marion Co., Iowa
Lampropeltis triangulum triangulum: N side of Maquoketa River, Jones Co., Iowa

Eimeria zapi sp. n., illus.
Gerard, G.; Chobotar, B.; and Ernst, J. V., 1977, J. Protozool., v. 24 (3), 362-363
Zapus hudsonius (feces): 3 km WNW of Berrien Springs, Berrien County, Michigan

Eimeria zuerni Martin, 1909
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (8), 68-70
Bos grunniens domesticus (feces): Tadzhikistan

Eimeria zurni
Cabaret, J., 1976, Rev. Elevage et Med. Vet. Pays Trop. v. 29 (3), 221-226
cattle, survey, treatment, economic importance: Kaedi area (Mauritania)

Eimeria zuernii
Ciordia, H.; et al., 1977, Am. J. Vet. Research, v. 38 (9), 1335-1339
gastrointestinal parasitism of cattle on fescue pastures fertilized with broiler litter vs. NH_4NO_3, prevalence, yearly and seasonal variation; parasite burden lower in calves raised on broiler litter-fertilized pastures (where available forage was greater), no significant differences in adult cows nor in calf weight gains

Eimeria zurnii
Danielli, Y.; and Neuman, M., 1975, Refuah Vet., v. 32 (4), 94-95, 153-154
mixed parasites, cattle, good results following repeated chemotherapy: Birkat Ata

Eimeria zurnii
Julian, R. J.; Harrison, K. B.; and Richardson, J. A., 1976, Mod. Vet. Pract., v. 57 (9), 711-712, 716, 718
Eimeria zurnii, E. bovis, cattle, nervous signs, pathology, severe enteritis, possibility of toxins from coccidia: Canada

Eimeria zuernii, illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves (caecum, colon, rectum): Veliko Turnovo Okrug, Bulgaria

Eimeria zuernii
Marquardt, W. C., 1976, J. Protozool., v. 23 (2), 287-290
host-parasite interactions in Coccidia, past-president's address with emphasis on: winter coccidiosis caused by Eimeria zuernii; pathogenesis of mixed infections; genetic basis of host specificity of E. separata

Eimeria zuernii
Miner, M. L.; and Jensen, J. B., 1976, Am. J. Vet. Research, v. 37 (9), 1043-1045
Eimeria bovis, E. zuernii, calves (exper.), decoquinate administered orally in grain mix suppressed oocyst discharge and bloody diarrhea, clinical signs did not recur for 23 days after treatment was discontinued

Eimeria zuernii
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Bos taurus: Chile

Eimeria zuernii
Pasternak, J.; et al., 1977, Parasitology, v. 74 (2), 199-203
Eimeria zuernii, cattle intestinal cells infected with first-generation schizonts, nuclear hypertrophy is dissociated from DNA replication

Eimeria zuerni
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Eimeria zuernii
Stockdale, P. H. G., 1976, Canad. J. Zool., v. 54 (7), 1064-1068
Eimeria contorta and E. nieschulzi in rats, E. zuernii in calves, production of patent infections using oocysts, sporocysts, or sporozoites

Eimeria zuernii (Rivolta, 1878) Martin, 1909, illus.
Stockdale, P. H. G., 1976, Vet. Parasitol., v. 1 (4), 367-376
Eimeria zuernii, schizogony and gametogony

Eimeria zuernii
Stockdale, P. H. G., 1977, Brit. Vet. J., v. 133 (5), 471-473
Eimeria zuernii, calves (exper.), life cycle apparently contains at least two schizogonous generations before gametogony

Eimeria zuernii, illus.
Stockdale, P. H. G., 1977, Canad. J. Comp. Med., v. 41 (3), 338-344
Eimeria zuernii, calves (exper.), pathological study of enteric lesions

Eimeria zuernii
Stockdale, P. H. G.; and Niilo, L., 1976, Canad. Vet. J., v. 17 (2), 35-37
Eimeria zuernii, production of bovine coccidiosis, treatment with dexamethasone enhances reproduction by parasite, produces acute disease rather than mild infection, possibly lowers host resistance

Eimeriasis. See Coccidiosis.

Eimeriidae Minchin 1903
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
diagnosis

Eimeriidae Minchin 1903
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
diagnosis
includes: Isospora; Toxoplasma; Besnoitia; Sarcocystis; Frenkelia

Eimeriidae Minchin, 1903
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
diagnosis

Eimeriidae Minchin, 1903 fam. emend.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
definition, key to genera

Eimeriinae Wenyon, 1926 subfam. emend.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
Eimeriidae fam. emend.
definition
includes: Eimeria Schneider, 1875; Isospora Schneider 1881 gen. emend.

Eleutheroschizon Brasil, 1906
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Eleutheroschizonidae; includes: E. duboscqui Brasil, 1906 (type species); E. murmanicum Averintsev, 1908

Eleutheroschizonidae Chatton & Villeneuve, 1936
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Protococcidiorida; includes: Eleutheroschizon

Embryocola diflagellatus sp. n., illus.
Michajlow, W.; and Wita, I., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 24 (10), 1976, 615-618
Embryocola diflagellatus sp. n., developmental cycle
Eucyclops serrulatus
Megacyclops viridis
all from Jablonna near Warsaw

Encephalitozoon [sp.], illus.
Buyukmihci, N.; et al., 1977, J. Am. Vet. Med. Ass., v. 171 (4), 355-357
cat (cornea)

Encephalitozoon sp., illus.
Kemp, R. L.; and Kluge, J. P., 1975, J. Protozool., v. 22 (4), 489-491
ultrastructure, first confirmed report of microsporidan infection in birds, focal hepatic necrosis around clusters of organisms, mouse inoculation negative
Agapornis personata (renal tubular epithelial cells, hepatocytes, bile duct epithelial cells, intestinal epithelial cells): Illinois aviary, purchased from Arizona aviary

Encephalitozoon cuniculi
  Barker, R. J., 1973, J. Protozool., v. 20 (4), 524
    Encephalitozoon cuniculi, no evidence of transmission from mother to young transplacentally, perinatally, or by post-natal contact

Encephalitozoon cuniculi
  Chalupsky, J.; Vavra, J.; and Bedrnik, P., 1975, J. Protozool., v. 22 (3), 60A-61A [Abstract]
    Encephalitozoon cuniculi, laboratory animals, diagnosis with indirect fluorescent antibody test

Encephalitozoon cuniculi
  Cox, J. C., 1977, Infect. and Immun., v. 15 (2), 392-395
    Encephalitozoon cuniculi-infected rabbits, depressed IgG response and elevated IgM response to Brucella abortus as immunogen

Encephalitozoon cuniculi
  Cox, J. C.; et al., 1977, Lab. Animal Sc., v. 27 (2), 204-209
    establishment of Encephalitozoon cuniculi-free rabbit colony, serological screening by immunofluorescence, isolation of seronegative rabbits, elimination of later positive ones

Encephalitozoon cuniculi
  Huebner, J.; et al., 1973, J. Protozool., v. 20 (4), 536
    Encephalitozoon cuniculi, serodiagnosis, indirect micro-haemagglutination test not successful, microprecipitation in agar gel may have potential

Encephalitozoon cuniculi
  Moffatt, R. E.; and Schiefer, B., 1973, Lab. Animal Sc., v. 23 (2), 282-284
    Encephalitozoon cuniculi (Nosema cuniculi) causing typical encephalitic and nephritic lesions in apparently healthy guinea pigs in a closed laboratory colony, implications for experimental use of these animals

Encephalitozoon cuniculi, illus.
  Pakes, S. P.; Shadduck, J. A.; and Cali, A., 1975, J. Protozool., v. 22 (4), 481-488
    Encephalitozoon cuniculi from rabbits, fine structure and development in rabbit choroid plexus cell cultures found to be identical with that of isolates from hamster and mouse, "proposed, therefore, that the 3 organisms represent the same species, Encephalitozoon cuniculi."

Encephalitozoon (Nosema) cuniculi
  Pye, D.; and Cox, J. C., 1977, Lab. Animals, v. 11 (4), 233-234
    Encephalitozoon cuniculi, isolation from urine of infected rabbits, use of human and canine cell cultures, advantages of urine over tissue as source

Encephalitozoon cuniculi (Nosema cuniculi)
  Waller, T., 1977, Lab. Animals, v. 11 (2), 93-97
    Encephalitozoon cuniculi, rabbits, rapid diagnosis by india-ink immunoreaction, comparison with indirect fluorescent antibody and skin hypersensitivity tests

Encephalitozoon cuniculi, illus.
  Weidner, E., 1975, Ztschr. Parasitenk., v. 47 (1), 1-9
    Encephalitozoon cuniculi, parasitophorous vacuoles within host peritoneal macrophages, growth by pinocytotic mechanism, movement of substances across vacuole boundary, absence of lysosomal fusion, electron microscopy

Encephalitozoon cuniculi Levaditi et al., 1923
  Weiser, J., 1975, J. Protozool., v. 22 (3), 70A [Abstract]
    microsporidians pathogenic to man

Encephalitozoon cuniculi
  Wosu, N. J.; et al., 1977, J. Infect. Dis., v. 135 (6), 944-948
    Encephalitozoon cuniculi, complement fixation test is sensitive and capable of detecting infection in rabbits (exper.) as early as 15 days after intracerebral infection

Encephalitozoon cuniculi, illus.
  Wosu, N. J.; et al., 1977, Lab. Animal Sc., v. 27 (2), 210-216
    Encephalitozoon cuniculi, rabbits (exper.), diagnosis by immunofluorescence and intradermal test, no cross reactions between E. cuniculi and experimentally induced Toxoplasma gondii, Eimeria perforans, or E. stiedai

Endamoeba histolytica. See Entamoeba histolytica.

Endamoeba muris. See Entamoeba muris.

Endolimax nana
  Alcasid, M. L. S.; et al., 1973, N. York State J. Med., v. 73 (13), 1786-1788
    human intestinal parasites, no significant correlation with bronchial asthma, results of study with control and asthmatic groups: New York

Endolimax nana
  Garrocho-Sandoval, C.; and Torres-Ruvalcaba, A., 1977, Am. J. Clin. Path., v. 67 (6), 603-605
    human intestinal parasites, comparison study of two different methods for collecting fecal samples for diagnostic purposes

Endolimax nana
  Giboda, M.; Hoffman, F.; and Rimar, P., 1971, Ceskoslov. Pediat., v. 26 (2), 88-89
    intestinal protozoal infections in children aged 1-15 years, clinical trials with metronidazole, results and parasites eliminated varied with age and parasite: Kosice, Czechoslovakia

Endolimax nana
Markell, E. K.; and Quinn, P. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1139-1142
comparison of immediate polyvinyl alcohol fixation with delayed Schaudinn's fixation for demonstrating protozoa in feces, concluded that immediate fixation results in preservation of larger number of organisms and recommends routine use prior to transportation to laboratory

Endolimax nana
Reyes, H.; Olea, M.; and Hernandez, R., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 115-116
survey of food handlers for intestinal parasites shows 50.5% infection rate: Santiago, Chile

Endolimax nana
Sagua, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 58-60
comparison of phenol-alcohol-formalin sedimentation and polyvinyl alcohol fixative tests in diagnosis of human intestinal helminths and protozoa

Endolimax nana
Schenone, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 104-105
metronidazole used for mass therapy of human intestinal protozoa: Chile

Endolimax nana
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Endolimax nana
Torres, P.; and Navarrete, N., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 90-95
modified Telemann's and phenol-alcohol-formalin methods compared for diagnosis of common intestinal protozoa of man

Endolimax nana
Vinayak, V. K.; and Sehgal, S. C., 1976, Indian J. Med. Research, v. 64 (9), 1347-1350
human helminthic and protozoan parasites, comparison of nigrosin-methylene blue diagnostic test with formol-ether method and direct examination

Endorimospora gen. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
Eimeriidae fam. emend., Eimeriinae subfam. emend.
key, key to species completing their sexual cycle in cats, dogs and man
tod: E. miescheriana comb. nov.

Endorimospora Tadros and Laarman
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
"Endorimospora cannot be accepted in place of Sarcocystis"

Endorimospora Tadros and Laarman, 1976
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
"synonymize Endorimospora Tadros and Laarman, 1976 with Sarcocystis in part and Frenkelia in part."

Endorimospora bertrami Doflein, 1901 comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Isospora bigemina (Stiles, 1891) Luhe, 1906 [pro parte]; Sarcocystis equicanis Rommel & Geisel, 1975

Endorimospora buteonis (Henry, 1932) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
hosts, morphology, life cycle, pathogenicity
Syns.: Isospora buteonis Henry, 1932 pro parte; Frenkelia glareoli Biocca, 1968; Frenkelia clethrionomyobuteonis Rommel & Krampitz, 1975

Endorimospora cruzi (Hasselmann, 1926) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Miescheria cruzi Hasselmann, 1926 pro parte; Sarcocystis cruzi Hasselmann, 1926 pro parte; S. fusiformis Railliet, 1897 (as used by numerous authors in later literature) pro parte; Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte; Sarcocystis fusiformis (bovifelis) Heydorn, Gestrich, Mehlhorn & Rommel, 1975

Endorimospora hirsuta (Moule, 1887) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Sarcocystis hirsuta Moule, 1887; Sarcocystis fusiformis Railliet, 1897 (as used by numerous authors in later literature) pro parte; Isospora bigemina (Stiles, 1891) Luhe, 1907 pro parte; Miescheria cruzi Hasselmann, 1926 pro parte; Sarcocystis bovicanis Heydorn, Gestrich, Mehlhorn & Rommel, 1975

Endorimospora hominis (Railliet & Lucet, 1891) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Psorospermium sp. Virchow, 1860 pro parte; Coccidium bigeminum var. hominis Railliet & Lucet, 1891 pro parte; Sarcocystis fusiformis Railliet, 1897 (as used by numerous authors in later literature) pro parte; Isospora hominis (Railliet & Lucet) Wenyon, 1923 pro parte; Miescheria cruzi Hasselmann, 1926 pro parte; Sarcocystis bovihominis Heydorn, Gestrich, Mehlhorn & Rommel, 1975; Lucetina hominis (Railliet & Lucet, 1891) Henry & Leblois, 1926 pro parte

Endorimospora miescheriana (Kuhn, 1865) comb. nov. (tod)
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Synchytrium miescherianum Kuhn, 1865 [pro parte]; Coccidium bigeminum Stiles,1891 pro parte; C. bigeminum var. cani (Stiles, 1891) Railliet & Lucet, 1891 pro parte; Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte; Sarcocystis miescheriana (Kuhn, 1865) Lankester, 1882 pro parte

Endorimospora muris (Blanchard, 1885) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Miescheria muris Blanchard, 1885; Sarcocystis muris (Blanchard, 1885) Labbe, 1899; Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte

Endorimospora ovicanis Heydorn, Gestrich, Mehlhorn & Rommel, 1975 [comb. nov.]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
synonymy, key, hosts, morphology, life cycle, pathogenicity

Endorimospora putorii (Railliet & Lucet, 1891) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
hosts, morphology, life cycle, pathogenicity
Syns.: Coccidium bigeminum var. putorii Railliet & Lucet, 1891; Lucetina putorii Henry & Leblois, 1926; Isospora putorii Becker, 1934

Endorimospora sebeki nom. nov. [i.e., n. sp.]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
hosts, morphology, life cycle, pathogenicity
"The same organism as that described by Sebek (1960) in Czechoslovakia, by Tadros (1970) in Britain and by Krasnova (1971) in Russia."

Endorimospora suihominis nom. nov. [i.e., n. sp.]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Psorospermium sp. Virchow, 1860 pro parte; Synchytrium miescherianum Kuhn, 1865 pro parte; Sarcocystis miescheriana (Kuhn, 1865) Lankester, 1882 pro parte; Coccidium bigeminum var. hominis Railliet & Lucet, 1891 pro parte; Isospora hominis (Railliet & Lucet, 1891) Wenyon, 1923 pro parte; Lucetina hominis (Railliet & Lucet, 1891) Henry & Leblois, 1926 pro parte

Endorimospora suihominis
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
"Tadros and Laarman [1976] also proposed 2 new organisms: Endorimospora (Sarcocystis) suihominis from the pig and Isospora heydorni from the dog. I [1976] named them Sarcocystis porcihominis and Isospora wallacei respectively. Because Tadros and Laarman's paper was published a month earlier than mine, the names proposed by Tadros and Laarman have priority over Sarcocystis porcihominis and Isospora wallacei."

Endorimospora tenella (Railliet, 1886) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Balbiana gigantia [sic] Railliet,1886; Sarcocystis tenella Railliet, 1886; Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte; Sarcocystis tenella (ovifelis) Heydorn, Gestrich, Mehlhorn & Rommel, 1975

Endorimosporinae subfam. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
Eimeriidae fam. emend.
includes: Endorimospora gen. nov.

Endotrypanum sp.
Zeledon, R.; Ponce, C.; and de Ponce, E., 1975, Am. J. Trop. Med. and Hyg., v. 24 (4), 706-707
Choloepus hoffmanni: Costa Rica

E[ndotrypanum] schaudinii
Operti, M. S., 1973, J. Protozool., v. 20 (4), 530 [Abstract]
E[ndotrypanum] schaudinii, effect of CO_2 in glucose and glycerol metabolism

Entamoeba
Lamy, L. H.; and Crignon, I., 1972, Protistologica, v. 8 (4), 435-438
Entamoeba, position of endosome inside nucleus of trophozoites, use as specific character in relation to number of cyst nuclei

Entamoeba
Trigg, P. I.; and Gutteridge, W. E., 1977, Symposia Brit. Soc. Parasitol., v. 15, 57-81
parasitic Protozoa, morphological, biochemical, and physiological changes occurring during life cycles, symposium presentation

Entamoeba spp.
McConnell, E. E.; et al., 1974, Onderstepoort J. Vet. Research, v. 41 (3), 97-168
pathological and parasitological survey of 100 free-ranging chacma baboons
Papio ursinus (large intestine): Kruger National Park, Transvaal

Entamoeba [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Siganus (digestive tract): Elat, Gulf of Aqaba, Red Sea

Entamoeba coli
Alcasid, M. L. S.; et al., 1973, N. York State J. Med., v. 73 (13), 1786-1788
human intestinal parasites, no significant correlation with bronchial asthma, results of study with control and asthmatic groups: New York

Entamoeba coli
Best, J. C.; et al., 1976, Med. J. Australia, v. 1 (1-2), 14-20
assessment of growth-rate and growth-retardation in Australian Aboriginal children before and after treatment for common intestinal parasites

Entamoeba coli
de Carneri, I.; Grassi, L.; and Trane, F., 1972, Parassitologia, v. 14 (1), 113
Entamoeba histolytica, E. coli, Dientamoeba fragilis, prevalence in an unselected hospital population: Milano

Entamoeba coli
Garrocho-Sandoval, C.; and Torres-Ruvalcaba, A., 1977, Am. J. Clin. Path., v. 67 (6), 603-605
human intestinal parasites, comparison study of two different methods for collecting fecal samples for diagnostic purposes

Entamoeba coli
Giboda, M.; Hoffman, F.; and Rimar, P., 1971, Ceskoslov. Pediat., v. 26 (2), 88-89
intestinal protozoal infections in children aged 1-15 years, clinical trials with metronidazole, results and parasites eliminated varied with age and parasite: Kosice, Czechoslovakia

Entamoeba coli type, illus.
Lamy, L. H.; and Crignon, I., 1972, Protistologica, v. 8 (4), 435-438
Entamoeba, position of endosome inside nucleus of trophozoites, use as specific character in relation to number of cyst nuclei

Entamoeba coli
McConnell, E. E.; et al., 1974, Onderstepoort J. Vet. Research, v. 41 (3), 97-168
pathological and parasitological survey of 100 free-ranging chacma baboons
Papio ursinus (large intestine): Kruger National Park, Transvaal

Entamoeba coli
Markell, E. K.; and Quinn, P. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1139-1142
comparison of immediate polyvinyl alcohol fixation with delayed Schaudinn's fixation for demonstrating protozoa in feces, concluded that immediate fixation results in preservation of larger number of organisms and recommends routine use prior to transportation to laboratory

Entamoeba coli
Mirza, M. Y.; and al-Rawas, A. Y., 1975, J. Protozool., v. 22 (1), 23-24
Tatera indica (feces): Baghdad district, Iraq

Entamoeba coli
Rajasekaran, P.; Dutt, P. R.; and Pisharoti, K. A., 1977, Indian J. Med. Research, v. 66 (2), 189-199
human intestinal parasites, survey of correlation between infection rate and source of water supply (well, street tap, home with tap water) as indication of control of water-borne diseases by public water supplies: Madurai district, Tamil Nadu, India

Entamoeba coli
Reyes, H.; Olea, M.; and Hernandez, R., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 115-116
survey of food handlers for intestinal parasites shows 50.5% infection rate: Santiago, Chile

Entamoeba coli, illus.
Rondanelli, E. G.; et al., 1972, Parassitologia, v. 14 (2-3), 357-363
Entamoeba coli, trophozoites, electron microscopy, comparison of strains from London and Milano in in vitro culture

Entamoeba coli, illus.
Rondanelli, E. G.; et al., 1973, Riv. Parassitol., Roma, v. 34 (4), 263-270
Entamoeba coli cysts, ultrastructure in monoxenic culture, basis for outlining an organization model

Entamoeba coli
Sagua, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 58-60
comparison of phenol-alcohol-formalin sedimentation and polyvinyl alcohol fixative tests in diagnosis of human intestinal helminths and protozoa

Entamoeba coli
Savanat, T.; et al., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (2), 149-154
determinations of total serum IgE levels in humans with amoebic liver abscess or other parasitic infections

Entamoeba coli
Schenone, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 104-105
metronidazole used for mass therapy of human intestinal protozoa: Chile

Entamoeba coli
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Entamoeba coli
Torres, P.; and Navarrete, N., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 90-95
modified Telemann's and phenol-alcohol-formalin methods compared for diagnosis of common intestinal protozoa of man

Entamoeba coli
Vinayak, V. K.; and Sehgal, S. C., 1976, Indian J. Med. Research, v. 64 (9), 1347-1350
human helminthic and protozoan parasites, comparison of nigrosin-methylene blue diagnostic test with formol-ether method and direct examination

Entamoeba dysenteriae
Renault, J.; Cavier, R.; and Renault, S., 1976, Compt. Rend. Acad. Sc., Paris, v. 282, s. D, (5), 509-511
Entamoeba dysenteriae, secondary aminoquinolines, influence of alkylamino chain length and position on activity in vitro and in vivo

Entamoeba gingivalis
Gall, Z.; and Hraste, J., 1969, Medicina, Rijeka, v. 6 (2), 157-163
survey of fauna of human oral cavity, Entamoeba gingivalis and Trichomonas elongata discovered mostly in presence of tooth decay and alkaline reactions: Rijeka, Yugoslavia

Entamoeba gingivalis
Murar, B.; and Merarova, M., 1977, J. Protozool., v. 24 (2), Suppl., 16A [Abstract]
Entamoeba gingivalis, Trichomonas tenax, occurrence in patients with mouth and dental problems

Entamoeba hartmanni
Artigas, J.; Otto, I.; and Kawada, M. E., 1966, Bol. Chileno Parasitol., v. 21 (4), 114-118
Entamoeba histolytica, pathologic effects of motile trophozoites on human leucocytes compared with non-motile E. moshkovskii and another free-living amoeba both of which showed no action in blood cells; possible use in differentiating E. histolytica from E. hartmanni

Entamoeba hartmanni
Chwirot, E.; and Kasprzak, W., 1975, Polski Tygod. Lekar., v. 30 (6), 261-263
Entamoeba histolytica, E. hartmanni, Dientamoeba fragilis, humans, clinical trials to evaluate effectiveness of metronidazole, good results with minimal side effects

Entamoeba hartmanni
Markell, E. K.; and Quinn, P. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1139-1142
comparison of immediate polyvinyl alcohol fixation with delayed Schaudinn's fixation for demonstrating protozoa in feces, concluded that immediate fixation results in preservation of larger number of organisms and recommends routine use prior to transportation to laboratory

Entamoeba hartmanni
Spillmann, R.; Ayala, S. C.; and de Sanchez, C. E., 1976, Am. J. Trop. Med. and Hyg., v. 25 (4), 549-551
double-blind test of metronidazole and tinidazole in the treatment of asymptomatic Entamoeba histolytica and E. hartmanni carriers; both drugs considered ineffective

Entamoeba hartmanni
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Entamoeba hartmanni
Vinayak, V. K.; and Sehgal, S. C., 1976, Indian J. Med. Research, v. 64 (9), 1347-1350
human helminthic and protozoan parasites, comparison of nigrosin-methylene blue diagnostic test with formol-ether method and direct examination

Entamoeba histolytica. [See also Amoebiasis]

Entamoeba histolytica
Abioye, A. A., 1976, J. Trop. Med. and Hyg., v. 79 (11), 252-255
Entamoeba histolytica, drug (emetine, metronidazole) and immuno-diagnostic (fluorescent antibody, gel diffusion and latex agglutination tests) resistant amoebic hepatic abscess in man, case report, blood-abscess cavity barrier postulated as possible mechanism for diagnostic failure: Nigeria

Entamoeba histolytica
Adetuyibi, A.; Akisanya, J. B.; and Onadeko, B. O., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 466-473
malaria, Entamoeba histolytica, humans, analysis of causes of death on hospital wards over 14-year period: Ibadan, Nigeria

Entamoeba histolytica
Alam, M.; and Ahmad, S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 370-373
Entamoeba histolytica antigen fractions, immunogenicity, experimental evidence of heterogeneity

Entamoeba histolytica
Albach, R. A.; et al., 1974, J. Protozool., v. 21 (3), 439 [Abstract]
Entamoeba histolytica, nuclear DNA synthesis

Entamoeba histolytica
Albach, R. A.; et al., 1977, Exper. Parasitol., v. 42 (1), 248-259
Entamoeba histolytica, nuclear sites of RNA synthesis, autoradiographic analysis

Entamoeba histolytica
Albach, R. A.; Booden, T.; and Downing, S. W., 1974, J. Protozool., v. 21 (3), 435 [Abstract]
Entamoeba histolytica, site of synthesis of nuclear RNA (nuclear stroma and especially peripheral chromatin)

Entamoeba histolytica
Albornoz Plata, A., 1975, Med. y Cirug., Bogota, v. 39 (3), 99-103
historical review of clinical findings and management of human Entamoeba histolytica since its discovery in 1875

Entamoeba histolytica
Alvarado Cerna, R., 1973, Prensa Med. Mexicana, v. 38 (5-6), 153-158
Entamoeba histolytica, human invasive cutaneous forms, clinical aspects, pathology, case reports

Entamoeba histolytica
Alvarez Chacon, R., 1975, Mod. Problems Paediat., v. 17, 259-261
Entamoeba histolytica, 4-month-old child with genital cutaneous amoebiasis following acute intestinal infection, case report, medical management: Mexico

Entamoeba histolytica
Ambroise-Thomas, P., 1975, Maroc Med. (588), v. 55, 130-136
human sero-diagnosis by immunofluorescence, review

Entamoeba histolytica
Ambroise-Thomas, P.; et al., 1973, Nouv. Presse Med., v. 2 (33), 2200 [Letter]
human Entamoeba histolytica, hepatic abscess simulating echinococcal cyst, differential diagnosis and post-therapy evaluations using indirect fluorescent antibody technique

Entamoeba histolytica
Ambroise-Thomas, P.; et al., 1975, Acta Trop., v. 32 (4), 365-370
Entamoeba histolytica, human, epidemic of autochthonous hepatic and intestinal amoebiasis in autumn 1974: Saint-Laurent-du-Pont, 30 km north of Grenoble

Entamoeba histolytica
Ambroise-Thomas, P.; and Meyer, H. A., 1975, Acta Trop., v. 32 (4), 359-364
Entamoeba histolytica, hepatic amoebiasis, human, diagnosis, agglutination and indirect fluorescent antibody tests, clinical trials with Tinidazole (Fasigyn), well tolerated with encouraging results but some cases required supplementary treatment with metronidazole: region of Kilimanjaro, northeast Tanzania

Entamoeba histolytica
Anderson, R. I.; and Buck, A. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 447-456
complement levels in residents of rural village in relation to wide variety of clinical, laboratory, and epidemiological factors including parasitic diseases: Ouli Bangala, Republic of Chad

Entamoeba histolytica
Andre, L. J.; and Condat, J., 1975, Medecine et Armees, v. 3 (4), 319-321
human intestinal amoebiasis, review of current clinical management

Entamoeba histolytica
Anh, D. H., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 34-38
Entamoeba histolytica, histopathologic examination of 4 amoebic granulomas excised surgically from human colons

Entamoeba histolytica, illus.
Apt, W.; Difilippi, C.; and Subiabre, V., 1972, Bol. Chileno Parasitol., v. 27 (1-2), 55-56
Entamoeba histolytica trophozoites in exudates of hepatic abscess in young girl, case report: Chile

Entamoeba histolytica
Arcilla-Latonio, A.; Arcilla, C. A.; and de Guzman-Matawaran, L., 1974, Philippine J. Int. Med., v. 12 (3), 131-143
Entamoeba histolytica, humans, comparison of 2 dosage regimens of teclozan in the treatment of mild forms of intestinal amoebiasis: Manila, Philippines

Entamoeba histolytica, illus.
Artigas, J.; Otto, I.; and Kawada, M. E., 1966, Bol. Chileno Parasitol., v. 21 (4), 114-118
Entamoeba histolytica, pathologic effects of motile trophozoites on human leucocytes compared with non-motile E. moshkovskii and another free-living amoeba both of which showed no action in blood cells; possible use in differentiating E. histolytica from E. hartmanni

Entamoeba histolytica
Atias, A.; et al., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 119-121
clinical trials using teclozan to treat human chronic intestinal amoebiasis, good tolerance and short duration of treatment: Chile

Entamoeba histolytica
Balamuth, W.; and Siddiqui, W. A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 439-468
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Entamoeba histolytica
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Entamoeba histolytica, case history of human pulmonary ameboma, radiological aspects, successful treatment with Ambilhar; review on localization and diffusion of E. histolytica

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Entamoeba histolytica, pharmacological aspects and clinical indications for treatment, drugs in current use, humans

Entamoeba histolytica, illus.
Goegler, H.; and Knight, R., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 177-185
Entamoeba histolytica, hamsters, effect of hepatic injury upon development of amoebic liver abscess

Entamoeba histolytica
Gorbach, S. L.; et al., 1975, N. England J. Med., v. 292 (18), 933-936
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Rattus rattus (caecum, rectum): Chandigarh, India

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Entamoeba histolytica hepatic abscess in Vietnam veteran, poor response to metronidazole therapy, possible drug resistance, apparent cure after emetine and chloroquine: Illinois

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Trichuris trichiura, Ascaris lumbricoides and Entamoeba histolytica, fecal survey of nursing staff and inmates of hospital for mentally retarded for presence of intestinal parasites: Groot Schuylenburg, Apeldoorn, Netherlands

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Entamoeba histolytica, humans in carrier and disease states, results of lymphocytic transformation in response to specific antigen and to mitogen, specific cellular immunodepression as possible factor in amoebic invasion of bowel mucosa

Entamoeba histolytica
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Entamoeba histolytica, illus.
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Entamoeba histolytica, trophozoites, freeze-etching electron microscopy, cell membrane surfaces, nuclei, cytoplasm

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Hernandez-Perez, E., 1975, Med. Cutan. Ibero-Latino-Am., v. 3 (1), 47-54
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Entamoeba histolytica in humans, differential diagnosis of imported disease for physicians in non-endemic areas: Netherlands

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Entamoeba histolytica, human venereal amoebiasis, epidemiology: New York City

Entamoeba histolytica, illus.
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Entamoeba histolytica, amoebic lung abscess in man who 5 years previously had been treated for intestinal amoebiasis, case report, differential diagnosis: New York City

Entamoeba histolytica, illus.
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Knight, R.; McCaul, T. F.; and Bird, R. G., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 1 [Demonstration]
Entamoeba histolytica, damage and changes in host morphology (rabbit kidney cell monolayers) in presence of parasites; evidence that direct amoebic cell contact needed for damage to occur, possible role of enzymes

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Lahav, M.; et al., 1975, J. Protozool., v. 22 (3), 72A [Abstract]
Entamoeba histolytica, human, detection of antibodies by counterelectrophoresis and passive haemagglutination technique

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Entamoeba histolytica, endocytosis and digestion by trophozoites, electron microscopy

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Lushbaugh, W. B.; Kairalla, A. B.; and Pittman, F. E., 1976, J. Protozool., v. 23 (2), 12A [Abstract]
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Entamoeba histolytica
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Entamoeba histolytica, humans, extensive epidemiologic review of human amoebiasis with comparative study of environmental and sociologic factors of 2 Amerindian reservations one of which had a high prevalence of infection and the other none

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Entamoeba histolytica
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patients with amebic abscess of liver, diminished cell-mediated immunity to Entamoeba histolytica antigens when tested by skin tests and for migration-inhibition factor production, skin reactions to unrelated antigen were normal, 10 days after hospital discharge cell-mediated immune responses to E. histolytica antigen were normal, antibodies were present in sera at all stages

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Entamoeba histolytica
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Entamoeba histolytica
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Entamoeba histolytica, human, metronidazole combined with diloxanide furoate cured liver abscess and eliminated intestinal infection in most subjects

Entamoeba histolytica, illus.
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Entamoeba histolytica
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Entamoeba histolytica axenic and monoxenic (in association with Crithidia sp.) cultures, survival after storage in liquid nitrogen, semiautomatic vs. automatic freezing, influence of specimen concentration and different plastic storage vessels

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Entamoeba histolytica in humans, acidic pH of amoebic hepatic abscess exudate, diagnostic significance

Entamoeba histolytica
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Entamoeba histolytica
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Entamoeba histolytica in rats (exper.), strain variations in pathogenicity, pathology of lesions, role of host nutrition in pathogenesis

Entamoeba histolytica
Rao, V. G.; and Padma, M. C., 1975, Indian J. Exper. Biol., v. 13 (2), 168-171
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Entamoeba histolytica
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Entamoeba histolytica
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Entamoeba histolytica
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Entamoeba histolytica
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Entamoeba histolytica
Reeves, R. E.; Serrano, R.; and South, D. J., 1976, J. Biol. Chem., v. 251 (10), 2958-2962
Entamoeba histolytica, 6-phosphofructokinase, physical properties, divalent cation requirement, mechanism of reaction it catalyzes

Entamoeba histolytica
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Entamoeba histolytica
Ridley, D. S.; and Hedge, E. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 522-525
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Entamoeba histolytica in human, amoebiasis of cecum presenting as pelvic mass or tubo-ovarian abscess, clinical case report, diagnosis from pathology found at surgery: New York

Entamoeba histolytica
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Entamoeba histolytica
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man, amoebiasis of rectum presenting as rectal carcinoma, case report, differential diagnosis, emetine: Burmese native living in London

Entamoeba histolytica
Rukmono, B.; Naumar, S. A.; and Talogo, R. W., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (2), 249-255
Ascaris lumbricoides, Entamoeba histolytica, Giardia lamblia implicated as possible pathogens in survey of infantile diarrhea in a population of low socio-economic group in Djakarta, Indonesia

Entamoeba histolytica
deSa, A. E., 1974, Progr. Drug Research, v. 18, 77-90
Entamoeba histolytica, humans, complications of hepatic or intestinal infections requiring surgical intervention, review

Entamoeba histolytica
Sagua, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 58-60
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Entamoeba histolytica
San Antonio Alvarez, J., 1972, Med. Trop., Madrid, v. 48 (3-4), 188-195
clinical symptoms and pathologic findings of 7 cases of Entamoeba histolytica cystitis, probable secondary infection resulting from amoebic dysentery

Entamoeba histolytica, illus.
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Entamoeba histolytica epidemic in institutionalized mental defectives, high associated incidence of Trichuris trichiura suggests it as factor contributing to Entamoeba's pathogenicity; comparative study of effectiveness of intestopan and humatin in treatment of amoebiasis: Queensland, Australia

Entamoeba histolytica
Sapunar, J.; Munoz, M. A.; and Castro, J., 1967, Bol. Chileno Parasitol., v. 22 (2), 60-65
Entamoeba histolytica, case report of hepatic abscess in man which ruptured into the bronchi after abscess aspiration, emetine and chloroquine therapy: Santiago, Chile

Entamoeba histolytica
Sargeaunt, P. G.; and Lumsden, W. H. R., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 54-56
Entamoeba histolytica in vitro, action of furazolidone and iodochlorhydroxyquin alone and as combination drug (dependal)

Entamoeba histolytica
Sargeaunt, P. G.; and Williams, J. E., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 385-386 [Demonstration]
Entamoeba histolytica, enzyme electrophoresis patterns established

Entamoeba histolytica
Satwik, A.; Sarin, G. S.; and Misra, R. C., 1977, J. Trop. Med. and Hyg., v. 80 (2), 40-42
Entamoeba histolytica, human, increased serum seromucoid levels in active colonic or hepatic disease, normal levels in asymptomatic cyst passers

Entamoeba histolytica
Savel, J., 1976, Ann. Pharm. Franc., v. 34 (7-8), 249-259
Entamoeba histolytica, Trichomonas vaginalis, ornidazole compared with metronidazole in vitro and in vivo, rats, hamsters

Entamoeba histolytica
Schenone, H.; et al., 1973, Bol. Chileno Parasitol., v. 28 (3-4), 104-105
metronidazole used for mass therapy of human intestinal protozoa: Chile

Entamoeba histolytica
Schenone, H.; et al., 1975, Bol. Chileno Parasitol., v. 30 (3-4), 76-79
Entamoeba histolytica, Giardia lamblia, children, tinidazole drug of choice for individual or mass therapy: Chile

Entamoeba histolytica
Scragg, J. N.; and Proctor, E. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 824-825
Entamoeba histolytica, amoebic dysentery in children successfully treated with tinidazole in weight related dosages, recommended as safe and simple form of therapy: South Africa

Entamoeba histolytica
Scragg, J. N.; and Proctor, E. M., 1977, Arch. Dis. Childhood, v. 52 (5), 408-410
Entamoeba histolytica in children, tinidazole highly effective in treatment of hepatic abscess, clinical trials: South Africa

E[ntamoeba] histolytica
Segovia, E.; et al., 1974, Prensa Med. Mexicana, v. 39 (11-12), 449-452
E[ntamoeba] histolytica, human amoebic hepatic abscess alone and associated with rectocolonic infections, therapy with tinidazole and RP 14539 showed good results

Entamoeba histolytica, illus.
Serrano, R.; Deas, J. E.; and Warren, L. G., 1977, Exper. Parasitol., v. 41 (2), 370-384
Entamoeba histolytica, cell fractionation study, isolation and electron microscopy of two membrane fractions, properties and distribution of marker enzymes, chemical analysis of fractions, electrophoretic patterns of membrane polypeptides, glucose transport

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Sher, A.; and Ahmad, S., 1977, Indian J. Exper. Biol., v. 15 (12), 1083-1088
Entamoeba histolytica, rabbits, humans, detection and characterization of specific humoral antibodies from immune sera

Entamoeba histolytica, illus.
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Entamoeba histolytica, electron microscopic study of autochthonous Romania strain with emphasis on phagocytosis

Entamoeba histolytica
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Entamoeba histolytica
Singh, K.; and Dutta, G. P., 1976, Indian J. Med. Research, v. 64 (8), 1185-1191
action of metabolic inhibitors on Entamoeba histolytica in axenic culture

Entamoeba histolytica
Singh, K.; and Dutta, G. P., 1977, Indian J. Med. Research, v. 65 (1), 51-57
Entamoeba histolytica, axenic cultures, effects on parasite growth of storage up to 21 days, different media constituents and dilution of medium

Entamoeba histolytica
Spencer, H. C., jr.; et al., 1976, Am. J. Epidemiol., v. 104 (1), 93-99
statistical epidemiologic survey of endemic human amoebiasis area, household crowding, poor sanitation and resulting fecal contamination implicated as cause of transmission

Entamoeba histolytica
Spencer, H. C.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 628-635
Entamoeba histolytica, epidemiologic statistics of endemic amebiasis in an extended family, two cousins of which had hepatic abscesses, random survey of remainder of community showed little evidence of infection, transmission among extended family members thought to be person to person: Bloomington, Texas and Florida

Entamoeba histolytica
Spillmann, R.; Ayala, S. C.; and de Sanchez, C. E., 1976, Am. J. Trop. Med. and Hyg., v. 25 (4), 549-551
double-blind test of metronidazole and tinidazole in the treatment of asymptomatic Entamoeba histolytica and E. hartmanni carriers; both drugs considered ineffective

Entamoeba histolytica
Stamm, W. P.; and Phillips, E. A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 490-492
human amoebiasis, cellulose acetate membrane precipitin (CAP) diagnostic test compared with gel diffusion precipitin; results suggest that CAP is slightly more sensitive and equally specific

Entamoeba histolytica
Stamm, W. P.; Phillips, E. A.; and Bell, K., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 433 [Demonstration]
Entamoeba histolytica, immunofluorescent staining of trophozoites in culture, in fresh and preserved feces and tissue sections

E[ntamoeba] histolytica
Stoopen, M.; De Stoopen, V. B.; and Landa, L., 1975, Prensa Med. Mexicana, v. 40 (3-4), 59-65
E[ntamoeba] histolytica, perfusion of emetine into hepatic artery over a 24-48 hour period in persons with severe toxic infection and hepatic failure, useful therapeutic resource in instances of severe infections: Mexico

Entamoeba histolytica
Stotka, V. L., 1975, North Carolina Med. J., v. 36 (10), 607-611
Entamoeba histolytica, hepatic abscesses in Vietnam veterans, diagnostic difficulties, clinical aspects: North Carolina

Entamoeba histolytica
Sunarwan, I.; and Rosekrans, P. C. M., 1976, Nederl. Tijdschr. Geneesk., v. 120 (8), 319-321
Entamoeba histolytica, extensive painful cutaneous ulcer around colostomy stoma of man undergoing treatment for ulcerative colitis finally diagnosed as cutaneous amoebiasis, impressive improvement after metronidazole: Netherlands (resident of Indonesia)

Entamoeba histolytica
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intestinal amoebiasis in children, tiberal a promising therapy in comparison clinical trials with metronidazole: Indonesia

Entamoeba histolytica, illus.
Takeuchi, A.; Jervis, H. R.; and Phillips, B. P., 1977, Virchows Arch., B Cell Path., v. 24 (4), 263-277
Entamoeba histolytica, guinea pig (exper.), histolysis of cecum, light and electron microscopy studies

Entamoeba histolytica, illus.
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mode of penetration of intestinal epithelium of germ-free guinea pigs by Entamoeba histolytica trophozoites, light and electron microscopic study

Entamoeba histolytica, illus.
Takeuchi, A.; and Phillips, B. P., 1976, Virchows Arch., B Cell Path., v. 20 (1), 1-13
Entamoeba histolytica, guinea pigs, early stages in development of acute amebic colitis, cellular and vascular changes accompanying invasion of lamina propria (continued epithelial shedding, polymorphonuclear leukocyte degeneration, endothelial damage and occlusive thrombosis in capillaries and venules)

Entamoeba histolytica, illus.
Takeuchi, T.; Weinbach, E. C.; and Diamond, L. S., 1977, Exper. Parasitol., v. 43 (1), 107-114
Entamoeba histolytica, phosphorylase, particulate glycogen, subcellular distribution, characterization, particulate glycogen may be polysaccharide-protein complex

Entamoeba histolytica, illus.
Takeuchi, T.; Weinbach, E. C.; and Diamond, L. S., 1977, Exper. Parasitol., v. 43 (1), 115-121
Entamoeba histolytica, phosphoglucomutase, uridine diphosphate glucose pyrophosphorylase, glycogen synthase, subcellular distribution, characterization

Entamoeba histolytica
Talis, B.; Stein, B.; and Gold, D., 1977, J. Protozool., v. 24 (2), Suppl., 18A-19A [Abstract]
hartmanelloid amoeba, invasiveness and antigenicity as compared to Entamoeba histolytica strains

Entamoeba histolytica
Tandon, B. N.; et al., 1975, Exper. and Molecular Path., v. 23 (2), 155-163
Entamoeba histolytica, human, amebic abscess, electron microscopic study of liver biopsies, evidence of diffuse parenchymal injury similar to that reported for patients with nonsuppurative hepatic amebiasis

Entamoeba histolytica
Tanowitz, H. B., 1974, Med. Aspects Human Sexual., v. 8 (9), 45-65
human parasitic gynecologic diseases, clinical aspects, epidemiology, sexual transmission, review

Entamoeba histolytica, illus.
Tanowitz, H. B.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 602-606
Entamoeba histolytica, 3 strains grown in axenic culture, effect of varying dosages of metronidazole

Entamoeba histolytica, illus.
Tanowitz, H. B.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (1), 19-28
metronidazole, suggested mechanism of antimicrobial action and selective toxicity for Entamoeba histolytica as opposed to mammalian cell types and Trypanosoma cruzi

Entamoeba histolytica
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Entamoeba histolytica
Torres, P.; and Navarrete, N., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 90-95
modified Telemann's and phenol-alcohol-formalin methods compared for diagnosis of common intestinal protozoa of man

Entamoeba histolytica, illus.
Trissl, D.; et al., 1977, J. Exper. Med., v. 145 (3), 652-665
Entamoeba spp., pathogenic strains show cell surface properties different from those of nonpathogenic strains as evidenced by concanavalin A-induced agglutination studies

Entamoeba histolytica-like, Laredo strain
Trissl, D.; et al., 1977, J. Exper. Med., v. 145 (3), 652-665
Entamoeba spp., pathogenic strains show cell surface properties different from those of nonpathogenic strains as evidenced by concanavalin A-induced agglutination studies

Entamoeba histolytica
Tubis, M.; et al., 1976, Nuklearmedizin, v. 15 (1), 36-38
Entamoeba histolytica in humans, demonstration of hepatic abscess by scintigraphy utilizing a radiopharmaceutical (bromometronidazole)

Entamoeba histolytica
Urai, L.; and Kali, A., 1976, Orvosi Hetilap, v. 117 (23), 1406-1407
Entamoeba histolytica in young man, case report of chronic severe amoebiasis in association with Raynaud syndrome, possible relationships: Hungary

Entamoeba histolytica
Vakil, B. J.; and Dalal, N. J., 1974, Progr. Drug Research, v. 18, 353-364
Entamoeba histolytica, humans, techniques for comparative evaluation of amoebicidal drugs: sites of action, efficacy, side effects, and toxicity

Entamoeba histolytica
Vega Franco, L.; et al., 1975, Prensa Med. Mexicana, v. 40 (7-8), 197-201
intestinal parasites, comparison of D-xylose intestinal absorption in infected children showed that only those with Giardia lamblia had statistically different absorption from non-infected children: Mexico

Entamoeba histolytica
Vijil, C., 1972, N. England J. Med., v. 286 (2), 106-107 [Letter]
Entamoeba histolytica in humans, frequent association with Shigella dysenteriae, need for improved research on pathogenic interaction, dysentery often not relieved without additional amoebicidal therapy with metronidazole: Nicaragua

Entamoeba histolytica
Villafane, H. A.; et al., 1972, Prensa Med. Argent., v. 59 (3), 124-126
Entamoeba histolytica, humans, successful clinical trials with teclozan: Cordoba, Argentina

Entamoeba histolytica
Villarejos, V. M.; et al., 1971, Am. J. Trop. Med. and Hyg., v. 20 (4), 602-607
metronidazole tested as chemoprophylaxis for diarrhea caused by Giardia lamblia and Entamoeba histolytica

Entamoeba histolytica, illus.
Villegas-Gonzalez, J.; and Fastag de Shor, A., 1974, Bol. Med. Hosp. Inf., v. 31 (1), 35-49
Entamoeba histolytica, children, invasive amoebiasis, pathologic review of autopsies; comparison axenic and monoxenic cultures, multiple enzymes present, relationship between acid phosphatase and invasive capacity of parasite

Entamoeba histolytica
Vinayak, V. K.; et al., 1976, Indian J. Med. Research, v. 64 (5), 661-667
value of immunoelectrophoresis in immunodiagnosis of human hepatic and intestinal Entamoeba histolytica, comparison with indirect agglutination and gel diffusion methods

Entamoeba histolytica
Vinayak, V. K.; and Chitkara, N. L., 1976, Indian J. Med. Research, v. 64 (10), 1443-1445
Entamoeba histolytica, 17 strains, selective ability to hemolyse red blood cells could be utilized as a parameter for differentiation of strains

Entamoeba histolytica
Vinayak, V. K.; and Chitkara, N. L., 1976, Indian J. Med. Research, v. 64 (10), 1446-1450
Entamoeba histolytica, Laredo strain rarely if ever encountered, 32 strains differentiated: India

Entamoeba histolytica
Vinayak, V. K.; and Sehgal, S. C., 1976, Indian J. Med. Research, v. 64 (9), 1347-1350
human helminthic and protozoan parasites, comparison of nigrosin-methylene blue diagnostic test with formol-ether method and direct examination

Entamoeba histolytica
Voller, A.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 431-437
human tropical parasitic diseases, comparison of enzyme-immunoassay and radio-immunoassay in detection of antibodies, both assays sensitive and reproducible and gave comparable results

Entamoeba histolytica
Voller, A.; Bartlett, A.; and Bidwell, D. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 98-106
application of enzyme immunoassay to diagnosis of human parasitic diseases, technique for microplate enzyme-linked immunosorbent assay

Entamoeba histolytica
Wadstroem, T.; et al., 1976, Arch. Dis. Childhood, v. 51 (11), 865-870
Ascaris, Entamoeba histolytica, Giardia lamblia, children with diarrheal disease, survey, no correlation with toxinogenic bacteria in stools: Ethiopia

Entamoeba histolytica
Weinbach, E. C.; et al., 1976, J. Parasitol., v. 62 (1), 127-128
Entamoeba histolytica, evidence that iron-sulfur proteins are functional in aerobic metabolism

Entamoeba histolytica
Weinbach, E. C.; et al., 1977, Exper. Parasitol., v. 41 (1), 186-197
Entamoeba histolytica, diaphorase activities

Entamoeba histolytica
Wessenberg, H., 1974, Perspect. Biol. and Med., v. 17 (2), 250-266
Entamoeba histolytica, possible role of heat stress in amoebic invasion of host tissues

Entamoeba histolytica
Wilde, H., 1973, N. England J. Med., v. 289 (7), 378 [Letter]
Entamoeba histolytica, case reports of poor response of hepatic abscess to metronidazole therapy, cure after emetine, chloroquine and di-iodohydroxyquin: Thailand

Entamoeba histolytica
Winkelmann, E.; et al., 1977, Arzneimittel-Forsch., v. 27 (12), 2251-2263
Entamoeba histolytica, Trichomonas foetus, Trypanosoma spp., in golden hamsters and mice (both exper.), laboratory trials with more than 135 new 2 methyl-5-nitroimidazoles substituted in the 1-position and 1-methyl-5-nitroimidazoles substituted in the 2-position, comparison of activity with that of metronidazole and tinidazole

Entamoeba histolytica
Wittnich, C., 1976, Canad. Vet. J., v. 17 (10), 259-263
Entamoeba histolytica, dog, clinical and laboratory findings, probable transmission from human, Flagyl, flumethasone, good results: southeastern Quebec

Entamoeba histolytica
Wonchee Hernandez, G., 1972, Prensa Med. Mexicana, v. 37 (9-10), 384
Entamoeba histolytica, cutaneous infection in young man successfully treated with nimorazole, case report: Mexico

Entamoeba histolytica
Yadava, J. N. S.; and Dutta, G. P., 1976, Indian J. Med. Research, v. 64 (4), 562-567
lethal effect of bile on axenic Entamoeba histolytica culture without increased effect on amoebicidal action of metronidazole or emetine, possible etiologic role of inadequate bile secretion as factor in clinical amoebiasis

Entamoeba histolytica
Zuidema, P. J., 1971, Nederl. Tijdschr. Geneesk., v. 115 (29), 1209-1212
man, case history of mixed infections of Entamoeba histolytica, Necator americanus, Plasmodium falciparum and Schistosoma mansoni, medical management: Netherlands (native of Surinam)

Entamoeba histolytica
Zuidema, P. J., 1975, Nederl. Tijdschr. Geneesk., v. 119 (35), 1339-1345
Entamoeba histolytica, human hepatic amoebic abscesses, treatment regimens using metronidazole, emetine and diloxanide furoate

Entamoeba histolytica
Zuidema, P. J., 1976, Nederl. Tijdschr. Geneesk., v. 120 (21), 901-906
human intestinal parasites, differential diagnosis of causes of diarrhea in hikers returning from visits to India: Netherlands

Entamoeba histolytica
Zuliani A., C.; and Garcia Cuestas, J., 1970, Pediatria, Bogota, v. 11 (3), 226-237
Entamoeba histolytica, review of pathologic findings and clinical symptoms from autopsies of 20 children who died from complications of intestinal perforation caused by amoebiasis: Colombia

Entamoeba histolytica histolytica
Duong, T. H.; and Barrabes, A., 1976, Compt. Rend. Soc. Biol., Paris, v. 170 (4), 908-915
Entamoeba h. histolytica (exper.), level of circulating antibodies (indirect immunofluorescence) is lower in female castrated golden hamsters implanted with pellet of oestradiol than in castrated control animals

Entamoeba histolytica histolytica
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Entamoeba histolytica f. minuta
Giboda, M.; Hoffman, F.; and Rimar, P., 1971, Ceskoslov. Pediat., v. 26 (2), 88-89
intestinal protozoal infections in children aged 1-15 years, clinical trials with metronidazole, results and parasites eliminated varied with age and parasite: Kosice, Czechoslovakia

Entamoeba histolytica minuta
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Entamoeba invadens, illus.
Bird, R. G.; and McCaul, T. F., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 81-93
presence of rhabdoviruses in Entamoeba histolytica and E. invadens, possible role in pathogenicity

Entamoeba invadens
Buro, N. C.; and Weller, D. L., 1974, J. Protozool., v. 21 (5), 796-802
Entamoeba invadens, malic enzyme, purification, characterization, 3 isoenzymes

Entamoeba invadens
Dolensek, E. P.; Bihn, J. P.; and Napolitano, R. L., 1976, J. Protozool., v. 23 (2), 19A [Abstract]
Entamoeba invadens, Naegleria, and Hartmanella isolated from exhibited reptiles

Entamoeba invadens
Donaldson, M.; et al., 1975, Am. J. Vet. Research, v. 36 (6), 807-817
reptilian amebiasis, fatal epizootic, possible cause of many previous snake deaths, pathologic changes, probable mode of introduction and spread, treatment (metronidazole successful) and prophylaxis, amebae identified as or presumed to be Entamoeba invadens
Boa constrictor (liver, intestine, stools) (nat. and exper.)
Python molurus bivittatus (feces)
Tiliqua scincoides
Boiga cynodon
Pelusios subniger (feces)
turtle (feces)
alligator (feces)
all from Steinhart Aquarium, California Academy of Sciences, Golden Gate Park, San Francisco

Entamoeba invadens
Dwyer, D. M., 1974, J. Protozool., v. 21 (1), 139-145
Trichomonas gallinae, Histomonas meleagridis, Dientamoeba fragilis, Entamoeba invadens, E. histolytica, antigenic relationships analyzed by immunoelectrophoretic techniques

Entamoeba invadens
Farquhar, J. A.; and Cannings, P. J. G., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 282 [Demonstration]
Entamoeba invadens, Trichomonas spp., Giardia muris, comparison of staining methods for production of permanent mounts of trophozoites

Entamoeba invadens
Gabrisch, K., 1976, Prakt. Tierarzt , v. 57, Sondernummer, 37-40
parasites of reptiles, diagnosis, treatment, brief review

Entamoeba invadens
McCaul, T. F.; Poston, R. N.; and Bird, R. G., 1977, Exper. Parasitol., v. 43 (2), 342-352
Entamoeba histolytica and E. invadens trophozoites cause chromium release from labeled human liver cells in culture, phospholipase inhibitor suppresses chromium release, results support belief that toxin affecting membrane permeability is operative factor in pathogenesis of amoebiasis

Entamoeba invadens
Mueller, M.; and Lindmark, D. G., 1976, Antimicrob. Agents and Chemotherapy, v. 9 (4), 696-700
Tritrichomonas foetus, Trichomonas vaginalis, Entamoeba invadens, uptake of metronidazole under anaerobic vs. aerobic conditions, effect on viability of T. foetus

Entamoeba invadens
Natarajan, P. N.; Wan, A. S.; and Zaman, V., 1974, Planta Med., v. 25 (3), 258-260
Plasmodium berghei in mice, Entamoeba invadens in monophasic medium, poor results in laboratory trials with gentianine and 3-hydroxymethyl-4-hydroxyethyl-5-vinyl-pyridine

Entamoeba invadens
Poston, R. N.; McCaul, T. F.; and Bird, R. G., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 112-113 [Demonstration]
Entamoeba histolytica, E. invadens, cytotoxic activity on labelled Chang cells, results indicate that amebic agent indirectly affects host cell membrane permeability by altering cell membrane architecture and strongly suggest that phospholipid alteration by phospholipase may be basis of loss of membrane integrity

Entamoeba invadens
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Entamoeba invadens, illus.
Trissl, D.; et al., 1977, J. Exper. Med., v. 145 (3), 652-665
Entamoeba spp., pathogenic strains show cell surface properties different from those of nonpathogenic strains as evidenced by concanavalin A-induced agglutination studies

Entamoeba invadens, illus.
van Vliet, H. H. D. M.; et al., 1976, J. Cell Biol., v. 71 (2), 357-369
Entamoeba invadens, procedure for subcellular fractionation, isolation of phagolysosomal and plasma membranes, characterization (enzymes, lipids, morphology)

Entamoeba invadens
van Vliet, H.H.D.M.; Op den Kamp, J.A.F.; and van Deenen, L.L.M., 1975, J. Protozool., v. 22 (3), 428-432
Entamoeba invadens, development of medium for large-scale axenic cultivation, lipid requirement for growth, fatty-acid composition of amebae grown with different fatty acids and cholesterol

Entamoeba invadens, illus.
Zaman, V., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 229-231
Entamoeba invadens cysts, ultrastructure

Entamoeba moshkovskii, illus.
Artigas, J.; Otto, I.; and Kawada, M. E., 1966, Bol. Chileno Parasitol., v. 21 (4), 114-118
Entamoeba histolytica, pathologic effects of motile trophozoites on human leucocytes compared with non-motile E. moshkovskii and another free-living amoeba both of which showed no action in blood cells; possible use in differentiating E. histolytica from E. hartmanni

Entamoeba moshkovskii, illus.
Pennisi, L.; Faraone, U.; and Todaro, F., 1972, Riv. Parassitol., Roma, v. 33 (3), 161-171
Entamoeba histolytica, 3 strains, E. moshkovskii, 2 strains, trophozoite surface ultrastructure

Entamoeba moshkovskii
Scott, H.; and McMillan, B., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 357 [Letter]
first isolation from sewage in Australia

Entamoeba moshkovskii
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Entamoeba moshkovskii, illus.
Trissl, D.; et al., 1977, J. Exper. Med., v. 145 (3), 652-665
Entamoeba spp., pathogenic strains show cell surface properties different from those of nonpathogenic strains as evidenced by concanavalin A-induced agglutination studies

Entamoeba muris, illus.
Grewal, M. S., 1975, Research Bull. Panjab Univ., v. 23 (3-4), 1972, 119-124
Rattus rattus: Chandigarh, India

Endamoeba muris
Mishra, G. S.; and Gonzalez, J. P., 1975, Arch. Inst. Pasteur Tunis, v. 52 (1-2), 71-87
Rattus norvegicus (intestin grele, gros intestin): Tunis, Tunisia

Entamoeba muris
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Rattus norvegicus
Mus musculus
all from Carshalton

Entamoeba polecki
Desowitz, R. S.; and Saave, J. J., 1965, Med. J. Malaya, v. 20 (2), 153
human (stool): Trobriand Islands

Entamoeba polecki
Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
amebas in fecal matter, techniques of fecal examination, characters for identification, review

Entamoeba testudinus
Kurashvili, B. E., 1975, Izvest. Akad. Nauk Gruzinsk. SSR, s. Biol., v. 1 (4), 317-320
antagonistic and synergetic interrelationships between intestinal parasites

Enterocystidae M. Codreanu, 1940
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
diagnosis; checklist of genera and species of 6 aseptate gregarine families

Enterocystis
Codreanu-Balcescu, D., 1976, J. Protozool., v. 23 (4), 7A [Abstract]
Enterocystis, role of Golgi apparatus in elaboration of paraglycogen

Enterocystis Tsvetkov, 1926
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Enterocystidae
diagnosis; checklist of species

Enterocystis bullis Noble, 1938
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Echiurocystis bullis (Noble, 1938) Levine, 1977

Enterocystis ephemerae (Frantzius, 1848) Desportes, 1963
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
synonymy

Enterocystis fungoides M. Codreanu
Desportes, I., 1974, J. Protozool., v. 21 (1), 83-94
Enterocystis fungoides, trophont, fine structure
Baetidae (tube digestif): Var, a 10 km de Vidauban

Enterocystis greeffi Noble, 1938
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Echiurocystis greeffi (Noble, 1938) Levine, 1977

Enterocystis hydrophili n. comb., illus.
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
Syn.: Sphaerocystis hydrophili Foerster, 1938
description
Hydrochara caraboides: Loire-Atlantique (Ouest de la France)

Enterocystis hydrophili (Foerster, 1938) Baudoin and Maillard, 1972
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Sphaerocystis hydrophili Foerster, 1938

Enterocystis rhithrogenae M. Codreanu, 1940
Codreanu, R.; and Codreanu-Balcescu, D., 1975, J. Protozool., v. 22 (3), 77A-78A [Abstract]
hyperparasitized by Pleistophora hyperparasitica
Rhithrogena semicolorata: environs de la Station Zoologique de Sinaia, Carpates Meridionales, Roumanie

Enterocystis rhithrogenae
Codreanu-Balcescu, D.; and Codreanu, R., 1976, J. Protozool., v. 23 (4), 8A [Abstract]
hyperparasitized by Pleistophora hyperparasitica
Rhithrogena semicolorata (intestin): Sinaia, Roumanie

Enterocystis umushii Iitsuka, 1933, lapsus calami
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Echiurocystis yumushii (Iitsuka, 1933) Levine, 1977

Enterocystis yumushii Iitsuka, 1933
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Echiurocystis yumushii (Iitsuka, 1933) Levine, 1977

Enteromonadina subord. nov.
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Diplomonadida
diagnosis; includes: Enteromonas (type genus)

Enteromonas (type genus of subord.), illus.
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Enteromonadina subord. nov.

Enteromonas sp.
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
ultrastructure
Oryctolagus cuniculi (crottes)

Enteromonas sp.
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
ultrastructure
Triturus vulgaris (intestin)

Enteromonas [sp.]
Brugerolle, G., 1976, J. Protozool., v. 23 (4), 5A [Abstract]
Box salpa (intestin posterieur)

Enteromonas sp.
Nohynkova, E., 1974, J. Protozool., v. 21 (3), 457-458 [Abstract]
Haemopis sanguisuga (intestine): Czechoslovakia

Enteromonas hominis
Best, J. C.; et al., 1976, Med. J. Australia, v. 1 (1-2), 14-20
assessment of growth-rate and growth-retardation in Australian Aboriginal children before and after treatment for common intestinal parasites

Enteromonas hominis
Giboda, M.; Hoffman, F.; and Rimar, P., 1971, Ceskoslov. Pediat., v. 26 (2), 88-89
intestinal protozoal infections in children aged 1-15 years, clinical trials with metronidazole, results and parasites eliminated varied with age and parasite: Kosice, Czechoslovakia

Enteromonas hominis
Vinayak, V. K.; and Sehgal, S. C., 1976, Indian J. Med. Research, v. 64 (9), 1347-1350
human helminthic and protozoan parasites, comparison of nigrosin-methylene blue diagnostic test with formol-ether method and direct examination

Eperythrozoon
Kreier, J. P.; and Gothe, R., 1976, Vet. Parasitol., v. 2 (1), 83-95
Aegyptianella, Eperythrozoon, Grahamella, Haemobartonella, morphology, host-parasite interactions, review

Eperythrozoon
Kreier, J. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 24-28
Rickettsiales, Anaplasmataceae

Eperythrozoon sp.
Adam, K. M. G.; et al., 1976, Parasitology, v. 73 (1), 1-11
antibody-positive blood from wild Cervus elaphus produced Babesia infections in splenectomized C. elaphus (also Eperythrozoon infection resembling wenyoni), possible transient deer Babesia infection produced in 1 of 6 bovine calves, indirect fluorescent antibody tests, "Despite their similarities, specific status for B. divergens and the red deer Babesia is probably justified; at present there is insufficient evidence to justify separation of the red deer Babesia from B. capreoli."

Eperythrozoon [sp.]
Oduye, O. O.; and Dipeolu, O. O., 1976, J. Small Animal Practice, v. 17 (5), 331-337
blood parasites of dogs, single and mixed infections, correlation between incidence and rainfall, degree of parasitaemia, infectivity rate within age groups, no significant difference in host susceptibility of local and exotic breeds to infection: Ibadan, Nigeria

Eperythrozoon [sp.]
Okon, E. D., 1976, Trop. Animal Health and Prod., v. 8 (2), 96
B[abesia] trautmanni, Eperythrozoon, Haematopinus suis, pigs, incidence of infection according to host age groups: Ibadan, Nigeria

Eperythrozoon sp.
Peters, W.; Howells, R. E.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 21 [Demonstration]
Aotus trivirgatus (attached to periphery of erythrocyte)

Eperythrozoon [sp.], illus.
Peters, W.; Molyneux, D. H.; and Howells, R. E., 1974, Ann. Trop. Med. and Parasitol., v. 68 (1), 47-50
Aotus trivirgatus: obtained from Shamrock Farms Ltd., Henfield, Sussex, imported from Colombia

Eperythrozoon sp.
Smith, M. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 18 [Demonstration]
cattle: Trinidad

Eperythrozoon coccoides
Cox, H. W.; and Calaf-Iturri, G., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 73-79
Haemobartonella muris in rats, Eperythrozoon coccoides in mice, production of essentially same disease characterized by anemia with splenomegaly and erythrophagocytosis associated with presence of cold-active haemagglutinin, serum antigen, and antibody to serum in blood

Eperythrozoon coccoides
Schottelius, J., 1977, Tropenmed. u. Parasitol., v. 28 (2), 202-204
latex Chagas test performed on sera of Bartonella bacilliformis-infected humans, Haemobartonella muris-infected rats, and Eperythrozoon coccoides-infected mice, positive reaction observed only with rat sera, nature of this cross-reaction has not been clarified

Eperythrozoon felis and/or Haemobartonella felis
Hayes, H. M.; and Priester, W. A., 1973, J. Small Animal Practice, v. 14 (12), 797-804
feline infectious anaemia, relative risk according to age, sex and breed of cats, prior disease, seasonal occurrence, mortality, pattern of risk suggests horizontal transmission probably by direct contact

Eperythrozoon ovis
Daddow, K. N., 1977, Austral. Vet. J., v. 53 (3), 139-143
Eperythrozoon ovis, sheep (nat. and exper.), complement fixation test, diagnosis

Eperythrozoon ovis
Daddow, K. N.; and Dunlop, L. B., 1976, Queensland Agric. J., v. 102 (1), 88-89
sheep, diagnosis, transmission, symptoms, review

Eperythrozoon ovis
Daddow, K. N.; and Dunlop, L. B., 1976, Queensland J. Agric. and Animal Sc., v. 33 (2), 233-236
Eperythrozoon ovis, sheep, detection by complement fixation test and stained thin blood smears, ewes appeared to be source of infection for lambs, possible role of sandflies and mosquitoes in mode of transmission

Eperythrozoon ovis
Ilemobade, A. A.; and Blotkamp, C., 1977, Vet. Rec., v. 101 (8), 153-154
Eperythrozoon ovis, sheep (nat. and exper.), anaemia, apparent endemic stability in local flocks: Shika Agricultural Research Station near Zaria, Nigeria

Eperythrozoon ovis
Laws, L.; and Daddow, K. N., 1976, Austral. Vet. J., v. 52 (5), 243 [Letter]
Eperythrozoon ovis, sheep (nat. and exper.), diagnosis, pathogenicity tests: Queensland

Eperythrozoon ovis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Eperythrozoon ovis
Semka, Z.; and Rogowska, W., 1976, Med. Wet., v. 32 (9), 523-524
Eperythrozoon ovis, imported sheep, blood values, anemia: Poland

Eperythrozoon ovis
Sutton, R. H., 1976, Austral. J. Exper. Biol. and Med. Sc., v. 54 (5), 449-458
Eperythrozoon ovis, carbohydrate metabolism, infected sheep erythrocytes, increased glucose utilization and lactic acid production

Eperythrozoon ovis
Sutton, R. H., 1977, Austral. Vet. J., v. 53 (10), 478-481
Eperythrozoon ovis, sheep, low venous blood glucose levels, increased blood lactic acid levels

Eperythrozoon teganodes
Hinaidy, H., 1976, Ztschr. Parasitenk., v. 50 (2), 212-213
Rind (Blut): Osterreich

Eperythrozoon teganodes
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Eperythrozoon teganodes
Trees, A. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 287 [Demonstration]
blood from healthy indigenous Nigerian cattle inoculated into splenectomized calf, calf became infected with Babesia bigemina, B. bovis, Anaplasma marginale, Borrelia theileri, Theileria mutans, Eperythrozoon teganodes, E. wenyoni; isolation of pure stabilate of B. bigemina

Eperythrozoon tuomi
Hinaidy, H., 1976, Ztschr. Parasitenk., v. 50 (2), 212-213
Rind (Blut): Osterreich

Eperythrozoon tuomii
Purnell, R. E.; Brocklesby, D. W.; and Sellwood, S. A., 1973, Isotopes and Radiation Parasitol. III, 169-171
Babesia major-infected bovine erythrocytes, ^{60}Co-irradiation and subsequent injection into splenectomized calves, irradiation at 30 or 50 kR appeared to destroy virulence without altering immunogenicity but results must be interpreted with caution because of the presence of Eperythrozoon tuomii in blood of calves after challenge

Eperythrozoon wenyoni
Adam, K. M. G.; et al., 1976, Parasitology, v. 73 (1), 1-11
antibody-positive blood from wild Cervus elaphus produced Babesia infections in splenectomized C. elaphus (also Eperythrozoon infection resembling wenyoni), possible transient deer Babesia infection produced in 1 of 6 bovine calves, indirect fluorescent antibody tests, "Despite their similarities, specific status for B. divergens and the red deer Babesia is probably justified; at present there is insufficient evidence to justify separation of the red deer Babesia from B. capreoli."

Eperythrozoon wenyoni, illus.
Craig, T. M., 1975, Southwest. Vet., v. 28 (3), 243-245
Eperythrozoon wenyoni, morphology, scanning electron microscope

Eperythrozoon wenyoni
Hinaidy, H., 1976, Ztschr. Parasitenk., v. 50 (2), 212-213
Rind (Blut): Osterreich

Eperythrozoon wenyoni
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Eperythrozoon wenyoni, illus.
Poole, D. B. R.; et al., 1976, Vet. Rec., v. 99 (24), 481
Eperythrozoon wenyoni, cattle, clinical signs associated with anaemia

Eperythrozoon wenyoni
Purnell, R. E.; Brocklesby, D. W.; and Young, E. R., 1976, Vet. Rec., v. 98 (20), 411
Eperythrozoon wenyoni, calf (blood), possible cause of anemia in British cattle

Eperythrozoon wenyoni
Trees, A. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 287 [Demonstration]
blood from healthy indigenous Nigerian cattle inoculated into splenectomized calf, calf became infected with Babesia bigemina, B. bovis, Anaplasma marginale, Borrelia theileri, Theileria mutans, Eperythrozoon teganodes, E. wenyoni; isolation of pure stabilate of B. bigemina

Epistylis [sp.], illus.
Hubert, W. A.; and Warner, M. C., 1975, J. Wildlife Dis., v. 11 (2), 241-244
Epistylis, channel catfish, bioassays of potential control chemicals, salt alone and combined with formalin most effective: experimental raceways, Gallatin, Tennessee

Epistylis sp., illus.
Overstreet, R. M.; and Howse, H. D., 1977, Ann. N. York Acad. Sc., v. 298, 427-462
helminths and protozoans of estuarine fishes, incidence and intensity; possible relationships with water pollutants; tumorous growths
Archosargus probatocephalus
Ictalurus punctatus
Lepomis macrochirus
L. microlophus (integument)
pumpkinseed
warmouth
largemouth bass
black drum
all from estuaries of Mississippi

Esarabdina synaptae Mingazzini, 1893
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Selenidium synaptae (Mingazzini, 1893) Levine, 1971

Esarabdina synaptae (Cuenot, 1891) Mingazzini, 1893
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora synaptae (Cuenot, 1891) Leger, 1892

Eucoccidium of Grell, 1953, preoccupied non Eucoccidium Luehe, 1902
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
as syn. of Grellia gen. n. [i.e., nom. nov.]

Eucoccidium dinophili Grell, 1953
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
as syn. of Grellia dinophili (Grell, 1953) nom. nov. [i.e., n. comb]

Eucoccidium ophryotrochae Grell, 1960
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
as syn. of Grellia ophryotrochae (Grell, 1960) nom. nov. [i.e., n. comb]

Eudrilophrya de Puytorac, 1971
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Eudrilophryinae

Eudrilophrya complanata de Puytorac, 1970, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eudrilus eugeniae

Eudrilophryinae de Puytorac, 1971
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam.
includes: Eudrilophrya

Eugregarinida (Doflein 1901)
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
Eugregarinida, ultrastructure of syzygial junction, epimerite and septum in gamonts, specificity and taxonomic significance of these structures

Eumonodontophrya kijenskyi Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Monodontophrya kijenskyi Cheissin, 1930

Euspora fallax Schneider, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Melolontha melolontha (intestine): Poland

Euspora fallax A. Schneider
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Allecula letestui: Gabon

Extremocystis Setna, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Extremocystis dendrostomi Setna, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Fabespora vermicola sp. n., illus.
Overstreet, R. M., 1976, J. Parasitol., v. 62 (5), 680-684
infection acts as biological control agent by stopping reproduction in digenean host Crassicutis archosargi (among parenchymal cells, surrounding reproductive organs, integument): lower portion of Escatawpa River, Jackson County, Mississippi

Farinocystis tribolii Weiser
Zizka, Z., 1973, J. Protozool., v. 20 (4), 537
Farinocystis tribolii, ultrastructure

Farinocystis tribolii Weiser
Zizka, Z., 1974, J. Protozool., v. 21 (3), 459 [Abstract]
Farinocystis tribolii, life cycle

Faucispora phryganeae (Berg von Emme, 1912) Baudoin, 1967
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Diplocystis phryganeae Berg von Emme, 1912

Ferraria Setna, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Bhatiella Setna, 1931

Ferraria cornucephali Setna, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Bhatiella marphysae Setna, 1931 emend.

Ferraria cornucephala iwamusi Hoshide, 1944
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Bhatiella iwamusi (Hoshide, 1935) comb. nov.

Filipodium Hukui, 1939
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Filipodium aspidosiphoni Tuzet & Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Filipodium ozakai Hukui, 1939
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Syn.: Lithocystis parquerae [sic] Jones, 1971

Filipodium ozakai Hukui, 1939
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Lithocystis parquerae [sic] Jones, 1971

Filipodium ozakii Hukui 1939
Theodorides, J., 1975, Ann. Parasitol., v. 50 (1), 141
Syn.: Lithocystis parguerae I. Jones 1971

Flagellates, epimastigote
Ashford, R. W., 1974, J. Med. Entom., v. 11 (5), 605-616
Phlebotomus duboscqi (mid-gut and hind-gut)
Sergentomyia affinis (mid-gut)
all from Ethiopia

Flagellate parasites
Ashford, R. W.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 265-271
Phlebotomus papatasi: Bir Ayyad, Libya

Flagellates, illus.
Ashford, R. W.; Bray, R. S.; and Bray, M. A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 385-386
trypanosomes from Riparia paludicola (blood and bone marrow) possibly the same species as the flagellates from Ornithomyia fur (hind and mid gut) that had been removed from the birds' nests, possible transmission to the birds when the flies are eaten during preening: Lake Koka, Ethiopia

Flagellates
Lainson, R.; et al., 1976, Acta Amazonica, v. 6 (4), 55-60
Lutzomyia yuilli: Municipio of Aripuana, Mato Grosso State, Brazil

Flagellates, resembling Trichomonas microti Wenrich and Saxe 1950
Winchell, E. J., 1977, J. Parasitol., v. 63 (4), 756-757
Microtus breweri (intestines): Muskeget Island, 5 miles west of Nantucket, Massachusetts

Frenkelia
Beier, T. V.; Siim, I. K.; and Hutchison, W. M., 1977, Tsitologiia, v. 19 (12), 1362-1368
Eimeriidae, Isosporinae

Frenkelia Biocca 1968
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Sarcocystidae, Sarcocystinae
diagnosis, life cycle pattern

Frenkelia Biocca 1968
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Eimeriidae
diagnosis
"synonymize Endorimospora Tadros and Laarman, 1976 with Sarcocystis in part and Frenkelia in part."

Frenkelia
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    Sarcocystis and related coccidian parasites, general review with discussion of biological aspects of life cycles and new proposal for classification

Frenkelia [sp.]
  Burtscher, H.; and Meingassner, J. G., 1976, Ztschr. Parasitenk., v. 50 (2), 220
    brief description
    Chinchilla laniger (Gehirn)

Frenkelia sp., illus.
  Hayden, D. W.; King, N. W.; and Murthy, A. S. K., 1976, Vet. Path., v. 13 (5), 337-342
    Frenkelia sp. cysts associated with granulomatous encephalitis, in laboratory-reared rats, spontaneous infection (brain, cervical spinal cord): Wilmington, Massachusetts

Frenkelia [sp.], illus.
  Krampitz, H. E.; and Rommel, M., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (1), 17-19
    Microtus agrestis (nat. and exper.) (brain): westlich Munchens
    Buteo buteo (exper.) (feces)
    Microtus arvalis (exper.)
    Apodemus sylvaticus (exper.)
    A. flavicollis (exper.)
    A. agrarius (exper.)
    golden hamster (exper.)
    white laboratory rat (exper.)

Frenkelia sp., illus.
  Meingassner, J. G.; and Burtscher, H., 1977, Vet. Path., v. 14 (2), 146-153
    Frenkelia sp. and Toxoplasma gondii, mixed, fatal infection in caged Chinchilla laniger (brain)

Frenkelia sp.
  Mueller, B. E. G.; Mehlhorn, H.; and Scholtyseck, E., 1973, J. Protozool., v. 20 (4), 505
    ultrastructure and origin of cyst wall in Toxoplasmatea

Frenkelia sp., illus.
  Scholtyseck, E.; Mehlhorn, H.; and Mueller, B.E.G., 1974, J. Protozool., v. 21 (2), 284-294
    Sarcocystis tenella, Besnoitia jellisoni, Frenkelia sp., Toxoplasma gondii, nature of cyst, fine structure of cyst wall

Frenkelia sp. (M-organism), illus.
  Senaud, J.; Chobotar, B.; and Scholtyseck, E., 1976, Tropenmed. u. Parasitol., v. 27 (2), 145-159
    comparative fine structure morphology of Plasmodium cathemerium, Eimeria ferrisi, E. stiedai, Besnoitia jellisoni, and Frenkelia sp. micropores, discussion of micropore function in parasite nutrition

Frenkelia clethrionomyobuteonis, illus.
  Krampitz, H. E.; et al., 1976, Ztschr. Parasitenk., v. 51 (1), 7-14
    Frenkelia clethrionomyobuteonis, asexual development in bank vole, Clethrionomys glareolus

Frenkelia clethrionomyobuteonis
  Rommel, M.; et al., 1976, Ztschr. Parasitenk., v. 50 (2), 204-205
    Frenkelia clethrionomyobuteonis, life cycle

Frenkelia clethrionomyobuteonis, illus.
  Rommel, M.; Krampitz, H. E.; and Geisel, O., 1977, Ztschr. Parasitenk., v. 51 (2), 139-146
    Frenkelia clethrionomyobuteonis, sexual development in small intestine of Buteo buteo (exper.) infected with cysts from Clethrionomys glareolus

Frenkelia clethrionomyobuteonis Rommel & Krampitz, 1975
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora buteonis (Henry, 1932) comb. nov.

Frenkelia glareoli
  Kepka, O.; and Rezaeian, M., 1976, Ztschr. Parasitenk., v. 50 (2), 210
    Frenkelia glareoli, Sarcocystis tenella, chemotaxonomic comparison, disc electrophoresis, serology, close but not identical species

Frenkelia glareoli (Erhardova, 1955) Tadros, Bird & Ellis, 1972
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    Syn.: Toxoplasma glareoli Erhardova, 1955

Frenkelia glareoli Biocca, 1968
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora buteonis (Henry, 1932) comb. nov.

Frenkelia microti (Findlay & Middleton, 1934) Biocca, 1968
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    Syn.: Toxoplasma microti Findlay & Middleton, 1934

Gamocystis ephemerae, illus.
  Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
    ultrastructure
    Ephemera vulgata
    E. danica

Gamocystis francisci Schneider, 1882
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    as syn. of Enterocystis ephemerae (Frantzius, 1848) Desportes, 1963

Gamocystis tenax Schneider, illus.
  Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
    description
    Ectobius lapponicus (intestine): Poland

Gamocystis tenax, illus.
  Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
    ultrastructure
    Ectobius silvestris

Ganapatiella n. gen.
  Kalavati, C., 1977, Arch. Protistenk., v. 119 (3), 217-223
    Adeleina, Adeleidae
    mt: Ganapatiella odontotermi n. sp.

Ganapatiella odontotermi n. sp. (mt), illus.
  Kalavati, C., 1977, Arch. Protistenk., v. 119 (3), 217-223
    life cycle
    Odontotermes obesus (adipose tissue, coelomic fluid): University Campus, India

Ganymedes Huxley, 1910
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    Ganymedidae
    diagnosis; checklist of species

Ganymedes
  Theodorides, J.; and Desportes, I., 1975, J. Protozool., v. 22 (3), 85A-86A [Abstract]
    gregarines of marine invertebrates, organization of epicyte

Ganymedes apsteini Theodorides & Desportes, 1972
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Paraophioidina apsteini (Theodorides & Desportes, 1972) comb. nov.

Ganymedes apsteini Theodorides and Desportes, 1972
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    as syn. of Paraophioidina apsteini (Theodorides & Desportes, 1972) Levine, 1977

Ganymedes haeckeli (Mingazzini, 1893) Theodorides & Desportes, 1972
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Paraophioidina haeckeli (Mingazzini, 1891) comb. nov.

Ganymedes haeckeli (Mingazzini, 1893) Theodorides and Desportes, 1972
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    as syn. of Paraophioidina haeckeli (Mingazzini, 1893) Levine, 1977

Ganymedes oaklandi Jones, 1968
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Paraophioidina oaklandi (Jones, 1968) comb. nov.

Ganymedes oaklandi Jones, 1968
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    as syn. of Paraophioidina oaklandi (Jones, 1968) Levine, 1977

Ganymedes vibiliae Theodorides & Desportes, 1972
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Paraophioidina vibiliae (Theodorides & Desportes, 1972) comb. nov.

Ganymedes vibiliae Theodorides and Desportes, 1972
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    as syn. of Paraophioidina vibiliae (Theodorides and Desportes, 1972) Levine, 1977

Ganymedidae Huxley, 1910
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    diagnosis; checklist of genera and species of 6 aseptate gregarine families

Gastrocystis besnoiti (Marotel 1912) Brumpt 1913
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Besnoitia besnoiti (Marotel 1912) Henry 1913

Georgevitchiella de Puytorac, 1957
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Maupasellidae, Maupasellinae

Georgevitchiella aculeata (Georgevitch, 1943) de Puytorac, 1957
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Syn.: Mesnilella aculeata Georgevitch, 1943
    Eiseniella

Giardia Kunstler
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Diplomonadina emend.

Giardia
Fortess, E.; and Meyer, E. A., 1976, J. Parasitol., v. 62 (5), 689
Giardia trophozoites, isolation from guinea pig, axenization, long-term culture, low temperature preservation

Giardia
Gheorghescu, P.; et al., 1976, Med. Interne, Bucarest, v. 14 (1), 31-38
Giardia infections alone or in combination with Strongyloides or Taenia solium, absorption studies before and after treatment

Giardia
Holberton, D. V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 29-30 [Demonstration]
Giardia, mechanism of attachment to wall of small intestine

Giardia
Meuwissen, J. H. E. T.; v. Tongeren, J. H. M.; and Werkman, H. P. T., 1977, Lancet, London (8027), v. 2, 32-33 [Letter]
human giardiasis, routine fecal examination of patients with abdominal complaints disclosed that presence of infection was not necessarily related to travel in endemic areas as previously had been reported: the Netherlands

Giardia
Thompson, A.; et al., 1977, J. Clin. Path., v. 30 (3), 292-294 [Letter]
giardiasis, human, immunoglobulin-bearing plasma cells in jejunal mucosae, results indicate that early immune response in jejunal lamina propria may frequently be restricted to synthesis of IgM to be followed by IgA and IgG

Giardia sp., illus.
Colley, F. C.; and Mullin, S. W., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (2), 257
description of cysts
Tragulus javanicus (feces): West Malaysia

Giardia sp.
Rajasekaran, P.; Dutt, P. R.; and Pisharoti, K. A., 1977, Indian J. Med. Research, v. 66 (2), 189-199
human intestinal parasites, survey of correlation between infection rate and source of water supply (well, street tap, home with tap water) as indication of control of water-borne diseases by public water supplies: Madurai district, Tamil Nadu, India

Giardia canis, illus.
Pfeiffer, H.; and Supperer, R., 1976, Wien. Tierarztl. Monatsschr., v. 63 (1), 1-6
Giardia canis, dogs, first report, case histories, successful treatment with Flagyl: Austria

Giardia canis
Yang, J.; and Scholten, T., 1977, Am. J. Clin. Path., v. 67 (3), 300-304
diagnosis of human intestinal parasites, fecal examination technique using Junod's fixative for concentration and permanent staining procedures, comparison with results using formalin-ether procedure

Giardia duodenalis
Brightman, A. H. II; and Slonka, G. F., 1976, J. Am. Animal Hosp. Ass., v. 12 (4), 492-497
Giardia duodenalis, cats (feces), gastrointestinal illness, quinacrine hydrochloride, good results

Giardia duodenalis
Holberton, D. V., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 284 [Demonstration]
Giardia duodenalis mastigont, dissection

Giardia intestinalis
Dellamonica, P.; et al., 1976, Nouv. Presse Med., v. 5 (30), 1913 [Letter]
Giardia intestinalis in woman with associated urticaria and dysentery, relief of symptoms after metronidazole therapy, clinical case report: Nice, France

Giardia intestinalis, illus.
Grewal, M. S., 1975, Research Bull. Panjab Univ., v. 23 (3-4), 1972, 119-124
Rattus rattus (jejunum, duodenum): Chandigarh, India

Giardia intestinalis
Kociecka, W.; Lisowska, M.; and Gerwel, M., 1974, Polski Tygod. Lekar., v. 29 (28), 1219-1220
Giardia intestinalis, outbreak of giardiasis in tourist group who had travelled to Lebanon and Syria, water supply of hotel thought to be source of infection: Poland

Giardia intestinalis (G. lamblia)
Lagardere, B., 1976, Medecine Infant., v. 83 (1), 55-60
clinical review of Giardia intestinalis infection in children

Giardia intestinalis, illus.
Lapierre, J.; et al., 1975, Medecine et Malad. Infect., v. 5 (2), 87-92
statistical clinical analysis of 368 cases of human giardiasis, diagnosis, epidemiology, metronidazole: Paris

Giardia lambli
Nitzulescu, V., [1976], Ann. Parasitol., v. 50 (5), 1975, 647-648
emendation of Giardia lamblia Stiles, 1915

Giardia lamblia
Anderson, R. I.; and Buck, A. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 447-456
complement levels in residents of rural village in relation to wide variety of clinical, laboratory, and epidemiological factors including parasitic diseases: Ouli Bangala, Republic of Chad

Giardia lamblia
Banki, F.; Farkas, A.; and Szanto, I., 1972, Orvosi Hetilap, v. 113 (21), 1295-1296
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Giardiasis
Radermecker, M.; et al., 1974, Internat. Arch. Allergy and Applied Immunol., v. 47 (2), 285-295
various human helminthic or protozoal infections, serum IgE concentration, IgE level often raised in parasitosis with prominent tissue phases and remains normal with helminths restricted to lumen of digestive tract, IgE level tends to increase significantly and rapidly following specific treatment and then to decrease slowly and return to normal in a few months

Giardiasis
Rigillo, N.; et al., 1969, Minerva Pediat., v. 21 (21), 960-965
giardiasis in children, secondary malabsorption and altered xylose test values return to normal after disappearance of parasite from stool

Giardiasis
Tomecki, R., 1974, Mater. Med. Pol. (21), v. 6 (4), 301-303
giardiasis, successful use of nifosept in human intestinal infection refractory to treatment with other drugs

Giardiasis
Vignola, G.; Mori, P. G.; and Cottafava, F., 1969, Minerva Pediat., v. 21 (33), 1526-1530
mixed giardiasis and Trichocephalus infections in young child resulting in severe malabsorption syndrome, pathology of intestinal mucosa

Gigaductus africanus n. sp., illus.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Plangiopsis sp. (paroi intestinale): Station d'Ecologie tropicale de Lamto situee entre N'Douci et Toumodi, a 200 km environ au nord d'Abidjan, Cote d'Ivoire

Globidium besnoiti (Marotel 1912) Wenyon 1926
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Besnoitia besnoiti (Marotel 1912) Henry 1913

Globidium besnoiti Wenyon, 1926
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora besnoiti (Marotel, 1912) comb. nov.

Globidium gilruthi
Porchet, E.; and Dubremetz, J. F., 1976, J. Protozool., v. 23 (4), 18A [Abstract]
Globidium gilruthi, formation of supernumerary conoids during course of schizogony

Globidium (Globidium) gilruthi, illus.
Tontis, A.; Koenig, H.; and Hoerning, B., 1977, Schweiz. Arch. Tierh., v. 119 (4), 161-165
Globidium gilruthi, pathology
Schaf (Magen-Darm-Trakt)
Ziege (Magen-Darm-Trakt)
Rind (Magen-Darm-Trakt)

Globidium leuckarti (Eimeria leuckarti), illus.
Wheeldon, E. B.; and Greig, W. A., 1977, Vet. Rec., v. 100 (6), 102-104
Globidium leuckarti, horse, clinical signs, diarrhea, case report, pathogenicity reviewed: United Kingdom

?Glossatella spp. (Butschlii, 1889)
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Perca fluviatilis: Loch Leven, Scotland

Glossatella sp.
Heckmann, R.; and Farley, D. G., 1973, J. Wildlife Dis., v. 9 (3), 221-224
Hesperoleucus symmetricus symmetricus (gills, body surface): foothill streams east of Fresno, California

Glugea
Weiser, J., 1975, J. Protozool., v. 22 (3), 70A [Abstract]
microsporidians in simuliids (including Pleistophora debaisieuxi in Odagmia ornata; and Glugea), provocation of neoplastic xenoma

Glugea [sp.]
Jakowska, S., 1964, An. 2. Cong. Latino-Am. Zool., v. 1, 265-273
Lophius americanus (celulas nervosas): Aquario de New York

Glugea anomala
Dykova, I.; and Lom, J., 1977, J. Protozool., v. 24 (4), 50A [Abstract]
Glugea anomala, tissue response in Gasterosteus aculeatus (exper.), site of infection differed according to route of spore administration, characteristics of xenomas

Glugea anomala Moniez
Landry, G., 1976, Naturaliste Canad., v. 103 (6), 583-584
Gasterosteus aculeatus: lac Temiscouata (Quebec)

Glugea anomala (Moniez, 1887)
Willemse, J. J., 1968, Bull. Zool. Mus. Univ. Amsterdam, v. 1 (8), 83-87
Anguilla anguilla (gonads): IJsselmeer
Pygosteus pungitius: Schellingwoude

Glugea fumiferana (Thomson)
McLaughlin, R. E., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 95-98
entomopathogenic protozoa as control agents of insect pests, methods of application, economic factors, further research needs, review

Glugea habrodesmi
Bouix, G.; et al., 1976, J. Protozool., v. 23 (4), 5A [Abstract]
Glugea habrodesmi, development, ultrastructure
Habrodesmus falx
Oxydesmus granulosus

Glugea pyraustae (Paillot)
McLaughlin, R. E., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 95-98
entomopathogenic protozoa as control agents of insect pests, methods of application, economic factors, further research needs, review

Glugea stephani
Olson, R. E., 1975, J. Protozool., v. 22 (3), 29A [Abstract]
Parophrys vetulus were exper. infected with Glugea stephani by feeding on Artemia salini and Corophium spinicorne that had been allowed to ingest spores, temperature found to be of critical importance in establishment and development of G. stephani

Glugea stephani (Hagenmuller, 1899) Woodcock, 1904, illus.
Olson, R. E., 1976, J. Protozool., v. 23 (1), 158-164
Glugea stephani in Parophrys vetulus, incidence and intensity of infection in different months, field and laboratory infections only occurred at temperatures above 15 C, successful transmission by Artemia salina or Corophium spinicorne vectors as well as direct ingestion of spores, heavy infections are potentially lethal
Parophrys vetulus (xenomas in lamina propria of intestinal tissue and in liver tissue) (nat. and exper.): upper Yaquina Bay estuary, Oregon
Artemia salina (exper.)
Corophium spinicorne (exper.)

Glugea stephani, illus.
Weidner, E., 1976, J. Protozool., v. 23 (2), 234-238
Glugea stephani-induced xenoma in Pseudopleuronectes americanus, ultrastructure of peripheral zone

Gonospora Schneider, 1875
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Urosporidae
synonymy, checklist of species

Gonospora arenicolae (Cunningham, 1907) Tregouboff, 1918
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
synonymy

Gonospora gonadipertha (Djakonov, 1923) new combination
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Syn.: Diplodina gonadipertha Djakonov, 1923

Gonospora goodrichae nomen novum
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Syn.: Gonospora minchinii Goodrich & Goodrich, 1920 non G. minchinii (Woodcock, 1904) Tregouboff, 1918

Gonospora intestinalis (Ssokoloff, 1913) Tregouboff, 1918
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Gonospora irregularis (Minchin, 1893) Tregouboff, 1918
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Gonospora longissima Caullery & Mesnil, 1898
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Urospora longissima (Caullery & Mesnil, 1898)

Gonospora minchini (Woodcock, 1904) Tregouboff, 1918
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Gonospora minchinii Goodrich & Goodrich, 1920 non G. minchinii (Woodcock, 1904) Tregouboff, 1918
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Gonospora goodrichae nomen novum

Gonospora minchinii Goodrich & Goodrich, 1920
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    not a syn. of Gonospora minchini (Woodcock, 1904) Tregouboff, 1918

Gonospora stichopi (Lutzen, 1967) new combination
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Cystobia stichopi Lutzen, 1967

Gonospora terebellae (von Koelliker) Schneider, 1875?
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Gonospora varia Leger, 1892

Gonospora testiculi (Tregouboff, 1916) Tregouboff, 1918
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Cystobia testiculi Tregouboff, 1916

Gonospora varia Leger, 1892
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Gonospora terebellae (von Koelliker) Schneider, 1875?

Grahamella
  Kreier, J. P.; and Gothe, R., 1976, Vet. Parasitol., v. 2 (1), 83-95
    Aegyptianella, Eperythrozoon, Grahamella, Haemobartonella, morphology, host-parasite interactions, review

Grahamella
  Kreier, J. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 24-28
    Eubacteriales, Bartonellaceae

Grahamella sp., illus.
  Wiger, R., 1973, J. Wildlife Dis., v. 9 (2), 125-128
    Lemmus lemmus (blood): Norway

Grahamella [sp.]
  Wiger, R., 1973, Norwegian J. Zool., v. 21 (4), 325 [Abstract]
    Trypanosoma [sp.] and Grahamella [sp.], morphophysiological responses of naturally infected Lemmus lemmus and Clethrionomys glareolus: Norway

Grahamella [sp.]
  Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
    Peromyscus maniculatus (blood): Petty Ranch near Frazier Mountain Park, Ventura County, California
    Perognathus fallax: Juniper Hills near Pearblossom, Los Angeles County, California
    Peromyscus truei: Juniper Hills near Pearblossom, Los Angeles County, California

Grebnickiella
  Ormieres, R.; and Marques, A., 1976, J. Protozool., v. 23 (4), 15A-16A [Abstract]
    Echinomera, Grebnickiella, Dactylophorus, mode of fixation to host

Grebneckiella pixellae, illus.
  Singotam, L.; and Dass, C. M. S., 1977, Indian J. Exper. Biol., v. 15 (9), 719-727
    Grebneckiella pixellae, scanning electron microscopy of various life cycle stages reveals: epimerite as attachment organ to host gut epithelium, epicytic folds of body wall at various regions of trophozoite changes in body folds during gamont formation and sexual dimorphism, syzygy, gamete formation, spore structure, possible role of epicytic folds in gregarine movement and absorption of nutrients
    Scolopendra morsitans (midgut): in and around Delhi

Gregarina spp.
  Carter, J. B., 1976, J. Applied Ecol., v. 13 (1), 103-122
    Tipula paludosa
    T. oleracea
    T. czizeki
    T. luna
    T. unca
    T. vernalis
    T. variipennis
    T. marmorata or alpium?
    Prionocera turcica
    all from north-east England

Gregarina sp., illus.
  Heller, G.; and Weise, R. W., 1973, J. Protozool., v. 20 (1), 61-64
    Gregarina sp. from Udeopsylla nigra, scanning electron microscopy, patterns of epicytic folds in old vs. young gamonts, relationship to motility and development

Gregarina sp.
  Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
    Passalid[ae]: Khao Yai, Thailande

Gregarina sp.
  Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
    Basidentitermes potens: Cote d'Ivoire

Gregarina sp., illus.
  Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
    Cryphaeus taurus: Gabon

Gregarina acridiorum Leger, 1893, illus.
  Corbel, J.-C., 1973, Acrida, v. 2 (2), 69-75
    Gregarina acridiorum, description, affinities with closely related species, geographical distribution, host specificity
    Anonconotus alpinus: France
    Platycleis tessellata: France
    Ephippiger perforatus: France
    E. terrestris: France
    Calliptamus italicus: France; Espagne
    C. i. var. marginellus: France
    C. itericus: France
    C. wattenwylianus: France
    Acrotylus patruelis: "
    Oedaleus decorus: France
    Oedipoda coerulescens: France
    O. fuscocincta var. coerulea: Espagne
    Sphingonotus coerulans: France
    Arcyptera brevipennis: "
    Myrmeleotettix maculatus: France
    Stenobothrus lineatus: France

Gregarina amarae, illus.
  Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
    ultrastructure
    Amara aulica

Gregarina amarae (Hammerschmidt) Frantzius
  Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
    Amara sp.: Coree du sud

Gregarina ausoniae Ghidini et Moriggi
  Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
    Cubitermes sp.: Cote d'Ivoire

Gregarina blaberae
  Philippe, M., 1974, J. Protozool., v. 21 (3), 474-475 [Abstract]
    Gregarina blaberae, trophozoite and cyst, analysis of polysaccharides

Gregarina blaberae
  Philippe, M.; and Cottin, R., 1976, J. Protozool., v. 23 (4), 17A [Abstract]
    Gregarina blaberae, cell surface membrane, carbohydrates

Gregarina blaberae
  Philippe, M.; and Schrevel, J., 1977, J. Protozool., v. 24 (4), 63A [Abstract]
    Gregarina blaberae, proteins and glycoproteins of cortical membranes

Gregarina blaberae
  Schrevel, J.; et al., J. Protozool., v. 23 (4), 19A-20A [Abstract]
    Gregarina blaberae, study of surface carbohydrates with lectins

Gregarina blaberae
  Schrevel, J.; Philippe, M.; and Tronchin, G., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
    Gregarina blaberae trophozoite, body wall, structural, biochemical, and physiological aspects

Gregarina blaberae
  Tronchin, G., 1974, J. Protozool., v. 21 (3), 475-476 [Abstract]
    Gregarina blaberae in Blaberus craniifer (exper.), relationship between life cycle of parasite and host

Gregarina blaberae Frenzel, illus.
  Tronchin, G.; and Schrevel, J., 1977, J. Protozool., v. 24 (1), 67-82
    Gregarina blaberae, development of sporozoite, growth and development of trophozoite, ultrastructural study
    Blaberus craniifer (exper.)

Gregarina blattarum Siebold, illus.
  Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
    synonymy, description
    Blatta orientalis
    Periplaneta americana
    (intestine of all): all from Poland

Gregarina bonelliae Frenzel, 1885
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Ophioidina bonelliae (Frenzel, 1885) Labbe, 1899

Gregarina chrysomelae sp. n., illus.
  Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
    Chrysomela polita (gastric caeca): Bialowieza, Poland

Gregarina cionae Frenzel, 1885
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891

Gregarina clavata von Kolliker, 1848
  Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
    as syn. of Enterocystis ephemerae (Frantzius, 1848) Desportes, 1963

Gregarina clavellinae von Kolliker, 1848
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria clavellinae (von Kolliker, 1848)

Gregarina coccinellae sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
[lapsus p. 177 as G. cocinellae sp. n.]
Syn.: Gregarina barbarara Watson sensu Foerster, 1938
Coccinella septempunctata: Poznan and Mielno Koszalinskie, Poland
Hippodamia tredecimpunctata: Bialowieza, Poland
(intestine of all)

Gregarina cocinellae n. sp. [lapsus p. 177 for G. coccinellae n. sp.]
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179

Gregarina crenata (Bhatia et Setna)
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Rhaphidopalpa similis: Bangkok (Bangkhen), Thailande

Gregarina crenata (Bhatia et Setna)
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Chrysolina staphylea: Coree du sud

Gregarina culicidis Ross, 1895 lapsus calami for G. culicis Ross, 1898
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis culicis (Ross, 1898) Grasse, 1953

Gregarina culicis Ross, 1898 lapsus calami as G. culicidis Ross, 1895
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis culicis (Ross, 1898) Grasse, 1953

G[regarina] cuneata
Harry, O. G., 1973, J. Protozool., v. 20 (4), 526
4 species of gregarines in Tenebrio molitor, incidence, effect of seasonal changes in temperature and humidity on infection: British Isles

Gregarina cuneata Stein, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Tenebrio molitor (intestine): Poland

Gregarina cuneata, illus.
Ruehl, H., 1976, Ztschr. Parasitenk., v. 48 (3-4), 199-214
Gregarina spp., cytochalasin B (phomin) inhibiting movement, changing cell fine structure and disorganizing fibrillar bundles or myonemes; fine structure of myonemes, skeleton-fibrils and pellicle; elements and mode of movement
Tenebrio molitor

Gregarina cuneata, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure

Gregarina cuneata Stein
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Tenebrio obscurus: Coree du sud

Gregarina darchenae n. sp., illus.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Cubitermes sp.: Station d'Ecologie tropicale de Lamto situee entre N'Douci et Toumodi, a 200 km environ au nord d'Abidjan, Cote d'Ivoire

Gregarina decourti n. sp., illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Pseudoblaps javana: Phuket and Hua Hin, Thailande

Gregarina delmasi Tuzet et Rambier
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Catoptropteryx apicalis: Gabon

Gregarina diabotrica Watson-Kamm, 1918: Theodorides, 1954
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina munieri (Schneider)

Gregarina dimorpha Filipponi, 1974
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
synonymy
Chlaeniellus inops: Coree du sud

Gregarina distapliae Bogoyavlensky, 1926
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria distapliae (Mingazzini, 1891) Ormieres, 1965

Gregarina embiae n. sp., illus.
Loubes, Cl.; Ormieres, R.; and Bouix, G., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 519-530
Gregarina embiae n. sp., life cycle, ultrastructure of cephalin
Monotylota ramburi
Haploembia solieri
(intestin of all): all from Midi de la France

Gregarina ephemerae (Frantzius, 1848) Diesing, 1859
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Enterocystis ephemerae (Frantzius, 1848) Desportes, 1963

Gregarina fallax n. sp., illus.
Ormieres, R., [1976], Ann. Parasitol., v. 50 (5), 1975, 521-529
Gregarina ovata, life cycle, production of macrosporocysts, compared with Gregarina fallax n. sp. life cycle producing microsporocysts, host specificity
Forficula auricularia (nat. and exper.): Alpes; Angleterre
Anechura bipunctata: Alpes

Gregarina forficulae sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Forficula auricularia (intestine): Poznan, Poland

Gregarina fusipara n. sp., illus.
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
Hydrochara caraboides: Loire-Atlantique (Ouest de la France)

Gregarina grassei Theod., Orm. et Jol.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Cosmoderus erinaceus: Gabon

Gregarina harpali sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Harpalus aeneus (intestine): Kobylniki, Poland

Gregarina holothuriae Schneider, 1858
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora holothuriae (Schneider, 1858) Tregouboff, 1918

Gregarina hylobii, illus.
Ruehl, H., 1976, Ztschr. Parasitenk., v. 48 (3-4), 199-214
Gregarina spp., cytochalasin B (phomin) inhibiting movement, changing cell fine structure and disorganizing fibrillar bundles or myonemes; fine structure of myonemes, skeleton-fibrils and pellicle; elements and mode of movement
Hylobius abieta

Gregarina hylobii, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Hylobius abietis

Gregarina hypophloei sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Hypophloeus unicolor (intestine): Rogalin, Poland

Gregarina irregularis Minchin, 1893
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora irregularis (Minchin, 1893) Tregouboff, 1918

Gregarina juli: Diesing, 1851
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora juli (Frantzius)

Gregarina juli marginati Leidy, 1853
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora julimarginati (Leidy)

Gregarina katherina Watson sensu Foerster, 1938
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina ruszkowskii sp. n.

Gregarina lagenoides, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Lepisma saccharina

Gregarina macrocephala (A. Schneider)
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Mecopoda elongata: Khao Yai, Thailande

Gregarina macrocephalia sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Aphodius depressus (intestine and gastric caeca): Chludowo, Poland

Gregarina miescheriana, Rivolta, 1878, pro parte
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
as syn. of Sarcocystis suihominis n. spec.

Gregarina minuta Ishii, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Tribolium confusum (intestine): Poland

Gregarina monoducta Hoshide, illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Diestrammena japonica: Coree du sud

Gregarina munieri (Schneider), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Chrysomela coerulans (intestine): Poland

Gregarina munieri (A. Schneider)
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Platycorinus peregrinus: Lam Narai, Thailande
Rhaphidopalpa similis: Bangkok (Bangkhen), Thailande

Gregarina munieri (A. Schneider)
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Chrysolina aurichalcea
C. staphylea
all from Coree du sud

Gregarina nigra M. E. Watson
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Patanga succincta: Lam Narai, Thailande
Aularches miliaris: Lam Narai and Hua Hin, Thailande

Gregarina nigra Watson
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Catantops spissus: Cote d'Ivoire

Gregarina oblongata (Hamm.) Diesing, 1851
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stylocephalus oblongatus (Hammerschmidt)

Gregarina ormierei Theod.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Mesomorphus villiger: Coree du sud

Gregarina ormierei Theod.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Taraxides laevigatus: Gabon

Gregarina ovata Dufour, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Forficula auricularia (intestine): Poland

Gregarina ovata Dufour, 1828, illus.
Ormieres, R., [1976], Ann. Parasitol., v. 50 (5), 1975, 521-529
Gregarina ovata, life cycle, production of macrosporocysts, compared with Gregarina fallax n. sp. life cycle producing microsporocysts, host specificity
Forficula auricularia (nat. and exper.): Sete; Angleterre
F. decipiens (nat. and exper.): Sete
Anechura bipunctata: Alpes

Gregarina ovata, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Forficula auricularia

Gregarina pachydrili Claparede, 1861
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Schaudinnella enchytraei (von Kolliker, 1848) Meier, 1956

Gregarina paradoxa: Gabriel, 1880
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora juli (Frantzius)

Gregarina paradoxa: Lankester, 1863
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Didymophyes paradoxa Stein

G[regarina] polymorpha
Harry, O. G., 1973, J. Protozool., v. 20 (4), 526
4 species of gregarines in Tenebrio molitor, incidence, effect of seasonal changes in temperature and humidity on infection: British Isles

Gregarina polymorpha: Lankester, 1863
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina cuneata Stein

Gregarina polymorpha, illus.
Ruehl, H., 1976, Ztschr. Parasitenk., v. 48 (3-4), 199-214
Gregarina spp., cytochalasin B (phomin) inhibiting movement, changing cell fine structure and disorganizing fibrillar bundles or myonemes; fine structure of myonemes, skeleton-fibrils and pellicle; elements and mode of movement
Tenebrio molitor

Gregarina polymorpha, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure

Gregarina polymorpha
Snigirevskaia, E. S.; et al., 1977, Tsitologiia, v. 14 (4), 342-349
Gregarina polymorpha, cell surface and septate membrane between two cells in syzygy, electron microscopy, measurement of electrical resistance; suggestion of plasma membrane involvement in cell recognition
Tenebrio molitor (intestine)

Gregarina polymorpha (Hamm.)
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Tenebrio obscurus: Coree du sud

Gregarina polymorpha var. cuneata: Labbe, 1899
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina cuneata Stein

Gregarina portuni Frenzel, 1885
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Paraophioidina portuni (Frenzel, 1885) comb. nov.

Gregarina pycnoceri n. sp., illus.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Pycnocerus sulcatus: Gabon

Gregarina rostrata Welmer, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
redescription
Lagria hirta (intestine): Poland

Gregarina ruszkowskii sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Syn.: Gregarina katherina Watson sensu Foerster, 1938
Coccinella septempunctata: Poznan, Poland
C. quinquepunctata: Kalek and Mielno Koszalinskie, Poland
(intestine of all)

Gregarina saenuridis von Kolliker, 1848 of Delphy (1922)
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Oligochaetocystis pachydrili (Lankester, 1863) Meier, 1956

Gregarina sagittata Leuckart, 1861
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ancora sagittata (Leuckart, 1861) Labbe, 1899

Gregarina sipunculi von Kolliker, 1845
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora sipunculi (von Kolliker, 1845) Leger, 1892

Gregarina spionis von Kolliker, 1848
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Polyrhabdina spionis (von Kolliker, 1848) Mingazzini, 1891

G[regarina] steini
Harry, O. G., 1973, J. Protozool., v. 20 (4), 526
4 species of gregarines in Tenebrio molitor, incidence, effect of seasonal changes in temperature and humidity on infection: British Isles

Gregarina steini Berndt, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Tenebrio molitor (intestine): Poland

Gregarina steini, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Tenebrio molitor

Gregarina typographi Fuchs, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Ips typographus (intestine): Poland

Gregarina ulomae Hoshide
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Taraxides laevigatus: Gabon

Gregarina xylopini Crawley 1903
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Gregarina cuneata Stein

Gregarines, illus.
Arvy, L.; and Sowa, R., 1976, Ann. Parasitol., v. 51 (1), 111-120
Ephemera danica
Baetis rhodani
all from region de Cracovie, Pologne

Gregarines
Ashford, R. W., 1974, J. Med. Entom., v. 11 (5), 605-616
Phlebotomus longipes: Ethiopia

Gregarines
Beaucournu, J. C.; and Deunff, J., [1976], Ann. Parasitol., v. 50 (6), 1975, 831-835
siphonapteres: France

Gregarines
Carter, J. B., 1976, J. Applied Ecol., v. 13 (1), 103-122
haemocoelic gregarines
Tipula paludosa
T. oleracea
T. czizeki
all from north-east England

Gregarines
Devauchelle, G.; and Vinckier, D., 1976, Ztschr. Parasitenk., v. 48 (3-4), 297-298 [Abstract]
Nosema vivieri in gregarines of nemerteans; bacteria in gregarines of Lithobia, earthworms and flourworms

Gregarine, schizont
Killick-Kendrick, R.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 22 [Demonstration]
Phlebotomus ariasi (fat of abdomen, thorax): Gard, southern France

Gregarine, probably Monocystis chagasi
Lainson, R.; et al., 1976, Acta Amazonica, v. 6 (4), 55-60
Psychodopygus davisi: Municipio of Aripuana, Mato Grosso State, Brazil

Gregarines
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
checklist of species of 6 aseptate gregarine families

Gregarines
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
gregarines of arthropods in Poland, host specificity, geographic distribution, host and parasite records

Gregarine
Sawyer, T. K.; Newman, M. W.; and Otto, S. V., 1973, J. Protozool., v. 20 (4), 511
gregarine-like intestinal parasite causing seasonal pathology in the American oyster: rivers and tributaries of Chesapeake Bay, Maryland

Gregarinidea [sp.], illus.
McGhee, R. B.; and Pursglove, S. R., jr., 1977, J. Parasitol., v. 63 (1), 159-160
gregarine with close similarity to Monocystis sp.
Philohela minor (liver): Canaan Valley, Tucker County, West Virginia

Grellia gen. n. [i.e., nom. nov.]
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Grelliidae fam. nov.; type species: Grellia dinophili (Grell, 1953) nom. nov. [i.e., n. comb.]; diagnosis
Syn.: Eucoccidium of Grell, 1953, preoccupied non Eucoccidium Luehe, 1902

Grellia dinophili (Grell, 1953) nom. nov. [i.e., n. comb.] (type species)
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Syn.: Eucoccidium dinophili Grell, 1953

Grellia ophryotrochae (Grell, 1960) nom. nov. [i.e., n. comb.]
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Syn.: Eucoccidium ophryotrochae Grell, 1960

Grelliidae fam. nov.
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Protococcidiorida; includes: Grellia gen.n. [i.e., nom. nov.]; Coelotropha; Myriosporides; Defretinella

Gurleya Doflein, 1898
Loubes, C.; and Maurand, J., 1975, J. Protozool., v. 22 (3), 42A [Abstract]
Gurleya, ultrastructure, developmental stages

Gurleya Doflein, 1898
Loubes, C.; and Maurand, J., 1975, J. Protozool., v. 22 (3), 83A [Abstract]
Gurleya, ultrastructure of some developmental stages

Gurleya [sp.]
Jakowska, S., 1964, An. 2, Cong. Latino-Am. Zool., v. 1, 265-273
Lophius americanus (celulas nervosas): Aquario de New York

Gurleya elegans
Vavra, J.; and Barker, R. J., 1977, J. Protozool., v. 24 (4), 54A [Abstract]
Gurleya elegans, demonstration of mucocalyx in scanning electron microscope
Cyclops strenus

Gymnodinioides inkystans Minkiewicz
Grimes, B. H., 1976, J. Protozool., v. 23 (2), 246-251
Uca sp.
U. minax
U. pugnax
U. pugilator
Sesarma cinereum
S. reticulatum
Gammarid[ea]
Panopeus herbstii
Callinectes sapidus
Pagurus longicarpus
all from North Carolina

Haemamoeba Simond, 1901, in part
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon Berestnev, 1904

Haemamoeba danilewskyi Ziemann, 1898
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon danilewskyi (Ziemann, 1898) Berestnev, 1904

Haemamoeba liothricis Laveran & Marullaz, 1914
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon liothricis (Laveran & Marullaz, 1914) Coatney, 1937

Haemamoeba neavei Balfour, 1906
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon neavei (Balfour, 1906) Wenyon, 1908

Haematoxenus [sp.], illus.
Young, A. S.; Irvin, A. D.; and Woodford, M. J., 1973, J. Wildlife Dis., v. 9 (1), 94-98
description
Syncerus caffer: Queen Elizabeth Park, Uganda

Haematoxenus separatus Uilenberg and Andreasen, 1974, illus.
Uilenberg, G.; and Schreuder, B. E. C., 1976, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 29 (2), 119-126
Haematoxenus separatus, course of infection in splenectomized sheep, experimental transmission by ticks, unsuccessful experimental attempt to transmit to splenectomized goat by injection of infected sheep blood
Rhipicephalus evertsi (exper.)

Haematoxenus separatus Uilenberg & Andreasen 1974, illus.
Young, A. S.; and Mchinja, S. J., 1977, Research Vet. Sc., v. 23 (3), 387-388
Haematoxenus separatus, sheep (blood), nature of veils associated with infected erythrocytes, phase-contrast microscopy: Kenya

Haematoxenus veliferus
Mpangala, C.; Uilenberg, G.; and Schreuder, B. E. C., 1976, Tropenmed. u. Parasitol., v. 27 (2), 192-196
serological characterization of Haematoxenus veliferus from cattle using the indirect fluorescent antibody test, no cross-reactions with either Theileria parva or T. mutans

Haematoxenus veliferus
Schreuder, B.E.C.; Uilenberg, G.; and Tondeur, W., 1977, Tropenmed. u. Parasitol., v. 28 (3), 367-371
Theileriidae, reinfection and retransmission experiments in cattle and African buffalo

Haematoxenus veliferus
Uilenberg, G.; and Schreuder, B. E. C., 1976, Tropenmed. u. Parasitol., v. 27 (1), 106-111
successful transstadial transmission of Haematoxenus veliferus by Amblyomma variegatum in exper. cattle; Theileria mutans and rickettsial organism transmitted at the same time

Haematoxenus veliferus, illus.
Young, A. S.; Irvin, A. D.; and Woodford, M. J., 1973, J. Wildlife Dis., v. 9 (1), 94-98
cow: Athi River, Kenya

Haematozoa lacertae Danilewsky, 1886
Svahn, K., 1975, Norwegian J. Zool., v. 23 (4), 277-295
as syn. of Karyolysus lacertae Danilewsky, 1886

Haematractidium [sp.], illus.
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses
Scomber scombrus (intraerythrocytic): Cardigan Bay, Wales

Haemobartonella
Kreier, J. P.; and Gothe, R., 1976, Vet. Parasitol., v. 2 (1), 83-95
Aegyptianella, Eperythrozoon, Grahamella, Haemobartonella, morphology, host-parasite interactions. review

Haemobartonella
Kreier, J. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 24-28
Rickettsiales, Anaplasmataceae

Haemobartonella [sp.]
Dissanaike, A. S., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 142-143 [Demonstration]
Rattus muelleri
R. rajah
(blood of all): all from Johore Lobis Forest Reserve

Haemobartonella [sp.]
Peters, W.; Howells, R. E.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 21 [Demonstration]
rhesus monkey (associated with surface of erythrocyte)

Haemobartonella [sp.], illus.
Peters, W.; Molyneux, D. H.; and Howells, R. E., 1974, Ann. Trop. Med. and Parasitol., v. 68 (1), 47-50
Macaca mulatta

Haemobartonella sp. Tyzzer 1939
Winchell, E. J., 1977, J. Parasitol., v. 63 (4), 756-757
Microtus breweri (blood): Muskeget Island, 5 miles west of Nantucket, Massachusetts

Haemobartonella [sp.]
Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
Perognathus a. apache (blood): Chaco Canyon National Monument, San Juan County, New Mexico
Reithrodontomys megalotis (blood): Griffith Park, Los Angeles County, California
Peromyscus californicus insignis (blood): Griffith Park, Los Angeles County, California
Peromyscus maniculatus (blood): Petty Ranch near Frazier Mountain Park, Ventura County, California; Juniper Hills near Pearblossom, Los Angeles County, California
Peromyscus boylii: Juniper Hills near Pearblossom, Los Angeles County, California

Haemobartonella bovis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Haemobartonella canis
Bundza, A.; et al., 1976, Canad. Vet. J., v. 17 (10), 267-270
Haemobartonella canis, dog, Coombs' positive anemia following splenectomy

Haemobartonella canis
Krakowka, S., 1977, J. Am. Vet. Med. Ass., 1977, v. 171 (8), 750-753
transplacentally acquired parasitic diseases of dogs, diagnostic features, review

Haemobartonella canis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Haemobartonella canis, illus.
Leeflang, P.; Pimentel, W. J.; and Blotkamp, J., 1974, Bull. Epizoot. Dis. Africa, v. 22 (1), 51-53
Babesia canis, Hepatozoon canis, Haemobartonella canis, dog, case report, also infested with Rhipicephalus sanguineus: Nigeria

Hemobartonella canis
Lyngset, A.; and Yri, T., 1975, Norsk Vet.-Tidsskr., v. 87 (3), 172-175
Hemobartonella canis, case history of first reported case in dog (erythrocytes): Norway

Haemobartonella canis
Oduye, O. O.; and Dipeolu, O. O., 1976, J. Small Animal Practice, v. 17 (5), 331-337
blood parasites of dogs, single and mixed infections, correlation between incidence and rainfall, degree of parasitaemia, infectivity rate within age groups, no significant difference in host susceptibility of local and exotic breeds to infection: Ibadan, Nigeria

Haemobartonella canis
Seneviratna, P.; Weerasinghe, N.; and Ariyadasa, S., 1973, Research Vet. Sc., v. 14 (1), 112-114
Haemobartonella canis, transmission to splenectomised dogs by all stages of Rhipicephalus sanguineus, stage-to-stage as well as transovarial transmission

Haemobartonella felis
Bradley, R. L., 1976, Vet. Med. and Small Animal Clin., v. 71 (9), 1233-1234
Haemobartonella felis, feline infectious anemia, mixed with leukemia virus, chloramphenicol, tetracycline, fair results

Haemobartonella felis
Delaitre, J.-L., 1976, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 29 (2), 115-117
Haemobartonella felis, case reports, tetracycline, chloramphenicol
chats: Tchad

Haemobartonella felis, illus.
Florio, R.; et al., 1977, Rev. Med. Vet., Toulouse, v. 128 (3), 313-322
Haemobartonella felis, cats, clinical reactions, oxytetracycline, six cases fatal despite treatment: Toulouse district

Haemobartonella felis, illus.
Harvey, J. W.; and Gaskin, J. M., 1977, J. Am. Animal Hosp. Ass., v. 13 (1), 28-38
Haemobartonella felis, cats (exper.), hematology studies, time course of disease

Haemobartonella felis and/or Eperythrozoon felis
Hayes, H. M.; and Priester, W. A., 1973, J. Small Animal Practice, v. 14 (12), 797-804
feline infectious anaemia, relative risk according to age, sex and breed of cats, prior disease, seasonal occurrence, mortality, pattern of risk suggests horizontal transmission probably by direct contact

Haemobartonella felis
Maede, Y., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (5), 269-272
Haemobartonella felis, cats, erythrocytes labeled with ^{51}Cr, reduced lifespan in infected cats measured by radioactivity

Hemobartonella felis, illus.
Wightman, S. R.; Kier, A. B.; and Wagner, J. E., 1977, Feline Pract., v. 7 (3), 23-26
Cytauxzoon, cats (blood), clinical features, pathology, diagnosis, morphology compared to Hemobartonella felis

Haemobartonella muris
Cox, H. W.; and Calaf-Iturri, G., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 73-79
Haemobartonella muris in rats, Eperythrozoon coccoides in mice, production of essentially same disease characterized by anemia with splenomegaly and erythrophagocytosis associated with presence of cold-active haemagglutinin, serum antigen, and antibody to serum in blood

Haemobartonella muris
Finch, S. C.; and Jones, A. M., 1973, J. Reticuloendothel. Soc., v. 13 (1), 20-26
Haemobartonella muris, rats, activation of latent infection with development of hemolytic anemia after parenteral administration of ethyl palmitate, effect probably due to development of acute splenic necrosis with loss of splenic sinusoidal function

Haemobartonella muris
Prosl, H., 1976, Ztschr. Parasitenk., v. 50 (2), 214
Ratte

Haemobartonella muris
Schottelius, J., 1977, Tropenmed. u. Parasitol., v. 28 (2), 202-204
latex Chagas test performed on sera of Bartonella bacilliformis-infected humans, Haemobartonella muris-infected rats, and Eperythrozoon coccoides-infected mice, positive reaction observed only with rat sera, nature of this cross-reaction has not been clarified

Haemococcidia
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis crassulus: Guatemala
A. tropidonotus: Honduras
Sceloporus variabilis: Costa Rica; Honduras
Basiliscus plumbifrons: Costa Rica
B. basiliscus: Panama
Anolis limifrons: Panama
A. lionotus: Panama
A. poecilopus: Panama
A. frenatus: Panama
A. tropidogaster: Panama
Iguana iguana: Panama
Polychrus gutturosus: Panama
Corytophanes cristatus: Panama
Sceloporus malachiticus: Panama

Haemoflagellate
Weinman, D.; et al., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (3), 443 [Demonstration]
morphology
Tupaia glis (blood): Perlis, West Malaysia

Haemoflagellate, illus.
Weinman, D.; et al., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (3), 443-445 [Demonstration]
closely related or identical sp. discovered in triatome bug and tree shrew
mice (exper.)
Triatoma sp. (nat. and exper.) (gut)
Tupaia glis (blood)
all from caves in Perlis, West Malaysia

Haemogregarina sp., illus.
Desser, S. S.; and Weller, I., 1973, J. Protozool., v. 20 (1), 65-73
Haemogregarina sp. from Rana berlandieri (imported from Northern Mexico), intra- and extracellular gametocytes studied by light and electron microscopy, locomotion in relation to certain structural features, cytochemistry (proteinaceous and polysaccharide inclusions)

Haemogregarina (sensu lato) [sp.]
McAuliffe, J. R., 1977, J. Parasitol., v. 63 (3), 580-581
varying Haemogregarina infection levels in 3 turtle species may be a function of their basking behavior which affects incidence of parasitism by the presumed leech vector (Placobdella parasitica)
Chrysemys picta bellii: Swan Lake and Whitman Marsh of the Nebraska Sandhills in Arthur and Grant counties
Chelydra serpentina: Swan Lake and Whitman Marsh of the Nebraska Sandhills in Arthur and Grant counties
Emydoidea blandingii: Swan Lake, Nebraska Sandhills

Haemogregarina "sensu lato" [sp.]
de Sousa, M. A.; and Borriello Filho, A., 1975, J. Protozool., v. 22 (3), 36A [Abstract]
description
Bufo crucifer (erythrocytes): Rio de Janeiro city

Hemogregarina spp.
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Rana erythraea
Leptobrachium hasselti
Rana cancrivora
R. limnocharis
R. macrodon
Rhacophorus leucomystax
Bufo melanostictus
all from Malayan Peninsula

Haemogregarina akodoni Carini & Maciel, 1915
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Hepatozoon akodoni

Haemogregarina balli sp. n., illus.
Paterson, W. B.; and Desser, S. S., 1976, J. Protozool., v. 23 (2), 294-301
Haemogregarina balli sp. n., life cycle and biology: leeches as presumed vectors; description of sporogonic development; no transovarial transmission in turtle; failure to exper. infect turtles or leeches
Chelydra serpentina serpentina (blood, liver, lung, spleen)
Placobdella parasitica (gastric and intestinal ceca)
P. ornata (gastric and intestinal ceca)
all from Lake Sasajewan, Algonquin Park, Ontario, Canada

Haemogregarina bigemina Laveran & Mesnil, 1901, illus.
Davies, A. J.; and Johnston, M. R. L., 1976, J. Protozool., v. 23 (2), 315-320
Haemogregarina bigemina in Blennius pholis, incidence in relation to length of host, structure and development, no evidence for leech Oceanobdella blennii as vector, circumstantial evidence for Gnathia maxillaris as vector: Wales

Haemogregarina ibra [i.e. ?, ibera]
Kurashvili, B. E., 1975, Izvest. Akad. Nauk Gruzinsk. SSR, s. Biol., v. 1 (4), 317-320
antagonistic and synergetic interrelationships between intestinal parasites

Haemogregarina moringa n. sp., illus.
Pessoa, S. B.; and de Biasi, P., 1975, Mem. Inst. Butantan, v. 39, 79-83
Gymnothorax moringa (sangue): Sao Sebastiao (litoral norte do Estado de Sao Paulo), Brasil

Haemogregarina pseudemydis n. sp., illus.
Acholonu, A. D., 1974, J. Protozool., v. 21 (5), 659-664
Chelydra serpentina serpentina
Graptemys kohni
Kinosternon subrubrum hippocrepis
Pseudemys floridana hoyi
P. scripta elegans
Terrapene carolina carolina
T. carolina triunguis
Trionyx spinifer
(blood of all): all from southeastern Louisiana

Haemogregarina ranarum
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon ranarum Carini, 1907

Haemogregarina salvelini
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon salvelini Fantham, Porter & Richardson, 1942

Haemogregarina serini Aragao, 1933
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Haemogregarines, similar to Hepatozoon triatomae
Barrett, T. V., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 382-383 [Demonstration]
haemogregarines described from triatome bugs, vertebrate hosts probably animals on which the bugs habitually feed
Triatoma brasiliensis (gut contents)
Panstrongylus geniculatus (gut contents)
T. tibiamaculata (gut, haemolymph and salivary glands)
all from Bahia, Brazil

Haemogregarine
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Tauraco livingstonii (blood): Zaire

Haemogregarines
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Erithacus rubecula (blood): Bramham, Yorkshire and Bramley, Hampshire, England
Parus caeruleus: Great Britain

Haemogregarine
Landau, I.; Colley, F. C.; and Yap, L. F., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 144 [Demonstration]
Rana limnocharis (red blood cells): Bukit Lanjan, Selangor

Hemogregarine-like sporozoon, presumably Hematractidium scombri
MacLean, S. A.; and Sawyer, T. K., 1976, Tr. Am. Micr. Soc., v. 95 (2), 269 [Abstract]
Scomber scombrus (blood): Chincoteague, Virginia

Haemogregarine
Manawadu, B. R., 1972, J. Protozool, v. 19 (4), 587-589
Varanus cepedianus
Calotes sp.
all from Ceylon

Hemogregarines
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Bambusicola thoracica sonorivox (blood): Taiwan

Haemogregarine 1 (probably Haemogregarina tigrinae Hoare (1918)), illus.
Miyata, A., 1974, Nettai Igaku (Trop. Med.), v. 16 (2), 71-84
Elaphe quadrivirgata
Rhabdophis tigrinus tigrinus
Agkistrodon halys
all from Nagasaki City and vicinities, Japan

Haemogregarine 2, illus.
Miyata, A., 1974, Nettai Igaku (Trop. Med.), v. 16 (2), 71-84
Elaphe quadrivirgata
E. clima corphora
Rhabdophis tigrinus tigrinus
all from Nagasaki City and vicinities, Japan

Haemogregarine 3, illus.
Miyata, A., 1974, Nettai Igaku (Trop. Med.), v. 16 (2), 71-84
Rhabdophis tigrinus tigrinus: Nagasaki City and vicinities, Japan

Haemogregarine, unidentified, illus.
Miyata, A.; and Tsukamato, M., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 113-130
Varanus salvator: Palawan Island, the Philippines

Haemogregarine
Peirce, M. A.; and Cooper, J. E., 1977, Vet. Rec., v. 100 (23), 493
Nyctea scandiaca (blood): Great Britain

Haemogregarines
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis limifrons: Costa Rica; Panama
A. lemurinus: Costa Rica
A. lionotus: Costa Rica; Panama
Basiliscus vittatus: Honduras
Ctenosaura similis: Costa Rica; Honduras
Gonatodes albogularis: Panama
Anolis poecilopus : Panama
A. biporcatus: Panama
A. frenatus: Panama
A. tropidogaster: Panama
Basiliscus basiliscus: Panama
Ameiva ameiva: Panama

Haemogregarines
Zakharian, V. Z.; and Vashetko, E. V., 1976, Dokl. Akad. Nauk UzSSR (9), 67
[Rana ridibunda]: Fergansk valley, Uzbekistan

Haemohormidium Henry, 1910
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses

Haemoproteus
Greiner, E. C.; et al., 1975, Canad. J. Zool., v. 53 (12), 1762-1787
avian hematozoa, prevalence with reference to distribution by geographic region, by host family, by vertical stratification of nesting sites and by feeding behavior of known vectors: North America north of Mexico
[Checklist includes 388 bird species and contains both published and unpublished records. For records from specific hosts, see entries in Supplement 22, Part 7, Hosts.]

Haemoproteus Kruse
Misra, K. K.; and Choudhury, A., 1977, Arch. Protistenk., v. 119 (4), 395-400
". . . haemoproteids of reptiles should be designated as species of the genus Haemoproteus . . ."

Haemoproteus [sp.]
Andrews, S. E.; and Threlfall, W., 1975, Proc. Helminth. Soc. Washington, v. 42 (1), 24-28
Corvus brachyrhynchos: insular Newfoundland

Haemoproteus [sp.]
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Ixobrychus minutus
Ardeola ralloides
Bostrychia hagedash
Alopochen aegyptiaca
Milvus migrans
Melierax metabates
Accipiter tachiro
Aquilla rapax
Numida meleagris
Porphyrio alleni
Tringa glareola
Gallinago gallinago
Columba guinea
Streptopelia turtur
S. semitorquata
S. decipiens
S. vinacea
S. capicola
S. senegalensis
Oena capensis
Turtur tympanistria
T. afer
T. chalcospilos
T. abyssinicus
Aplopelia larvata
Treron australis
Clamator jacobinus
Centropus superciliosus
Tyto alba
Otus scops
O. leucotis
Caprimulgus clarus
Ceryle rudis
Alcedo cristata
Halcyon chelicuti
Merops nubicus
M. albicollis
Coracias abyssinica
Eurystomus glaucurus
Lybius bidentatus
L. guifsobalito
L. undatus
L. leucomelas
Indicator variegatus
I. indicator
Jynx torquilla
Campethera nubica
Thripias namaguus
Eremopterix leucotis
Riparia paludicola
Hirundo aethiopica
H. senegalensis
Motacilla flava
Anthus cervinus
Pycnonotus barbatus
Phyllastrephus strepitans
Prionops plumata
Tchagra minuta
Laniarius aethiopicus
L. erythrogaster
L. funebris
Malaconotus sulfureopectus
Lanius collurio
L. nubicus
Saxicola rubetra
Myrmecocichla semirufa
Monticola rufocinerea
Phoenicurus phoenicurus

Haemoproteus [sp.].-- Continued.
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Cossypha semirufa
Luscinia megarhynchos
Luscinia luscinia
Turdoides leucopygius
T. rubiginosus
Acrocephalus schoenobaenus
A. scirpaceus
A. arundinaceus
A. baeticatus
A. gracilirostris
Chloropeta natalensis
Hippolais pallida
Sylvia nisoria
S. borin
S. atricapilla
S. communis
S. curruca
S. mystacea
Phylloscopus trochilus
P. sibilatrix
Cisticola erythrops
C. galactotes
C. natalensis
C. brachyptera
Prinia subflava
Phyllolais pulchella
Camaroptera brevicaudata
Eremomela icteropygialis
E. canescens
Sylvietta brachyura
S. whytii
Muscicapa striata
Melaenornis edolioides
Bradornis pallidus
Hyliota flavigaster
Anthreptes orientalis
Anthreptes platurus
Nectarinia olivacea
N. senegalensis
N. habessinica
N. pulchella
Zosterops abyssinica
Z. senegalensis
Serinus mozambicus
S. atrogularis
S. leucopygius
S. citrinelloides
Pytelia phoenicoptera
Estrilda paludicola
E. rhodopyga
E. astrild
Uraeginthus ianthinogaster
U. bengalus
Lagonosticta larvata
L. senegala
L. rhodopareia
L. rubricata
Lonchura fringilloides
L. cucullata
Amblyospiza albifrons
Ploceus baglafecht
P. luteolus
P. galbula
P. taeniopterus
P. intermedius
P. velatus
P. cucullatus
Malimbus rubriceps
Quelea erythrops
Q. quelea
Euplectes afer
E. albonotatus

Haemoproteus [sp.].-- Continued.
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
E. ardens
E. axillaris
E. gierowii
E. hordeaceus
E. macrourus
E. franciscanus
Bubalornis niger
Passer griseus
P. eminibey
Petronia pyrgita
P. dentata
Lamprotornis chalybaeus
Spreo superbus
Creatophora cinerea
Oriolus oriolus
Dicrurus adsimilis
Tockus erythrorhnchus
all from Ethiopia

Haemoproteus [sp.]
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Crotophaga ani
Galbula ruficauda
Chloropipo holochlora
Chlorothraupis olivacea
Tachyphonus delatrii
Tangara nigro-cincta
Thraupis virens
Contopus virens
Mionectes olivaceous
Myiozetetes cayanensis
(blood of all): all from Colombia

Haemoproteus sp.
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Trachyphonus erythrocephalus: Kenya
Accipiter tachiro: Tanzania
Aquila sp.: Zaire
Ptonoprogne fuligula: Tanzania
Bradornis pumilus: Kenya
Anthropetes collaris: Tanzania
Nectarinia senegalensis: Kenya
N. venusta: Kenya
Phyllastrephus fischeri: Kenya
Pycnonotus barbatus: Kenya; Zaire
P. xanthopygus: Tanzania

Haemoproteus [sp.] or Plasmodium [sp.]
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Dioptrornis fischeri
Phylloscapys trochilus
all from Kenya

Haemoproteus [sp.]
Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
survey, avian hematozoa, seasonal prevalence
Tympanistria tympanistria
Lonchura bicolor
L. cucullata

Haemoproteus [sp.].-- Continued.
Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
Mandingoa nitidula
Cecropis abyssinica
Hirundo rustica
Indicator indicator
Aerops albicollis
Melittophagus variegatus
Motacilla flava
Alseonax griseigularis
Hylia prasina
Nectarinia cuprea
N. senegalensis
Melanopteryx sp.
Passer griseus
Ploceus aurantius
P. cucullatus
P. melanocephalus
P. nigerrimus
P. nigricollis
P. ocularis
P. pelzelni
P. weynsi
Quelea erythrops
Pycnonotus barbatus
Ciccaba woodfordii
Acrocephalus sp.
Camaroptera brevicaudata
Phylloscopus trochilus
Sylvia borin
Zosterops senegalensis
(blood of all): all from Uganda

Haemoproteus sp.
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
Falco peregrinus
Pica pica
Carduelis chloris
(blood of all): all from Great Britain

Haemoproteus sp.
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Fringilla coelebs (blood): Berkshire, England

Haemoproteus sp.
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Sylvia atricapilla (blood): Bramham, Yorkshire, England

Haemoproteus sp.
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Prunella modularis (blood): Bramham, Yorkshire, England

Haemoproteus sp.
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Sylvia borin (blood): Bramham, Yorkshire, England

Haemoproteus sp.
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Passer domesticus (blood): Berkshire, England

Haemoproteus sp.
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Turdus philomelos (blood): Bramham, Yorkshire, England
T. merula (blood): Bramham, Yorkshire and Bramley, Hampshire, England

Haemoproteus sp., resembled H. orizivorae
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Parus major (blood): Berkshire, England

Haemoproteus [sp.]
Choudhury, A.; and Misra, K. K., 1973, J. Protozool., v. 20 (4), 514
Lanius schach collurio (blood): India

Haemoproteus sp.
Fallis, A. M.; Jacobson, R. L.; and Raybould, J. N., 1973, J. Protozool., v. 20 (3), 438-442
Guttera pucherani (blood): Amani, Tanzania
Numida mitrata (blood): vicinity of Handeni, Tanzania

Haemoproteus sp., morphologically similar to H. antigone
Forrester, D. J.; et al., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 55-59
Grus canadensis tabida (blood): Florida

Haemoproteus sp.
Lewis, J. C.; Carpenter, J. W.; and Morrison, J. A., 1975, J. Wildlife Dis., v. 11 (4), 537-539
Mimus polyglottos (blood): near Fairview, Major County, Oklahoma

Haemoproteus [sp.]
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Streptopelia chinensis formosae
Hypsipetes madagascariensis nigerrimus
Turdus c. chrysolaus
Zosterops japonica simplex
(blood of all): all from Taiwan

Haemoproteus [sp.]
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Erythrura prasina
Coeriba flaviola
Gracula religiosa indica
(blood of all): all imported to U. S., origin unknown

Haemoproteus sp.
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Turdoides striatus: Orissa, India

Haemoproteus [sp.]
Noblet, R.; Moore, H. S. IV; and Noblet, G. P., 1976, Poultry Science, v. 55 (1), 447-449
Quiscalus quiscula
Cyanocitta c. cristata
(blood of all): all from South Carolina

Haemoproteus sp.
Pav, J.; and Zajicek, D., 1974, Veterinarstvi, v. 24 (11), 517-520
Lyrus tetrix: CSSR

Haemoproteus sp.
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Eremopteryx signata (blood): Kenya

Haemoproteus [sp.]
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Pytelia melba (blood): Kenya

Haemoproteus [sp.], resembled H. coraciae
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Coracias garrulus (blood): Kenya

Haemoproteus [sp.], probably H. handai
Peirce, M. A.; and Bevan, B. J., 1977, Vet. Rec., v. 100 (14), 282-283
Cacatua galerita
Psittacus erithacus
Loriculus g. galgulus
(blood of all): all imported to London from Indonesia

Haemoproteus sp., resembles H. sanguinis
Peirce, M. A.; and Cheke, A. S., 1977, J. Protozool., v. 24 (2), Suppl., 33A [Abstract]
Hypsipetes crassirostris moheliensis: Comoro Islands

Haemoproteus [sp.]
Peirce, M. A.; and Cooper, J. E., 1977, Vet. Rec., v. 100 (23), 493
Accipiter gentilis (blood): Great Britain

Haemoproteus sp., illus.
Stabler, R. M.; and Kitzmiller, N. J., 1976, J. Parasitol., v. 62 (4), 539-544
Dendragapus obscurus
Lophortyx gambelii
Oreortyx pictus
(blood of all): all from Colorado

Haemoproteus sp.
Subkhanov, M.; and Mirzobakhadurov, R., 1976, Izvest. Akad. Nauk Tadzhiksk. SSR, Otdel. Biol. Nauk (63 (2)), 108-109
[Erythrina rhodochlamys]
[Phylloscopus griseolus]
[Sylvia nisoria]
[Motacilla cinerea]
(blood of all): all from Tegirmansai, Kirgizia

Haemoproteus [sp.]
Thomas, S. E.; and Dobson, L. D., 1975, Onderstepoort J. Vet. Research, v. 42 (1), 67-68
Threskiornis aethiopicus
Anas erythrorhyncha
Francolinus swainsonii
Numida meleagris
Columba guinea
Streptopelia senegalensis
(blood of all): all from vicinity of Onderstepoort, Republic of South Africa

Haemoproteus accipiter sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Accipiter]: north-east Azerbaidzhan

Haemoproteus alaudae Celli et Sanfelise, 1891
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Alauda arvensis]
[Galerida cristata]
all from north-east Azerbaidzhan

Haemoproteus anthi de Mello, 1935
Subkhanov, M., 1975, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 18 (3), 62-63
birds (erythrocytes): Tadzhikistan

Haemoproteus antigonis de Mello, 1935
Forrester, D. J.; Bush, A. O.; and Williams, L. E., jr., 1975, J. Parasitol., v. 61 (3), 547-548
Grus canadensis pratensis (blood): Florida

Haemoproteus archilocus
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Eutoxeres aquila (blood): Colombia

Haemoproteus asio sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Asio otus]: north-east Azerbaidzhan

Haemoproteus balearicae sp. nov., illus.
Peirce, M. A., 1973, Bull. Epizoot. Dis. Africa, V. 21 (4), 467-475
Balearica p. pavonina (blood, lungs): Surrey, England (captive)
B. p. gibbericeps (blood, lungs): Surrey, England (imported from East Africa)

Haemoproteus bambusicolae sp. n. [? nomen nudum], illus.
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Bambusicola thoracica sonorivox (blood): Taiwan

Haemoproteus beckeri
Lewis, J. C.; Carpenter, J. W.; and Morrison, J. A., 1975, J. Wildlife Dis., v. 11 (4), 537-539
Toxostoma rufum (blood): near Fairview, Major County, Oklahoma

Haemoproteus bennetti sp. n., illus.
Greiner, E. C.; Mandal, A. K.; and Nandi, N.C., 1977, J. Parasitol., v. 63 (4), 651-656
Picus flavinucha (erythrocytes): Sukua, Darjeeling W.B., India
P. erythropygius (blood): central Thailand

Haemoproteus borgesi Tendeiro 1947, illus.
Greiner, E. C.; Mandal, A. K.; and Nandi, N.C., 1977, J. Parasitol., v. 63 (4), 651-656
redescription
Picus vittatus (blood): Rantan Panjang, Malaysia
Jynx torquilla: India
Picus canus: India

Haemoproteus borgesi Tendeiro, 1947
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Picoides m. mahrattensis: Godavari River Basin, Maharashtra

Haemoproteus canachites
Eve, J. H.; and Davidson, W. R., 1976, J. Parasitol., v. 62 (1), 142-144
Bonasa umbellus (blood): Kentucky; Michigan; West Virginia

Haemoproteus chloris Ortega et Berenguer, 1950
Subkhanov, M.; and Mirzobakhadurov, R., 1976, Izvest. Akad. Nauk Tadzhiksk. SSR, Otdel. Biol. Nauk (63 (2)), 108-109
[Chloris chloris] (blood): Tegirmansai, Kirgizia

Haemoproteus chloris Ortega and Berenguer, 1950
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Chloris chloris]: north-east Azerbaidzhan

Haemoproteus columbae, illus.
Ahmed, F. E.; and Mohammed, A. H. H., 1977, J. Protozool., v. 24 (3), 389-393
Haemoproteus columbae, schizogony
Columba livia domestica (lungs) (exper.)
Pseudolynchia canariensis (exper.)

Haemoproteus columbae
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Columba guinea: Kenya
Streptopelia semitorquata: Kenya
Tympanistra tympanistra: Tanzania

Haemoproteus columbae Kruse, illus.
Gallucci, B. B., 1974, J. Protozool., v. 21 (2), 254-263
Haemoproteus columbae, macrogametogenesis, fertilization, fine structure
Columba livia (blood): Raleigh, North Carolina

Haemoproteus columbae Kruse, illus.
Gallucci, B. B., 1974, J. Protozool., v. 21 (2), 264-275
Haemoproteus columbae, differentiating ookinete, fine structure, presence of conoid reported for first time in Haemosporina
Pseudolynchia maura (exper.) (gut)

Haemoproteus columbae Kruse
Klei, T. R.; and DeGiusti, D. L., 1975, J. Wildlife Dis., v. 11 (1), 130-135
Haemoproteus columbae and Pseudolynchia canariensis in Columba livia, seasonal occurrence of H. columbae highest during fall and winter, correlating with changes in vector population: Detroit, Michigan

Haemoproteus columbae Kruse, 1890
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Columba livia intermedia: Godavari River Basin, Maharashtra

Haemoproteus columbae Kruse, 1890
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Streptopelia chinensis: Orissa. India

Haemoproteus columbae
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
"L[averania] is a synonym pro parte of both H. passeris and H. columbae."

Haemoproteus columbae
Stabler, R. M.; Kitzmiller, N. J.; and Braun, C. E., 1977, J. Wildlife Management, v. 41 (1), 128-130
Columba fasciata (blood): Colorado; Mexico

Haemoproteus columbae, illus.
Sterling, C. R.; and Aikawa, M., 1973, J. Protozool., v. 20 (1), 81-92
Plasmodium gallinaceum, Haemoproteus columbae, Leucocytozoon simondi, gametocyte ultrastructure compared in detail

Haemoproteus columbae Kruse, 1890
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Columba livia]
[pigeon]
all from north-east Azerbaidzhan

Haemoproteus coraciae
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Coracias caudata: Kenya
C. spatulata: Zaire
Eurystomus glaucurus: Zaire

Haemoproteus coraciae de Mello and Afonso, 1935
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Coracias b. benghalensis: Godavari River Basin, Maharashtra

Haemoproteus coracitis Tartakovskyi, 1913
Subkhanov, M.; and Mirzobakhadurov, R., 1976, Izvest. Akad. Nauk Tadzhiksk. SSR, Otdel. Biol. Nauk (63 (2)), 108-109
[Coracias garrulus] (blood): Tegirmansai, Kirgizia

Haemoproteus coracitis Tartakowski, 1913
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Coracias garrulus]: north-east Azerbaidzhan

Haemoproteus danilewski Kruse, 1890
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Corvus corone cornix]: north-east Azerbaidzhan

Haemoproteus danilewskii var. urbanensis Sachs (1953)
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
as syn. of Haemoproteus passeris Kruse (1890)

Haemoproteus dicruri de Mello, 1935
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Dicrurus adsimilis: Orissa, India

Haemoproteus emberiza Berson, 1964
Subkhanov, M.; and Mirzobakhadurov, R., 1976, Izvest. Akad. Nauk Tadzhiksk. SSR, Otdel. Biol. Nauk (63 (2)), 108-109
[Emberiza calandra] (blood): Tegirmansai, Kirgizia

Haemoproteus emberiza Berson, 1964
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Emberiza calandra]
[Emberiza melanocephala]
all from north-east Azerbaidzhan

Haemoproteus enucleator
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Ceyx picta: Ethiopia

Haemoproteus enucleator
Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
survey, avian hematozoa, seasonal prevalence
Ispidina picta (blood): Uganda

Haemoproteus fallisi
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Monticola saxatilis
Saxicola torquata
(blood of all): all from Kenya

Haemoproteus fallisi
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Turdus merula (blood): Berkshire, England

Haemoproteus fallisi (?)
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Turdus merula (blood): Sussex, England

Haemoproteus fallisi Bennett & Campbell 1972
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Copsychus saularis: Orissa, India

Haemoproteus fringillae
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Emberiza cabanisi: Tanzania
Estrilda paludicola: Zaire
Lonchura cucullata: Zaire; Tanzania
Mandingoa nitidula: Tanzania
Othypantes stuhlmanni: Tanzania
Euplectes ardens: Zaire
Ploceus reichenowi: Kenya

Haemoproteus fringillae/orizivora
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Hyphanturgis ocularis: Tanzania
Quelea quelea: Zaire

Haemoproteus fringillae
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Fringilla coelebs (blood): Bramley, Hampshire, England

Haemoproteus fringillae Labbe, 1894
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Copsychus saularis: Orissa, India

Haemoproteus fringillae Labbe, 1894
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Fringilla coelebs]
[Fringilla montifringilla]
all from north-east Azerbaidzhan

Haemoproteus garnhami
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
"I believe that H. garnhami is probably a synonym of H. orizivorae."

Haemoproteus garrulus sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Garrulus glandarius]: north-east Azerbaidzhan

Haemoproteus glaucidii de Mello, 1937
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Glaucidium radiatum: Orissa, India

Haemoproteus globulosus Ortega and Berenguer, 1950
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Carduelis carduelis]: north-east Azerbaidzhan

Haemoproteus granulosum Rey Vila (1945)
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
as syn. of Haemoproteus passeris Kruse (1890)

Haemoproteus halcyonis
Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
survey, avian hematozoa, seasonal prevalence
Ceryle rudis (blood): Uganda

Haemoproteus handai, illus.
Peirce, M. A.; and Bevan, B. J., 1977, Vet. Rec., v. 100 (14), 282-283
Cacatua sulphurea
Psittacula cyanocephala
P. krameri borealis
(blood of all): all imported to London from Indonesia

Haemoproteus hirundinis Sergent et Sergent, 1905
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Hirundo rustica]: north-east Azerbaidzhan

Haemoproteus lanii
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Dryoscopus cubla: Zaire
Lanius collaris: Kenya
L. collurio: Tanzania
Tchagara minuta: Zaire

Haemoproteus lanii de Mello, 1937
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Lanius schach: Orissa, India

Haemoproteus lanii De Mello, 1935
Subkhanov, M.; and Mirzobakhadurov, R., 1976, Izvest. Akad. Nauk Tadzhiksk. SSR, Otdel. Biol. Nauk (63 (2)), 108-109
[Lanius minor] (blood): Tegirmansai, Kirgizia

Haemoproteus lanii De Mello, 1935
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Lanius minor]
[Lanius cristatus]
all from north-east Azerbaidzhan

Haemoproteus larae Iakuin, 1972
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Larus argentatus]: north-east Azerbaidzhan

Haemoproteus maccallumi
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Columbigallina talpacoti (blood): Colombia

Haemoproteus maccallumi
Greiner, E. C., 1975, J. Wildlife Dis., v. 11 (2), 150-156
Haemoproteus in Zenaida macroura, prevalence with respect to year, season, host age; no correlation between parent and nestling infections; collection of potential vectors (Culicidae and Hippoboscidae): Lancaster County, Nebraska

Haemoproteus maccallumi
Lewis, J. C.; Carpenter, J. W.; and Morrison, J. A., 1975, J. Wildlife Dis., v. 11 (4), 537-539
Zenaida macroura (blood): near Fairview, Major County, Oklahoma

Haemoproteus majoris Laveran, 1902
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon majoris (Laveran, 1902) Coatney, 1937

Haemoproteus majoris sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Parus major]: north-east Azerbaidzhan

Haemoproteus meleagridis Levine, 1961
Forrester, D. J.; et al., 1974, J. Protozool., v. 21 (4), 494-497
prevalence in relation to host age, host sex, geographic distribution, and year
Meleagris gallopavo (blood): Florida

Haemoproteus meleagridis
Noblet, R.; and Moore, H. S., IV, 1975, J. Wildlife Dis., v. 11 (4), 516-518
Leucocytozoon smithi and Haemoproteus meleagridis in Meleagris gallopavo, wild turkeys as important reservoir hosts for Leucocytozoon smithi: South Carolina

Haemoproteus meropi Zargar, 1945
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Merops orientalis: Orissa, India

Haemoproteus meropis Tartakowski, 1913
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Merops apiaster]: north-east Azerbaidzhan

Haemoproteus merulla sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Turdus merula]: north-east Azerbaidzhan

Haemoproteus metchnikovi, illus.
Sterling, C. R.; and DeGiusti, D. L., 1974, J. Protozool., v. 21 (2), 276-283
Haemoproteus metchnikovi in Chrysops callidus (midgut, salivary glands), differentiating oocysts and mature sporozoites, fine structure

Haemoproteus montezi
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Tauraco hartlaubi: Kenya
T. livingstonii: Zaire
Cyanomitra olivacea: Tanzania
Nectarinia amethystina: Zaire

Haemoproteus motasillae sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Motacilla alba]: north-east Azerbaidzhan

Haemoproteus (Parahaemoproteus) nettionis
Bennett, G. F.; et al., 1974, J. Wildlife Dis., v. 10 (4), 442-451
survey, prevalence of hematozoa in anatids, infection rate increases with host age, seasonal distribution
Anas rubripes
Anas platyrhynchos
Anas platyrhynchos x A. rubripes
Anas discors
Anas carolinensis
Aix sponsa
Lophodytes cucullatus
Mareca penelope
Aythya collaris
Branta canadensis
(blood of all): all from Massachusetts

Haemoproteus (Parahaemoproteus) nettionis
Bennett, G. F.; et al., 1975, J. Wildlife Dis., v. 11 (2), 280-289
hematozoa of Anatidae, prevalence survey
Anas rubripes rubripes
Anas platyrhynchos platyrhynchos
Anas crecca carolinensis
Anas discors
Anas acuta
Aix sponsa
Aythya collaris
Glaucionetta clangula
Mareca americana
Branta canadensis
all from Maritime provinces of Canada

Haemoproteus (Parahaemoproteus) nettionis
Herman, C. M.; and Bennett, G. F., 1976, Canad. J. Zool., v. 54 (7), 1038-1043
exposure of captive sentinel Anatidae to natural infection, seasonal transmission
Anas platyrhynchos: Nova Scotia-New Brunswick border area

Haemoproteus noctuae
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Bubo africanus (blood): Zaire

Haemoproteus noctuae Celli et Sanfelise, 1891
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Athene noctua]: north-east Azerbaidzhan

Haemoproteus orizivora
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Ploceus baglafecht (blood): Tanzania

Haemoproteus orizivora/fringillae
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Hyphanturgis ocularis: Tanzania
Quelea quelea: Zaire

Haemoproteus oryzivorae Anschutz, 1909
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Copsychus saularis: Orissa, India

Haemoproteus orizivorae
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Sylvia communis (blood): Kenya

Haemoproteus orizivorae
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
"I believe that H. garnhami is probably a synonym of H. orizivorae."

Haemoproteus passeris Kruse, 1890
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Passer domesticus: Orissa, India

Haemoproteus passeris Kruse (1890), illus.
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
only valid species of haemoproteid parasite from Passer, synonymy, redescription, first description of schizogonic stages, does not belong to subgenus Parahaemoproteus
Passer domesticus: England

Haemoproteus passeris Kruse, 1890
Subkhanov, M.; and Mirzobakhadurov, R., 1976, Izvest. Akad. Nauk Tadzhiksk. SSR, Otdel. Biol. Nauk (63 (2)), 108-109
[Passer montanus]
[P. domesticus]
(blood of all): all from Tegirmansai, Kirgizia

Haemoproteus passeris Kruse, 1890
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Passer domesticus]
[Passer montanus]
all from north-east Azerbaidzhan

Haemoproteus picae Coatney et Roudabush, 1937
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Pica pica]: north-east Azerbaidzhan

Haemoproteus plataleae de Mello, 1935
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Pseudibis p. papillosa: Godavari River Basin, Maharashtra

Haemoproteus pratezi [i.e. ?, pratasi]
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Numida meleagris (blood): Zaire

Haemoproteus quiscalus
Lewis, J. C.; Carpenter, J. W.; and Morrison, J. A., 1975, J. Wildlife Dis., v. 11 (4), 537-539
Icterus galbula (blood): near Fairview, Major County, Oklahoma

Haemoproteus sacharovi
Greiner, E. C., 1975, J. Wildlife Dis., v. 11 (2), 150-156
Haemoproteus in Zenaida macroura, prevalence with respect to year, season, host age; no correlation between parent and nestling infections; collection of potential vectors (Culicidae and Hippoboscidae): Lancaster County, Nebraska

Haemoproteus sacharovi
Lewis, J. C.; Carpenter, J. W.; and Morrison, J. A., 1975, J. Wildlife Dis., v. 11 (4), 537-539
Zenaida macroura (blood): near Fairview, Major County, Oklahoma

Haemoproteus sacharovi Novy & MacNeal, 1905
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Streptopelia chinensis: Orissa, India

Haemoproteus sacharovi
Stabler, R. M.; Kitzmiller, N. J.; and Braun, C. E., 1977, J. Wildlife Management, v. 41 (1), 128-130
Columba fasciata (blood): Colorado; California; Mexico

Haemoproteus sittae sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Sitta europaea]: north-east Azerbaidzhan

Haemoproteus sternae sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Sterna hirundo]: north-east Azerbaidzhan

Haemoproteus sturnii
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Cinnyricinclus leucogaster: Zaire
Lamprotornis chloropterus: Zaire
L. chalybaeus: Zaire
Unichognathus walleri: Tanzania

Haemoproteus sturni de Mello, 1935
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Acridotheres t. tristis
Sturnus pagodarum
all from Godavari River Basin, Maharashtra

Haemoproteus sturni De Mello, 1935
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Sturnus vulgaris]: north-east Azerbaidzhan

Haemoproteus syrnii
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Bubo vosseleri (blood): Tanzania

Haemoproteus syrnii
Peirce, M. A.; and Cooper, J. E., 1977, Vet. Rec., v. 100 (23), 493
Strix aluco (blood): Great Britain

Haemoproteus telfordi
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Lissotis melanogaster (blood): Zaire

Haemoproteus tinnunculus Wasielewski & Wulker, 1918
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Falco t. tinnunculus: Godavari River Basin, Maharashtra

Haemoproteus tinnunculus
Peirce, M. A.; and Cooper, J. E., 1977, Vet. Rec., v. 100 (23), 493
Falco cherrug (blood): Great Britain

Haemoproteus tinnunculus Wasilewski and Wulker, 1918
Subkhanov, M., 1975, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 18 (3), 62-63
birds (erythrocytes): Tadzhikistan

Haemoproteus tinnunculus Wasielewski et Wulker, 1918
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Falco tinnunculus]
[Falco naumanni]
all from north-east Azerbaidzhan

Haemoproteus trionyxii n. sp., illus.
Misra, K. K.; and Choudhury, A., 1977, Arch. Protistenk., v. 119 (4), 395-400
Trionyx gangeticus (blood): river Ganges and its estuary

Haemoproteus turtur
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Streptopelia turtur (blood): Bramham, Yorkshire, England

Haemoproteus turtur Ortega and Berenguer, 1950
Subkhanov, M., 1975, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 18 (3), 62-63
birds (erythrocytes): Tadzhikistan

Haemoproteus turtur Ortega et Berenguer, 1950
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Streptopelia turtur]: north-east Azerbaidzhan

Haemoproteus upupae De Mello
Subkhanov, M.; and Mirzobakhadurov, R., 1976, Izvest. Akad. Nauk Tadzhiksk. SSR, Otdel. Biol. Nauk (63 (2)), 108-109
[Upupa epops] (blood): Tegirmansai, Kirgizia

Haemoproteus upupae De Mello, 1935
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Upupa epops]: north-east Azerbaidzhan

Haemoproteus velans Coatney and Roudabush 1937, illus.
Greiner, E. C.; Mandal, A. K.; and Nandi, N.C., 1977, J. Parasitol., v. 63 (4), 651-656
redescription
Dendrocopos mahrattensis: Maharashtra, India
Jynx torquilla: Rajasthan, India
Picus chlorolophus: Chiengmai, Thailand
P. flavinucha: Chiengmai, Thailand
P. canus: Thailand
Chrysocolaptes lucidus: Malaya

Haemoproteus velans Coatney and Roudabush, 1937
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Picoides m. mahrattensis: Godavari River Basin, Maharashtra

Haemoproteus wenyoni Sergent & Sergent (1948)
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
as syn. of Haemoproteus passeris Kruse (1890), also homonym of H. wenyoni de Mello, Bras de Sa, de Sousa, Dias & Noronha (1916)

Haemoproteus zosteropsi
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Zosterops virens (blood): Kenya

Haemosporidia
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Haemotrichomonas gallinarum
Kulda, J.; Suchankova, E.; and Svoboda, S., 1974, Acta Vet. Brno, v. 43 (1), 53-64
as syn. of Tetratrichomonas gallinarum (Martin & Robertson 1911) Honigberg 1963

Hammondia
Beier, T. V.; Siim, I. K.; and Hutchison, W. M., 1977, Tsitologiia, v. 19 (12), 1362-1368
Eimeriidae, Isosporinae

Hammondia Frenkel and Dubey 1975
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Sarcocystidae, Toxoplasmatinae
diagnosis, life cycle pattern

Hammondia species
Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
Vulpes vulpes (exper.) (feces)

Hammondia hammondi
Christie, E.; and Dubey, J. P., 1977, Infect. and Immun., v. 18 (2), 412-415
cross-immunity between Toxoplasma gondii and 6 strains of Hammondia hammondi in mice and hamsters

Hammondia hammondi Frenkel and Dubey 1975
Christie, E.; Dubey, J. P.; and Pappas, P. W., 1977, J. Parasitol., v. 63 (5), 929-931
Hammondia hammondi, Toxoplasma gondii, prevalence, life cycle comparisons
cats (nat. and exper.): Ohio
mice (exper.)

Hammondia hammondi, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Hammondia hammondi
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (5), 541-543
attempted transmission of feline coccidia from chronically infected queens to their kittens, results indicate congenital transmission of coccidia unlikely

Hammondia hammondi Frenkel and Dubey, 1975
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Hammondia hammondi
Dubey, J. P.; and Streitel, R. H., 1976, J. Parasitol., v. 62 (4), 548-551
Hammondia hammondi, life cycle and transmission: tachyzoites are noninfectious to cats, infectivity being associated with cysts formed by the 10th day of mouse infection; more cysts found in mouse muscle than in brain, less frequent occurrence in spleen, liver, lungs, lymph nodes; absence of congenital transmission in mice; oocysts noninfectious to chickens

Hammondia hammondi Frenkel 1974
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma hammondi (Frenkel 1974) Levine and Nye (in press) [?n. comb.]

Hammondia hammondi Frenkel & Dubey, 1975
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma hammondi (Frenkel & Dubey, 1975) comb. nov.

Hammondia hammondi (Frenkel und Dubey 1975), illus.
Rommel, M.; and v. Seyerl, F., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (20), 398-399
Hammondia hammondi, isolation from cat feces, exper. mouse infection, epidemiological significance for distinguishing from Toxoplasma gondii: Germany

Hammondia hammondi, illus.
Sheffield, H. G.; Melton, M. L.; and Neva, F. A., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 217-225
Hammondia hammondi, artificially excysted sporozoites, development in cell culture, ultrastructure, endodyogeny only type division observed, characteristics to differentiate from Toxoplasma gondii

Hammondia hammondi Frenkel, 1974
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora hammondi (Frenkel, 1974) comb. nov.

Hammondia heydorni (Tadros and Laarman, 1976) Dubey, 1977 [n. comb.]
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
Syn.: Isospora heydorni Tadros and Laarman
"Tadros and Laarman also proposed 2 new organisms: Endorimospora (Sarcocystis) suihominis from the pig and Isospora heydorni from the dog. I named them Sarcocystis porcihominis and Isospora wallacei respectively. Because Tadros and Laarman's paper was published a month earlier than mine, the names proposed by Tadros and Laarman have priority over Sarcocystis porcihominis and Isospora wallacei."

Haplosporidan
Lichtenfels, J. R.; et al., 1976, Tr. Am. Micr. Soc., v. 95 (2), 265-266 [Abstract]
anisakid larvae, resembles Paranisakiopsis, nearly 100% hyperparasitized: coastal waters from New Jersey to North Carolina

Haplozoon axiothellae Siebert 1973, illus.
Siebert, A. E.; and West, J. A., 1974, Protoplasma, v. 81 (1), 17-35
Haplozoon axiothellae, fine structure morphology, attachment cell, penetration stylet, reproduction, characteristics of theca
Axiothella rubrocincta: Mitchell Bay, San Juan Island, Washington and Bodega Bay, California

Haptophryoidea n. superfam. [nomen nudum]
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Astomatida

Hartmanella
Dolensek, E. P.; Bihn, J. P.; and Napolitano, R. L., 1976, J. Protozool., v. 23 (2), 19A [Abstract]
Entamoeba invadens, Naegleria, and Hartmanella isolated from exhibited reptiles

Hartmannella
Jadin, J. B.; and Willaert, E., 1972, Protistologica, v. 8 (4), 505-508
presence of limax group amoebae in swimming pools and drinking water system, possible control methods: Anvers

Hartmannella-Acanthamoeba group
Lyons, T. B. III.; and Kapur, R., 1977, Applied and Environment. Microbiol., v. 33 (3), 551-555
limax amoebae in public swimming pools, concentration, correlations, and public health significance, preliminary tests for the human pathogen Naegleria fowleri were inconclusive: Albany, Schenectady, and Rensselaer counties, New York

Hartmanella
Martinez, A. J.; Fultz, D. G.; and Armstrong, C. E. V., 1976, Proc. 34. Ann. Meet. Electron Microsc. Soc. America (Miami Beach, Florida, Aug. 9-13), 226-227
amebic meningoencephalitides, electron microscopy, differential diagnosis, embedding and staining technique

Hartmannella sp., illus.
Takano, J.; et al., 1976, Patologia, v. 14 (4), 287-297
Hartmannella sp. causing fatal primary subacute amoebic meningoencephalitis in young man previously without symptoms of infection, clinical case report: Lima, Peru

Hartmannella astronyxis
Hadas, E.; Kasprzak, W.; and Mazur, T., 1977, Tropenmed. u. Parasitol., v. 28 (1), 35-43
Naegleria fowleri, N. gruberi and strains of Hartmannella spp., taxonomic differentiation by means of disc electrophoresis of protein patterns

Hartmannella castellanii
Ambrosioni, P.; and Bernagozzi, M., 1972, Parassitologia, v. 14 (1), 45-51
Hartmannella castellanii, solid medium for axenic culture

Hartmannella castellanii
Hadas, E.; Kasprzak, W.; and Mazur, T., 1977, Tropenmed. u. Parasitol., v. 28 (1), 35-43
Naegleria fowleri, N. gruberi and strains of Hartmannella spp., taxonomic differentiation by means of disc electrophoresis of protein patterns

Hartmannella culbertsoni, illus.
Allen, D. J.; and Didio, L. J. A., 1976, Ohio J. Sc., v. 76 (4), 167-171
Hartmannella culbertsoni, morphology and pleomorphism during active growth, scanning electron microscopy

Hartmannella culbertsoni
Hadas, E.; Kasprzak, W.; and Mazur, T., 1977, Tropenmed. u. Parasitol., v. 28 (1), 35-43
Naegleria fowleri, N. gruberi and strains of Hartmannella spp., taxonomic differentiation by means of disc electrophoresis of protein patterns

Hartmanella culbertsoni
Kaushal, D. C.; Verma, A. K.; and Raizada, M. K., 1976, Current Sc., Bangalore, v. 45 (5), 181-183
Hartmanella culbertsoni, cyclic nucleotide phosphodiesterase activity in cells and culture medium, cells starving or differentiating into cysts

Hartmannella culbertsoni Singh and Das, 1970
Maitra, S. C.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (3), 229-235
Hartmannella culbertsoni, ultrastructural differences in mouse brain and under different cultural conditions

Hartmannella culbertsoni, illus.
Maitra, S. C.; et al., 1976, Internat. J. Parasitol., v. 6 (6), 489-493
Naegleria aerobia, Hartmannella culbertsoni, comparison of ultrastructural characteristics in brain of mice with exper. primary amoebic meningoencephalitis, possible relationship of structure to pathogenicity

Hartmannella polyphaga
Hadas, E.; Kasprzak, W.; and Mazur, T., 1977, Tropenmed. u. Parasitol., v. 28 (1), 35-43
Naegleria fowleri, N. gruberi and strains of Hartmannella spp., taxonomic differentiation by means of disc electrophoresis of protein patterns

Hartmannella vermiformis Page
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
Acanthamoeba, Naegleria, Hartmannella, comparative studies on free-living and pathogenic amebae: cyst structure; nutrition; protein composition; immunology; cell free plaques and other cytopathic effects; phospholipase liberation; sensitivity to amphotericin B

Hartmanelloid amoeba
Talis, B.; Stein, B.; and Gold, D., 1977, J. Protozool., v. 24 (2), Suppl., 18A-19A [Abstract]
hartmanelloid amoeba, invasiveness and antigenicity as compared to Entamoeba histolytica strains

Helella Kaczanowski, 1961
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Radiophryinae nov. comb.

Helella intermedia (de Puytorac, 1955) Kaczanowski, 1961
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Radiophrya intermedia de Puytorac, 1955
Enchytraeoides

Helella rhizodrili Kaczanowski, 1961, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Rhizodrilus pilosus

Helicosporidium sp., near parasiticum
Fukuda, T.; Lindegren, J. E.; and Chapman, H. C., 1976, Mosquito News, v. 36 (4), 514-517
Helicosporidium sp. in Culex nigripalpus, susceptibility of various mosquito and other insect hosts, possible biological control agent
Aedes aegypti (exper.)
Aedes epactius (exper.)
Aedes taeniorhynchus (exper.)
Anopheles albimanus (exper.)
Anopheles quadrimaculatus (exper.)
Culex peccator (exper.)
Culex salinarius (exper.)
Culex tarsalis (exper.)
Culex territans (exper.)
Culiseta inornata (exper.)
Psorophora ferox (exper.)
Psorophora horrida (exper.)
Psorophora varipes (exper.)
Uranotaenia lowii (exper.)
Culicoides sp. (exper.)
Carpophilus hemipterus (exper.)
Lasioderma serricorne (exper.)
Trogoderma variabile (exper.)
Galleria mellonella (exper.)
Paramyelois transitella (exper.)
Culex nigripalpus: near Lake Charles, Louisiana

Hematodinium sp., illus.
Newman, M. W.; and Johnson, C. A., 1975, J. Parasitol., v. 61 (3), 554-557
Hematodinium sp., "agrees in all characteristics with Hematodinium perezi.", causing disease in Callinectes sapidus (hemolymph, tissues): North Carolina; Georgia; Florida

Hemogregarines. See Haemogregarines.

Henneguya spp.
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Salmo trutta
Gasterosteus aculeatus
all from Loch Leven, Scotland

Henneguya sp. (resembles H. exilis), illus.
McCraren, J. P.; et al., 1975, J. Wildlife Dis., v. 11 (1), 2-7
Henneguya sp. in Ictalurus punctatus, 7 disease manifestations described

Henneguya sp.
Meyers, T. R.; and Sawyer, T. K., 1976, Tr. Am. Micr. Soc., v. 95 (2), 269 [Abstract]
description of cyst and spore
Pomatomus saltatrix (surface and lumen of bulbous arteriosus): Raritan Bay, New Jersey; Chesapeake Bay, Maryland

Henneguya sp. similar to H. sebasta Moser and Love 1975, illus.
Meyers, T. R.; Sawyer, T. K.; and MacLean, S. A., 1977, J. Parasitol., v. 63 (5), 890-896
Henneguya sp. in Pomatomus saltatrix (bulbus and truncus arteriosus), gross and microscopic pathology, size of host: Atlantic Ocean near Montauk Point, Long Island, New York; Raritan Bay, New Jersey; Chesapeake Bay, Maryland

Henneguya [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Mugil cephalus (truncus arteriosus): Elat, Gulf of Aqaba, Red Sea

Henneguya adiposa sp. n., illus.
Minchew, C. D., 1977, J. Protozool., v. 24 (2), 213-220
development
Ictalurus punctatus (between connective tissue bands of adipose fin): pond in Lee Co., Mississippi

Henneguya diversis sp. n., illus.
Minchew, C. D., 1977, J. Protozool., v. 24 (2), 213-220
development
Ictalurus punctatus (near base of dorsal barbel; pectoral fins; along the isthmus; liver and kidney; necrotic muscle tissue near anus; dorsal and caudal fins): catfish farm in Perry Co., Alabama

Henneguya exilis Kudo, 1929
Baker, J. C.; and Crites, J. L., 1976, Proc. Helminth. Soc. Washington, v. 43 (1), 37-39
Ictalurus punctatus (gills): island region of western Lake Erie

Henneguya exilis, illus.
Current, W. L.; and Janovy, J., jr., 1976, J. Parasitol., v. 62 (6), 975-981
Henneguya exilis, interlamellar infections in Ictalurus punctatus, ultrastructural aspects (plasmodium wall structure, interactions between plasmodium and adjacent basal cells of host, apparent parasite-induced cytoarchitectural rearrangements of basal cells): Lake Keystone, Keith County, Nebraska

Henneguya exilis Kudo, 1929, illus.
Minchew, C. D., 1977, J. Protozool., v. 24 (2), 213-220
description
Ictalurus punctatus (filament of gill arch): commercial catfish farm in Humphreys Co., Missisippi

Henneguya longicauda sp. n., illus.
Minchew, C. D., 1977, J. Protozool., v. 24 (2), 213-220
development
Ictalurus punctatus (lamellar connective tissue and capillaries of gill filaments): pond in Neshoba Co., Mississippi

Henneguya pellis sp. n., illus.
Minchew, C. D., 1977, J. Protozool., v. 24 (2), 213-220
development
Ictalurus furcatus (dermis): Lee County, Alabama

Henneguya postexilis sp. n., illus.
Minchew, C. D., 1977, J. Protozool., v. 24 (2), 213-220
development
Ictalurus punctatus (lamellar connective tissue and capillaries of gill filaments): pond in Saline Co., Missouri

Henneguya psorospermica Thelohan, illus.
  Reichenbach-Klinke, H. H.; and Sturm, G., 1975, Fisch u. Umwelt (1), 51-57

Henneguya psorospermica Thelohan, 1895
  Willemse, J. J., 1968, Bull. Zool. Mus. Univ. Amsterdam, v. 1 (8), 83-87
    Esox lucius (gills, gonads): Ankeveen; Exloerveen; Haarlemmermeerpolder; Heemskerkerbroekpolder

Henneguya sebasta sp. n., illus.
  Moser, M.; and Love, M. S., 1975, J. Parasitol., v. 61 (3), 481-483
    Sebastes paucispinis
    S. jordani
    S. chlorostictus
    S. diploproa
    S. miniatus
    S. vexillaris
    S. serranoides
    (bulbus and truncus arteriosus, and heart chambers of all): all from Santa Barbara to Morro Bay, California

Henneguya sebasta Moser & Love 1975
  Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
    Sebastes chlorostictus (gallbladder, urinary bladder): off southern California
    S. flavidus (heart, gallbladder): off northern California
    S. goodei (heart, urinary bladder): off southern California
    S. levis (heart): off southern California
    S. melanops (heart): off central California
    S. rufus (heart): off southern California

Henneguya waltairensis n. sp., illus.
  Narasimhamurti, C. C.; and Kalavati, C., 1975, Riv. Parassitol., Roma, v. 36 (4), 255-259
    Ophiocephalus punctatus (gills): Viskhaptanm (Andhra Pradesh, India)

Henneguya zschokkei (Gurley), illus.
  Reichenbach-Klinke, H. H.; and Sturm, G., 1975, Fisch u. Umwelt (1), 51-57
    Coregonus acronius

Hentschelia Mackinnon & Ray, 1931
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Lecudinidae, revision and checklist of species
    diagnosis

Hentschelia thalassemae Mackinnon & Ray, 1931
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Hepatocystis
  Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
    Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis sp.
  Canning, E. U.; et al., 1977, J. Protozool., v. 24 (2), Suppl., 6A [Abstract]
    merocyst, fine structure
    Callosciurus nigrovittatus (liver): near Kuala Lumpur

Hepatocystis sp.
  Canning, E. U.; Killick-Kendrick, R.; and Garnham, P. C. C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 8 [Demonstration]
    Hepatocystis sp. from Callosciurus nigrovittatus, nuclear activity in macrogametocytes

Hepatocystis [sp.]
  Ewers, W. H., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 88-89 [Demonstration]
    Dobsonia inermis resea
    D. moluccensis
    D. minor
    Melonycteris woodfordi
    Syconycteris crassa
    Macroglossus lagochilus
    Pteropus rayneri grandis
    P. macrotis
    P. conspiculatus
    Paranyctimene raptor
    Nyctimene aello
    N. papuanus
    all from New Guinea

Hepatocystis n. sp.
  Killick-Kendrick, R.; Garnham, P.C.C.; and Rajapaksa, N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 2 [Demonstration]
    Callosciurus nigrovittatus (blood, liver): Kota Belud, Sabah

Hepatocystis [sp., ? rodhaini n. sp.]
  Landau, I.; et al., 1976, Ann. Parasitol., v. 51 (3), 303-307
    Hipposideros bicolor (sang): Limestone Cave (Ulu Gombak), grotte des environs de Kuala Lumpur, Malaisie

Hepatocystis sp.
  Landau, I.; and Adam, J. P., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 6-7 [Demonstration]
    2 types of schizonts
    Rhinolophus
    Miniopterus minor minor
    (liver, lungs of all): all from caves in Congo-Brazzaville

Hepatocystis bainae n. sp., illus.
  Mialhe, E.; and Landau, I., 1977, Ann. Parasitol., v. 52 (4), 385-390
    Hipposideros galeritus (hematies, foie): Ulu Gombak, Etat de Selangor, Malaisie

Hepatocystis brayi n. sp., illus.
  Miltgen, F.; et al., 1976, Ann. Parasitol., v. 51 (3), 287-297
    development
    Callosciurus notatus: Ulu Gombak, Bukit Mandol and Bukit Lanjan, Malaisie
    C. nigrovittatus: Ulu Gombak and Bukit Lanjan, Malaisie
    C. caniceps: Ulu Gombak, Malaisie
    Lariscus insignis: Bukit Lanjan, Malaisie

Hepatocystis brayi, illus.
  Canning, E. U.; et al., [1977], Ann. Parasitol., v. 51 (6), 1976, 607-623
    Hepatocystis malayensis, H. brayi, merocyst, gametocytes, gametogenesis, fertilization, ultrastructural observations

Hepatocystis brayi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis brayi, illus.
Miltgen, F.; et al., 1976, Ann. Parasitol., v. 51 (3), 299-302
Hepatocystis brayi, development in Culicoides spp.
Culex fatigans ("seuls des ookinetes ont ete observes dans le contenu stomacal quelques heures apres qu'ils se soient gorges")
Anopheles stephensi ("seuls des ookinetes ont ete observes dans le contenu stomacal quelques heures apres qu'ils se soient gorges")
Culicoides nebeculosus (paroi de l'estomac)
C. variipennis (entre l'epithelium stomacal et la basale)
(all exper.)

Hepatocystis brosseti n. sp., illus.
Miltgen, F.; et al., 1977, Ann. Parasitol., v. 52 (6), 589-596
Epomops franqueti (foie, rate): alentours de Makokou, Gabon

Hepatocystis epomophori
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis fieldi, illus.
Hoo, C. C.; and Sandosham, A. A., 1968, Med. J. Malaya, v. 22 (4), 299-301
morphology of early forms of Hepatocystis fieldi and exper. Plasmodium traguli in Tragulus javanicus

Hepatocystis hippopotami
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis kochi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis malayensis Field et Edeson, 1950 new rank, illus.
Landau, I.; et al., 1976, Ann. Parasitol., v. 51 (3), 271-286
redescription, development, immunological tissue reaction
Syn.: Hepatocystis vassali malayensis Field and Edeson, 1950
Callosciurus nigrovittatus: Ulu Gombak and Bukit Lanjan, Malaisie
Sundasciurus tenuis: Ulu Gombak, Malaisie

Hepatocystis malayensis, illus.
Canning, E. U.; et al., [1977], Ann. Parasitol., v. 51 (6), 1976, 607-623
Hepatocystis malayensis, H. brayi, merocyst, gametocytes, gametogenesis, fertilization, ultrastructural observations

Hepatocystis malayensis
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis oriheli Garnham, 1977
Garnham, P. C. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 386 [Demonstration]
Miopithecus talapoin (blood, liver): Rio Muni

Hepatocystis perronae
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis pteropi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis pteropi, illus.
Miyata, A.; and Tsukamato, M., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 113-130
Pteropus leucotis obscurus
P. vampyrus lanensis
all from Palawan Island, the Philippines

Hepatocystis rayi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis rayi wui, illus.
Pal, N.; and Dasgupta, B., 1974, Indian J. Physiol. and Allied Sc., v. 28 (2), 61-65
Hepatocystis rayi wui, composition of protoplasmic rim and colloidal mass of merocyst, amyloids and nucleic acids, fluorescent microscopy
Petaurista magnificus (spleen)
P. elegans caniceps (lung)

Hepatocystis rodhaini n. sp., illus.
Landau, I.; et al., 1976, Ann. Parasitol., v. 51 (3), 303-307
Hipposideros galeritus (sang, foie): Limestone Cave (Ulu Gombak), grotte des environs de Kuala Lumpur, Malaisie

Hepatocystis rodhaini
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis semnopitheci
Garnham, P. C. C.; and Rajapaksa, N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 2 [Demonstration]
Presbytis aygula (blood): Kota Belud, Sabah

Hepatocystis simiae, illus.
McConnell, E. E.; et al., 1974, Onderstepoort J. Vet. Research, v. 41 (3), 97-168
pathological and parasitological survey of 100 free-ranging chacma baboons
Papio ursinus (liver, blood): Kruger National Park, Transvaal

Hepatocystis taiwanensis
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Hepatocystis vassali (Laveran, 1905), illus.
Miyata, A.; and Tsukamato, M., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 113-130
"not identical with the Malayan subspecies but rather similar to the Taiwan subspecies, H.v. yokogawai Wu."
Callosciurus juvencus: Palawan Island, the Philippines

Hepatocystis vassali malayensis Field and Edeson, 1950
Landau, I.; et al., 1976, Ann. Parasitol., v. 51 (3), 271-286
as syn. of Hepatocystis malayensis Field et Edeson, 1950 new rank

Hepatocystis vassali yokogawai Wu
Miyata, A.; and Tsukamato, M., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 113-130
Hepatocystis vassali (Laveran, 1905), "not identical with the Malayan subspecies but rather similar to the Taiwan subspecies, H. v. yokogawai Wu."

Hepatozoon
Landau, I., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 403-407
analysis of analogy between Toxoplasma and Hepatozoon life cycles; importance and role of cyst formation in the Coccidia

Hepatozoon sp., illus.
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Lutzomyia vexatrix occidentis
Sceloporus occidentalis (nat. and exper.) (blood)
S. graciosus (blood)
Crotalus viridis (nat. and exper.) (blood)
Gerrhonotus [sp.] (exper.) (blood)
Lampropeltus getulus (nat. and exper.) (blood)
Pituophis melanoleucus (blood)
Thamnophis sirtalis (blood)
T. couchi (blood)
Colubra constrictor (nat. and exper.) (blood)
all from central California

Hepatozoon sp., illus.
Beaucournu, J. C.; and Deunff, J., [1976], Ann. Parasitol., v. 50 (6), 1975, 831-835
Ctenophthalmus baeticus arvernus: France

Hepatozoon spp., illus.
Clark, K. A., 1973, J. Wildlife Dis., v. 9 (2), 182-193
review of spp. from rodents and reptiles with literature references

Hepatozoon spp., resembling H. canis, illus.
Ewing, G. O., 1977, Feline Pract., v. 7 (6), 37-40
cat (liver), granulomatous cholangiohepatitis: California, originally from Oahu, Hawaii

Hepatozoon sp., illus.
McCully, R. M.; et al., 1975, Onderstepoort J. Vet. Research, v. 42 (4), 117-133
Hepatozoon sp. in wild carnivores and H. canis in dogs compared, location in host, developmental stages, lesions, pathogenesis; attempt to transmit Hepatozoon from jackal to dogs by means of Rhipicephalus sanguineus was inconclusive
Hyaena crocuta
Panthera leo
P. pardus
Acinonyx jubatus
Canis mesomelas
C. familiaris (exper.)
Rhipicephalus simus
R. sanguineus (exper.)
all from South Africa

Hepatozoon sp.
Presidente, P. J. A.; and Karstad, L. H., 1975, J. Wildlife Dis., v. 11 (4), 479-481
Mustela vison (lungs; pulmonary parenchyma; peribronchiolar or perivascular sites): southwestern Ontario

Hepatozoon sp., illus.
Wiger, R., 1973, J. Wildlife Dis., v. 9 (2), 125-128
Lemmus lemmus (blood): Norway

Hepatozoon [sp.]
Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
Neotoma a. albigula (blood): Tonto National Monument, near Roosevelt, Gila County, Arizona
N. fuscipes macrotis (blood): Griffith Park, Los Angeles County, California

Hepatozoon akodoni
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
synonymy

Hepatozoon canis
Bordoski, A.; and Savin, Z., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 33-36
dog: Bar district, Montenegro

Hepatozoon canis
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon canis James, 1905

Hepatozoon canis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Hepatozoon canis
Leeflang, P.; Pimentel, W. J.; and Blotkamp, J., 1974, Bull. Epizoot. Dis. Africa, v. 22 (1), 51-53
Babesia canis, Hepatozoon canis, Haemobartonella canis, dog, case report, also infested with Rhipicephalus sanguineus: Nigeria

Hepatozoon canis James, 1905, illus.
McCully, R. M.; et al., 1975, Onderstepoort J. Vet. Research, v. 42 (4), 117-133
Hepatozoon sp. in wild carnivores and H. canis in dogs compared, location in host, developmental stages, lesions, pathogenesis; attempt to transmit Hepatozoon from jackal to dogs by means of Rhipicephalus sanguineus was inconclusive: South Africa

Hepatozoon canis, illus.
Novilla, M. N.; Kwapien, R. P.; and Peneyra, R. S., 1977, Proc. Helminth. Soc. Washington, v. 44 (1), 98-101
Hepatozoon canis, dogs, 3 case reports, histopathology: Philippines

Hepatozoon domerguei
Vivier, E., 1975, J. Protozool., v. 22 (3), 69A [Abstract]
Hepatozoon domerguei, ultrastructural observations of fertilization and meiosis

Hepatozoon domerguei, Landau, Chabaud, Michel, et Brygoo (1970), illus.
Vivier, E.; Petitprez, A.; and Landau, I., 1972, Protistologica, v. 8 (3), 315-334
Hepatozoon domerguei, sporoblastogenesis, ultrastructure

Hepatozoon domerguei
Vivier, E.; and Porchet, E., 1976, J. Protozool., v. 23 (4), 22A
Aggregata eberthi, Hepatozoon domerguei, Dehornia sthenelais, crystalline inclusions

Hepatozoon erhardovae Krampitz 1964
Krampitz, H. E.; Wongchari, V.; and Geisel, O., 1976, Ztschr. Parasitenk., v. 50 (2), 202
cell parasitism in vertebrates and invertebrates

Hepatozoon felis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Hepatozoon felisdomesticae
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon felis-domesticae Patton, 1908

Hepatozoon funambuli
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon funambuli Patton, 1908

Hepatozoon griseisciuri
Davidson, W. R., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 211-217
epizootiologic and pathologic study of endoparasites of selected populations of gray squirrels
Sciurus carolinensis (blood, lung): southeastern United States

Hepatozoon griseisciuri, illus.
Davidson, W. R.; and Calpin, J. P., 1976, J. Wildlife Dis., v. 12 (1), 72-76
Hepatozoon griseisciuri, location and pathogenicity of schizogonic stages, age and sex of host (gray squirrel), seasonal distribution, pathology
Sciurus carolinensis (blood, lungs): Alabama; Georgia; South Carolina; Mississippi; Maryland; North Carolina; Arkansas; Kentucky; Tennessee; Virginia; West Virginia

Hepatozoon griseisciuri Clark 1958, illus.
Hendricks, L. D., 1975, J. Parasitol., v. 61 (3), 458-461
Hepatozoon griseisciuri, schizogonic development in the gray squirrel, distribution and concentration of stages throughout host reticuloendothelial system, wide divergence in number of merozoites, comparison of in vivo and in vitro forms
Sciurus carolinensis: Maryland

Hepatozoon griseisciuri
  Hendricks, L. D.; and Fayer, R., 1973, J. Protozool., v. 20 (5), 550-554
    Hepatozoon griseisciuri, development in cultured squirrel cells

Hepatozoon leporis
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    Syn.: Leucocytozoon leporis Patton, 1908

Hepatozoon leptosoma
  Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
    Peromyscus maniculatus (blood): Petty Ranch near Frazier Mountain Park, Ventura County, California
    Peromyscus boylii: Juniper Hills near Pearblossom, Los Angeles County, California

Hepatozoon muris
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    synonymy

Hepatozoon musculi
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    Syn.: Leucocytogregarina musculi (Porter, 1908) Sangiorgi, 1912

Hepatozoon neophrontis (Todd & Wolbach, 1912) Bray, 1964
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    Syn.: Toxoplasma neophrontis (Todd & Wolbach, 1912) Wenyon, 1926

Hepatozoon procyonis
  Clark, K. A., 1973, J. Wildlife Dis., v. 9 (2), 182-193
    Hepatozoon procyonis, high prevalence in Procyon lotor (blood, myocardium, skeletal muscle, spleen), probable accidental infection of Odocoileus virginianus (myocardium), increased numbers of gametocytes following splenectomy or corticosteroid treatment of raccoons, lesions associated with hepatozoonosis: Texas

Hepatozoon ratti
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    Syn.: Leucocytozoon ratti Adie, 1906

Herpetomonas
  Daggett, P. M.; and Decker, J. E., 1974, J. Protozool., v. 21 (3), 429 [Abstract]
    comparison of excreted factors of insect trypanosomatids at 25 C vs. 37 C, results indicate secretion products are primarily under parasite not host control

Herpetomonas
  Schnur, L. F., 1977, J. Protozool., v. 24 (2), Suppl., 5A-6A [Abstract]
    Leptomonas, Crithidia, Herpetomonas, L[eishmania] tropica, L[eishmania] tropica major, L[eishmania] tropica minor, L[eishmania] donovani, excreted metabolic factors (EF) and homologous antisera, cross-reactions, no EF apparently produced by Trypanosoma cruzi, suggests closer relationship between insect flagellates and leishmanias than between these and trypanosomes like Trypanosoma cruzi

Herpetomonas [sp.]
  Hommel, M.; and Croft, S. L., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 431 [Demonstration]
    Herpetomonas [sp.], naturally occurring in colony of Culicoides variipennis bred at Animal Virus Research Centre, morphological aspects: Pirbright, England

Herpetomonas sp.
  Lushbaugh, W. B.; Rowton, E. D; and McGhee, R. B., 1975, J. Protozool., v. 22 (3), 10A-11A [Abstract]
    Drosophila melanogaster (intestine)

Herpetomonas sp. (Leptomonas pessoai), illus.
  de Souza, W.; et al., 1976, Canad. J. Microbiol., v. 22 (2), 197-203
    Leptomonas pessoai, "...this insect flagellate is a member of the genus Herpetomonas"; ultrastructure, electron microscopy of promastigotes

Herpetomonas ampelophilae
  Rowton, E. D.; and McGhee, R. B., 1975, J. Protozool., v. 22 (3), 29A [Abstract]
    Herpetomonas ampelophilae, population dynamics in feral Drosophila melanogaster maintained in the laboratory and in a population of clean Drosophila to which infected flies were added

Herpetomonas megaseliae
  Daggett, P. M.; and Decker, J. E., 1973, J. Protozool., v. 20 (4), 512
    6 insect trypanosomatids, increased survival in mice with repeated passage associated with alteration in secretion

Herpetomonas megaseliae
  Janovy, J., jr.; et al., 1973, J. Protozool., v. 20 (4), 502
    Herpetomonas megaseliae, differentiation in culture, morphological and physiological changes

Herpetomonas megaseliae
  Janovy, J., jr.; et al., 1975, Proc. Oklahoma Acad. Sc., v. 55, 130-135
    Herpetomonas megaseliae, cytodifferentiation in culture, changes in population composition (promastigotes, paramastigotes, opisthomastigotes), physiological changes (organisms became increasingly cyanide-sensitive, anaerobic stimulation of glucose uptake was doubled, and anaerobic acid production was halved as makeup of culture population shifted), differentiation was postulated to involve metabolic shift from anaerobic to aerobic metabolism

Herpetomonas megaseliae Daggett, Dollahon and Janovy, 1972, illus.
Janovy, J., jr.; Lee, K. W.; and Brumbaugh, J. A., 1974, J. Protozool., v. 21 (1), 53-59
Herpetomonas megaseliae, ultrastructure of both undifferentiated (promastigote and paramastigote) and differentiated (opisthomastigote) forms

Herpetomonas megaseliae Daggett, Dollahon, and Janovy 1972, illus.
Keppel, A. D.; and Janovy, J., jr., 1977, J. Parasitol., v. 63 (5), 879-882
Herpetomonas megaseliae and Crithidia harmosa, growth on blood-agar plates as discrete clone colonies, colony morphology differs in the two genera and species, possible applications

Herpetomonas megaseliae, illus.
Knight, S. A., 1976, J. Parasitol., v. 62 (4), 515-522
Herpetomonas megaseliae, cultures grown with hydroxyurea (inhibitor of DNA synthesis), differences in population number, kinetoplast number and position, and pellicular morphology, net effects of hydroxyurea are enhanced differentiation and abortive cytokinesis

Herpetomonas muscarum
Carvalho, A. L. M.; and Deane, M. P., 1974, J. Protozool., v. 21 (1), 5-8
Trypanosomatidae isolated from Zelus leucogrammus, only Blastocrithidia thought to be a parasite proper of this host spread by cannibalism and ingestion of cysts, other trypanosomatids acquired by feeding on insect prey: Brazil

Herpetomonas muscarum
Daggett, P. M.; and Decker, J. E., 1973, J. Protozool., v. 20 (4), 512
6 insect trypanosomatids, increased survival in mice with repeated passage associated with alteration in secretion

Herpetomonas muscarum
Muse, K. E.; and Roberts, J. F., 1973, J. Protozool., v. 20 (4), 502
ultrastructure
Hippelates pusio (midgut) (nat. and exper.): laboratory colonies

Herpetomonas muscarum ingenoplastis
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Herpetomonas muscarum ingenoplastis, illus.
Wallace, F. G.; Wagner, M.; and Rogers, W. E., 1973, J. Protozool., v. 20 (2), 218-222
Herpetomonas m. muscarum vs. H. m. ingenoplastis, very different structure of kinetoplast nucleoid DNA; ultrastructural nature of doubling of flagellum

Herpetomonas muscarum muscarum, illus.
Wallace, F. G.; Wagner, M.; and Rogers, W. E., 1973, J. Protozool., v. 20 (2), 218-222
Herpetomonas m. muscarum vs. H. m. ingenoplastis, very different structure of kinetoplast nucleoid DNA; ultrastructural nature of doubling of flagellum

Herpetomonas oestrorum Rodhain & Bequaert
Pester, F. R. N.; and Laurence, B. R., 1974, J. Zool., London, v. 174 (3), 397-406
Oestrus
Kirkioestrus
Gedoelstia cristata
G. hassleri
Oestrus aureoargenteus
Gedoelstia
all from Kenya

Herpetomonas pachelabrae, Froilano de Mello, 1921
Nicoli, R. M.; Penaud, A.; and Timon-David, P., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 415-419
as syn. of Malacozoomonas pachylabrae (Froilano de Mello, 1921) emend. [n. comb.]

Herpetomonas patellae A. Porter, 1914
Nicoli, R. M.; Penaud, A.; and Timon-David, P., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 415-419
as syn. of Malacozoomonas patellae (A. Porter, 1914) [n. comb.]

Herpetomonas samuelpessoai sp. n. [i.e., nom. nov.], illus.
Roitman, I.; et al., 1976, J. Protozool., v. 23 (2), 291-293
"Leptomonas pessoai may be regarded as a thermophilic species of Herpetomonas. . .and we suggest that it be designated Herpetomonas samuelpessoai sp. n.", differentiation of original strain and clones from promastigote to opisthomastigote faster at high temperature and in defined medium

Herpetomonas samuelpessoai, illus.
Angluster, J.; Bunn, M. M.; and de Souza, W., 1977, J. Parasitol., v. 63 (5), 922-924
Herpetomonas samuelpessoai grown in defined medium, differentiation, influence of 2-deoxy-D-glucose, temperature, and time of cultivation

Herpetomonas samuelpessoai, illus.
de Carvalho, T. U.; and de Souza, W., 1977, J. Parasitol., v. 63 (6), 1116-1117
Herpetomonas samuelpessoai, electron-dense granules, fine structure and x-ray microanalysis

Herpetomonas samuelpessoai
Mattei, D. M.; et al., 1977, FEBS Letters, v. 74 (2), 264-268
Trypanosoma cruzi, biochemical strain characterization by restriction endonuclease cleavage of kinetoplast-DNA, comparison to Herpetomonas samuelpessoai, possible application in kinetoplastid taxonomy

Herpetomonas samuelpessoai
Pereira, N. M.; et al., 1977, J. Protozool., v. 24 (4), 511-514
Crithidia fasciculata, Herpetomonas samuelpessoai, Leishmania tarentolae, isolation of flagella, 3 types of flagella give similar electrophoretic pattern of proteins, H. samuelpessoai and to a lesser extent C. fasciculata flagella confer protection against Trypanosoma cruzi infections in mice

Hexamastix batrachorum (Alexeieff 1911)
Nohynkova, E., 1974, J. Protozool., v. 21 (3), 457-458 [Abstract]
Haemopis sanguisuga (intestine): Czechoslovakia

Hexamita Dujardin, 1838 (type genus)
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Diplomonadina emend., Hexamitidae

Hexamita sp., illus.
Leibovitz, L., 1973, Am. J. Vet. Research, v. 34 (8), 1053-1061
Protozoa contributing to pathogenesis of necrotic enteritis in commercial white Pekin breeder ducks; necrotic enteritis also observed in other wild and domestic Anseriformes

Hexamita [sp.]
Nohynkova, E., 1974, J. Protozool., v. 21 (3), 457-458 [Abstract]
Haemopis sanguisuga (intestine): Czechoslovakia

Hexamita sp.
Overstreet, R. M., 1976, J. Parasitol., v. 62 (5), 702-708
Crassicutis archosargi (ceca): Grand Isle, Louisiana; Mississippi Sound and adjacent areas

Hexamita [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Siganus (digestive tract)
Mugil cephalus
all from Elat, Gulf of Aqaba, Red Sea

Hexamita intestinalis, illus.
Herkner, H., 1975, Fisch u. Umwelt (1), 139-146
parasites as cause of disease in aquarium fish, review

Hexamita intestinalis
Lank, D. R., jr., 1971, Proc. Indiana Acad. Sc., v. 81 (2), 359-364
Rana catesbeiana: Indiana

Hexamita muris, illus.
Boorman, G. A.; et al., 1973, Lab. Animal Sc., v. 23 (2), 187-193
Hexamita muris, Giardia muris, potentially pathogenic in newly weaned mice, causing enteritis and mortality in association with normal intestinal bacterial flora, quinacrine dihydrochloride not effective in reducing mortality, dimetridazole effective

Hexamita muris (Grassi, 1881)
Kunstyr, I.; and Ammerpohl, E., 1976, Ztschr. Parasitenk., v. 50 (2), 212
Hexamita muris, isolation, establishment of infection in thymusless mice, possible experimental model for immunological studies

Hexamita muris, illus.
Matthiesen, T.; Kunstyr, I.; and Tuch, K., 1976, Ztschr. Versuchstierk., v. 18 (3), 113-120
Hexamita muris infection in laboratory mice and hamsters, clinical and pathohistological findings, pathogenicity resulting from stress conditions
Syn.: Octomitus muris

Hexamita muris
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Rattus norvegicus
Mus musculus
all from Carshalton

Hexamita parva
Gabrisch, K., 1976, Prakt. Tierarzt , v. 57, Sondernummer, 37-40
parasites of reptiles, diagnosis, treatment, brief review

Hexamita parva, illus.
Zwart, P.; and Truyens, E. H. A., 1975, Vet. Parasitol., v. 1 (2), 175-183
pathogenic properties, kidneys especially affected
Testudo horsfieldii
T. marginata
T. elegans
T. carbonaria
Cuora amboinensis
Terrapene ornata
Geoemyda p. punctularia
Clemmys caspica leprosa

Hexamita salmonis, illus.
Rothenbacher, H. J.; and Bohl, M., 1975, Fisch u. Umwelt (1), 13-23
Hexamita salmonis, brief review of morphology, pathogenicity, diagnosis and therapy; trial of Enheptin, good results

Hexamita truttae (Dujardin) Schmidt, 1920
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Salmo trutta: Loch Leven, Scotland

Hexamitidae Kent
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
diagnosis

Histomonas meleagridis
Al-Khateeb, G. H.; and Hansen, M. F., 1973, Avian Dis., v. 17 (2), 269-273
Histomonas meleagridis, turkeys (exper.), plasma glutamic oxalacetic transaminase level, correlation with number of liver lesions, salfuride treatment reduced number of liver lesions and lowered plasma GOT levels, useful as indicator of course of disease and for screening potential histomonastats

Histomonas meleagridis
Al-Khateeb, G. H.; and Hansen, M. F., 1974, Avian Dis., v. 18 (4), 507-514
Histomonas meleagridis, turkeys and chickens (both exper.), plasma enzyme levels used to evaluate susceptibility according to breed and age of host, duration of infection in chickens, effect of route of inoculation, and effect on virulence of the age of in vitro subcultures

Histomonas meleagridis
Bradley, R. E.; and Panitz, E., 1976, J. Parasitol., v. 62 (4), 643-644
Histomonas meleagridis, turkey poults (exper.), prophylactic efficacy of (5-nitro-1-thienyl)-N-acetylamine

Histomonas meleagridis
Chappel, L. R., 1975, Avian Dis., v. 19 (4), 679-682
HmL1 strain of Histomonas meleagridis, chicks, pathogenicity; three 1,2-disubstituted 5-nitroimidazoles tested to demonstrate applicability of disease model to chemotherapy

Histomonas meleagridis
Chute, A. M.; and Lund, E. E., 1972, J. Protozool., v. 19 (4), 639-643
Histomonas meleagridis in Numida meleagris (exper.), severe infections following inoculation by rectal route, much less severe response to natural route of infection (ingestion of Heterakis gallinarum eggs or earthworms carrying H. gallinarum and Histomonas meleagridis)

Histomonas meleagridis
Cuckler, A. C., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 371-397
coccidiosis and histomoniasis in avian hosts, host-parasite relationships, immunity, review

Histomonas meleagridis
Dwyer, D. M., 1974, J. Protozool., v. 21 (1), 139-145
Trichomonas gallinae, Histomonas meleagridis, Dientamoeba fragilis, Entamoeba invadens, E. histolytica, antigenic relationships analyzed by immunoelectrophoretic techniques

Histomonas meleagridis
Hirsch, R. P.; and Hirsch, D. S., 1977, J. Parasitol., v. 63 (2), 386-388
Histomonas meleagridis, in vitro culture, millipore-sterilized DeVolt's medium supports better growth than does autoclaved DeVolt's medium

Histomonas meleagridis
Kemp, R. L.; and Franson, J. C., 1975, Avian Dis., v. 19 (4), 741-744
Histomonas meleagridis, experimental transmission to chickens and turkeys by earthworms recovered from pheasant yard soil; earthworms may serve as transport hosts for Heterakis eggs and thus ultimately for H. meleagridis

Histomonas meleagridis
Kulda, J.; Suchankova, E.; and Svoboda, S., 1974, Acta Vet. Brno, v. 43 (1), 53-64
Tetratrichomonas gallinarum, synonymy; axenic culture of 10 strains isolated from poultry, virulence determined by mouse assay did not reflect pathogenicity for turkey poults, results indicate T. gallinarum should not be considered serious pathogen; enterohepatitis caused by mixed infection with Histomonas meleagridis from contaminated culture

Histomonas meleagridis
Long, P. L., 1971, Symposia Brit. Soc. Parasitol., v. 9, 65-75
maintenance of Eimeria and Histomonas in vivo, extensive review: maintenance of Eimeria (purification of species, exper. infections in chickens, factors affecting susceptibility of chickens and chicken embryos to infection; freeze-preservation); maintenance of Histomonas meleagridis (infection with ground-up infected tissues; use of in vitro cultures to obtain in vivo infections; use of embryonated Heterakis gallinarum ova to induce histomoniasis in vivo)

Histomonas meleagridis
Lund, E. E.; and Chute, A. M., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 73-76
larval Heterakis gallinarum, reciprocal transfer between Phasianus colchicus and Coturnix coturnix japonica, influence of early development in incompatible host on subsequent development; effect on liberation of Histomonas meleagridis and Parahistomonas wenrichi and transmission of H. meleagridis

Histomonas meleagridis (Smith, 1895) Tyzzer, 1920
Madsen, H., 1976, Norwegian J. Zool., v. 24 (4), 460 [Abstract]
epidemiology
Syn.: Parahistomonas wenrichi

Histomonas meleagridis (Smith, 1895) Tyzzer, 1920
Marx, D. J., 1973, J. Protozool., v. 20 (4), 519
White Wrolsted turkey (bursa of Fabricius) (exper.)

Histomonas meleagridis
Mayaudon T., H., 1974, Rev. Med. Vet. y Parasitol., Maracay, v. 25 (1-8), 191-196
very brief review of disease and prevalence, case history in Meleagris gallopavo: Venezuela

Histomonas meleagridis
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Histomonas meleagridis
Robinson, R. M., 1975, Proc. 3. National Wild Turkey Symp., 33-35
infection as possible source of disaster in wild turkey management

Histomonas meleagridis, illus.
Rybicka, K.; Honigberg, B. M.; and Holt, S. C., 1972, Protistologica, v. 8 (1), 107-120
Histomonas meleagridis culture forms, mastigont system, fine structure

Histomonas meleagridis
Sullivan, T. W.; Grace, O. D.; and Aksoy, A., 1977, Poultry Science, v. 56 (2), 571-576
Histomonas meleagridis, turkeys (exper.), influence of level, timing and duration of ronidazole water medication

Histomonas meleagridis
Voogd, C. E.; van der Stel, J. J.; and Jacobs, J. J. J. A. A., 1974, Mutation Research, v. 26 (6), 483-490
possible mutagenic action of metronidazole and nimorazole used to treat human trichomoniasis and dimetridazole and ronidazole used prophylactically against Histomonas meleagridis in poultry

Histomonas meleagridis
Wilkins, D.; and Lee, D. L., 1974, J. Comp. Path., v. 84 (4), 589-597
Histomonas meleagridis, topography of caecal mucosa of infected and uninfected turkeys studied with scanning electron microscope

Histomonas meleagridis
Wilkins, D.; and Lee, D. L., 1976, Parasitology, v. 72 (1), 51-63
Histomonas meleagridis, caecal wall and liver of infected turkey poults, changes in amount and distribution of acid and alkaline phosphatase, non-specific esterase, glycogen, lipid, and acid mucopolysaccharide

Histomoniasis
Soltys, M. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 309-322
immunization against protozoan diseases of animals, review

Hoarella
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Hoareosporidium Pande, Bhatia, and Chauhan 1972
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis Lankester 1882

Hoareosporidium pellerdyi Pande, Bhatia, and Chauhan 1972
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
probably as syn. of Sarcocystis ovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975

Hoareosporidium pellerdyi Pande, Bhatia, and Chauhan 1972
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
perhaps as syn. of Sarcocystis bertrami Doflein 1901

Hoplitophrya Stein, 1859
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Hoplitophryinae

Hoplitophrya acanthodes de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eminoscolex: Gabon

Hoplitophrya armata Stein, 1860
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Hoplitophrya brasili p. p. Hentschel, 1925
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Durchoniella brasili (Leger et Duboscq, part. 1894) de Puytorac, 1954

Hoplitophrya cheissini de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eminoscolex: Gabon

Hoplitophrya clavata (part.) Schweier, 1898
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella clavata (Leidy, 1855) Cepede, 1910

Hoplitophrya criodrili Miyashita, 1933
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Criodrilus sp.

Hoplitophrya eminoscolecis de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eminoscolex: Gabon

Hoplitophrya fastigata Mobius, 1888
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella fastigata (Mobius, 1888) Cepede, 1910

Hoplitophrya fissispiculata (Cheissin, 1930) Heidenreich, 1935, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Protoradiophrya fissispiculata Cheissin, 1930
Styloscolex sp.: Baikal

Hoplitophrya georgevitchi de Puytorac, 1958, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Lumbriculus sp.: Ochrid

Hoplitophrya hamata Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Hoplitophrya henleae Meier, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Henlea nasuta
H. ventriculosa

Hoplitophrya libyodrili de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Libyodrilus violaceus: Gabon

Hoplitophrya lituiformis Georg., 1943
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lituiformis (Georgevitch, 1943)

Hoplitophrya lumbrici (Stein, 1860) in Kent, 1882 and in Buetschli, 1887
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Hoplitophrya lumbrici Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin. 1841)

Hoplitophrya lumbrici (part.) in Schewiakoff, 1896 and in Schweier, 1898
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Hoplitophrya lumbriculi Katashima, 1952
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Lumbriculus sp.

Hoplitophrya lumbriculi Kijenskij, 1925
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Hoplitophrya minuta (Miyashita), 1933
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Protoradiophrya minuta, Miyashita, 1933
Criodrilus sp.

Hoplitophrya pungens Stein, 1861
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya pungens (Stein, 1861) Heidenreich, 1935

Hoplitophrya secans (Stein 1859) Stein, 1861, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Lumbriculus variegatus

Hoplitophrya secans part. in Schewiakoff, 1896
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Hoplitophrya secans part. in Schweier, 1898
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Hoplitophrya secans part. Schewiakoff, 1896
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella spiculata (Warpachowsky, 1886) Cepede, 1930

Hoplitophrya secans part. Schouteden
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella spiculata (Warpachowsky, 1886) Cepede, 1930

Hoplitophrya secans part. Schweier, 1898
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella spiculata (Warpachowsky, 1886) Cepede, 1930

Hoplitophrya tubificis Mac Kinnon et Adam, 1924
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya pungens (Stein, 1861) Heidenreich, 1935

Hoplitophrya variabilis Katashima, 1952
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubifex sp.

Hoplitophrya werestschagini (Cheissin, 1930) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Protoradiophrya werestchagini [sic], Cheissin, 1930
Styloscolex sp.: Baikal
Lamprodrilus sp.: Baikal

Hoplitophryidae (Hoplitophryidae, Cheissin, 1930) nov. comb.
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryoidea nv. super-fam.
includes: Hoplitophryinae; Jirovecellinae n. s. fam.; Juxtaradiophryinae n. s. fam.; Mixtophryinae n. s. fam.; Mesnilellinae

Hoplitophryinae Cepede, 1910
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae
includes: Hoplitophrya; Protoradiophryopsis

Hoplitophryoidea nv. super-fam.
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Astomatida
includes: Hoplitophryidae (Hoplitophryidae) nov. comb.; Radiophryidae n. fam.; Contophryidae n. fam.; Maupasellidae; Intoshellinidae

Hoplorhynchus ozakii Hukui ssp. coreica nov., illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Otocryptops rubiginosus: Seoul, Coree du sud

Hovasseiella de Puytorac, 1955
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Durchoniellinae n. s. fam.

Hovasseiella polydorae de Puytorac, 1955, illus.
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Polydora giardi

Hyalophysa Bradbury, emend.
  Bradbury, P. C.; and Clamp, J. C., 1973, J. Protozool., v. 20 (2), 210-213
    diagnosis

Hyalophysa chattoni, illus.
  Bradbury, P. C., 1973, J. Protozool., v. 20 (3), 405-414
    Hyalophysa chattoni, fine structure of organelles concerned with ingestion of exuvial fluid by trophont

Hyalophysa chattoni, illus.
  Bradbury, P. C., 1974, J. Protozool., v. 21 (1), 112-120
    Hyalophysa chattoni, phoront, fine structure

Hyalophysa chattoni Bradbury, emend.
  Bradbury, P. C.; and Clamp, J. C., 1973, J. Protozool., v. 20 (2), 210-213
    diagnosis

Hyalophysa chattoni Bradbury
  Grimes, B. H., 1976, J. Protozool., v. 23 (2), 246-251
    Palaemonetes pugio
    P. vulgaris
    Panopeus herbstii
    Callinectes sapidus
    Clibanarius vittatus
    Pagurus longicarpus
    Upogebia affinis
    all from North Carolina

Hyalophysa lwoffi sp.n., illus.
  Bradbury, P. C.; and Clamp, J. C., 1973, J. Protozool., v. 20 (2), 210-213
    Palaemonetes paludosus (exoskeleton, molt): Yates Pond, Raleigh, North Carolina, U.S.A.

Hyalophysa lwoffi Bradbury & Clamp
  Grimes, B. H., 1976, J. Protozool., v. 23 (2), 246-251
    Palaemonetes paludosus
    Cambarus sp.
    both from North Carolina

Hyalophysa trageri sp. n., illus.
  Grimes, B. H., 1976, J. Protozool., v. 23 (2), 246-251
    life cycle
    Uca sp.: Walden Creek, Old Brunswick, and Sunset Beach, North Carolina, U.S.A.
    U. pugnax: Walden Creek, Southport; and Old Brunswick, North Carolina, U.S.A.
    U. minax: Walden Creek, Old Brunswick, and Sunset Beach, North Carolina, U.S.A.
    Sesarma reticulatum: Walden Creek and Old Brunswick, North Carolina, U.S.A.
    S. cinereum: Walden Creek and Old Brunswick, North Carolina, U.S.A.
    (gill lamellae and molt of crab of all)

Hyperidion Mackinnon & Ray, 1931
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Lecudinidae, revision and checklist of species
    diagnosis

Hyperidion neptuni Reichenow, 1932
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Hyperidion thalassemae Mackinnon & Ray, 1931

Hyperidion thalassemae Mackinnon & Ray, 1931
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Hyperidion neptuni Reichenow, 1932

Hypotrichomonadinae Honigberg, 1963
  Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
    Trichomonadida Kirby, emend.; Monocercomonadidae Kirby, emend.

Hypotrichomonas Lee
  Honigberg, B. M.; Daniel, W. A.; and Mattern, C. F. T., 1972, J. Protozool., v. 19 (3), 446-453
    Trichomitus batrachorum, fine structure, evolutionary significance; evidence that Hypotrichomonas is valid genus and not abnormal acostate form of Trichomitus

Hypotrichomonas hemidactyli n. sp., illus.
  Bhaskar Rao, T.; Bhaskar Rao, T. S.; and Devi, A., 1976, Current Sc., Bangalore, v. 45 (7), 263-264
    Hemidactylus sp.: Warangal region, A.P., India

Ichthyobodo necatrix Pinto, 1928, illus.
Bohl, M., 1975, Fisch u. Umwelt (1), 25-33
distribution in pond fish, description, pathology, diagnosis, therapy, review

Ichthyophthirius sp.
Rawstron, R. R., 1971, Calif. Fish and Game, v. 57 (4), 253-256
Salmo gairdneri (gills): Darrah Springs Hatchery, Shasta County, California

Ichthyophthirius multifiliis, illus.
Antychowicz, J., 1977, Bull. Vet. Inst. Pulawy, v. 21 (1-2), 35-43
Ichthyophthirius multifiliis, influence of copper sulphate, malachite green, methylene blue, trypan blue and Neguvon on development

Ichthyophthirius multifiliis Fouquet, 1876
Baker, J. C.; and Crites, J. L., 1976, Proc. Helminth. Soc. Washington, v. 43 (1), 37-39
Ictalurus punctatus (gills): island region of western Lake Erie

Ichthyophthirius multifiliis Fouquet
Cloutman, D. G., 1976, Southwest Nat., v. 21 (1), 67-70
Campostoma anomalum pullum (skin): White River, Arkansas

Ichthyophthirius multifiliis Fouquet, 1876
Dabrowska, Z., 1970, Acta Parasitol. Polon., v. 17 (20-38), 189-193
Barbus barbus
Leuciscus cephalus
(gills of all): all from Vistula River near Warsaw

Ichthyophthirius multifiliis
Heckmann, R.; and Farley, D. G., 1973, J. Wildlife Dis., v. 9 (3), 221-224
Hesperoleucus symmetricus symmetricus (gills, body surface): foothill streams east of Fresno, California

Ichthyophthirius multifiliis, illus.
Herkner, H., 1975, Fisch u. Umwelt (1), 139-146
parasites as cause of disease in aquarium fish, review

Ichthyophthirius multifiliis
Kozel, T. R., 1976, Tr. Kentucky Acad. Sc., v. 37 (1-2), 41-44
Ichthyophthirius multifiliis, distribution on body of Fundulus notatus, younger fish more heavily infested than older ones, host behavioral changes: area surrounding confluence of Pope Lick Creek and Floyds Fork Creek, near Pope Lick Road bridge, southeastern Jefferson County, Kentucky

Ichthyophthirius multifiliis
Lom, J.; and Cerkasovova, A., 1974, J. Protozool., v. 21 (3), 457 [Abstract]
Ichthyophthirius multifiliis theronts, host-finding

Ichthyophthirius multifiliis
Loyacano, H. A., jr.; and Crane, J. S., 1977, Progr. Fish Cult., v. 39 (1), 39-40
Ichthyophthirius multifiliis in Anguilla rostrata, malachite green alone and combined with formalin (good results), Dylox (ineffective)

Ichthyophthirius multifiliis
Munday, B. L., 1976, Austral. Vet. Practitioner, v. 6 (1), 46-50
brief review of protozoan and other infectious diseases, farmed trout: Australia

Ichthyophthirius multifiliis, illus.
Negele, R. D., 1975, Fisch u. Umwelt (1), 59-65
description, life cycle, localization, pathology, epidemiology, prophylaxis, therapy, review

Ichthyosporidium sp. Schwartz, 1963, illus.
Sprague, V.; and Vernick, S. H., 1974, J. Protozool., v. 21 (5), 667-677
Ichthyosporidium sp., apparently identical with I. giganteum, fine structure of cyst and some sporulation stages, cyst apparently a defensive reaction of host origin but living and metabolically active, new interpretation of Golgi complex
Leiostomus xanthurus: Chesapeake Bay, Maryland

Ichthyosporidium giganteum (Thelohan, 1895) Swarczewsky, 1914, illus.
Sprague, V.; and Vernick, S. H., 1974, J. Protozool., v. 21 (5), 667-677
Ichthyosporidium sp., apparently identical with I. giganteum, fine structure of cyst and some sporulation stages, cyst apparently a defensive reaction of host origin but living and metabolically active, new interpretation of Golgi complex
Leiostomus xanthurus: Chesapeake Bay, Maryland

Immanoplasma Neumann, 1909, illus.
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses

Infundibulorium cameli
Gill, H. S., 1976, Indian Vet. J., v. 53 (11), 897-898
Camelus dromedarius (feces): States of Punjab, Haryata, and Rajasthan

Intoshellina Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Intoshellinidae

Intoshellina sp. de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Intoshellina cheissini n. sp.

Intoshellina cheissini n. sp., illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Intoshellina sp. de Puytorac, 1954
Tubifex

Intoshellina limnodrili Katashima, 1952
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Limnodrilus

Intoshellina maupasi Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubifex tubifex

Intoshellina ochridana de Puytorac, 1960, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Euilyodrilus ochridanus

Intoshellina poljanskyi Cheissin, 1930, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Limnodrilus arenarius

Intoshellina sapkarevi de Puytorac, 1960, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Euilyodrilus isochaetus

Intoshellina sicyodes Georgevitch, 1943
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Limnodrilide

Intoshellina wautieri de Puytorac et Juget, 1964 illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Euilyodrilus heuscheri

Intoshellinidae Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryoidea nv. super-fam.
includes: Intoshellina; Monodontophrya; Spirobuetschliella

Iodamoeba spp.
McConnell, E. E.; et al., 1974, Onderstepoort J. Vet. Research, v. 41 (3), 97-168
pathological and parasitological survey of 100 free-ranging chacma baboons
Papio ursinus (large intestine): Kruger National Park, Transvaal

Iodamoeba butschli
Giboda, M.; Hoffman, F.; and Rimar, P., 1971, Ceskoslov. Pediat., v. 26 (2), 88-89
intestinal protozoal infections in children aged 1-15 years, clinical trials with metronidazole, results and parasites eliminated varied with age and parasite: Kosice, Czechoslovakia

Iodamoeba buetschlii
Markell, E. K.; and Quinn, P. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1139-1142
comparison of immediate polyvinyl alcohol fixation with delayed Schaudinn's fixation for demonstrating protozoa in feces, concluded that immediate fixation results in preservation of larger number of organisms and recommends routine use prior to transportation to laboratory

Iodamoeba buetschlii
Savanat, T.; et al., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (2), 149-154
determinations of total serum IgE levels in humans with amoebic liver abscess or other parasitic infections

Iodamoeba buetschlii
Vinayak, V. K.; and Sehgal, S. C., 1976, Indian J. Med. Research, v. 64 (9), 1347-1350
human helminthic and protozoan parasites, comparison of nigrosin-methylene blue diagnostic test with formol-ether method and direct examination

Iorella genus novum (type genus)
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Iorellidae familia nova
tod: Iorella wegoreki g. n., sp. n.

Iorella wegoreki g. n., sp. n. (tod), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Syns.: Bothriopsides histrio (Schneider) pro parte sensu Baudoin, 1961; Legeria agilis (Schneider) pro parte sensu Baudoin, 1961; Bothriopsides graphoderi Baudoin, 1961 pro parte
Dytiscus marginalis (intestine): Poznan, Poland

Iorellidae familia nova
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Eugregarinaria
includes: Iorella genus novum (type genus)

Isospora
Beier, T. V.; Siim, I. K.; and Hutchison, W. M., 1977, Tsitologiia, v. 19 (12), 1362-1368
Eimeriidae, Isosporinae

Isospora
Duszynski, D. W.; and Speer, C. A., 1976, Ztschr. Parasitenk., v. 48 (3-4), 191-197
Isospora arctopitheci, I. bigemina, sporozoite excystation, lack of Stieda body, comparison with other Isospora, Eimeria and Sarcocystis species

Isospora Schneider 1881
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Eimeriidae
diagnosis

Isospora Schneider 1881
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Eimeriidae
diagnosis

Isospora
Loeffler, K., 1974, Prakt. Tierarzt, v. 55, Sondernummer, 78-80
cat diseases, prophylaxis and therapy, brief review

Isospora
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Isospora
Markus, M. B., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 535 [Letter]
human malaria, support for hypothesis that sporozoites remain dormant after entering host body and are mechanisms responsible for relapse phenomenon and extended prepatent period; likened to similar behavior of some Isospora spp.

Isospora
Romero-Rodriguez, J., 1975, Rev. Iber. Parasitol., v. 35 (1-2), 113-116
Isospora schwetzi, description, sporulation, differentiation of species of Isospora found in genus Corvus

Isospora
Speer, C. A.; and Duszynski, D. W., 1974, J. Protozool., v. 21 (3), 424 [Abstract]
relationship between sporocyst structure and process of excystation in coccidia

Isospora Schneider 1881 gen. emend.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
Eimeriidae fam. emend., Eimeriinae subfam. emend.
definition, key, key to species completing their sexual cycle in cats and dogs and forming tissue stages in intermediate hosts

Isospora
Werner, H.; and Janitschke, K., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 57-64
Toxoplasma gondii, morphological variations in developmental stages, details of sexual forms and oocysts, life cycle completed only in domestic cat, partial common antigenicity demonstrated between Toxoplasma and Isospora

Isospora sp.
Bejsovec, J., 1973, J. Protozool., v. 20 (4), 535
Phasianus colchicus: Central Bohemia

Isospora sp.
Box, E. M., 1973, J. Protozool., v. 20 (4), 510
comparative development of Atoxoplasma and Isospora species in canaries

Isospora sp.
Duszynski, D. W.; and File, S. K., 1973, J. Protozool., v. 20 (4), 510
in vitro excystation
Callimico goeldii (feces): Tulane University Delta Primate Center, Covington

Isospora sp.
Faust, B. S.; and Pappas, P. W., 1977, J. Zoo Animal Med., v. 8 (1), 18-23
Chen caerulescense caerulescense
Rhea americana
Anas platyrhynchos
Gallus gallus
Haliaeetus leucocephalus
Passer domesticus
(feces of all): all from Columbus (Ohio) Zoo

Isospora sp. (resembling I. lacazei)
Fisher, J. W.; and Wacha, R. S., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 226-227
Phasianus colchicus (feces): private game farm, Jasper County, Iowa

Isospora sp.
Joseph, T., 1974, J. Protozool., v. 21 (1), 12-15
Didelphis virginiana (feces from rectum, intestine): Northern Indiana, U.S.A.

Isospora sp.
Joseph, T., 1974, Proc. Indiana Acad. Sc., v. 83, 1973, 467 [Abstract]
Didelphis virginiana: St. Joseph's, Elkhart, La Porte and Marshall Counties, Indiana

Isospora [sp.], illus.
Kiurtov, N., 1975, Vet. Med. Nauk., v. 12 (8), 60-66
coccidia, incidence, localization and pathogenicity in calves: Veliko Turnovo Okrug, Bulgaria

Isospora sp. n. Trayser and Todd (in preparation)
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Canis familiaris

Isospora sp.
Markus, M. B., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 7-8 [Demonstration]
Isospora sp. in Cynictis penicillata (epithelial cells of small intestine), developmental stages, gametogony: western Transvaal, South Africa

Isospora sp., illus.
Ovezmukhammedov, A.; and Berdyev, A. S., 1975, Izvest. Akad. Nauk Turkmensk. SSR, s. Biol. Nauk (6), 55-59
brief description
Felis margarita: Turkmenistan

Isospora spp.
Pavlasek, I., 1975, J. Protozool., v. 22 (3), 68A [Abstract]
coccidiosis, calves, incidence in relation to age

Isospora A
Peters, W.; Tennant, L.; and McDermott, S. N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 8-10 [Demonstration]
lions: Knowsley Safari Park, Prescot, Lancashire

Isospora B
Peters, W.; Tennant, L.; and McDermott, S. N., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 8-10 [Demonstration]
lions: Knowsley Safari Park, Prescot, Lancashire

Isospora [sp.]
Powell, E. C.; and McCarley, J. B., 1975, J. Parasitol., v. 61 (5), 928-931
murine sarcocysts fed to cats produced isosporan infection, resulting oocysts fed to mice produced sarcocysts, sarcocysts from exper.-infected mice fed to other cats again produced Isospora felis-like infection

Isospora [sp.]
Tharaldsen, J., 1973, Norwegian J. Zool., v. 21 (4), 327-328 [Abstract]
dogs (feces): quarantine station, Oslo, Norway

Isosporan oocysts
Wacha, R. S.; and Christiansen, J. L., 1975, J. Protozool., v. 22 (3), 46A [Abstract]
Agkistrodon contortrix
Arizona elegans
Coluber constrictor
Pituophis melanoleucus
Sistrurus catenatus
all from North America

Isospora anatolicum sp. n., illus.
Sayin, F.; Dincer, S.; and Meric, I., 1977, J. Protozool., v. 24 (2), 210-212
Spalax leucodon (intestine): Lalahan district, Ankara, Turkey

Isospora arctopitheci Rodhain, 1933, illus.
Duszynski, D. W.; and Speer, C. A., 1976, Ztschr. Parasitenk., v. 48 (3-4), 191-197
Isospora arctopitheci, I. bigemina, sporozoite excystation, lack of Stieda body, comparison with other Isospora, Eimeria and Sarcocystis species
Saguinus geoffroyi

Isospora arctopitheci Rodhain, 1933, illus.
Hendricks, L. D., 1974, Proc. Helminth. Soc. Washington, v. 41 (2), 229-233
redescription
Cebus capucinus (nat. and exper.)
Saguinus geoffroyi (nat. and exper.)
all from Republic of Panama

Isospora arctopitheci Rodhain 1933
Hendricks, L. D., 1977, J. Parasitol., v. 63 (1), 32-35
Aotus trivirgatus
Ateles fusciceps
Cebus capucinus
Alouatta villosa
Saimiri sciureus
Saguinus geoffroyi
Canis familiaris
Nasua nasua
Potos flavus
Eira barbara
Felis catus
Didelphis marsupialis
(all exper.)

Isospora belli
Campos, R.; Amato Neto, V.; and Campos, L. L., 1969, Bol. Chileno Parasitol., v. 24 (3-4), 127-129
children in orphanage, clinical aspects of infection: Sao Paulo

Isospora belli
Giboda, M., 1973, J. Protozool., v. 20 (4), 536
human children: Czechoslovakia

Isospora belli, illus.
Janitschke, K.; Palme, G.; and Ziegler, U., 1976, Med. Welt., v. 27 (19), 927-929
Isospora belli oocysts discovered in feces of man ill 10 months with severe diarrhea and weight loss: Germany

Isospora belli, illus.
Misic, S., 1973, Acta Parasitol. Iugoslavica, v. 4 (1), 37-41
Isospora belli, human (stool), Janeckso-Urbanyi flotation technique found most suitable for fecal examination; first record: Yugoslavia

Isospora belli
Pietrushka, D., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
Isospora belli, various stages, contents of glycogen and fats

Isospora belli
Pietrushka, D.; et al., 1974, J. Protozool., v. 21 (3), 465 [Abstract]
Isospora belli causing severe coccidiosis in a 51-year-old woman, partial remission with relapse obtained with Entobex, radical cure with Septrin: Israel

Isospora belli
Whitehead, R., 1973, Major Problems Path., v. 3, 105-110
human intestinal infection, diagnosis, pathological appearance of mucosal biopsy of gastrointestinal tract

Isospora belli, illus.
Zaman, V., 1968, Med. J. Malaya, v. 22 (3), 230
first reported cases of human Isospora belli discovered during stool survey in Singapore

Isospora besnoiti (Marotel, 1912) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syns.: Sarcocystis besnoiti Marotel, 1912; Globidium besnoiti Wenyon, 1926; Besnoitia besnoiti Henry, 1913

Isospora bigemina (Stiles)
Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
as syn. of Sarcocystis tenella (Railliet, 1886)

Isospora bigemina
Birchall, L.; Brown, G.; and Hansman, D., 1972, Med. J. Australia, v. 1 (24), 1269
life cycle of Toxoplasma gondii and differential morphology and diagnosis from Isospora spp.

Isospora bigemina small race
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
as syn. of Isospora wallacei n. sp.

Isospora bigemina, illus.
Dubey, J. P.; and Fayer, R., 1976, Parasitology, v. 73 (3), 371-380
Isospora bigemina "small race", life cycle in dogs (nat. and exper.) which can serve as both definitive and intermediate host, sporulated oocysts from dog not infectious to cattle, cats, or mice but structurally identical oocysts were shed by dogs after ingesting hearts and diaphragms from naturally infected cattle

Isospora bigemina (Stiles, 1891) Luhe, 1906, illus.
Duszynski, D. W.; and Speer, C. A., 1976, Ztschr. Parasitenk., v. 48 (3-4), 191-197
Isospora arctopitheci, I. bigemina, sporozoite excystation, lack of Stieda body, comparison with other Isospora, Eimeria and Sarcocystis species
Lynx rufus

Isospora bigemina
Fameree, L.; and Cotteleer, C., 1976, J. Protozool., v. 23 (4), 10A [Abstract]
coccidiosis, cats, prevalence: Belgium

Isospora bigemina, Stiles 1891, Luehe 1906, pro parte
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
as syn. of Sarcocystis suihominis n. spec.

Isospora bigemina, Stiles, 1891; Luhe 1906, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovicanis n. spec.

Isospora bigemina, Stiles 1891, Luhe 1906, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovifelis n. spec.

Isospora bigemina, Stiles, 1891, Luhe, 1906, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis ovicanis n. spec.

Isospora bigemina, Stiles, 1891; Luhe, 1906, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis ovifelis n. spec.

Isospora bigemina, illus.
Heydorn, A. O.; Gestrich, R.; and Ipczynski, V., 1975, Berl. u. Munchen. Tierarztl. Wchnschr., v. 88 (23), 449-453
small race of Isospora bigemina, life cycle, developmental stages, intestine of dog

Isospora bigemina, illus.
Heydorn, A. O.; Mehlhorn, H.; and Gestrich, R., 1975, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 233 (1), 123-137
Sarcocystis fusiformis, calves (exper.), infected with oocysts and sporocysts of large form of Isospora bigemina from dogs, fine structure of cyst stages

Isospora bigemina
Ito, S.; et al., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 229-234
Toxoplasma, strains from cats infected with Isospora bigemina, oocysticidal effect of disinfectants, alcohols and other chemicals, only Lomasept, Neo Kurehasol, ethanol, methanol, peracetic acid, ammonia water, ammonium sulfide, and formalin were effective

Isospora bigemina Swellengrebel 1914
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora felis Wenyon 1923

Isospora bigemina (Stiles 1891) Luhe 1906 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis bigemina (Stiles 1891) comb. n.

Isospora bigemina (Stiles 1891) Luhe 1906 of Levine and Ivens (1965) (non) Isospora bigemina (Stiles 1891) Luhe 1906
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora heydorni Tadros and Laarman, 1976

Isospora bigemina "'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) and Heydorn, Mehlhorn, and Gestrich (1975) pro parte"
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis miescheriana (Kuhn 1865) Labbe 1899

Isospora bigemina "'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) and Heydorn, Mehlhorn, and Gestrich (1975) pro parte"
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Isospora bigemina "'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) and Heydorn, Mehlhorn, and Gestrich (1975) pro parte"
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis ovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975

Isospora bigemina "'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) and Heydorn, Mehlhorn, and Gestrich (1975) pro parte"
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis bertrami Doflein 1901

Isospora bigemina of Golemansky and Ridzhakov (1975) in red fox
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    perhaps as syn. of Isospora ohioensis Dubey 1975

Isospora bigemina
  Lodal, J., 1973, Norwegian J. Zool., v. 21 (4), 326 [Abstract]
    survey for Toxoplasma antibodies by Sabin-Feldman dye-test; oocysts of T. gondii and Isospora bigemina are morphologically identical (of 2 cats infected with I. bigemina oocysts only 1 was positive for Toxoplasma antibodies)
    Felis domestica (colon, rectum): Copenhagen area, Denmark

Isospora bigemina
  Loeffler, K., 1974, Prakt. Tierarzt, v. 55, Sondernummer, 68-72
    parasites, possible transmission from small domestic animals to man, brief review

Isospora bigemina
  Mehlhorn, H.; Heydorn, O.; and Gestrich, R., 1975, J. Protozool., v. 22 (3), 83A [Abstract]
    calves infected with Isospora hominis sporocysts or I. bigemina large forms from dog or cat, development of Sarcocystis fusiformis cysts in musculature, each of 3 species of Isospora forms characteristic cysts, "Ainsi, il semble prouve que les kystes de Sarcocystis fusiformis Railliet, 1897 font partie des cycles de developpement d'au moins trois especes differentes de Coccidies."

Isospora bigemina, illus.
  Rakovec, R.; and Senk, L., 1975, Zborn. Bioteh. Fak. Univ. Ljubljani, v. 12 (1), 165-171
    Isospora bigemina, sporulated oocysts from cats infecting mice, resulting in cysts in brain similar to Toxoplasma, structure; negative results in reinfecting other mice with brain cysts; negative Sabin-Feldman test results in all mice

Isospora bigemina
  Sreemannarayana, O., 1976, Indian Vet. J., v. 53 (11), 883-894
    Isospora bigemina, dogs, Quixalin, case reports

Isospora bigemina (Stiles, 1891) Luhe, 1906 [pro parte]
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora bertrami Doflein, 1901 comb. nov.

Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora cruzi (Hasselmann, 1926) comb. nov.

Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora hirsuta (Moule, 1887) comb. nov.

Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora miescheriana (Kuhn, 1865) comb. nov.

Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora muris (Blanchard, 1885) comb. nov.

Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora ovicanis Heydorn, Gestrich, Mehlhorn & Rommel, 1975 [comb. nov.]

Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora tenella (Railliet, 1886) comb. nov.

Isospora bigemina Swellengrebel, 1914
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Isospora felis (Wasielewski, 1904) Wenyon, 1923

Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte: the so-called small form of I. bigemina of dogs Wenyon, 1926 [et auct.]
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Isospora heydorni nom. nov. [i.e., n. sp.]

Isospora bigemina
  Yang, J.; and Scholten, T., 1977, Am. J. Clin. Path., v. 67 (3), 300-304
    diagnosis of human intestinal parasites, fecal examination technique using Junod's fixative for concentration and permanent staining procedures, comparison with results using formalin-ether procedure

Isospora bigemina
  Zukovic, M.; et al., 1973, Acta Parasitol. Iugoslavica, v. 4 (1), 43-45
    Isospora bigemina, small form from dogs given to mice orally or peritoneally, single or repeated dose; toxoplasmosis did not develop; no identity with Toxoplasma gondii found

Isospora boughtoni
  Joseph, T., 1974, Proc. Indiana Acad. Sc., v. 83, 1973, 467 [Abstract]
    "resembled Isospora boughtoni"
    Didelphis virginiana: St. Joseph's, Elkhart, La Porte and Marshall Counties, Indiana

Isospora buriatica
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Vulpes fulva]
[Alopex lagopus]
all from Kazakhstan fur farms

Isospora buteonis Henry 1932
Ashford, R. W., 1975, J. Protozool., v. 22 (3), 48A [Abstract]
canary fed on feces from Accipiter nisus infected with sporocysts resembling Isospora buteonis was infected with young Sarcocystis cysts when examined 2 months later, postulated that raptor-passerine cycle may exist

Isospora buteonis
Cerna, Z., 1977, J. Protozool., v. 24 (2), Suppl., 14A [Abstract]
Tyto alba
Asio otus
Falco tinnunculus
Accipiter gentilis
Mus musculus (cysts in muscle, schizonts in liver) (exper.)
Microtus arvalis (cysts in muscle, schizonts in liver) (exper.)

Isospora buteonis
Cerna, Z.; and Senaud, J., 1976, J. Protozool., v. 23 (4), 6A-7A [Abstract]
Isospora buteonis oocysts caused sarcosporidian infection in mice (exper.), ultrastructure of asexual stages

Isospora buteonis Henry, 1932 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora buteonis (Henry, 1932) comb. nov.

Isospora canaria sp. n., illus.
Box, E. D., 1975, J. Protozool., v. 22 (2), 165-169
description of exogenous stages
Serinus canarius (intestinal epithelium) (nat. and exper.)

Isospora canaria Box, illus.
Box, E. D., 1977, J. Protozool., v. 24 (1), 57-67
life cycle, endogenous stages, periodicity of oocyst discharge, patency and pathogenicity
Serinus canarius (exper.) (epithelium of small intestine)

Isospora canaria Box, 1975, illus.
Speer, C. A.; and Duszynski, D. W., 1975, J. Protozool., v. 22 (4), 476-481
Isospora serini, I. canaria, oocyst walls, fine structure; I. serini, excystation

Isospora canis Nemeseri
Dubey, J. P., 1975, J. Protozool., v. 22 (3), 416-417
cats became infected with Isospora canis but did not shed oocysts, dogs became infected after consuming infected mice; cats and mice can act as intermediate hosts

Isospora canis, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1073
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Isospora canis
Heil, R., 1974, Hamatologische Untersuchungen an Saugetieren und Kuken nach experimentellen Kokzidien-Infektionen, 137 pp.
Canis familiaris, experimental infection, haemoglobin and haematocrit values, differential blood cell counts

Isospora canis Nemeseri, 1959, illus.
Lepp, D. L.; and Todd, K. S., jr., 1974, J. Protozool., v. 21 (2), 199-206
Isospora canis in dogs (exper.), description, endogenous cycle, localization in small intestine; failure to reinfect previously infected dogs

Isospora canis Nemeseri 1959
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syn.: Isospora felis Wenyon 1923 of auctores in the dog
Canis familiaris

Isospora canis
Levine, N. D.; and Cechner, P. E., 1973, J. Protozool., v. 20 (4), 510-511
Eimeria spp. and Isospora canis oocysts fed to pigs, no evidence that Sarcocystis infections resulted

Isospora canis
Levine, N. D.; Cechner, P. E.; and Meyer, R. C., 1974, Indian Vet. J., v. 51 (1), 57-59
pigs fed oocysts of porcine Eimeria spp. and of Isospora canis, "Neither Sarcocystis nor any other protozoon was found, except for Eimeria spp. in the intestinal epithelium of pigs that had been inoculated with porcine Eimeria spp."

Isospora canis
Loveless, R. M.; and Andersen, F. L., 1975, J. Parasitol., v. 61 (3), 546-547
Canis latrans (exper.)

Isospora canis Nemeseri, 1959
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity

Isospora canis
Yang, J.; and Scholten, T., 1977, Am. J. Clin. Path., v. 67 (3), 300-304
diagnosis of human intestinal parasites, fecal examination technique using Junod's fixative for concentration and permanent staining procedures, comparison with results using formalin-ether procedure

Isospora canivelocis
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Vulpes fulva]
[Alopex lagopus]
all from Kazakhstan fur farms

Isospora cati Marotel 1921
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora felis Wenyon 1923

Isospora cati Marotel, 1922 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora felis (Wasielewski, 1904) Wenyon, 1923

Isospora chloridis Anwar 1966, illus.
Romero Rodriguez, J., 1973, Rev. Iber. Parasitol., v. 33 (2-3), 235-242
Isospora lacazei, I. chloridis, morphology, sporulation, endogenous cycle
Serinus canarius (intestino delgado): Spain

Isospora choudari Bhatia et al., 1972
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Gallus sonneratii (feces): Lucknow Zoo

Isospora cnemidophori Carini, 1942
Arcay de Peraza, L.; and Bastardo de San Jose, T., 1975, Acta Biol. Venez., v. 9 (1), 77-91
Isospora cnemidophori in Cnemidophorus lemniscatus lemniscatus (nat. and exper.) and Ameiva ameiva (exper.), endogenous cycle of development in liver, redescription: Venezuela

Isospora communis passerum Sjobring, 1897 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora lacazei (Labbe, 1893)

Isospora communis passerum Sjobring, 1897 [pro parte]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Isospora dasguptai n. sp.
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
Syns.: Isospora rivolta (Grassi, 1879) of Knowles and Das Gupta (1931); small Isospora garnhami Bray, 1954 of Dubey and Pande (1963)
Herpestes auropunctatus
H. edwardsi
(feces of all): all from India

Isospora decipiens sp.n., illus.
Lainson, R.; and Shaw, J. J., 1973, J. Protozool., v. 20 (3), 358-362
Philodryas olfersi (feces, small intestine, subepithelial connective tissue of villi): Barcarena, Para State, north Brazil

Isospora endocallimici n. sp., illus.
Duszynski, D. W.; and File, S. K., 1974, Tr. Am. Micr. Soc., v. 93 (3), 403-408
Isospora endocallimici, sporozoites, excystation, oocyst structure, comparisons with other isosporans
Callimico goeldii (feces): Tulane University Delta Regional Primate Research Center (some imported from Peru)

Isospora endocallimici, illus.
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Isospora endocallimici, illus.
Speer, C. A.; et al., 1976, J. Parasitol., v. 62 (6), 984-987
Isospora endocallimici, sporocyst wall during excystation, ultrastructural changes

Isospora eversmanni
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Mustela]: Kazakhstan fur farms

Isospora felis (Wasielewski, 1904) Wenyon, 1923
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Panthera tigris (feces): Lucknow Zoo

Isospora felis
Birchall, L.; Brown, G.; and Hansman, D., 1972, Med. J. Australia, v. 1 (24), 1269
life cycle of Toxoplasma gondii and differential morphology and diagnosis from Isospora spp.

Isospora felis
Chessum, B. S., 1972, Brit. Vet. J., v. 128 (7), xxiii-xxxvi
Isospora felis, cats (exper.), reactivation of latent Toxoplasma gondii infection

Isospora felis (Wenyon)
Dubey, J. P., 1975, J. Protozool., v. 22 (3), 416-417
dogs became infected with Isospora felis but did not shed oocysts, cats became infected after consuming extra-intestinal organs of infected dogs; dogs can act as intermediate hosts

Isospora felis, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Isospora felis
Dubey, J. P., 1976, Nature, London (5565), v. 262, 213-214
Toxoplasma gondii, reshedding of oocysts by chronically infected cats in absence of exogenous reinfection, effect of superinfection with Isospora rivolta or I. felis, public health implications

Isospora felis
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (5), 541-543
attempted transmission of feline coccidia from chronically infected queens to their kittens, results indicate congenital transmission of coccidia unlikely

Isospora felis
Dubey, J. P.; and Streitel, R. H., 1976, Brit. Vet. J., v. 132 (6), 649-651
Isospora felis, I. rivolta, cats fed infected mouse tissue or oocysts, prepatent and patent periods, results indicate Isospora not a serious feline pathogen

Isospora felis
Fameree, L.; and Cotteleer, C., 1976, J. Protozool., v. 23 (4), 10A [Abstract]
coccidiosis, cats, prevalence: Belgium

Isospora felis
Gregory, G. G.; and Munday, B. L., 1976, Austral. Vet. J., v. 52 (7), 317-320
feral cats: Tasmanian Midlands

Isospora felis
Guterbock, W. M.; and Levine, N. D., 1977, J. Am. Vet. Med. Ass., v. 170 (12), 1411-1413
cats (feces): east central Illinois

Isospora felis Wenyon 1923
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Diplospora bigemina von Wasielewski 1904 pro parte; Isospora bigemina Swellengrebel 1914; I. rivolta Dobell and O'Connor 1921; I. cati Marotel 1921; Lucetina felis (Wenyon 1923) Henry and Leblois 1926
Felis catus

Isospora felis Wenyon 1923 of auctores in the dog
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora canis Nemeseri 1959

Isospora felis
Lodal, J., 1973, Norwegian J. Zool., v. 21 (4), 326 [Abstract]
Felis domestica (colon, rectum): Copenhagen area, Denmark

Isospora felis, illus.
Mehlhorn, H.; and Markus, M. B., 1976, Ztschr. Parasitenk., v. 51 (1), 15-24
Isospora felis, mouse, stages in mesenteric lymph nodes, parasitophorous vacuole in host cell, electron microscopy; considered to be waiting stage

Isospora felis Wenyon, 1923, illus.
Ovezmukhammedov, A.; and Berdyev, A. S., 1975, Izvest. Akad. Nauk Turkmensk. SSR, s. Biol. Nauk (6), 55-59
brief description
Felis catus dom.: Turkmenistan

Isospora felis-like
Powell, E. C.; and McCarley, J. B., 1975, J. Parasitol., v. 61 (5), 928-931
murine sarcocysts fed to cats produced isosporan infection, resulting oocysts fed to mice produced sarcocysts, sarcocysts from exper.-infected mice fed to other cats again produced Isospora felis-like infection

Isospora felis, illus.
Rasin, K., 1973, Vet. Med., Praha, v. 46, v. 18 (10), 593-618
Toxoplasma gondii (14 strains), Isospora felis, cats, natural and exper. infections, chronology of oocyst elimination; mixed infections; no cross immunity

Isospora felis
Rifaat, M. A.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (3), 67-70
seroepidemiolgoic survey of stray cats for Toxoplasma gondii antibodies and role in epidemiology of human infection, no cross-immunity with Isospora spp. in relationship study, high percentage of cats surveyed shedding oocysts in feces: Cairo, Egypt

Isospora felis (Wasielewski, 1904) Wenyon, 1923
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
synonymy, hosts, morphology, life cycle, pathogenicity, key

Isospora felis
Tharaldsen, J., 1973, Norwegian J. Zool., v. 21 (4), 327-328 [Abstract]
cats (feces): quarantine station, Oslo, Norway

Isospora felis
Tsang, C. L.; and Lee, Y. C., 1975, J. Chinese Soc. Vet. Sc., v. 1 (1), 37-41
Isospora felis, puppies, preventive and therapeutic effects of immune serum from adult dogs

Isospora felis
Wilkinson, G. T., 1977, Vet. Rec., v. 100 (8), 156-157
Isospora felis, cats, good results using sulphadimethoxine, no effect on number of oocysts in faeces of one cat treated with sulphadimidine; results indicate need to keep closed cat colony closed for disease prevention

Isospora garnhami Bray, 1954
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
description, review

Isospora garnhami Bray, 1954 of Dubey and Pande (1963), small form
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
as syn. of Isospora dasguptai n. sp.

Isospora garnhami Bray, 1954 of Dubey and Pande (1963), large form
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
as syn. of Isospora mungoi n. sp.

Isospora gondii (Nicolle & Manceaux, 1908) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
hosts, morphology, life cycle, pathogenicity, key
Syns.: Leishmania gondii Nicolle & Manceaux, 1908; Toxoplasma gondii Nicolle & Manceaux, 1909; T. canis Mello, 1910; T. cuniculi Splendore, 1910; T. musculi Sangiorgi, 1913; T. pyrogenes Castellani, 1913; T. caviae Carini & Migliano, 1916

Isospora graculai n. sp., illus.
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Gracula religiosa (feces): Delhi Zoo

Isospora hammondi (Frenkel, 1974) comb. nov.
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syn.: Hammondia hammondi Frenkel, 1974

Isospora herpestei n. sp., illus.
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
Herpestes auropunctatus (feces): imported from India, to Virgin Islands, thence to Georgia

Isospora heydorni nom. nov. [i.e., n. sp.]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, live cycle, pathogenicity
Syn.: Isospora bigemina (Stiles, 1891) Luhe, 1906 pro parte: the so-called small form of I. bigemina of dogs Wenyon, 1926 [et auct.]

Isospora heydorni Tadros and Laarman, 1976
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
as syn. of Hammondia heydorni (Tadros and Laarman, 1976) [n. comb.]
"Tadros and Laarman also proposed 2 new organisms: Endorimospora (Sarcocystis) suihominis from the pig and Isospora heydorni from the dog. I named them Sarcocystis porcihominis and Isospora wallacei respectively. Because Tadros and Laarman's paper was published a month earlier than mine, the names proposed by Tadros and Laarman have priority over Sarcocystis porcihominis and Isospora wallacei."

Isospora heydorni Tadros and Laarman, 1976
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syn.: Isospora bigemina (Stiles 1891) Luhe 1906 of Levine and Ivens (1965) (non) Isospora bigemina (Stiles 1891) Luhe 1906
Canis familiaris
Vulpes vulpes

Isospora hoarei Bray, 1954
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
description, review

Isospora hominis, illus.
Aryeetey, M. E.; and Piekarski, G., 1976, Ztschr. Parasitenk., v. 50 (2), 109-124
Sarcocystis fusiformis-infected raw meat fed to humans, Isospora hominis sporocysts excreted in feces; no immunity to reinfection manifested as reduced number of sporocysts; antibodies detected by indirect immunofluorescence test, also present in survey of population (common in adults, present in children over 6 months of age); rats fed infected sausage showed negative reaction to Sarcocystis antigen

Isospora hominis
Dymowska, Z.; et al., 1974, Med. Dosw. i Mikrobiol., v. 26 (3), 247-250
Toxoplasma gondii antibody levels determined by immunofluorescence in persons with Isospora hominis infection, no essential correlation found

Isospora hominis, Railliet und Lucet, 1891, Wenyon 1923, pro parte
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
as syn. of Sarcocystis suihominis n. spec.

Isospora hominis, Railliet, Lucet, 1891, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovihominis n. spec.

Isospora hominis (Railliet and Lucet 1891) Wenyon 1923 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hominis (Railliet and Lucet 1891) comb. n.

Isospora hominis
Mehlhorn, H.; Heydorn, O.; and Gestrich, R., 1975, J. Protozool., v. 22 (3), 83A [Abstract]
calves infected with Isospora hominis sporocysts or I. bigemina large forms from dog or cat, development of Sarcocystis fusiformis cysts in musculature, each of 3 species of Isospora forms characteristic cysts, "Ainsi, il semble prouve que les kystes de Sarcocystis fusiformis Railliet, 1897 font partie des cycles de developpement d'au moins trois especes differentes de Coccidies."

Isospora hominis (Railliet & Lucet, 1891) Wenyon, 1923 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hominis (Railliet & Lucet, 1891) comb. nov.

Isospora hominis (Railliet & Lucet, 1891) Wenyon, 1923 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora suihominis nom. nov. [i.e., n. sp.]

Isospora hominis, illus.
Vasallo Matilla, F., 1971, Med. Trop., Madrid, v. 47 (1), 37-47
morphology, life cycle, first reports of human infection with Isospora hominis recorded in Spain

Isospora hylomysis n. sp., illus.
Colley, F. C.; and Mullin, S. W., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 289-291
Hylomys suillus (intestinal contents): Janda Baik, Bentong, Pahang, Malaysia

Isospora ichneumonis n. sp.
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
Syn.: Isospora rivolta (Grassi, 1879) of Balozet (1933)
Herpestes ichneumon (feces): Tunisia

Isospora knowlesi Dubey and Pande, 1963, non Isospora knowlesi Ray and Das Gupta, 1937
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
as syn. of Isospora pellerdyi Dubey and Pande, 1964

Isospora koreani Bhatia et al., 1972
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Phasianus colchicus
P. versicolor
(feces of all): all from Delhi Zoo

Isospora lacazei Labbe, 1893
Anpilogova, N. V., 1970, Parazitologiia, Leningrad, v. 4 (3), 288-295
cross-infection experiments between sparrows and chickens indicate that Isospora oocysts found in feces of chickens (and possibly other poultry) are oocysts of I. lacazei ingested with feed or water contaminated by Passer montanus pallidus and not a true infection: Tadzhikistan poultry farms

Isospora lacazei, illus.
Cerna, Z., 1974, J. Protozool., v. 21 (4), 481-482
Isospora lacazei oocysts, anomalous sporulation, resemblance to Caryospora
Passer domesticus domesticus: Prague, Czechoslovakia

Isospora lacazei (Labbe), illus.
Mandal, A. K., 1977, Indian J. Animal Health, v. 16 (1), 5-8
Isospora lacazei, Eimeria labbeana, distribution patterns of DNA and RNA in various stages of life cycle

Isospora lacazei Labbe 1898, illus.
Romero Rodriguez, J., 1973, Rev. Iber. Parasitol., v. 33 (2-3), 235-242
Isospora lacazei, I. chloridis, morphology, sporulation, endogenous cycle
Serinus canarius (intestino delgado): Spain

Isospora lacazei, illus.
Sperhake, P.; and Kaleta, E. F., 1974, Prakt. Tierarzt, v. 55, Sondernummer, 29-31
Isospora lacazei, canaries, clinical aspects, pathology, diagnosis, differential diagnosis, therapy with Amprolvet Super in drinking water

Isospora lacazei (Labbe, 1893)
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
synonymy, hosts, morphology, life cycle, pathogenicity

Isospora lacazei (Labbe, 1893) pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Isospora laidlawi
Foreyt, W. J.; and Todd, A. C., 1976, J. Parasitol., v. 62 (3), 496
prevalence survey in domestic mink: mink ranches, Wisconsin

Isospora laidlawi
Foreyt, W. J.; Todd, A. C.; and Hartsough, G. R., 1977, Am. J. Vet. Research, v. 38 (3), 391-394
Isospora laidlawi, Eimeria vison and Eimeria sp. in Mustela vison (exper.), anticoccidial activity of eight compounds

Isospora laidlawi
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Mustela]: Kazakhstan fur farms

Isospora lyruri
Bejsovec, J., 1977, J. Protozool., v. 24 (4), 48A [Abstract]
persistence of Coccidia despite changing land use and decreasing numbers of host animals
Lyrurus tetrix: Bohemia

Isospora melopsittacusi n. sp., illus.
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Melopsittacus undulatus (feces): Delhi Zoo

Isospora mungoi n. sp.
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
Syn.: large Isospora garnhami Bray, 1954 of Dubey and Pande (1963)
Herpestes edwardsi (feces): India

Isospora novocati
Dubey, J. P., 1975, J. Parasitol., v. 61 (3), 462-465
"Pellerdy (1974) proposed a new name, I. novocati, for I. rivolta of the cat on the assumption that I. rivolta was discovered first in the dog by Grassi (1879). Wenyon (1923), who reviewed all reports of I. rivolta, concluded that I. rivolta was first discovered in the cat and later in the dog. I agree with Wenyon. Therefore, I. novocati becomes a nomen nudum in favor of I. rivolta. The new name, I. ohioensis, is proposed for the dog form and the name I. rivolta is retained for cats."

Isospora novocati Pellerdy 1974
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora rivolta (Grassi 1879) Wenyon 1923

Isospora ohioensis sp. n., illus.
Dubey, J. P., 1975, J. Parasitol., v. 61 (3), 462-465
Isospora ohioensis sp. n., proposed for I. rivolta-like oocysts from dog feces on basis of cross-infection experiments between canine and feline isolates of I. rivolta, I. rivolta retained for cat species
dogs (nat. and exper.): Columbus, Ohio
mice (exper.)

Isospora ohioensis, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Isospora ohioensis Dubey 1975
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Isospora rivolta (Grassi 1879) Wenyon 1923 of auctores in the dog; perhaps Isospora bigemina of Golemansky and Ridzhakov (1975) in red fox
Canis familiaris
Vulpes vulpes

Isospora ohioensis Dubey, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syn.: Isospora rivolta (Grassi, 1879) Wenyon, 1923 pro parte

Isospora papionis McConnell, A.J. de Vôs, Basson & V. de Vos, 1971, illus.
McConnell, E. E.; et al., 1972, Onderstepoort J. Vet. Research, v. 39 (2), 113-115
Isospora papionis, numerous oocysts found in skeletal muscles of Papio ursinus, possible importance of this localization in regard to parasite life cycle
Papio ursinus (longissimus dorsi, quadriceps, masseter, small intestine, faeces): Kruger National Park, Republic of South Africa

Isospora papionis
McConnell, E. E.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (6), 851-855
Papio ursinus (intestine, skeletal muscle); Kruger National Park, Republiç of South Africa

Isospora papionis, illus.
McConnell, E. E.; et al., 1974, Onderstepoort J. Vet. Research, v. 41 (3), 97-168
pathological and parasitological survey of 100 free-ranging chacma baboons
Papio ursinus (small intestine, skeletal muscle): Kruger National Park, Transvaal

Isospora passerum Sjobring, 1897 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora lacazei (Labbe, 1893)

Isospora passerum Sjobring, 1897 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Isospora pavlodarica
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Vulpes fulva]
[Alopex lagopus]
all from Kazakhstan fur farms

Isospora pellerdyi Dubey and Pande, 1964
Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
Syn.: Isospora knowlesi Dubey and Pande, 1963, non Isospora knowlesi Ray and Das Gupta, 1937

Isospora pellerdyi
Utebaeva, M. K., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 81-84
Pavo cristatus: Kazakhstan

Isospora putorii Becker, 1934
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora putorii (Railliet & Lucet, 1891) comb. nov.

Isospora pycnonotusi n. sp., illus.
Bhatia, B. B.; et al., 1974, Indian J. Animal Sc., v. 43 (10), 1973, 944-947
Pycnonotus jocosus (feces): Delhi Zoo

Isospora raggianai sp. n., illus.
Varghese, T., 1977, J. Parasitol., v. 63 (5), 887-889
Paradisaea raggiana (feces): Wau Highlands, Papua New Guinea

Isospora rivolta
Birchall, L.; Brown, G.; and Hansman, D., 1972, Med. J. Australia, v. 1 (24), 1269
life cycle of Toxoplasma gondii and differential morphology and diagnosis from Isospora spp.

Isospora rivolta (Grassi, 1879) Wenyon, 1923
  Dubey, J. P., 1975, J. Parasitol., v. 61 (3), 462-465
    Isospora ohioensis sp. n., proposed for I. rivolta-like oocysts from dog feces on basis of cross-infection experiments between canine and feline isolates of I. rivolta, I. rivolta retained for cat species
    cats (nat. and exper.): Kansas City, Kansas
    mice (exper.)
    dogs (exper., but did not shed oocysts)

Isospora rivolta, illus.
  Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
    Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Isospora rivolta
  Dubey, J. P., 1976, Nature, London (5565), v. 262, 213-214
    Toxoplasma gondii, reshedding of oocysts by chronically infected cats in absence of exogenous reinfection, effect of superinfection with Isospora rivolta or I. felis, public health implications

Isospora rivolta
  Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (5), 541-543
    attempted transmission of feline coccidia from chronically infected queens to their kittens, results indicate congenital transmission of coccidia unlikely

Isospora rivolta
  Dubey, J. P.; and Streitel, R. H., 1976, Brit. Vet. J., v. 132 (6), 649-651
    Isospora felis, I. rivolta, cats fed infected mouse tissue or oocysts, prepatent and patent periods, results indicate Isospora not a serious feline pathogen

Isospora rivolta
  Fameree, L.; and Cotteleer, C., 1976, J. Protozool., v. 23 (4), 10A [Abstract]
    coccidiosis, cats, prevalence: Belgium

Isospora rivolta
  Gregory, G. G.; and Munday, B. L., 1976, Austral. Vet. J., v. 52 (7), 317-320
    feral cats: Tasmanian Midlands

Isospora rivolta
  Guterbock, W. M.; and Levine, N. D., 1977, J. Am. Vet. Med. Ass., v. 170 (12), 1411-1413
    cats (feces): east central Illinois

Isospora rivolta (Grassi 1879) Wenyon 1923
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    Syns.: Coccidium rivolta Grassi 1879; Diplospora bigemina von Wasielewski 1904 pro parte; Isospora rivoltae Dobell 1919; Lucetina rivolta (Grassi 1879) Henry and Leblois 1926; Isospora novocati Pellerdy 1974
    Felis catus

Isospora rivolta Dobell and O'Connor 1921
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Isospora felis Wenyon 1923

Isospora rivolta (Grassi 1879) Wenyon 1923 of auctores in the dog
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Isospora ohioensis Dubey 1975

Isospora rivolta "'Isospora rivolta' sporocysts of Gassner (1940) and Levine and Ivens (1965) pro parte"
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Sarcocystis bertrami Doflein 1901

Isospora rivolta "'Isospora rivolta' sporocysts of Gassner (1940) and Levine and Ivens (1965) pro parte"
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Sarcocystis miescheriana (Kuhn 1865) Labbe 1899

Isospora rivolta "'free sporocysts of Isospora rivolta' of Gassner (1940) and Levine and Ivens (1965) pro parte"
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Isospora rivolta "'free sporocysts of Isospora rivolta' of Gassner (1940) and Levine and Ivens (1965) pro parte"
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Sarcocystis ovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975

Isospora rivolta (Grassi, 1879) of Knowles and Das Gupta (1931)
  Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
    as syn. of Isospora dasguptai n. sp.

Isospora rivolta (Grassi, 1879) of Balozet (1933)
  Levine, N. D.; Ivens, V.; and Healy, G. R., 1975, Proc. Oklahoma Acad. Sc., v. 55, 150-153
    as syn. of Isospora ichneumonis n. sp.

Isospora rivolta
  Lodal, J., 1973, Norwegian J. Zool., v. 21 (4), 326 [Abstract]
    Felis domestica (colon, rectum): Copenhagen area, Denmark

Isospora rivolta
  Loveless, R. M.; and Andersen, F. L., 1975, J. Parasitol., v. 61 (3), 546-547
    Canis latrans (exper.)

Isospora rivolta
Rifaat, M. A.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (3), 67-70
seroepidemiolgoic survey of stray cats for Toxoplasma gondii antibodies and role in epidemiology of human infection, no cross-immunity with Isospora spp. in relationship study, high percentage of cats surveyed shedding oocysts in feces: Cairo, Egypt

Isospora rivolta (Grassi, 1879) Wenyon, 1923
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
synonymy, hosts, morphology, life cycle, pathogenicity, key

Isospora rivolta (Grassi, 1879) Wenyon, 1923 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora ohioensis Dubey, 1975

Isospora rivolta
Thornton, J. E.; Bell, R. R.; and Reardon, M. J., 1974, J. Wildlife Dis., v. 10 (3), 232-236
Canis latrans (feces): Nueces County, Texas

Isospora rivolta
Yang, J.; and Scholten, T., 1977, Am. J. Clin. Path., v. 67 (3), 300-304
diagnosis of human intestinal parasites, fecal examination technique using Junod's fixative for concentration and permanent staining procedures, comparison with results using formalin-ether procedure

Isospora rivoltae Dobell 1919
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Isospora rivolta (Grassi 1879) Wenyon 1923

Isospora rivoltae Dobel, 1919
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora rivolta (Grassi, 1879) Wenyon, 1923

Isospora rivoltae (Grassi, 1879) Brumpt, 1922
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora rivolta (Grassi, 1879) Wenyon, 1923

Isospora schwetzi Yakimoff and Matschoulsky, 1939, illus.
Romero-Rodriguez, J., 1975, Rev. Iber. Parasitol., v. 35 (1-2), 113-116
Isospora schwetzi, description, sporulation, differentiation of species of Isospora found in genus Corvus
Corvus monedulae (heces fecales): Granada, Spain

Isospora serini (Aragao), illus.
Box, E. D., 1975, J. Protozool., v. 22 (2), 165-169
Isospora serini, description of exogenous stages, asexual stages in macrophages are indistinguishable from parasites previously called avian Toxoplasma, Atoxoplasma, and Lankesterella
Serinus canarius (mononuclear phagocytes, intestinal epithelium) (nat. and exper.)

Isospora serini
Box, E. M., 1975, J. Protozool., v. 22 (3), 28A [Abstract]
Isospora serini in Serinus canarius, explanation for chronic infection in contrast to usual self-limited coccidian life cycle

Isospora serini Aragao, illus.
Box, E. D., 1977, J. Protozool., v. 24 (1), 57-67
life cycle, endogenous stages, periodicity of oocyst discharge, patency and pathogenicity; forms resembling parasites called Atoxoplasma or Lankesterella were seen
Serinus canarius (exper.) (mononuclear phagocytes in intestinal loop, liver, spleen, lung; intestinal epithelium)

Isospora serini (Aragao, 1933), illus.
Speer, C. A.; and Duszynski, D. W., 1975, J. Protozool., v. 22 (4), 476-481
Isospora serini, I. canaria, oocyst walls, fine structure; I. serini, excystation

Isospora serini (Aragao, 1933) Box, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
synonymy, hosts, morphology, life cycle, pathogenicity

Isospora suis, illus.
Martinez-Gomez, F.; Hernandez-Rodriguez, G.; and Calero-Carretero, R., 1974, Rev. Iber. Parasitol., v. 34 (3-4), 171-176
pigs (feces): Cordoba (Spain)

Isospora suis
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Isospora suis
Onawunmi, O. A.; and Todd, A. C., 1976, Am. J. Vet. Research, v. 37 (6), 657-660
porcine coccidiosis (exper.), suppression and control with chlortetracycline combination, buquinolate, and lincomycin hydrochloride

Isospora suis
O'Neill, P. A.; and Parfitt, J. W., 1976, Vet. Rec., v. 98 (16), 321-323
Isospora suis, minimal disease pig herd, observations under non-experimental conditions, oocysts with sporoblasts recovered in infected pig feces 24 hours after collection, prepatent and patent periods, development of active acquired immunity following infection not affected by stress of farrowing

Isospora suis, illus.
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian J. Animal Sc., v. 46 (10), 567-570
coccidia, pigs (exper.), measurements of 4 types of mature schizonts recovered from single pig (small intestine)

Isospora suis
Upadhyay, A. N.; and Ahluwalia, S. S., 1977, Indian J. Animal Sc., v. 47 (9), 597-600
E[imeria] debliecki, E. neodebliecki, E. perminuta, Isospora suis, pigs (feces), sporulation time: Aligarh

Isospora suis (Biester, 1934), illus.
Yvore, P., 1976, Rev. Med. Vet., v. 152 (1), 25-32
pigs: France

Isospora thavari sp. n., illus.
Else, J. G.; and Colley, F. C., 1975, J. Protozool., v. 22 (4), 455-457
Gehyra mutilata (feces, epithelial cells of small intestine): Kuala Lumpur, Malaysia

Isospora triffitti
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Vulpes fulva]
[Alopex lagopus]
all from Kazakhstan fur farms

Isospora vulpina Nieschulz & Bos, 1933, illus.
Bledsoe, B., 1976, J. Protozool., v. 23 (3), 365-367
synonymy, diagnosis, transmission from fox to dog
Vulpes vulpes: Fromm Bros. Fox Farm, Hamburg, Wisconsin
Canis familiaris (exper.)

Isospora vulpina
Nukerbaeva, K. K.; and Svanbaev, S. K., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 92-95
[Vulpes fulva]
[Alopex lagopus]
all from Kazakhstan fur farms

Isospora vulpina Nieschulz and Bos, 1933, illus.
Ovezmukhammedov, A.; and Berdyev, A. S., 1975, Izvest. Akad. Nauk Turkmensk. SSR, s. Biol. Nauk (6), 55-59
brief description
Vulpes vulpes: Turkmenistan

Isospora vulpina Nieschulz & Bos, 1933
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
hosts, morphology, life cycle, pathogenicity

Isospora vulpina var. aprutina Mantovani, 1965
Bledsoe, B., 1976, J. Protozool., v. 23 (3), 365-367
as syn. of Isospora vulpina Nieschulz & Bos, 1933

Isospora vulpina var. vulpina Mantovani, 1965
Bledsoe, B., 1976, J. Protozool., v. 23 (3), 365-367
as syn. of Isospora vulpina Nieschulz & Bos, 1933

Isospora wallacei n. sp., illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Syn.: Isospora bigemina small race
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Isospora wallacei
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
"Tadros and Laarman [1976] also proposed 2 new organisms: Endorimospora (Sarcocystis) suihominis from the pig and Isospora heydorni from the dog. I [1976] named them Sarcocystis porcihominis and Isospora wallacei respectively. Because Tadros and Laarman's paper was published a month earlier than mine, the names proposed by Tadros and Laarman have priority over Sarcocystis porcihominis and Isospora wallacei."

Isospora wallacei nom. nov. [i.e., n. sp.]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key, hosts, morphology, life cycle, pathogenicity
Syn.: Besnoitia sp., Wallace & Frenkel, 1975

Isosporinae
Beier, T. V.; Siim, I. K.; and Hutchison, W. M., 1977, Tsitologiia, v. 19 (12), 1362-1368
Eimeriidae
includes: Toxoplasma; Isospora; Besnoitia; Hammondia; Sarcocystis; Frenkelia

Isosporosis
Markus, M. B., 1973, N. England J. Med., v. 289 (18), 980-981 [Letter]
human Toxoplasma gondii, no evidence for cross-reactions of routine serologic tests for toxoplasmosis with sarcosporidiosis and isosporosis

Jirovecella Lom, 1958
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Jirovecellinae n. s. fam.

Jirovecella hegemonis Lom, 1957, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Fridericia hegemon

Jirovecellinae n. s. fam.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae
includes: Jirovecella; Juxtamesnilelella; Akikodes; Delphyella

Juxtamesnilella de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Jirovecellinae n. s. fam.

Juxtamesnilella depressa (Rossolimo, 1926), illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Mesnilella depressa Rossolimo, 1926
Styloscolex sp.
Lamprodilus sp.
Styloscolex baikalensis

Juxtamesnilella deronis de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Dero digitata

Juxtamesnilella maritui (Cheissin 1930), illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Mesnilella maritui Cheissin, 1930
Styloscolex sp.
Teleuscolex korotneffi
Lamprodrilus sp.

Juxtamesnilella ocevskiji (de Puytorac, 1959)
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Juxtaradiophrya ocevskiji de Puytorac 1959
Rhynchelmis komarecki f. brevirostra

Juxtamesnilella ochridana (de Puytorac, 1959)
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Juxtaradiophrya ochridana de Puytorac 1959
Stylodrilus leucocephalus

Juxtamesnilella taupodrili (Kijenskìj, 1925) de Puytorac
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Rhyacodrilus coccineus
Tubifex tubifex

Juxtamesnilella variabilis (Rossolimo, 1926)
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Peloscolex inflatus
Lumbriculidae

Juxtamesnilella vermicularis (Cheissin, 1928), illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Styloscolex sp.
Teleuscolex korotneffi
Lamprodrilus sp

Juxtaradiophrya de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Juxtaradiophryinae n. s. fam.

Juxtaradiophrya elongata (Delphy, 1922) de Puytorac, 1957, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Pachydrilus lineatus
P. pagenstecheri

Juxtaradiophrya enchytraeoidei de Puytorac, 1955
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Enchytraeoides

Juxtaradiophrya kochi (Heidenreich) de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya elongata (Delphy, 1922) de Puytorac, 1957

Juxtaradiophrya multispiculata (Cheissin, 1930) de Puytorac, 1957, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Mesnilella multispiculata Cheissin, 1930
Lamprodrilus nigrescens

Juxtaradiophrya naisensis de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Nais obtusa

Juxtaradiophrya ocevskiji de Puytorac, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella ocevskiji (de Puytorac, 1959)

Juxtaradiophrya ochridana de Puytorac, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella ochridana (de Puytorac, 1959)

Juxtaradiophrya radiata (Cheissin, 1930) de Puytorac, 1957
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Mesnilella radiata, Cheissin, 1930
Clitellio korotneffi

Juxtaradiophrya rossolimoi (Cheissin, 1930)
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Radiophrya rossolimoi, Cheissin, 1930
Branchyura coccinea var. inequalis

Juxtaradiophrya styloscolecis (Heidenreich, 1935)
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Radiophrya prolifera Rossolimo, 1926
Styloscolex baikalensis

Juxtaradiophrya variabilis (Rossolimo, 1926) de Puytorac, 1957
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella variabilis (Rossolimo, 1926)

Juxtaradiophrya vermicularis (Chessin, 1928) de Puytorac, 1957
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella vermicularis (Cheissin, 1928)

Juxtaradiophryinae n. s. fam.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae
includes: Juxtaradiophrya; Radiophryoides; Buetschliellopsis

Kala azar. [See also Leishmaniasis]

Kala-azar
Ghose, A. C.; and Chowdhury, A. B., 1977, Indian J. Med. Research, v. 66 (4), 566-569
human malaria and kala-azar, immunoglobulin levels of persons with known infections compared with normal controls

Kala-azar
Woodruff, A. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 313-328
mechanisms involved in anemia associated with infection (schistosomiasis, kala-azar, malaria, trypanosomiasis) and splenomegaly in tropics, complement activation leading to hemolysis and splenomegaly due to erythrophagocytosis, review

Kalpidorhynchus Cunningham, 1907
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora Schneider, 1875

Kalpidorhynchus arenicolae Cunningham, 1907
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Gonospora arenicolae (Cunningham, 1907) Tregouboff, 1918

Karotomorpha
Brugerolle, G.; and Joyon, L., 1975, J. Protozool., v. 22 (3), 59A [Abstract]
Proteromonas, Karotomorpha, ultrastructural cytology

Karyolysus [sp.]
Svahn, K., 1976, Norwegian J. Zool., v. 24 (1), 1-6
Lacerta agilis (blood)

Karyolysus lacazei Labbe, 1894, illus.
Svahn, K., 1975, Norwegian J. Zool., v. 23 (4), 277-295
description, development
Lacerta agilis (nat. and exper.) (blood): Skillingaryd, Smaland; Tein, Bornholm; Melsted, Bornholm; west of Vordingborg, Zealand; Svanninge, Funen; Anholt
L. vivipara (nat. and exper.) (blood): Melsted, Bornholm
Sauronyssus saurarum (exper.)

Karyolysus lacertae
Kurashvili, B. E., 1975, Izvest. Akad. Nauk Gruzinsk. SSR, s. Biol., v. 1 (4), 317-320
antagonistic and synergetic interrelationships between intestinal parasites

Karyolysus lacertae Danilewsky, 1886, illus.
Svahn, K., 1975, Norwegian J. Zool., v. 23 (4), 277-295
synonymy, description, development
Lacerta agilis (nat. and exper.) (blood): Tein, Bornholm
Sauronyssus saurarum (exper.)

Karyolysus lacertarum Labbe, 1894
Svahn, K., 1975, Norwegian J. Zool., v. 23 (4), 277-295
as syn. of Karyolysus lacertae Danilewsky, 1886

Karyolysus latus sp. nov., illus.
Svahn, K., 1975, Norwegian J. Zool., v. 23 (4), 277-295
development
Lacerta agilis (nat. and exper.) (blood): Snogeholm, Scania; Maglehem, Scania; Degeberga, Scania; Hallamolla, Scania; Brosarp, Scania; Gilleleje, Denmark
L. vivipara (nat. and exper.) (blood): Degeberga, Scania
Sauronyssus saurarum (exper.)

Karyolysus minor sp. nov., illus.
Svahn, K., 1975, Norwegian J. Zool., v. 23 (4), 277-295
development
Lacerta agilis (nat. and exper.) (blood): Mols, Jutland; Svanninge, Funen; Degeberga, Scania

Kinetoplastida
Kallinikova, V. D., 1977, Molec. and Cell. Biochem., v. 14 (1-3), Feb. 4, 55-58
Kinetoplastida, kinetoplast genetic system, implications for concept of cell differentiation and phylogeny, relationship to phenotype of these flagellates

Kinetoplastida [sp.] "kinetoplast-containing biflagellate parasite"
Pokorny, K. S.; Ruggieri, G. D.; and Nigrelli, R. F., 1975, J. Protozool., v. 22 (3), 10A [Abstract]
fine structure
Cyclopterus lumpus (upper intestinal tract)

Klossia helicina
Fleischmann, B.; and Scholtyseck, E., 1976, Ztschr. Parasitenk., v. 50 (2), 193
Klossia helicina, motile stages, fine structure

Klossia pachyleparon n. sp., illus.
Colley, F. C.; and Else, J. G., [1976], Ann. Parasitol., v. 50 (6), 1975, 669-673
Varanus nebulosus (feces): Malaysia

Klossiella [sp.], illus.
Edgcomb, J. H.; Walker, D. H.; and Johnson, C. M., 1976, Vet. Path., v. 13 (4), 315-318
Didelphis marsupialis (kidneys): near Achiote, Panama

Klossiella sp. sporonts, illus.
Kusewitt, D. F.; Wagner, J. E.; and Harris, P. D., 1977, Vet. Parasitol., v. 3 (4), 365-369
Myotis sodalis (kidneys): Sequiota Cave, near Round Springs, south-central Missouri, U.S.A.

Klossiella cobaye, illus.
Dominiczak, K., 1975, Patol. Polska, v. 26 (2), 217-226
Klossiella muris in mice, K. cobaye in guinea pigs, morphology and development, little pathology observed

Klossiella equi
Brown, T. T., jr.; and Patton, C. S., 1977, J. Am. Vet. Med. Ass., v. 171 (6), 492 [Letter]
Klossiella equi, presence in horse urine and kidneys, similar organisms in nervous tissue, may be causative agent of equine protozoal encephalomyelitis

Klossiella equi, illus.
Lee, C. G.; and Ross, A. D., 1977, Austral. Vet. J., v. 53 (6), 287-288
description
horse (kidney): North Queensland, Australia

Klossiella killicki n. sp., illus.
Boulard, Y., 1975, Bull. Mus. National Hist. Nat., Paris, 3. s. (284), Zool. (194), 83-89
Microchiropter[a] (kidney): Sierra Leone

Klossiella muris, illus.
Brown, R. J.; Stafford, E. E.; and Sudomo, M., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (2), 274
Rattus exulans
Mus musculus
(kidney tubules of all): all from Sunar beach, Bali, Indonesia

Klossiella muris, illus.
Dominiczak, K., 1975, Patol. Polska, v. 26 (2), 217-226
Klossiella muris in mice, K. cobaye in guinea pigs, morphology and development, little pathology observed

Klossiella muris Smith and Johnson, 1902
Wilson, V. C. L. C.; and Edrissian, G. H., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 8 [Demonstration]
light microscopy of sporogonic stages (mother-sporocyst and sporozoites)
Mus musculus (kidney): Zagrass Mountains, Fars Province, Iran

Klossiella tejerai Scorza, 1957, illus.
Boulard, Y., 1975, Bull. Mus. National Hist. Nat., Paris, 3. s. (284), Zool. (194), 83-89
Klossiella tejerai, schizogony described, sporogony redescribed
Marmosa cinerea demararae: French Guyana

Kofoidina Henry, 1933
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Kofoidina ovata Henry, 1933
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Kudoa [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Mugil cephalus (gut wall): Elat, Gulf of Aqaba, Red Sea

Kudoa cerebralis sp. n., illus.
Paperna, I.; and Zwerner, D. E., 1974, J. Protozool., v. 21 (1), 15-19
host specificity, pathology, sex of host
Morone saxatilis (connective tissue of nervous system): York River (Cheatham Annex, and Gloucester Point, Virginia); Rappahannock River (Sharps, Virginia); Atlantic Ocean (littoral, near Chincoteague, Virginia)

Kudoa clupeidae (Hahn 1917)
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes elongatus (muscles)
S. paucispinis (muscles)
all from off southern California

Kudoa eleotrisi n. sp., illus.
Siau, Y., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 563-570
Eleotris kribensis (gills): Porto Novo (Dahomey)

Labyrinthula spp.
Li, M. F., 1976, Canad. Fed. Biol. Soc., Programme and Proc. 19. Ann. Meet., v. 19, 158 [Abstract]
Labyrinthula spp., possible causative agent of Malpeque disease of oysters, in vitro culture, oyster hemolymph (20%) as a supplement to medium appeared to enhance growth

Lainsonia
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243
definition

Lamblia
Bulucea, D., 1971, Pediatria, Bucuresti, v. 20 (2), 163-168
mixed ascariasis and lambliasis in young child with persistent eosinophilia which was not relieved by cure of parasitemia, clinical case report: Romania

Lamblia
Petrescu-Coman, V.; et al., 1970, Pediatria, Bucuresti, v. 19 (4), 313-324
lambliasis in children as cause of prolonged diarrhea, clinical and diagnostic aspects

Lamblia intestinalis
Dancescu, P.; et al., 1970, Maroc Med. (534), v. 50, 247-248
human infection, fecal examination, cyst differentiation using lugol solution, etiology of "blue cysts"

Lamblia intestinalis
Giboda, M.; Hoffman, F.; and Rimar, P., 1971, Ceskoslov. Pediat., v. 26 (2), 88-89
intestinal protozoal infections in children aged 1-15 years, clinical trials with metronidazole, results and parasites eliminated varied with age and parasite: Kosice, Czechoslovakia

Lamblia intestinalis
Hanczycowa, H.; Kociecka, W.; and Lubczynska-Kowalska, W., 1973, Polski Tygod. Lekar., v. 28 (47), 1864-1866
Lamblia intestinalis, taeniarhynchosis, significant rise in muramidase activity in serum and gastric juices of infected persons compared to normal controls, possible allergic manifestation

Lamblia intestinalis
Iwanowski, H.; and Herman, T., 1977, Pediat. Polska, v. 52 (6), 649-654
Ascaris lumbricoides in children, Soprunova urine test used successfully to diagnose ascariasis and differentiate from infections of Trichuris trichiura, Enterobius vermicularis and Lamblia intestinalis

Lamblia intestinalis
Jacobson, E.; Kubalska, J.; and Zytkiewicz, B., 1972, Pediat. Polska, v. 47 (6), 689-694
Lamblia intestinalis in children, D-xylose test to determine extent of intestinal malabsorption associated with infection

Lamblia intestinalis
Kvasz, L., 1971, Ceskoslov. Pediat., v. 26 (4), 183-184
Lamblia intestinalis, gastrointestinal X-ray examination of infected children revealed ulceration of duodenal mucosa and functional disorders and irritation of upper digestive tract, possible relationships to lambliasis, metronidazole therapy of parasites resulted in cure of ulceration

Lamblia intestinalis
Kvasz, L.; et al., 1973, Ceskoslov. Pediat., v. 28 (6), 326-327
Lamblia intestinalis in institutionalized children, intestopan therapy, recurrence of cysts in stool 4 weeks after treatment, reinfection from personnel not ruled out: Bratislava

Lamblia intestinalis
Obtulowicz, K., 1972, Polski Tygod. Lekar., v. 27 (8), 296-298
human intestinal parasites, possible role in etiology of asthma and other allergic conditions

Lamblia intestinalis, illus.
Pelikan, L.; Kod'ousek, R.; and Malinsky, J., 1972, Ceskoslov. Pediat., v. 27 (8), 378-380
Lamblia intestinalis in humans presenting with malabsorption syndrome, diagnosis using enterobiopsy, pathology, medical management: Czechoslovakia

Lamblia intestinalis
Rudobielska, M.; et al., 1971, Pediat. Polska, v. 46 (9), 1103-1110
significant reduction in serum vitamin B 12 levels in children infected with Lamblia intestinalis, no variation seen in children infected with Trichocephalus dispar when compared with normal controls

Lamblia intestinalis
Rudobielska, M.; et al., 1972, Polski Tygod. Lekar., v. 27 (42), 1629-1631
Lamblia intestinalis, changes in vitamin B_{12} serum levels of infected children, comparison with normal controls

Lamblia intestinalis
Shikuwa, K., 1968, Nettai Igaku (Trop. Med.), v. 10 (2), 81-94
human infection associated with Muneseki-Byo syndrome: Nagasaki

Lamblia intestinalis
Tuszkiewicz-Misztal, E., 1975, Pediat. Polska, v. 50 (4), 417-422
Lamblia intestinalis in children, changes in butyric dehydrogenase-lactic hydrogenase ratio useful in diagnosing lambliasis, possibly indicative of hepatocellular damage

Lambliasis. See Giardiasis.

Lankesterella
Box, E. D., 1975, J. Protozool., v. 22 (2), 165-169
Isospora serini, description of exogenous stages, asexual stages in macrophages are indistinguishable from parasites previously called avian Toxoplasma, Atoxoplasma, and Lankesterella

Lankesterella
Box, E. D., 1977, J. Protozool., v. 24 (1), 57-67
forms resembling parasites called Atoxoplasma or Lankesterella were seen in Serinus canarius infected with Isospora serini

Lankesterella
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243
definition

Lankesterella [sp.]
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Dendrocygna viduata
Zosterops senegalensis
Serinus atrogularis
all from Ethiopia

Lankesterella [sp.]
Bennett, G. F.; and Blancou, J., 1974, J. Wildlife Dis., v. 10 (3), 239-240
Acridotheres tristis (blood): Madagascar

Lankesterella [sp.]
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Tachyphonus delatrii (blood): Colombia

Lankesterella sp.
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Pycnonotus xanthopygus (blood): Tanzania

Lankesterella [sp.]
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
Prunella modularis (blood): Great Britain

Lankesterella spp.
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Rana cancrivora
R. erythraea
R. macrodon
all from Malayan Peninsula

Lankesterella adiei (Aragao, 1911) Levine, 1966 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora lacazei (Labbe, 1893)

Lankesterella adiei (Aragao, 1911) Levine, 1966 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Lankesterella alencari
Goncalves da Costa, S. C.; de Sousa, M. A.; and Weigl, D. R., 1975, J. Protozool., v. 22 (3), 36A [Abstract]
Lankesterella alencari, ultrastructure confirms that these forms are not Toxoplasma gondii

Lankesterella alencari da Costa & Pereira, 1971
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma alencari (da Costa & Pereira, 1971) Levine & Nye, 1976

Lankesterella alencari da Costa & Pereira, 1971
Levine, N. D.; and Nye, R. R., 1976, J. Protozool., v. 23 (4), 488-490
as syn. of Toxoplasma alencari (da Costa & Pereira, 1971) comb. nov.

Lankesterella butasturis (de Mello, 1935) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
Syn.: Toxoplasma butasturis de Mello, 1935

Lankesterella garnhami Lainson, 1959 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora lacazei (Labbe, 1893)

Lankesterella garnhami Lainson, 1959 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Lankesterella garnhami Lainson, 1959
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Passer domesticus]: north-east Azerbaidzhan

Lankesterella serini Lainson, 1959
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora serini (Aragao, 1933) Box, 1975

Lankesterellidae
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243
definition

Lankesteria Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Lankesteria [sp.], illus.
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Lutzomyia vexatrix occidentis (abdomen, hemocoel, accessory glands): central California

Lankesteria sp. Labbe, 1899 (?)
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria sp. Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria ormieresi n. sp.

Lankesteria sp. Swarczewsky, 1910
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella swarczewskyi Codreanu & Balcesco, 1971

Lankesteria sp. Vavra
Sanders, R. D.; and Poinar, G. O., jr., 1973, J. Protozool., v. 20 (5), 594-602
as syn. of Lankesteria clarki sp. n.

Lankesteria acutissima Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria amaroucii (Giard, 1873) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Lankesteria amaroucii var. magna Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
"Lankesteria amaroucii var. magna Ormieres, 1965. Emended to Lankesteria magna Ormieres, 1965."

Lankesteria arndti Valkanov, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella arndti (Valkanov, 1931) Valkanov, 1935

Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
synonymy

Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Lankesteria ascidiae (Lankester) of Harant (1931) non L. ascidiae (Lankester, 1872) Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria styelae Ormieres, 1965

Lankesteria ascidiellae Duboscq & Harant, 1923
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria barretti Vavra, 1969
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis barretti (Vavra, 1969) nov. comb.

Lankesteria botrylli Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria buetschlii (Mingazzini, 1891) Ormieres, 1965 emend.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Lankesteria butschlii (sic) (Mingazzini, 1891) Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria buetschlii (Mingazzini, 1891) Ormieres, 1965 emend.

Lankesteria clarki sp. n., illus.
Sanders, R. D.; and Poinar, G. O., jr., 1973, J. Protozool., v. 20 (5), 594-602
fine structure, life cycle, little potential as biological control agent
Syn.: Lankesteria sp. Vavra
Aedes sierrensis (midgut epithelial cells, Malpighian tubules)(nat. and exper.): California (Treten Canyon, Fresno County; Strawberry Canyon, University of California, Berkeley; Blodgett Forest, University of California Forestry Research Station, El Dorado County; Novato, Marin County)
Aedes aegypti (exper.)

Lankesteria clarki Sanders & Poinar, 1973
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis clarki (Sanders & Poinar, 1973) comb. nov.

Lankesteria clavellinae (von Kolliker, 1848) Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Lankesteria claviformis Geus, 1967 [proposed by Geus as new name for Monocystis neodendrocoeli Georgevitch, 1951]
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella neodendrocoeli (Georgevitch, 1951) de Puytorac and Grain, 1960

Lankesteria compacta Geus, 1967
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella compacta (Geus, 1967) comb. nov.

Lankesteria culicis (Ross, 1898) Wenyon, 1911
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis culicis (Ross, 1898) Grasse, 1953

Lankesteria culicis, illus.
Pillai, J. S.; Neill, H. J. C.; and Sone, P. F., 1976, Mosquito News, v. 36 (2), 150-154
Lankesteria culicis, infection rates according to habitats, histopathology, heavily infected mosquito larvae (Aedes polynesiensis) showed minimal or negligible effects from parasitism
Aedes polynesiensis
A. aegypti
(epithelial cells of gut, Malpighian tubules of all): all from Western Samoa

Lankesteria culicis
Sanders, R. D.; and Poinar, G. O., jr., 1973, J. Protozool., v. 20 (5), 594-602
Aedes sierrensis (exper.)
Aedes aegypti (exper.)

Lankesteria culicis (Ross)
Walsh, R. D.; and Olson, J. K., 1976, Mosquito News, v. 36 (2), 154-160
Lankesteria culicis, host susceptibility to infection, parasite mortality results from its inability to penetrate peritrophic membrane of mosquito to gain access to epithelial cells
Aedes aegypti
A. epactius
A. sollicitans
A. stimulans
A. vexans
Culiseta inornata
Psorophora columbiae
(all exper.)

Lankesteria cyclopori Poisson, 1921
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella cyclopori (Poisson, 1921) comb. nov.

Lankesteria diazonae (Mingazzini, 1891) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Cytomorpha diazonae Mingazzini, 1891

Lankesteria distapliae (Mingazzini, 1891) Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Lankesteria galliardi Garnham, 1973
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis galliardi (Garnham, 1973) comb. nov.

Lankesteria galliardi Garnham, 1973
Theodorides, J.; and Desportes, I., [1975], Ann. Parasitol., v. 49 (6), 1974, 787
may belong to new genus (as yet unnamed)

Lankesteria giardi Mingazzini, 1893
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria amaroucii (Giard, 1873) Labbe, 1899

Lankesteria gigantea Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria globosa Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria gracilis Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria gyriniformis Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria leuckarti (Mingazzini, 1893) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Tricystis leuckarti (Mingazzini, 1893) Ormieres, 1965

Lankesteria mackiei (Shortt & Swaminath, 1927) Bhatia, 1938
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis mackiei (Shortt & Swaminath, 1927) Ormieres, 1965

Lankesteria maculata Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Lankesteria magna Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
"Lankesteria amaroucii var. magna Ormieres, 1965. Emended to Lankesteria magna Ormieres, 1965."

Lankesteria molgulidarum Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria monstrosa Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria morchellii Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria ormieresi n. sp.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Lankesteria sp. Ormieres, 1965

Lankesteria parascidiae Duboscq & Harant, 1932
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria perophoropsis Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria phlebotomi mackiei Missiroli, 1932
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis mackiei (Shortt & Swaminath, 1927) Ormieres, 1965

Lankesteria plagiostomae (Georgevitch, 1951) Geus, 1967
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella plagiostomae (Georgevitch, 1951) comb. nov.

Lankesteria planariae (Mingazzini, 1893) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella planariae (Mingazzini, 1893) comb. nov.

Lankesteria siedleckii Duboscq & Harant, 1923 [emend.?]
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Lankesteria siedlickii Duboscq & Harant, 1923, lapsus calami

Lankesteria siedlickii Duboscq & Harant, 1923, lapsus calami
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria siedleckii Duboscq & Harant, 1923 [emend.?]

Lankesteria striata Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria styelae Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Lankesteria ascidiae (Lankester) *of* Harant (1931) *non* L. ascidiae (Lankester, 1872) Mingazzini, 1891

Lankesteria tripteroidesi Bhatia, 1938
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis tripteroidesi (Bhatia, 1938) Ormieres, 1965

Lankesteria tuzetae Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lankesteria zonata Ormieres, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Lateroprotomeritus n. g. [nomen nudum]
Theodorides, J.; and Desportes, I., 1975, J. Protozool., v. 22 (3), 85A-86A [Abstract]
gregarines of marine invertebrates, organization of epicyte

Laverania danilewsky
Peirce, M. A., 1976, Parasitology, v. 73 (3), 407-415
"L. danilewsky is a synonym pro parte of both H. passeris and H. columbae."

Lecudina Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae

Lecudina bogolepovae (Levine, 1971) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Polyrhabdina bogolepovae Levine, 1971; Polyrhabdina cirratuli Bogolepova, 1953

Lecudina catalinensis sp. n., illus.
Levine, N. D., 1974, J. Protozool., v. 21 (1), 10-12
Lumbrineris inflata (free in lumen of posterior and middle intestine): Santa Catalina Island, California

Lecudina elongata (Mingazzini, 1891) Kamm, 1922
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Ophioidina elongata Mingazzini, 1891

Lecudina eunicae (Lankester, 1866) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Polyrhabdina eunicae (Lankester, 1866) Labbe, 1899

Lecudina heterocephala (Mingazzini, 1891) Kamm, 1922
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Ophioidina heterocephala Mingazzini, 1891

Lecudina linei Vinckier
Vinckier, D., 1975, J. Protozool., v. 22 (2), 170-184
Lineus viridis (intestinal): littoral boulonnais, Station de Biologie maritime, Wimereux

Lecudina longissima Hoshide, 1944, illus.
Levine, N. D., 1974, J. Protozool., v. 21 (1), 10-12
description of gamonts
Lumbrineris zonata (free in lumen of posterior and middle intestine): Little Harbor, Santa Catalina Island, California

Lecudina nereicola (Bogolepova, 1953) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Polyrhabdina nereicola Bogolepova, 1953

Lecudina pherusae sp. n., illus.
Levine, N. D., 1974, J. Protozool., v. 21 (1), 10-12
Pherusa capulata (free in lumen of intestine): Big Fisherman's Cove, Santa Catalina Island, California

Lecudina polydorae (Leger, 1893) Kamm, 1922
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Polyrhabdina polydorae (Leger, 1893) Caullery & Mesnil. 1914

Lecudina zimmeri sp. n., illus.
Levine, N. D., 1974, J. Protozool., v. 21 (1), 10-12
Pherusa capulata (free in lumen of intestine): Big Fisherman's Cove, Santa Catalina Island, California

Lecudinidae Kamm, 1922
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Apicomplexa, Sporozoea, Gregarinia, Eugregarinida, Aseptatina
revision and checklist of species (excluding Lecudina)
Syn.: Doliocystidae Labbe, 1899
includes: Lecudina; Polyrhabdina; Ulivina; Sycia; Pontesia; Bhatiella; Viviera; Cochleomeritus; Ancora; Hentschelia; Lecythion; Hyperidion; Zygosoma; Filipodium; Diplauxis; Chlamydocystis; Contortiocorpa; Lankesteria; Monocystella; Ascocystis; Kofoidina; Ophioidina; Paraophioidina gen. n.; Extremocystis; Lobiancoella

Lecythion Mackinnon & Ray, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Lecythion thalassemae Mackinnon & Ray, 1931
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Legeria agilis (Schneider) pro parte sensu Baudoin, 1961
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Iorella wegoreki g. n., sp. n.

Legeria agilis, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Dytiscus marginalis

Legerozoon de Mello & Alfonso, 1935
Hsu, C.-K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon Berestnev, 1904

Leidyana ephestiae Daviault, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Ephestia kuehniella (intestine): Poland

Leidyana gryllorum, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Nemobius silvestris

Leidyana leptoplanae Bhatia & Setna, 1924
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella leptoplanae (Bhatia & Setna, 1924) comb. nov.

Leiotrocha
Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Urceolariidae

Leishmania
Abdalla, R. E., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1135-1138
human Sudan mucosal leishmaniasis, diagnosis comparing use of immunodiffusion, counterimmunoelectrophoresis, and immunofluorescence

Leishmania
Buys, J.; and Ruitenberg, E. J., 1977, Trop. and Geogr. Med., v. 29 (2), 205 [Abstract]
Trypanosoma brucei, T. rhodesiense, comparative application of enzyme-linked immunosorbent assay (ELISA) and immunofluorescence for serodiagnosis of human trypanosomiasis; ELISA represents good alternate method particularly suited for mass screening purposes; cross-reaction only in one person in whom antibodies to Leishmania were detected

Leishmania, illus.
Campo-Aasen, I.; Aleman, C.; and Torrealba, J. W., 1975, Acta Cien. Venezolana, v. 26 (5), 159-169
Leishmania, hairless mouse, paravertebral muscle, ultrastructure, pathology, mitochondria alteration, cellular defensive activity

Leishmania
Chance, M. L.; Schnur, L. F.; and Thomas, S. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 113 [Demonstration]
African rodent leishmanias, biochemical taxonomic identification of 4 strains distinct from L. donovani

Leishmania
Crofts, M. A. J., 1976, J. Trop. Med. and Hyg., v. 79 (5), 111-113
promising results in treating human mucocutaneous Leishmania with amphotericin B: Peru

Leishmania
Diamond, L. S.; and Mattern, C. F. T., 1976, Advances Virus Research, v. 20, 87-112
protozoal viruses, review

Leishmania, illus.
Ferry, A. P., 1977, Am. J. Ophth., Chicago, v. 84 (3), 349-354
cutaneous leishmaniasis of right lower eyelid of man who had earlier resided in area of Mediterranean Sea, diagnosis only after excision of lesion thought to be tumor or cyst, clinical diagnostic review: New York

Leishmania, illus.
Giauffret, A.; Sanchis, R.; and Vitu, C., 1976, Rev. Med. Vet., Toulouse, v. 127 (6), 913-930
Leishmania, canine, diagnosis by immunofluorescence, prognosis, treatment with Glucantime, review

Leishmania
Kallinikova, V. D., 1977, Molec. and Cell. Biochem., v. 14 (1-3), Feb. 4, 55-58
Kinetoplastida, kinetoplast genetic system, implications for concept of cell differentiation and phylogeny, relationship to phenotype of these flagellates

Leishmania
Martaresche, B.; et al., 1975, Medecine Trop., v. 35 (4), 309-310
human Plasmodium falciparum, positive immunofluorescence test for leishmaniasis

Leishmania
Mauel, J.; and Behin, R., 1974, Transplant. Rev., v. 19, 121-146
cell-mediated and humoral immunity to protozoan infections (leishmaniasis, malaria, trypanosomiasis), review

Leishmania
Ruitenberg, E. J.; and Buys, J., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 31-36
Trypanosoma brucei, enzyme-linked immunosorbent assay (ELISA) for serodiagnosis of human African sleeping sickness, comparison tests with immunofluorescence technique showed good results in rabbits (exper.) and serum from infected humans, cross-reactions only in person with Leishmania antibodies, possible application to epidemiologic surveys

Leishmania, illus.
Sanchis-Bayarri Lahoz, V.; and Sanchis-Bayarri Vaillant, V., 1974, Med. Espan. (426), an. 37, v. 72, 162-164
case report and pathologic findings of "Oriental boil" leishmanial infection in young child: Spain

Leishmania
Schnur, L. F.; and Chance, M. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 14 [Demonstration]
Leishmania strains from Sudan, Kenya and Ethiopia, comparison of strain differentiation methods

Leishmania
Schnur, L. F.; and Zuckerman, A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 15 [Demonstration]
Israeli leishmanial strains, excreted factor serotypes

Leishmania
Schnur, L. F.; Zuckerman, A.; and Greenblatt, C. L., 1973, J. Protozool., v. 20 (4), 534
Leishmania, relationship between clinical types and serotypes not as simple and direct as previously thought

Leishmania
Simpson, L., 1973, J. Protozool., v. 20 (1), 2-8
salivarian and stercorarian trypanosomes, kinetoplast DNA, structure and function, review

Leishmania
Stauber, L. A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 739-765
leishmaniasis, immunology, review

Leishmania
Tikasingh, E. S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (3), 261-262 [Letter]
human leishmaniasis, guinea pigs to be used as sentinel animals for study of Leishmania should first be immunized against locally occurring arboviruses to which they are normally very sensitive

Leishmania
Trigg, P. I.; and Gutteridge, W. E., 1977, Symposia Brit. Soc. Parasitol., v. 15, 57-81
parasitic Protozoa, morphological, biochemical, and physiological changes occurring during life cycles, symposium presentation

Leishmania sp.
Arias, J. R.; Menezes, H.; and de Freitas, R. A., 1976, Acta Amazonica, v. 6 (4), 483-485
Leishmania sp., 2 strains, golden hamsters immunized with anti-Chagas "PF" vaccine (Trypanosoma cruzi after 400 passages), no significant reduction of infection

Leishmania [sp.]
Ashford, R. W.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 265-271
cutaneous leishmaniasis, rural zoonotic infection with probable transmission by Phlebotomus papatasi between rodent spp., ecological and prevalence survey
Psammomys obesus
Meriones libycus
all from Libya

Leishmania possibly n. sp., illus.
Barbosa, W.; et al., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 389-399
characterization (clinical, immunological, histopathological, cultural, respiratory, DNA buoyant density, malic dehydrogenase isoenzyme pattern), does not fit into basic parameters of either L. mexicana or L. braziliensis complexes
human (cutaneous): Barra do Garcas, Mato Grosso, east central region of Brazil

Leishmania sp.
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania sp.
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania sp.
Corbeil, L. B.; et al., 1976, Clin. Immunol. and Immunopathol., v. 6 (2), 165-173
canine visceral leishmaniasis with amyloidosis, immunopathological case report, inappropriate increase in humoral response and decrease in cell-mediated immunity: New York State, imported from Greece

Leishmania sp.
Edeson, J. F. B.; and Himo, J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 27 [Demonstration]
presence of amastigotes in blood films
Agama stellio: Lebanon

Leishmania [sp.], morphologically similar to L. tropica major, illus.
Edrissian, G. H.; Ghorbani, M.; and Tahvildar-Bidruni, G., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 517-519
Meriones persicus (sore on tip of nose)
Rhombomys opimus (ear scrapings)
all from Iran

Leishmania sp.
El-Adhami, B., 1976, Am. J. Trop. Med. and Hyg., v. 25 (5), 759-761
white mice (exper.) (liver and spleen)
Rattus rattus (blood and liver cultures): Baghdad area, Iraq

Leishmania sp.
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania [sp.], illus.
George, J. W.; et al., 1976, Vet. Path., v. 13 (5), 365-373
Leishmania [sp.] in Great Dane dog, case report, mixed infection with amyloidosis: New York, imported from Greece

Leishmania [sp.]
Grove, S. S.; and Ledger, J. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 523-524
Phlebotomus
Procavia capensis
all from South West Africa

Leishmania [sp.]
Haile, T. T.; and Lemma, A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 180-181 [Letter]
possible new reservoir host
Arvicanthis niloticus (liver, spleen): Humera area, Ethiopa

Leishmania [sp.]
Haile, T. T.; and Lemma, A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 180-181 [Letter]
strain similar to that causing human cutaneous leishmaniasis, possible reservoir host
Arvicanthis sp. (liver): Omo River Valley, Ethiopia

Leishmania sp.
Kilgour, V.; et al., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 245-246
Leishmania spp., aminotransferases, demonstration of electrophoretic variation, potential for differentiating isolates and deriving taxonomic characters

Leishmania [sp.]
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 171-172
cutaneous leishmaniasis, Lutzomyia auduzei implicated as major vector: North Brazil

Leishmania sp.
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex
Proechimys guyannensis (liver, spleen): Transamazon highway, Itaituba-Altamira, Para State, Brazil

Leishmania sp.
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex
Rattus rattus (skin): Transamazon highway, Jacareacanga-Humaita, Para State, Brazil

Leishmania sp., illus.
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex
Didelphis marsupialis (liver, spleen): Transamazon highway, Itaituba-Altamira, Para State, Brazil

Leishmania spp.
Lumsden, W. H. R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 74-75 [Letter]
Leishmania spp., classification of taxonomic characters using morphology, chemical structure, immunologic effect, behaviour and clinical outcome in man as criteria

Leishmania sp.
Peters, W.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 501-502
Leishmania aethiopica, Leishmania adleri, Leishmania sp., enzyme variants, DNA buoyant densities and excreted factor serotypes as means of differentiation of human and animal isolates from Kenya

Leishmania sp.
Ranque, P.; Quilici, M.; and Debaene, A., 1972, Medecine Trop., v. 32 (4), 499-503
Arvicanthis niloticus, exper. infections with varying strains of Leishmania tropica, L. donovani and L. sp., minimal skin infections with no secondary visceral invasion

Leishmania [spp.]
Schnur, L. F.; and Zuckerman, A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 434-435 [Demonstration]
Leishmania spp. from Sudan, Ethiopia and Kenya, differentiation of leishmanial serotypes

Leishmania sp.
Schnur, L. F.; and Zuckerman, A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 273-294
Sudanese, Kenyan and Ethiopian leishmanial isolates from human visceral and cutaneous infections, various wild animals and sandflies, leishmanial excreted factor (EF) serotypes determined by gel diffusion, discussion of distribution and relationships

Leishmania [sp.]
Shaw, P. K.; et al., 1976, Am. J. Trop. Med. and Hyg., v. 25 (6), 788-796
Leishmania [sp.], human, 2 autochthonous cases, epidemiologic data suggest that dermal leishmaniasis is endemic in south-central Texas

Leishmania spp.
Steck, E. A., 1974, Progr. Drug Research, v. 18, 289-351
Leishmania spp., extensive review (history, etiology, pathology, epidemiology, immunology, cultivation, biochemistry, chemotherapy)

Leishmania adleri
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania adleri
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania adleri
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania adleri
Lewis, D. H., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 327-336
Leishmania mexicana mexicana, infection of dog sarcoma cells in presence of cytochalasin B indicates parasites can infect host cells without agency of active phagocytosis by latter; differing abilities of various Leishmania spp. promastigotes to infect same cell type suggest active contribution of parasites themselves to infection

Leishmania adleri, illus.
Lewis, D. H., 1975, J. Protozool., v. 22 (3), 344-352
Leishmania spp., 4 reptilian vs. 1 mammalian isolates, ultrastructure of promastigotes, possible characters for use in taxonomy and species or strain differentiation

Leishmania adleri
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania adleri
Peters, W.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 501-502
Leishmania aethiopica, Leishmania adleri, Leishmania sp., enzyme variants, DNA buoyant densities and excreted factor serotypes as means of differentiation of human and animal isolates from Kenya

Leishmania adleri
Schnur, L. F.; and Zuckerman, A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 273-294
Sudanese, Kenyan and Ethiopian leishmanial isolates from human visceral and cutaneous infections, various wild animals and sandflies, leishmanial excreted factor (EF) serotypes determined by gel diffusion, discussion of distribution and relationships

Leishmania aethiopica n. sp.
Bray, R. S.; Ashford, R. W.; and Bray, M. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 345-348
hamster (exper., with difficulty)
Procavia habessinica
Heterohyrax brucei
Phlebotomus longipes
P. pedifer
human
all from highlands of Ethiopia and/or Mt. Elgon, Kenya

Leishmania aethiopica
Ashford, R. W.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 568-601
parasites indistinguishable from those causing cutaneous leishmaniasis in man found in Procavia habessinica and Heterohyrax brucei and in Phlebotomus longipes and Phlebotomus pedifer (Leishmania aethiopica), close association between hyrax and sandflies, man is incidental host: Ethiopia

Leishmania aethiopica
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania aethiopica
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania aethiopica
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania aethiopica
Matossian-Rogers, A.; Lumsden, W. H. R.; and Dumonde, D. C., 1976, Immunology, v. 31 (1), 1-19
Leishmania enriettii, L. tropica major, L. aethiopica, L. mexicana amazonensis, numerical immunotaxonomy, differentiation according to reactivity and cross-reactivity in tests of parasite agglutination, indirect immunofluorescence, and passive cutaneous anaphylaxis

Leishmania aethiopica
Peters, W.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 501-502
Leishmania aethiopica, Leishmania adleri, Leishmania sp., enzyme variants, DNA buoyant densities and excreted factor serotypes as means of differentiation of human and animal isolates from Kenya

Leishmania aethiopica Bray et al., 1973
Schnur, L. F.; and Zuckerman, A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 273-294
Sudanese, Kenyan and Ethiopian leishmanial isolates from human visceral and cutaneous infections, various wild animals and sandflies, leishmanial excreted factor (EF) serotypes determined by gel diffusion, discussion of distribution and relationships

Leishmania agamae
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania agamae
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania agamae
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania agamae
Lewis, D. H., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 327-336
Leishmania mexicana mexicana, infection of dog sarcoma cells in presence of cytochalasin B indicates parasites can infect host cells without agency of active phagocytosis by latter; differing abilities of various Leishmania spp. promastigotes to infect same cell type suggest active contribution of parasites themselves to infection

Leishmania agamae, illus.
Lewis, D. H., 1975, J. Protozool., v. 22 (3), 344-352
Leishmania spp., 4 reptilian vs. 1 mammalian isolates, ultrastructure of promastigotes, possible characters for use in taxonomy and species or strain differentiation

Leishmania agamae
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania braziliensis
Avila, J. L.; et al., 1976, Biochem. Soc. Tr., v. 4 (4), 680-681 [Abstract]
Trypanosoma cruzi, Leishmania braziliensis, mice, infectivity loss after in vitro exposure to polyenzymic 'PIGO' (peroxidase, iodide, glucose, type-II oxidase and oxygen)

Leishmania braziliensis
Berens, R. L.; and Marr, J. J., 1977, J. Protozool., v. 24 (2), 340-344
Leishmania donovani, L. braziliensis, phosphofructokinase probably does not play an important role in glycolysis

Leishmania brasiliensis
Berens, R. L.; and Marr, J. J., 1977, Exper. Parasitol., v. 43 (2), 407-413
Leishmania donovani, L. brasiliensis, pyruvate kinase, regulatory properties in relation to regulation of glycolysis, comparison with Crithidia fasciculata

Leishmania braziliensis
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania braziliensis
Brazil, R. P., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 359 [Letter]
rebuttal to previously published letter on the identification of Brazilian Leishmania strains and on the use of the name L. braziliensis; need for thorough differentiation from L. mexicana complex

Leishmania brasiliensis, illus.
Campo-Aasen, I.; Aleman, C.; and Convit, J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 122-124
Leishmania brasiliensis, observation of new organelle (4 microtubule structure near and running parallel to flagellar pocket)

Leishmania braziliensis
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania braziliensis
Dawidowicz, K.; et al., 1975, J. Parasitol., v. 61 (5), 950-953
Leishmania braziliensis, infective amastigotes and promastigotes and noninfective promastigotes, agglutination by various lectins, results suggest presence of complex carbohydrate groups on surface membrane

Leishmania braziliensis
Decker, J. E.; and Schrot, J. R., 1976, J. Protozool., v. 23 (2), 18A-19A [Abstract]
Leishmania donovani, L. braziliensis, L. tropica, rapid identification by radiorespirometry

Leishmania brasiliensis
Decker, J. E.; Schrot, J. R.; and Levin, G. V., 1977, J. Protozool., v. 24 (3), 463-470
Leishmania spp., radiorespirometric technique has potential for rapid identification of species and strains

Leishmania braziliensis
Dollahon, N. R., 1974, J. Protozool., v. 21 (3), 422 [Abstract]
Anolis carolinensis (exper.)

Leishmania braziliensis
Ebert, F., 1974, Tropenmed. u. Parasitol., v. 25 (3), 259-266
Leishmania spp. of the New World, comparative electrophoretic studies on proteins, esterases, relationships to Leishmania donovani and L. tropica, use of species-specific enzyme patterns in differentiating species and strains

Leishmania braziliensis
Herrer, A.; and Christensen, H. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (1), 54-58
Leishmania braziliensis, human, endemic persistence of cutaneous leishmaniasis with occurrence of disease mainly in children, Choloepus hoffmanni as principal reservoir host with infections also found in Canis familiaris and Aotus trivirgatus, collection of potential phlebotomine sandfly vectors: El Aguacate, Panama

Leishmania braziliensis
Herrer, A.; and Christensen, H. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (1), 59-63
Leishmania braziliensis, dogs, survey for natural cutaneous leishmaniasis, may serve as incidental reservoir host and/or liason of infection between jungle and forest settlements: Panama

Leishmania braziliensis
Herrer, A.; and Christensen, H. A., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 59-65
Leishmania braziliensis, epidemiological description of an outbreak of cutaneous leishmaniasis among a small group of settlers entering the forest to establish a homestead, natural infections found in dogs but not in 41 feral mammals, potential sandfly vectors: Loma de Mercurio, foothills of Cerro Azul, 50 km east of Panama City, Panama

Leishmania braziliensis
Herrer, A.; Christensen, H. A.; and Beumer, R. J., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 67-71
Leishmania braziliensis, epidemiological description of incidental occurrence of cases in a non-endemic settlement, men and dogs apparently acquired infections while hunting in nearby forest which was an endemic focus, infection also found in Bradypus infuscatus, potential phlebotomine vectors collected: Majecito Arriba, Bayano basin, Panama

Leishmania brasiliensis
Huebsch, R. M.; Sulzer, A. J.; and Kagan, I. G., 1976, J. Parasitol., v. 62 (4), 523-527
evaluation of sensitivity and specificity of indirect immunofluorescence test for autoimmune-type EVI antibodies in sera of patients with Chagas disease (Trypanosoma cruzi), leishmaniasis (Leishmania brasiliensis, L. donovani), malaria, and several other nonparasitic diseases; second type of staining of heart tissue also reported for patients with leishmaniasis and malaria but not Chagas disease

Leishmania braziliensis
Marr, J. J.; and Berens, R. L., 1977, J. Infect. Dis., v. 136 (6), 724-732
Leishmania spp., in vitro study of antileishmanial effects of allopurinol and its metabolic derivative oxipurinol, suggested sites of action, reversal of antileishmanial effects by adenine and its derivatives

Leishmania brasiliensis
Martin, E.; et al., 1976, J. Protozool., v. 23 (4), 600-607
Leishmania, 4 human spp. compared, promastigotes, occurrence and levels of activity of various enzymes of carbohydrate catabolism

Leishmania brasiliensis, Vianna 1911
Medina, H.; et al., 1955, Arq. Biol. e Tecn., v. 10, 121-165
Astaril, tartar emetic and Repodral, effect on respiration and anaerobic glycolysis of Leishmania brasiliensis and isolated mouse diaphragm

Leishmania brasiliensis, Vianna 1911
Medina, H.; Amaral, D.; and Bacila, M., 1955, Arq. Biol. e Tecn., v. 10, 97-119
Leishmania brasiliensis, pathways of glucose and acetate oxidation

Leishmania brasiliensis Vianna 1911
Medina, H.; and Bacila, M., 1955, Arq. Biol. e Tecn., v. 10, 81-96
Leishmania brasiliensis, L. enriettii, cultured in new diphasic medium containing spinach and glucose; L. enriettii grew only in medium containing added guinea-pig blood

Leishmania brasiliensis, illus.
Merino, F.; et al., 1977, Internat. Arch. Allergy and Applied Immunol., v. 55 (1-6), 487-495
Leishmania brasiliensis, in vitro infection of murine macrophages, mechanism of penetration, endocytosis with active participation of both cells

Leishmania braziliensis
Miles, C. T.; Foster, W. A.; and Christensen, H. A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 531-532 [Letter]
Leishmania braziliensis, evidence that some vector Lutzomyia spp. may use host stimuli for sexual aggregation as well as blood feeding

Leishmania braziliensis
Minoprio, J. L., 1972, Prensa Med. Argent., v. 59 (8), 293-308
human Chagas disease, pathologic findings, comparison with characteristics of other Trypanosoma spp. and Leishmania spp., extensive review

Leishmania braziliensis, illus.
Muehlpfordt, H., 1975, Tropenmed. u. Parasitol., v. 26 (4), 385-389
Leishmania donovani, L. tropica. L. braziliensis, comparative electron morphology of developing promastigote forms using cationized ferritin to determine labelling of negative charges on cell surfaces

Leishmania brasiliensis
Oelerich, S., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 296-304
Leishmania spp., Trypanosoma cruzi, Crithidia sp., Mycobacterium smegmatis, antigenic analyses and cross reactions, double gel-diffusion, complement-fixation, indirect hemagglutination

Leishmania brasiliensis
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Leishmania braziliensis
Scholten, T.; et al., 1977, Canad. Med. Ass. J., v. 117 (4), 368-369
Leishmania braziliensis, ulcer on nose of Canadian man after stay in Mexico

Leishmania braziliensis
Steiger, R. F.; and Meshnick, S. R., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 441-443
Leishmania donovani, L. braziliensis, amino-acid and glucose utilization by parasites

Leishmania braziliensis
Steiger, R. F.; and Steiger, E., 1976, J. Parasitol., v. 62 (6), 1010-1011
Leishmania donovani, L. braziliensis, chemically defined medium for in vitro cultivation

Leishmania braziliensis
Steiger, R. F.; and Steiger, E., 1977, J. Protozool., v. 24 (3), 437-441
Leishmania donovani, L. brasiliensis, promastigotes, nutritional requirements studied in modifications of simple defined culture medium, special emphasis placed on glucose and amino acid requirements

Leishmania braziliensis
Walton, B. C.; Intermill, R. W.; and Hajduk, M. E., 1977, Am. J. Trop. Med. and Hyg., v. 26 (5, part 1), 850-855
Leishmania isolates from patients with espundia compared with reference strain of L. b. braziliensis, clinical features, growth in culture, pathogenesis in hamster, location in gut of sandflies (Lutzomyia gomezi and L. trapidoi), autochthonous Texas strain clearly falls into L. mexicana complex, Peru isolate has intermediate pattern of characteristics but primarily those of L. braziliensis

Leishmania braziliensis
Wilson, H. R.; and Fairchild, D. G., 1977, Stain Tech., v. 52 (2), 105-111
modified Whipf's polychrome staining to demonstrate protozoans and tissue components in cutaneous lesions of infected animals

Leishmania braziliensis
Zeledon, R.; and Alfaro, M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 416-417 [Letter]
Leishmania braziliensis strain isolated from Lutzomyia ylephiletor and L. shannoni in Costa Rica, data suggest that injection of small numbers may produce abortive infection followed by solid immunity

Leishmania braziliensis subsp.
Zeledon, R.; Ponce, C.; and de Ponce, E., 1975, Am. J. Trop. Med. and Hyg., v. 24 (4), 706-707
hamsters (exper.)
Bradypus griseus (spleen, liver, blood, dorsal skin): Costa Rica
Choloepus hoffmanni (spleen, liver, blood, skin): Costa Rica

Leishmania braziliensis
Zeledon, R.; de Ponce, E.; and Ponce, C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 536-537 [Letter]
comparison of Montenegro skin test for delayed hypersensitivity and macrophage migration inhibition test in 3 persons with cured leishmaniasis and woman with chronic Leishmania braziliensis

Leishmania braziliensis braziliensis
Brazil, R. P., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 89 [Letter]
metastatic spread of infection to extremities of hamsters (exper.)

Leishmania b. braziliensis
Chance, M. L.; and Gardener, P. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 9 [Demonstration]
Leishmania spp., morphological and biochemical differentiation into 4 significantly different groups within the genus

Leishmania braziliensis braziliensis
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania braziliensis braziliensis
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania brasiliensis brasiliensis
Childs, G. E.; McRoberts, M. J.; and Foster, K. A., 1976, J. Parasitol., v. 62 (5), 676-679
Leishmania spp. amastigotes, recovery from cutaneous lesions of host animals by a method adapted from primary tissue culture techniques, pathogenic characteristics retained, comparatively free of host cells

Leishmanis braziliensis braziliensis
Croft, S. L.; and Schnur, L. F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 114 [Demonstration]
Leishmania b. braziliensis; L. hertigi, morphological comparisons of excreted factor-producing strain and non-excreted factor-producing strain, possible relationships between excreted factor and Noguchi-Adler phenomenon

Leishmania braziliensis braziliensis
Gardener, P. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 9 [Demonstration]
Leishmania braziliensis braziliensis, unidentified cytoplasmic granule characteristic of amastigote stages of this sp., visible with light microscope

Leishmania braziliensis braziliensis
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania braziliensis braziliensis (H9 isolate), illus.
Gardener, P. J.; Shchory, L.; and Chance, M. L., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 147-155
Leishmania species differentiation by morphometric comparisons of ultrastructural features of leishmanial amastigotes

Leishmania braziliensis braziliensis
Hommel, M., 1975, J. Protozool., v. 22 (3), 52A [Abstract]
Leishmania braziliensis braziliensis, 2 "slow strains", pattern of infection in cyclophosphamide-pretreated hamsters differs with route of infection, heavy bloodstream infection produced in nude mice

Leishmania braziliensis braziliensis
Hommel, M.; Peters, W.; and Chance, M. L., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 9-10 [Demonstration]
Leishmania braziliensis braziliensis, growth and development in immunosuppressed hamsters and nude mice, pathology

Leishmania braziliensis braziliensis, illus.
Killick-Kendrick, R.; et al., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1131), v. 198, 191-199
Leishmania braziliensis in Lutzomyia longipalpis (exper.), development and attachment of parasites in ileum and pylorus

Leishmania braziliensis braziliensis
Killick-Kendrick, R.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 381 [Demonstration]
Leishmania braziliensis braziliensis promastigotes, localization in gut wall of Psychodopygus wellcomei vector flies in Serra dos Carajas, North Brazil

Leishmania braziliensis braziliensis, illus.
Lainson, R.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 184-196
Leishmania braziliensis braziliensis, descriptions of infections in 17 persons from endemic area, promastigotes "most likely" L. b. braziliensis found in Psychodopygus wellcomei, P. paraensis, and P. amazonensis, no infections found in reservoir hosts: Serra dos Carajas, Para, north Brazil

Leishmania braziliensis braziliensis
Lainson, R.; and Shaw, J. J., 1977, J. Trop. Med. and Hyg., v. 80 (2), 29-35
cross-immunity trials with Cebus apella apella and observations on natural and experimental human infections confirm separate identity of Leishmania mexicana mexicana, L. m. amazonensis, L. braziliensis braziliensis, L. b. guyanensis and L. b. panamensis

Leishmania braziliensis braziliensis, illus.
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex
man (cutaneous lesion): Serra dos Carajas, Para State, Brazil

Leishmania braziliensis braziliensis
Lewis, D. H.; and Peters, W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 295-312
Leishmania spp., promastigote forms, relationship between parasites and host macrophages and their relevance to intracellular survival of parasites, resistance of intracellular Leishmania to digestion by lysosomal enzymes

Leishmania braziliensis braziliensis
Schnur, L. F., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 277-278 [Demonstration]
Leishmania braziliensis braziliensis (slow strain), L. mexicana amazonensis (fast strain); dissemination in hamsters

Leishmania braziliensis braziliensis
Walton, B. C.; Intermill, R. W.; and Hajduk, M. E., 1977, Am. J. Trop. Med. and Hyg., v. 26 (5, part 1), 850-855
Leishmania isolates from patients with espundia compared with reference strain of L. b. braziliensis, clinical features, growth in culture, pathogenesis in hamster, location in gut of sandflies (Lutzomyia gomezi and L. trapidoi), autochthonous Texas strain clearly falls into L. mexicana complex, Peru isolate has intermediate pattern of characteristics but primarily those of L. braziliensis

Leishmania brasiliensis brasiliensis
Walton, B. C.; Shaw, J. J.; and Lainson, R., 1977, J. Parasitol., v. 63 (6), 1118-1119
Leishmania brasiliensis brasiliensis, in vitro cultivation

Leishmania braziliensis braziliensis Vianna
Ward, R. D., 1977, Proc. 15. Internat. Cong. Entom. (Washington, D.C., Aug. 19-27, 1976), 505-522
New World leishmaniasis, incidence, factors causing epidemiological changes in last 3 decades

Leishmania braziliensis braziliensis
Ward, R. D.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 174-183
survey of phlebotomine fauna in area highly endemic for Leishmania braziliensis braziliensis: forest of Serra dos Carajas, Para State, Brazil

Leishmania braziliensis guyanensis
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania braziliensis guyanensis
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania braziliensis guyanensis
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania braziliensis guyanensis, illus.
De Souza, W.; and Brasil, R. P., 1976, Ztschr. Parasitenk., v. 50 (1), 1-9
Leishmania braziliensis guyanensis, electron microscopic and cytochemical studies of concanavalin A receptors on cell membranes; agglutination; cell surface polysaccharides, glycoproteins and glycolipids

Leishmania braziliensis guyanensis
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cross-immunity trials with Cebus apella apella and observations on natural and experimental human infections confirm separate identity of Leishmania mexicana mexicana, L. m. amazonensis, L. braziliensis braziliensis, L. b. guyanensis and L. b. panamensis

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Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

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Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania braziliensis panamensis
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cross-immunity trials with Cebus apella apella and observations on natural and experimental human infections confirm separate identity of Leishmania mexicana mexicana, L. m. amazonensis, L. braziliensis braziliensis, L. b. guyanensis and L. b. panamensis

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Leishmania donovani, simple monophasic medium for axenic culture which favors amastigote to promastigote transformation, promastigote size distribution and glucose consumption during cell cycle; medium also supports high population growth of L. tarentolae promastigotes and Trypanosoma cruzi (Costa Rica but not Corpus Christi strain) epimastigotes

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Trypanosoma cruzi, Leishmania spp., amastigotes, comparative electron microscope study, pellicle-associated structures

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Leishmania donovani, L. mexicana amazonensis, enzyme differences between life-cycle stages, malate dehydrogenase occurred in promastigote culture stage as different isoenzyme variants but not in amastigote intracellular form, lactate dehydrogenase could not be demonstrated in either stage

Leishmania donovani
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Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

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Leishmania donovani cultivated at 25 C vs. 37 C, activities of glucose-6-phosphate, 6-phosphogluconate, and isocitrate dehydrogenase

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Leishmania donovani, mice, cyclophosphamide suppresses acquired immunity if given when infection becomes chronic, from experiments using such cyclophosphamide-immunosuppressed mice it appears that neither specific humoral anti-leishmanial antibody nor 'immune' macrophages per se is directly responsible for the acquired immunity

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Leishmania donovani
Ilardi, A.; Guglielmi, S.; and Proietti, A.M., 1972, Parassitologia, v. 14 (2-3), 317-324
X-irradiation of Leishmania and leptomonas forms of strain 2S of Leishmania donovani, hamsters, protection increases as irradiation increases

Leishmania donovani
Jadin, J. M.; et al., 1977, J. Protozool., v. 24 (4), 61A [Abstract]
Kinetoplastidae, culture in mouse peritoneal macrophages

Leishmania donovani
Janovy, J., jr.; Greiner, E. C.; and Decker, J. E., 1974, J. Protozool., v. 21 (3), 430 [Abstract]
Leishmania donovani vs. L. mexicana, anaerobic glycolysis and CO_2 fixation over the range 25-37 C, effect of pentostam vs. stibophen

Leishmania donovani, illus.
Jones, T. C., 1974, J. Reticuloendothel. Soc., v. 15 (5), 439-450
interactions of intracellular parasites (including protozoa with special emphasis on Toxoplasma gondii) with macrophages, review: endocytosis, the entry process; survival within vacuolar system; effects of immunity on intracellular parasitism

Leishmania donovani
Keithly, J. S., 1976, J. Protozool., v. 23 (2), 244-245
Leishmania donovani, improved infectivity assay using hamsters confirms greater infectivity of amastigotes vs. promastigotes

Leishmania donovani
Kilgour, V.; et al., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 245-246
Leishmania spp., aminotransferases, demonstration of electrophoretic variation, potential for differentiating isolates and deriving taxonomic characters

Leishmania donovani
Krassner, S. M.; and Flory, B., 1972, J. Protozool., v. 19 (4), 682-685
Leishmania donovani promastigotes, proline metabolism

Leishmania donovani, illus.
Kutar, K.; Birol, K.; and Kilic, Y., 1974, Mikrobiyol. Bul., v. 8 (3), 277-292
Leishmania donovani, adult type kala-azar in children, increasing incidence, differential diagnosis, clinical management, glucantime: Turkey

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Leishmania donovani, Trypanosoma cruzi, T. brucei group, comparisons of life cycle, morphology, reproduction and development, metabolism, extensive review

Leishmania donovani
La Placa, M.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 396-398
partially purified protein and polysaccharide antigens obtained from Leishmania donovani, both fractions induce positive intradermal skin reaction, but only the protein fraction detects humoral antibody in complement fixation tests

Leishmania donovani
Mahmoud, A. A. F.; and Warren, K. S., 1977, J. Infect. Dis., v. 136 (1), 160-163
Leishmania donovani, algorithms in diagnosis and management of human visceral leishmaniasis

Leishmania donovani
Manson-Bahr, P. E. C.; and Wilson, V. L. C., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 20 [Demonstration]
Leishmania donovani, Ethiopian strain in monkeys, production of self-healing hepatic granulomas and leishmaniomas

Leishmania donovani
Marr, J. J.; and Berens, R. L., 1977, J. Infect. Dis., v. 136 (6), 724-732
Leishmania spp., in vitro study of antileishmanial effects of allopurinol and its metabolic derivative oxipurinol, suggested sites of action, reversal of antileishmanial effects by adenine and its derivatives

Leishmania donovani
Martin, E.; et al., 1976, J. Protozool., v. 23 (4), 600-607
Leishmania, 4 human spp. compared, promastigotes, occurrence and levels of activity of various enzymes of carbohydrate catabolism

Leishmania donovani
Matossian, R. M.; Kurban, A. K.; and Malak, J. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 450-452
Leishmania tropica, humans, circulating antibodies, detection by immunofluorescence, cross reactivity with L. donovani and Trypanosoma cruzi antigens, possible cross reactivity in mixed tuberculosis infections

Leishmania donovani
Mattock, N. M., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 345-348
Leishmania mexicana mexicana and L. donovani amastigote forms grown in cell culture, failure to detect lactate dehydrogenase and malate dehydrogenase, presence of glucose-6-phosphate dehydrogenase indicated

Leishmania donovani
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania donovani, illus.
Mattock, N. M; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 349-357
Leishmania mexicana mexicana, L. tropica major, L. donovani amastigotes in dog sarcoma cells and in hamster peritoneal exudate cell line, techniques for studying sensitivity of parasites and hosts to drugs, concluded that tissue culture system is valid way of determining baseline sensitivity of a Leishmania to chemotherapy, illustrative data with amphotericin B and 7'-trifluormethyl-dihydrocinchonine dihydrochloride

Leishmania donovani
Mattock, N. M.; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 359-371
Leishmania spp. in tissue culture model system, variety of compounds used in treatment of parasitic or bacterial infections tested for activity, concluded that model may be of use in screening of drugs for leishmanicidal action

Leishmania donovani
Mattock, N. M.; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 449-462
Leishmania spp. in tissue culture model system, leishmanicidal action of over 100 compounds examined

Leishmania donovani
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epidemiologic and clinical review of Leishmania donovani in Tchad

Leishmania donovani
Minoprio, J. L., 1972, Prensa Med. Argent., v. 59 (8), 293-308
human Chagas disease, pathologic findings, comparison with characteristics of other Trypanosoma spp. and Leishmania spp., extensive review

Leishmania donovani, illus.
Muehlpfordt, H., 1975, Tropenmed. und Parasitol., v. 26 (4), 385-389
Leishmania donovani, L. tropica, L. braziliensis, comparative electron morphology of developing promastigote forms using cationized ferritin to determine labelling of negative charges on cell surfaces

Leishmania donovani
Musumeci, S.; Fischer, A.; and Pizzarelli, G., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 176-177
Leishmania donovani, persistence of dysproteinaemia in persons treated and clinically cured of infections, possible explanations discussed

Leishmania donovani, illus.
Naik, N. G.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 328-332
two autochthonous cases of kala-azar, first report from Zambia (indeed from Central and Southern Africa), both co-existing with tuberculosis, skin lesions were main presenting feature

Leishmania donovani
Oelerich, S., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 296-304
Leishmania spp., Trypanosoma cruzi, Crithidia sp., Mycobacterium smegmatis, antigenic analyses and cross reactions, double gel-diffusion, complement-fixation, indirect hemagglutination

L[eishmania] donovani
Pampiglione, S.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (6), 447-453
L[eishmania] donovani, serologic screening of population during outbreak of visceral leishmaniasis showed 6 persons with high complement fixing and positive skin tests for leishmanin without clinical symptoms: Northern Italy

Leishmania donovani
Pampiglione, S.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 60-68
Leishmania donovani, statistics of leishmanin skin test survey of old endemic focus and new outbreak area of human Mediterranean leishmaniasis, useful tool for epidemiologic studies: Italy

Leishmania donovani
Pampiglione, S.; La Placa, M.; and Schlick, G., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 349-359
Leishmania donovani, epidemiological, clinical and laboratory data on outbreak of visceral leishmaniasis affecting 60 persons, liver biopsies revealed granulomatous hepatitis, outbreak possibly related to exceptionally dry summer: Emilia-Romagna, Italy

Leishmania donovani, illus.
Pennisi, L.; and Faraone, U., 1973, Riv. Parassitol., Roma, v. 34 (4), 249-262
Leishmania donovani promastigotes, scanning electron microscopy

Leishmania donovani
Piekarski, G.; Saathoff, M.; and Nouri-Nekoui, M. H., 1973, Ztschr. Tropenmend. u. Parasitol., v. 24 (2), 161-173
Leishmania donovani, rats and hamsters (exper.), patients with kala-azar, indirect immunofluorescence and complement fixation tests for detection of antibodies, failure to apply principle of Sabin-Feldman test

Leishmania donovani
Pieragostini, E.; et al., 1974, Atti Soc. Ital. Sc., v. 28, 854-857
Leishmania donovani (3 strains), Leishmania tropica, analysis of sterols, comparison; cholesterol possibly from culture medium

Leishmania donovani
Ranque, P.; Quilici, M.; and Debaene, A., 1972, Medecine Trop., v. 32 (4), 499-503
Arvicanthis niloticus, exper. infections with varying strains of Leishmania tropica, L. donovani and L. sp., minimal skin infections with no secondary visceral invasion

Leishmania donovani
Rezai, H. R.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 149-151
Leishmania donovani, techniques of indirect immunofluorescence and counter current immunoelectrophoresis compared in diagnosing kala-azar in children, immunoelectrophoretic test found useful tool for epidemiologic surveys and diagnosis

Leishmania donovani
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Leishmania donovani, illus.
Rowecka-Trzebicka, K.; and Touhami, M., 1977, Pediat. Polska, v. 52 (4), 381-386
Leishmania donovani, difficulty in diagnosing visceral leishmaniasis in children, case reports of children with atypical clinical manifestations and negative results from diagnostic bone marrow biopsies: Poland

Leishmania donovani
Sanchis, R.; Vitu, C.; and Giauffret, A., 1976, Rev. Med. Vet., Toulouse, v. 127 (8-9), 1191-1202
Leishmania donovani, experimentally infected dogs, symptoms, clinical tests (haemogram, sedimentation rate, blood proteins, immunofluorescence), changes during course of disease, post mortem examination; treatment of one case with glucantime, effect on clinical data

L[eishmania] donovani
Schnur, L. F., 1977, J. Protozool., v. 24 (2), Suppl., 5A-6A [Abstract]
Leptomonas, Crithidia, Herpetomonas, L[eishmania] tropica, L[eishmania] tropica major, L[eishmania] tropica minor, L[eishmania] donovani, excreted metabolic factors (EF) and homologous antisera, cross-reactions, no EF apparently produced by Trypanosoma cruzi, suggests closer relationship between insect flagellates and leishmanias than between these and trypanosomes like Trypanosoma cruzi

Leishmania donovani
Schnur, L. F.; and El-On, J., 1974, J. Protozool., v. 21 (3), 465-466 [Abstract]
Leishmania donovani, L. tropica, evidence of haptenic nature of leishmanial excreted factors

Leishmania donovani
Schnur, L. F.; and Zuckerman, A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 434-435 [Demonstration]
Leishmania spp. from Sudan, Ethiopia and Kenya, differentiation of leishmanial serotypes

Leishmania donovani
Schnur, L. F.; and Zuckerman, A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 273-294
Sudanese, Kenyan and Ethiopian leishmanial isolates from human visceral and cutaneous infections, various wild animals and sandflies, leishmanial excreted factor (EF) serotypes determined by gel diffusion, discussion of distribution and relationships

Leishmania donovani
Semprevivo, L. H., 1975, J. Protozool., v. 22 (3), 17A [Abstract]
Leishmania donovani, technique for study of amastigote cell cycle in mice

Leishmania donovani
Smrkovski, L. L.; and Larson, C. L., 1977, Infect. and Immun., v. 16 (1), 249-257
Leishmania donovani, BALB/c mice, BCG immunization, prophylactic and therapeutic effect, influence of timing and of route of injection

Leishmania donovani
Smrkovski, L. L.: and Larson, C. L., 1977, Infect. and Immun., v. 18 (2), 561-562
Leishmania donovani, antigenic cross-reactivity with Mycobacterium bovis demonstrated using delayed hypersensitivity as a criterion

Leishmania donovani
Steiger, R. F.; and Meshnick, S. R., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 441-443
Leishmania donovani, L. braziliensis, amino-acid and glucose utilization by parasites

Leishmania donovani
Steiger, R. F.; and Steiger, E., 1976, J. Parasitol., v. 62 (6), 1010-1011
Leishmania donovani, L. braziliensis, chemically defined medium for in vitro cultivation

Leishmania donovani
Steiger, R. F.; and Steiger, E., 1977, J. Protozool., v. 24 (3), 437-441
Leishmania donovani, L. brasiliensis, promastigotes, nutritional requirements studied in modifications of simple defined culture medium, special emphasis placed on glucose and amino acid requirements

Leishmania donovani
Szarfman, A.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 19-24
evaluation of specificity of EVI factor observed in Trypanosoma cruzi cases using indirect fluorescent antibody test on sera from patients with other parasitic diseases, immunofluorescence of anti-skeletal muscle antibody from leishmaniasis

Leishmania donovani
Trigari, G.; et al., 1974, Atti Soc. Ital. Sc. Vet., v. 28, 846-849
Leishmania donovani, Leishmania tropica, analysis of phospholipids, comparison

Leishmania donovani, illus.
Tryphonas, L.; et al., 1977, Canad. J. Comp. Med., v. 41 (1), 1-12
Leishmania donovani, dog, hematological and pathological observations, significance of bone marrow evaluation and electron microscopic examination in diagnosis, potential public health hazard for North America: Canada, imported from Spain

Leishmania donovani
Voller, A.; Bartlett, A.; and Bidwell, D. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 98-106
application of enzyme immunoassay to diagnosis of human parasitic diseases, technique for microplate enzyme-linked immunosorbent assay

Leishmania donovani
Weissberger, H.; Spira, D. T.; and Zuckerman, A., 1973, J. Protozool., v. 20 (4), 534-535
Leishmania enriettii-infected guinea pigs, delayed hypersensitivity reactions to homologous and heterologous (L. tropica, L. donovani, L. tarentolae) antigens, skin test, blast transformation, macrophage migration inhibition

Leishmania donovani
Zivkovic, V.; and Miscevic, Z., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 107-116
survey of sandflies, Phlebotomus major considered to be major vector of Leishmania donovani: Montenegro and Dalmatia, Yugoslavia

Leishmania donovani chagasi
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania donovani chagasi
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania enrietti
Alexander, J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 10 [Demonstration]
unusual arrangements of microtubules in flagellar axoneme of Leishmania enrietti and L. mexicana not present in L. tropica major, possible use as taxonomic marker for differentiation between New and Old World species

Leishmania enriettii
Ardehali, S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 266-267 [Demonstration]
Leishmania enriettii, L. tropica major, culture forms, possible species differentiation by enumeration of free moving promastigotes growing in presence of rabbit antisera, trials with homologous and heterologous antisera

Leishmania enrietti
Behforouz, N.; Rezai, H. R.; and Gettner, S., 1976, Ann. Trop. Med. and Parasitol., v. 70 (3), 293-301
Leishmania spp., immunofluorescence used to detect antibodies in humans, mice and guinea pigs using heterologous and homologous antigens; immunoelectrophoretic identification of active fraction of guinea pig antibody

Leishmania enriettii
Behin, R.; Mauel, J.; and Rowe, D. S., 1977, Clin. and Exper. Immunol., v. 29 (2), 320-325
Leishmania enriettii, guinea pigs, inhibition of leishmanial lesion by delayed hypersensitivity reaction to unrelated antigens, concluded that cell-mediated immunity plays important role in healing leishmanial lesions

Leishmania enriettii
Belehu, A.; Poulter, L. W.; and Turk, J. L., 1976, Clin. and Exper. Immunol., v. 24 (1), 125-132
Leishmania enriettii, guinea pigs, pretreatment with cyclophosphamide, increased intensity of initial lesion and increased incidence of widespread metastases, decreased levels of circulating antibody, possible differential roles of cell-mediated immunity and humoral antibody in cutaneous leishmaniasis

Leishmania enriettii
Belehu, A.; Poulter, L. W.; and Turk, J. L., 1976, Infect. and Immun., v. 14 (2), 457-462
hamsters, effect of Plasmodium berghei or P. yoelii malaria on concurrent Leishmania enriettii infection depended on relative timing of two infections

Leishmania enriettii
Belehu, A.; and Turk, J. L., 1976, Infect. and Immun., v. 13 (4), 1235-1241
Leishmania enriettii, establishment of self-healing type of cutaneous leishmaniasis in hamsters, course of infection, immunopathological response, useful model

Leishmania enriettii
Blewett, T. M.; Kadivar, D. M. H.; and Soulsby, E. J. L., 1971, Am. J. Trop. Med. and Hyg., v. 20 (4), 546-551
cell-mediated immunity in cutaneous leishmaniasis in guinea pig, mitogenic stimulation by antigens on peripheral lymphocytes and inhibition of macrophage migration

Leishmania enriettii, illus.
Bryceson, A. D. M., 1975, Symposia Brit. Soc. Parasitol., v. 13, 85-100
mechanisms of disease in leishmaniasis, extensive review with some previously unpublished results: host-parasite specificity; prevention or evasion of immune response; role of immune response in production of disease; healing; immunity to reinfection

Leishmania enriettii
Bryceson, A. D. M.; Bray, R. S.; and Dumonde, D. C., 1974, Clin. and Exper. Immunol., v. 16 (2), 189-201
Leishmania enriettii, guinea pigs inoculated with graded doses, relationship between clinical course of infection and immunological response, selective suppression of cell-mediated immunity, extent of delayed hypersensitivity closely related to degree of host resistance, role of humoral antibody less clear

Leishmania enriettii
Chance, M. L.; and Gardener, P. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 9 [Demonstration]
Leishmania spp., morphological and biochemical differentiation into 4 significantly different groups within the genus

Leishmania enriettii
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania enriettii
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania enriettii
Childs, G. E.; McRoberts, M. J.; and Foster, K. A., 1976, J. Parasitol., v. 62 (5), 676-679
Leishmania spp. amastigotes, recovery from cutaneous lesions of host animals by a method adapted from primary tissue culture techniques, pathogenic characteristics retained, comparatively free of host cells

Leishmania enriettii
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania enriettii, illus.
Gardener, P. J.; Shchory, L.; and Chance, M. L., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 147-155
Leishmania species differentiation by morphometric comparisons of ultrastructural features of leishmanial amastigotes

Leishmania enriettii
Ghose, A. C., 1976, Experientia, v. 32 (8), 1059-1061
sera from guinea pigs infected with Leishmania enriettii showed higher hemagglutination titres for neuraminidase-treated human erythrocytes than those of normal guinea pigs

Leishmania enriettii
Ghose, A. C.; and Rowe, D. S., 1977, Immunochemistry, v. 14 (6), 459-465
Leishmania enriettii, subcellular fractionation, antigenic activity of fractions determined by micro-complement fixation, indirect radioimmunoassay, skin testing, and in vitro lymphocyte transformation, results indicate antigenic heterogeneity and suggest that major humoral and cell-mediated components of immune response in infected guinea-pigs are directed against different antigenic determinants of the parasite

Leishmania enriettii Muniz & Medina, illus.
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex

Leishmania enriettii
Lewis, D. H.; and Peters, W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 295-312
Leishmania spp., promastigote forms, relationship between parasites and host macrophages and their relevance to intracellular survival of parasites, resistance of intracellular Leishmania to digestion by lysosomal enzymes

Leishmania enriettii
Matossian-Rogers, A.; Lumsden, W. H. R.; and Dumonde, D. C., 1976, Immunology, v. 31 (1), 1-19
Leishmania enriettii, L. tropica major, L. aethiopica, L. mexicana amazonensis, numerical immunotaxonomy, differentiation according to reactivity and cross-reactivity in tests of parasite agglutination, indirect immunofluorescence, and passive cutaneous anaphylaxis

Leishmania enriettii
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania enriettii Muniz and Medina 1948
Medina, H.; and Bacila, M., 1955, Arq. Biol. e Tecn., v. 10, 81-96
Leishmania brasiliensis, L. enriettii, cultured in new diphasic medium containing spinach and glucose; L. enriettii grew only in medium containing added guinea-pig blood

Leishmania enrietti
Neal, R. A.; and Miles, R. A., 1976, J. Trop. Med. and Hyg., v. 79 (2), 32-37
recommendations for use of various leishmanin antigens in field surveys and seroepidemiologic study based on guinea pig reactions to Leishmania enrietti

Leishmania enriettii
Neal, R. A.; and Miles, R. A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 21-27
Leishmania enriettii in guinea pigs (exper.), relationships between sodium stibogluconate therapy and developing immunity in host

Leishmania enriettii
Pardoe, G. I.; et al., 1975, Behring Inst. Mitt. (58), 30-39
Leishmania enriettii, immunochemistry of surface antigens

Leishmania enriettii
Poulter, L. W., 1976, Cellular Immunol., v. 27 (1), 17-25
Leishmania enriettii, guinea pigs, effect of lymphokine contact in vitro on level of glucose oxidation and migrating ability of macrophages taken from animals during infection and after re-infection, relation to role of macrophages in recovery from infection and subsequent immunity

Leishmania enriettii, illus.
Radwanski, Z. K.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 124-132
Leishmania enriettii in guinea pigs (exper.), use of indirect immunofluorescence to demonstrate antibody response during infection, to detect presence of parasites, gamma-globulin and complement during active infection and during healing process; antibody detected in primary lesions unrelated to healing process or pathogenesis of tissue damage

Leishmania enriettii
Rezai, H. R.; Ardehali, S.; and Gettner, S., 1975, Ann. Trop. Med. and Parasitol., v. 69 (1), 29-33
Leishmania enriettii, L. tropica, quantification of growth inhibition activity of normal sera from humans, dogs, rabbits, guinea pigs, rats, sheep, cats, mice, and chickens

Leishmania enriettii
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Leishmania enriettii
Weissberger, H.; Spira, D. T.; and Zuckerman, A., 1973, J. Protozool., v. 20 (4), 534-535
Leishmania enriettii-infected guinea pigs, delayed hypersensitivity reactions to homologous and heterologous (L. tropica, L. donovani, L. tarentolae) antigens, skin test, blast transformation, macrophage migration inhibition

Leishmania gondii Nicolle and Manceaux 1908
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Leishmania gondii Nicolle & Manceaux, 1908
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Leishmania gondii Nicolle & Manceaux, 1908
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora gondii (Nicolle & Manceaux, 1908) comb. nov.

Leishmania hertigi
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania hertigi
Croft, S. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 13 [Demonstration]
Leishmania hertigi, electron microscopy of mitotic spindle in promastigote

Leishmania hertigi
Croft, S. L., 1977, J. Protozool., v. 24 (4), 42A-43A [Abstract]
Leishmania hertigi, nucleus, fine structure

Leishmania hertigi
Croft, S. L.; and Schnur, L. F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 114 [Demonstration]
Leishmania b. braziliensis, L. hertigi, morphological comparisons of excreted factor-producing strain and non-excreted factor-producing strain, possible relationships between excreted factor and Noguchi-Adler phenomenon

Leishmania hertigi
Croft, S. L.; Schnur, L. F.; and Chance, M. L., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 113 [Demonstration]
Leishmania hertigi, morphologic and biochemical characterization of 8 strains

Leishmania hertigi
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania hertigi
Molyneux, D. H., 1976, Ztschr. Parasitenk., v. 48 (3-4), 296 [Abstract]
Leishmania hertigi, virus-like particles in promastigotes, brief description

Leishmania hertigi, illus.
Zeledon, R.; Ponce, C.; and de Ponce, E., 1977, J. Parasitol., v. 63 (5), 924-925
Coendou mexicanum laenatum: Naranjal de Acosta, Province of San Jose, Costa Rica
hamster (exper.)

Leishmania hertigi deanei Lainson & Shaw
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex

Leishmania hertigi hertigi Herrer
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex

Leishmania hoogstraali
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania hoogstraali
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania hoogstraali
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania hoogstraali McMillan
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex

Leishmania hoogstraali
Lewis, D. H., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 327-336
Leishmania mexicana mexicana, infection of dog sarcoma cells in presence of cytochalasin B indicates parasites can infect host cells without agency of active phagocytosis by latter; differing abilities of various Leishmania spp. promastigotes to infect same cell type suggest active contribution of parasites themselves to infection

Leishmania hoogstraali, illus.
Lewis, D. H., 1975, J. Protozool., v. 22 (3), 344-352
Leishmania spp., 4 reptilian vs. 1 mammalian isolates, ultrastructure of promastigotes, possible characters for use in taxonomy and species or strain differentiation

Leishmania hoogstraali
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania hoogstraali
Schnur, L. F.; and Zuckerman, A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 273-294
Sudanese, Kenyan and Ethiopian leishmanial isolates from human visceral and cutaneous infections, various wild animals and sandflies, leishmanial excreted factor (EF) serotypes determined by gel diffusion, discussion of distribution and relationships

Leishmania infantum
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania infantum
Busuttil, A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (3), 236-240
Leishmania infantum, incidence of human kala-azar in the Maltese Islands, 25-year review

Leishmania infantum
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania infantum
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

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susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania infantum
Schnur, L. F.; and Zuckerman, A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 434-435 [Demonstration]
Leishmania spp. from Sudan, Ethiopia and Kenya, differentiation of leishmanial serotypes

Leishmania major Yakimov, 1915 new rank
Bray, R. S.; Ashford, R. W.; and Bray, M. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 345-348
Syn.: Leishmania tropica major

Leishmania major
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania major
Sharma, J. K.; Anaraki, F.; and Ala, F., 1977, Scand. J. Immunol., v. 6 (11), 1101-1106
in vitro lymphoblast transformation of unsensitized lymphocytes to Leishmania major antigen in presence of leishmania-specific and -nonspecific transfer factor, results clearly substantiate in vitro specificity of transfer factor

Leishmania mexicana
Alexander, J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 6 [Demonstration]
Leishmania mexicana, inhibition of macrophage invasion using cytochalasin B

Leishmania mexicana
Alexander, J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 273 [Demonstration]
Leishmania mexicana, amastigotes in cultured mouse macrophages

Leishmania mexicana, illus.
Alexander, J., 1975, J. Protozool., v. 22 (2), 237-240
differential effect of antiphagocytic agent cytochalasin B on macrophage invasion by Leishmania mexicana promastigotes vs. Trypanosoma cruzi epimastigotes suggests that L. mexicana enters macrophages by being phagocytosed whereas T. cruzi can actively penetrate these cells

Leishmania mexicana
Alexander, J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 10 [Demonstration]
unusual arrangements of microtubules in flagellar axoneme of Leishmania enrietti and L. mexicana not present in L. tropica major, possible use as taxonomic marker for differentiation between New and Old World species

Leishmania mexicana
Alexander, J.; and Phillips, R. S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 273 [Demonstration]
Trypanosoma brucei in mice suppresses growth of cutaneous nodules caused by Leishmania spp., subcurative treatment of T. brucei with Berenil allowed nodule to recommence growth but when trypanosome infection relapsed the lesion again stopped growing, possible mechanism, no evidence of cross-immunity

Leishmania mexicana
Alexander, J.; and Phillips, R. S., 1975, J. Protozool., v. 22 (3), 48A [Abstract]
Leishmania mexicana, immunity in mice, cross-protection between L. tropica major and L. mexicana

Leishmania mexicana complex
Brazil, R. P., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 359 [Letter]
rebuttal to previously published letter on the identification of Brazilian Leishmania strains and on the use of the name L. braziliensis; need for thorough differentiation from L. mexicana complex

Leishmania mexicana
Chance, M. L.; and Gardener, P. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 9 [Demonstration]
Leishmania spp., morphological and biochemical differentiation into 4 significantly different groups within the genus

Leishmania mexicana
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania mexicana
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania mexicana
Childs, G. E.; McRoberts, M. J.; and Foster, K. A., 1976, J. Parasitol., v. 62 (5), 676-679
Leishmania spp. amastigotes, recovery from cutaneous lesions of host animals by a method adapted from primary tissue culture techniques, pathogenic characteristics retained, comparatively free of host cells

Leishmania mexicana
Decker, J. E.; and Daggett, P. M., 1974, J. Protozool., v. 21 (3), 429 [Abstract]
Leishmania donovani, L. mexicana, comparison of composition of excretion factors

Leishmania mexicana
Ebert, F., 1974, Tropenmed. u. Parasitol., v. 25 (3), 259-266
Leishmania spp. of the New World, comparative electrophoretic studies on proteins, esterases, relationships to Leishmania donovani and L. tropica, use of species-specific enzyme patterns in differentiating species and strains

Leishmania mexicana
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania mexicana (1746 isolate)
Gardener, P. J.; Shchory, L.; and Chance, M. L., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 147-155
Leishmania species differentiation by morphometric comparisons of ultrastructural features of leishmanial amastigotes

Leishmania mexicana
Jadin, J. M.; et al., 1977, J. Protozool., v. 24 (4), 61A [Abstract]
Kinetoplastidae, culture in mouse peritoneal macrophages

Leishmania mexicana
Janovy, J., jr.; Greiner, E. C.; and Decker, J. E., 1974, J. Protozool., v. 21 (3), 430 [Abstract]
Leishmania donovani vs. L. mexicana, anaerobic glycolysis and CO_2 fixation over the range 25-37 C, effect of pentostam vs. stibophen

Leishmania 'mexicana'
Kilgour, V.; et al., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 245-246
Leishmania spp., aminotransferases, demonstration of electrophoretic variation, potential for differentiating isolates and deriving taxonomic characters

Leishmania mexicana
Marr, J. J.; and Berens, R. L., 1977, J. Infect. Dis., v. 136 (6), 724-732
Leishmania spp., in vitro study of antileishmanial effects of allopurinol and its metabolic derivative oxipurinol, suggested sites of action, reversal of antileishmanial effects by adenine and its derivatives

Leishmania mexicana
Martin, E.; et al., 1976, J. Protozool., v. 23 (4), 600-607
Leishmania, 4 human spp. compared, promastigotes, occurrence and levels of activity of various enzymes of carbohydrate catabolism

Leishmania mexicana
Mattock, N. M.; and Peters, W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 22 [Demonstration]
Leishmania mexicana amastigotes in tissue culture cells as basis of technique for rapid screening of compounds for leishmanicidal activity

Leishmania mexicana
Neal, R. A., 1976, Ann. Trop. Med. and Parasitol., v. 70 (2), 252
Leishmania mexicana, mice (exper.), suppression of developing lesions by sodium stibogluconate but not by pyrimethamine

Leishmania mexicana
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Leishmania mexicana
Walton, B. C.; Intermill, R. W.; and Hajduk, M. E., 1977, Am. J. Trop. Med. and Hyg., v. 26 (5, part 1), 850-855
Leishmania isolates from patients with espundia compared with reference strain of L. b. braziliensis, clinical features, growth in culture, pathogenesis in hamster, location in gut of sandflies (Lutzomyia gomezi and L. trapidoi), autochthonous Texas strain clearly falls into L. mexicana complex, Peru isolate has intermediate pattern of characteristics but primarily those of L. braziliensis

Leishmania mexicana
Wilson, H. R.; and Fairchild, D. G., 1977, Stain Tech., v. 52 (2), 105-111
modified Whipf's polychrome staining to demonstrate protozoans and tissue components in cutaneous lesions of infected animals

Leishmania mexicana amazonensis
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania mexicana amazonensis
Chance, M. L.; and Gardener, P. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 9 [Demonstration]
Leishmania spp., morphological and biochemical differentiation into 4 significantly different groups within the genus

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Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), pp. 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania mexicana amazonensis
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania mexicana amazonensis, illus.
Childs, G. E.; McRoberts, M. J.; and Foster, K. A., 1976, J. Parasitol., v. 62 (5), 676-679
Leishmania spp. amastigotes, recovery from cutaneous lesions of host animals by a method adapted from primary tissue culture techniques, pathogenic characteristics retained, comparatively free of host cells

Leishmania mexicana amazonensis, illus.
Gardener, P. J., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 167-176
Trypanosoma cruzi, Leishmania spp., amastigotes, comparative electron microscope study, pellicle-associated structures

Leishmania mexicana amazonensis
Gardener, P. J.; and Chance, M. L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 23 [Demonstration]
Leishmania donovani, L. mexicana amazonensis, enzyme differences between life-cycle stages, malate dehydrogenase occurred in promastigote culture stage as different isoenzyme variants but not in amastigote intracellular form, lactate dehydrogenase could not be demonstrated in either stage

Leishmania mexicana amazonensis
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania mexicana amazonensis
Gardener, P. J.; Howells, R. E.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 23 [Demonstration]
Leishmania tropica major, L. mexicana amazonensis, L. chagasi, amastigotes, comparison revealed no distinct ultrastructural characteristics attributable to any of the strains studied

Leishmania mexicana amazonensis (M1845 isolate), illus.
Gardener, P. J.; Shchory, L.; and Chance, M. L., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 147-155
Leishmania species differentiation by morphometric comparisons of ultrastructural features of leishmanial amastigotes

Leishmania mexicana amazonensis
Kilgour, V.; et al., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 245-246
Leishmania spp., aminotransferases, demonstration of electrophoretic variation, potential for differentiating isolates and deriving taxonomic characters

Leishmania mexicana amazonensis
Killick-Kendrick, R.; et al., 1977, Proc. Roy. Soc., London, s. B, Biol. Sc. (1122), v. 196, 105-115
Leishmania mexicana amazonensis, cyclic transmission to hamsters by bite of infected Lutzomyia longipalpis (exper.), ultrastructure of L. mexicana amazonensis in Lutzomyia foregut may explain difficulty of obtaining regular transmissions

Leishmania mexicana amazonensis
Killick-Kendrick, R.; Molyneux, D. H.; and Ashford, R. W., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 269 [Demonstration]
Leishmania mexicana amazonensis, ultrastructure, attachment in mid-gut and proventricular fold of experimentally infected Lutzomyia longipalpis

Leishmania mexicana amazonensis
Killick-Kendrick, R.; Sinden, R. E.; and Leaney, A. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 432 [Demonstration]
Leishmania mexicana amazonensis, scanning electron microscopy of promastigotes in midgut of experimentally infected Lutzomyia longipalpis

Leishmania mexicana amazonensis
Lainson, R.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 184-196
Oryzomys macconnelli: Serra dos Carajas, Para, north Brazil

Leishmania mexicana amazonensis
Lainson, R.; and Shaw, J. J., 1977, J. Trop. Med. and Hyg., v. 80 (2), 29-35
cross-immunity trials with Cebus apella apella and observations on natural and experimental human infections confirm separate identity of Leishmania mexicana mexicana, L. m. amazonensis, L. braziliensis braziliensis, L. b. guyanensis, and L. b. panamensis

Leishmania mexicana amazonensis
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex

Leishmania mexicana amazonensis
Leaney, A. J., 1977, Parasitology, v. 75 (2), xxviii-xxix [Abstract]
Leishmania mexicana amazonensis in Lutzomyia longipalpis (exper.) maintained at 3 different temperatures (22, 24, and 28 °C.), distribution of parasites and degree of infection at 24h intervals from 2 to 8 days after infective feed compared, appears that small changes in temperature may have marked effect on development and behavior of Leishmania in sandflies and therefore on transmission

Leishmania mexicana amazonensis, illus.
Lewis, D. H.; and Peters, W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 295-312
Leishmania spp., promastigote forms, relationship between parasites and host macrophages and their relevance to intracellular survival of parasites, resistance of intracellular Leishmania to digestion by lysosomal enzymes

Leishmania mexicana amazonensis
Matossian-Rogers, A.; Lumsden, W. H. R.; and Dumonde, D. C., 1976, Immunology, v. 31 (1), 1-19
Leishmania enriettii, L. tropica major, L. aethiopica, L. mexicana amazonensis, numerical immunotaxonomy, differentiation according to reactivity and cross-reactivity in tests of parasite agglutination, indirect immunofluorescence, and passive cutaneous anaphylaxis

Leishmania mexicana amazonensis
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania mexicana amazonensis
Schnur, L. F., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 277-278 [Demonstration]
Leishmania braziliensis braziliensis (slow strain), L. mexicana amazonensis (fast strain), dissemination in hamsters

Leishmania mexicana amazonensis, illus.
Shaw, J. J.; and Lainson, R., 1977, J. Parasitol., v. 63 (2), 384-385
Leishmania mexicana amazonensis, simply prepared amastigote antigen for use in indirect fluorescent antibody test

Leishmania mexicana amazonensis Lainson & Shaw
Ward, R. D., 1977, Proc. 15. Internat. Cong. Entom. (Washington, D.C., Aug. 19-27, 1976), 505-522
New World leishmaniasis, incidence, factors causing epidemiological changes in last 3 decades

Leishmania mexicana amazonensis
Ward, R. D.; Lainson, R.; and Shaw, J. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 608-609 [Letter]
Leishmania mexicana amazonensis, further evidence of role of Lutzomyia flaviscutellata as vector in Brazil

Leishmania mexicana amazonensis Lainson & Shaw
Ward, R. D.; Lainson, R.; and Shaw, J. J., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (3), 265-266
Leishmania mexicana amazonensis, disease transmission from infected to non-infected guinea pigs (exper.) by the bite of Lutzomyia flaviscutellata (exper.); increased evidence for L. flaviscutellata as major vector in nature

Leishmania mexicana goularti nov. s.sp. [nom. nud.]
Brazil, R. P., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 340 [Letter]
classification of strain No. 4 isolated in 1960 from human with chronic nodular diffuse leishmaniasis: Fundacao Goncalo Muniz (Bahia)

Leishmania mexicana mexicana
Alexander, J.; and Vickerman, K., 1975, J. Protozool., v. 22 (4), 502-508
Leishmania mexicana mexicana-infected mouse peritoneal macrophages, fusion of host cell secondary lysosomes with parasitophorous vacuole, may be pathway for nutrients, drugs, antibodies

Leishmania mexicana mexicana
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
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Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

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Leishmania spp., aminotransferases, demonstration of electrophoretic variation, potential for differentiating isolates and deriving taxonomic characters

Leishmania mexicana mexicana
Lainson, R.; and Shaw, J. J., 1977, J. Trop. Med. and Hyg., v. 80 (2), 29-35
cross-immunity trials with Cebus apella apella and observations on natural and experimental human infections confirm separate identity of Leishmania mexicana mexicana, L. m. amazonensis, L. braziliensis braziliensis, L. b. guyanensis, and L. b. panamensis

Leishmania mexicana mexicana
Lainson, R.; Ward, R. D.; and Shaw, J. J., 1977, Proc. Roy. Soc. London, s. B, Biol. Sc. (1135), v. 199, 309-320
Leishmania spp., presence or absence of development in hindgut of sandfly Lutzomyia longipalpis (exper.), importance of hindgut development in distinguishing between parasites of Leishmania mexicana and L. braziliensis complex

Leishmania mexicana mexicana, illus.
Lewis, D. H., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 327-336
Leishmania mexicana mexicana, infection of dog sarcoma cells in presence of cytochalasin B indicates parasites can infect host cells without agency of active phagocytosis by latter; differing abilities of various Leishmania spp. promastigotes to infect same cell type suggest active contribution of parasites themselves to infection

Leishmania mexicana mexicana
Lewis, D. H., 1975, J. Protozool., v. 22 (3), 53A [Abstract]
Leishmania mexicana mexicana, in vitro studies on intracellular relationship with lysosomal system of normal vs. sensitized host macrophages

Leishmania mexicana mexicana, illus.
Lewis, D. H., 1975, J. Protozool., v. 22 (3), 344-352
Leishmania spp., 4 reptilian vs. 1 mammalian isolates, ultrastructure of promastigotes, possible characters for use in taxonomy and species or strain differentiation

Leishmania mexicana mexicana
Lewis, D. H.; and Besso, A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (2), 113-114 [Demonstration]
Leishmania m. mexicana, possible inactivation of host cell lysosomal enzyme activity within parasitophorous vacuoles by intracellular Leishmania in order to avoid digestion

Leishmania mexicana mexicana, illus.
Lewis, D. H.; and Peters, W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 295-312
Leishmania spp., promastigote forms, relationship between parasites and host macrophages and their relevance to intracellular survival of parasites, resistance of intracellular Leishmania to digestion by lysosomal enzymes

Leishmania mexicana mexicana
Mattock, N. M., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 345-348
Leishmania mexicana mexicana and L. donovani amastigote forms grown in cell culture, failure to detect lactate dehydrogenase and malate dehydrogenase, presence of glucose-6-phosphate dehydrogenase indicated

Leishmania mexicana mexicana
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania mexicana mexicana, illus.
Mattock, N. M; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 349-357
Leishmania mexicana mexicana, L. tropica major, L. donovani amastigotes in dog sarcoma cells and in hamster peritoneal exudate cell line, techniques for studying sensitivity of parasites and hosts to drugs, concluded that tissue culture system is valid way of determining baseline sensitivity of a Leishmania to chemotherapy, illustrative data with amphotericin B and 7'-trifluormethyl-dihydrocinchonine dihydrochloride

Leishmania mexicana mexicana
Mattock, N. M.; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 359-371
Leishmania spp. in tissue culture model system, variety of compounds used in treatment of parasitic or bacterial infections tested for activity, concluded that model may be of use in screening of drugs for leishmanicidal action

Leishmania mexicana mexicana
Mattock, N. M.; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 449-462
Leishmania spp. in tissue culture model system, leishmanicidal action of over 100 compounds examined

Leishmania mexicana mexicana Biagi
Ward, R. D., 1977, Proc. 15. Internat. Cong. Entom. (Washington, D.C., Aug. 19-27, 1976), 505-522
New World leishmaniasis, incidence, factors causing epidemiological changes in last 3 decades

Leishmania mexicana pifanoi Medina & Romero
Ward, R. D., 1977, Proc. 15. Internat. Cong. Entom. (Washington, D.C., Aug. 19-27, 1976), 505-522
New World leishmaniasis, incidence, factors causing epidemiological changes in last 3 decades

Leishmania peruviana
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania peruviana
Ward, R. D., 1977, Proc. 15. Internat. Cong. Entom. (Washington, D.C., Aug. 19-27, 1976), 505-522
New World leishmaniasis, incidence, factors causing epidemiological changes in last 3 decades

Leishmania pifanoi
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania pifanoi (Medina und Romero, 1962)
Ebert, F., 1974, Tropenmed. u. Parasitol., v. 25 (3), 259-266
Leishmania spp. of the New World, comparative electrophoretic studies on proteins, esterases, relationships to Leishmania donovani and L. tropica, use of species-specific enzyme patterns in differentiating species and strains

Leishmania pifanoi
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania tarentolae
Berens, R. L.; Brun, R.; and Krassner, S. M., 1976, J. Parasitol., v. 62 (3), 360-365
Leishmania donovani, simple monophasic medium for axenic culture which favors amastigote to promastigote transformation, promastigote size distribution and glucose consumption during cell cycle; medium also supports high population growth of L. tarentolae promastigotes and Trypanosoma cruzi (Costa Rica but not Corpus Christi strain) epimastigotes

Leishmania tarentolae, illus.
Braly, P.; Simpson, L.; and Kretzer, F., 1974, J. Protozool., v. 21 (5), 782-790
Leishmania tarentolae, kinetoplast-mitochondrial complexes, isolation technique, partial characterization

Leishmania tarentolae
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania tarentolae
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania tarentolae
Englund, P. T.; DiMaio, D. C.; and Price, S. S., 1977, J. Biol. Chem., v. 252 (17), 6208-6216
Leishmania tarentolae, nicked form of kinetoplast DNA

Leishmania tarentolae
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania tarentolae
Leon, W.; Brun, R.; and Krassner, S. M., 1976, J. Protozool., v. 23 (2), 7A [Abstract]
Leishmania tarentolae promastigotes, Berenil, induction of dyskinetoplasty, inhibition of cell growth

Leishmania tarentolae, illus.
Leon, W.; Brun, R.; and Krassner, S. M., 1977, J. Protozool., v. 24 (3), 444-448
Leishmania tarentolae, effect of Berenil on growth, on buoyant density of kinetoplast DNA, on dyskinetoplasty at ultrastructural level, and on cell respiration, results suggest that Berenil adversely affects mitochondrial respiratory activity

Leishmania tarentolae
Leon, W.; and Krassner, S. M., 1976, J. Parasitol., v. 62 (1), 115
Leishmania tarentolae, promastigotes, inhibition of cell growth by 2 carcinogenic nitroquinoline derivatives

Leishmania tarentolae
Mattock, N. M.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 18 [Demonstration]
susceptibility of dog sarcoma and hamster peritoneal cells to various Leishmania spp.

Leishmania tarentolae
Pereira, N. M.; et al., 1977, J. Protozool., v. 24 (4), 511-514
Crithidia fasciculata, Herpetomonas samuelpessoai, Leishmania tarentolae, isolation of flagella, 3 types of flagella give similar electrophoretic pattern of proteins, H. samuelpessoai and to a lesser extent C. fasciculata flagella confer protection against Trypanosoma cruzi infections in mice

Leishmania tarentolae
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Leishmania tarentolae
Sheets, E. G.; and Krassner, S. M., 1974, J. Protozool., v. 21 (5), 742-744
Leishmania tarentolae in chemically defined medium containing taurine as sole sulfur source, chromate, selenate, and vanadate accelerated growth and enhanced incorporation of [^{35}S] taurine label into different cell fractions

Leishmania tarentolae
Simpson, A. M.; and Simpson, L., 1976, J. Protozool., v. 23 (4), 583-587
Crithidia fasciculata, Leishmania tarentolae, pulse-labeling of kinetoplast DNA, localization of 2 sites of synthesis within networks and kinetics of labeling of closed minicircles

Leishmania tarentolae, illus.
Simpson, L., 1973, J. Protozool., v. 20 (1), 2-8
salivarian and stercorarian trypanosomes, kinetoplast DNA, structure and function, review; brief mention of some unpublished results on biophysical properties of L. tarentolae minicircles

Leishmania tarentolae, illus.
Simpson, L.; and Berliner, J., 1974, J. Protozool., v. 21 (2), 382-393
Leishmania tarentolae, technique for isolation of kinetoplast DNA networks, preliminary physical characterization

Leishmania tarentolae
Wagner, K. P.; and Krassner, S. M., 1976, Exper. Parasitol., v. 39 (2), 186-194
Leishmania tarentolae, promastigotes, substances in Trager's Defined Medium capable of acting as proline precursors, proline synthesis unaffected by presence of preformed proline indicating absence of end product feedback inhibition

Leishmania tarentolae
Wagner, K. P.; and Krassner, S. M., 1976, Exper. Parasitol., v. 39 (2), 222-233
Leishmania tarentolae promastigotes, replica-plating techniques, isolation of stable mutant strains resistant to chloramphenicol, isolation of cell lines stress-adapted to streptomycin and to high culture temperatures, factors influencing resistance, mode of action of chloramphenicol (inhibition of protein synthesis and proline oxidation)

Leishmania tarentolae
Weissberger, H.; Spira, D. T.; and Zuckerman, A., 1973, J. Protozool., v. 20 (4), 534-535
Leishmania enriettii-infected guinea pigs, delayed hypersensitivity reactions to homologous and heterologous (L. tropica, L. donovani, L. tarentolae) antigens, skin test, blast transformation, macrophage migration inhibition

Leishmania tropica
Alexander, J.; and Phillips, R. S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 273 [Demonstration]
Trypanosoma brucei in mice suppresses growth of cutaneous nodules caused by Leishmania spp., subcurative treatment of T. brucei with Berenil allowed nodule to recommence growth but when trypanosome infection relapsed the lesion again stopped growing, possible mechanism, no evidence of cross-immunity

Leishmania tropica
Behforouz, N.; Rezai, H. R.; and Gettner, S., 1976, Ann. Trop. Med. and Parasitol., v. 70 (3), 293-301
Leishmania spp., immunofluorescence used to detect antibodies in humans, mice and guinea pigs using heterologous and homologous antigens; immunoelectrophoretic identification of active fraction of guinea pig antibody

Leishmania tropica
Borgatti, A. R.; et al., 1974, Atti Soc. Ital. Sc. Vet., v. 28, 849-854
Leishmania donovani (3 strains), Leishmania tropica, analyses of long chain fatty acids, non polar lipids, phospholipids and phospholipid fractions, comparisons, possible use in evolutionary studies

Leishmania tropica
Bray, R. S., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 65-77
zoonoses in leishmaniasis, review

Leishmania tropica
Bryceson, A. D. M., 1975, Symposia Brit. Soc. Parasitol., v. 13, 85-100
mechanisms of disease in leishmaniasis, extensive review with some previously unpublished results: host-parasite specificity; prevention or evasion of immune response; role of immune response in production of disease; healing; immunity to reinfection

Leishmania tropica
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania tropica
Decker, J. E.; and Schrot, J. R., 1976, J. Protozool., v. 23 (2), 18A-19A [Abstract]
Leishmania donovani, L. braziliensis, L. tropica, rapid identification by radiorespirometry

Leishmania tropica
Decker, J. E.; Schrot, J. R.; and Levin, G. V., 1977, J. Protozool., v. 24 (3), 463-470
Leishmania spp., radiorespirometric technique has potential for rapid identification of species and strains

Leishmania tropica
Ebert, F., 1974, Tropenmed. u. Parasitol., v. 25 (1), 49-53
Leishmania tropica strains, electrophoretic patterns for proteins and enzymes which made characterization of strains possible, taxon-specific esterase bands which could be used in differentiating from L. donovani

Leishmania tropica
Ebert, F., 1974, Tropenmed. u. Parasitol., v. 25 (3), 259-266
Leishmania spp. of the New World, comparative electrophoretic studies on proteins, esterases, relationships to Leishmania donovani and L. tropica, use of species-specific enzyme patterns in differentiating species and strains

Leishmania tropica
El-On, J.; and Schnur, L. F., 1974, J. Protozool., v. 21 (3), 463-464 [Abstract]
Leishmania donovani, L. tropica, excreted antigens, purification and preliminary characterization

Leishmania tropica
Farah, F. S.; Lazary, S.; and De Weck, A., 1976, Immunology, v. 30 (5), 629-634
Leishmania tropica and its products are capable of inhibition of the stimulation of normal mouse and guinea-pig lymphocytes by phytohaemagglutinin, inhibition is dose-dependent and not dependent on competition for nutrients in medium nor on neutralization of phytohaemagglutinin, inhibition observed on lymphocytes of species susceptible to leishmanial infection but not operative in resistant species

Leishmania tropica
Handman, E.; et al., 1974, J. Biol. Standardization, v. 2 (3), 223-229
Leishmania tropica vaccine, production and properties, standardization and quality control

Leishmania tropica
Handman, E.; et al., 1977, J. Protozool., v. 24 (2), Suppl., 20A-21A [Abstract]
Leishmania tropica, mice, protection by vaccination with non-living antigenic preparation

Leishmania tropica
Handman, E.; Spira, D. T.; and Zuckerman, A., 1977, J. Protozool., v. 24 (2), Suppl., 21A [Abstract]
Leishmania tropica amastigotes, growth in macrophages from normal and immune mice

Leishmania tropica, illus.
Hentzer, B.; and Kobayasi, T., 1977, Acta Path. et Microbiol., Scand., v. 85B (2), 153-160
Leishmania tropica, amastigote stage in skin lesions, fine structure of basal body, prokinosome and flagellum

Leishmania tropica, illus.
Hentzer, B.; and Kobayasi, T., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 157-166
Leishmania tropica, effects of pentamidine therapy on fine structure of amastigote stage

Leishmania tropica
Inferrera, C.; et al., 1973, Morgagni, v. 6 (2), 15-22
Leishmania tropica, fine structure morphology; histopathology of host cutaneous lesions

Leishmania tropica
Jadin, J. M.; et al., 1977, J. Protozool., v. 24 (4), 61A [Abstract]
Kinetoplastidae, culture in mouse peritoneal macrophages

Leishmania tropica
Kalra, N. L.; and Lewis, D. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 522
Leishmania tropica, Phlebotomus salehi identified as probable vector in rodents: India

Leishmania tropica
Kilgour, V.; et al., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 245-246
Leishmania spp., aminotransferases, demonstration of electrophoretic variation, potential for differentiating isolates and deriving taxonomic characters

Leishmania tropica
Koufman, Z.; et al., 1977, J. Protozool., v. 24 (2), Suppl., 18A [Abstract]
Leishmania tropica, human, immunization trials: Israel

Leishmania tropica
Leon, L. A., 1975, Medicina, Mexico (1202), an. 61, v. 55, 370-376
epidemiology, diagnosis and treatment of furunculous lesions of tropical diseases, Trypanosoma cruzi, Leishmania tropica, Dermatobia hominis

Leishmania tropica
Martin, E.; et al., 1976, J. Protozool., v. 23 (4), 600-607
Leishmania, 4 human spp. compared, promastigotes, occurrence and levels of activity of various enzymes of carbohydrate catabolism

Leishmania tropica
Matossian, R. M.; Kurban, A. K.; and Malak, J. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 450-452
Leishmania tropica, humans, circulating antibodies, detection by immunofluorescence, cross reactivity with L. donovani and Trypanosoma cruzi antigens, possible cross reactivity in mixed tuberculosis infections

Leishmania tropica
Minoprio, J. L., 1972, Prensa Med. Argent., v. 59 (8), 293-308
human Chagas disease, pathologic findings, comparison with characteristics of other Trypanosoma spp. and Leishmania spp., extensive review

Leishmania tropica, illus.
Mishra, A. K.; and Lodha, K. R., 1977, Indian Vet. J., v. 54 (6), 475-476
Leishmania tropica, dogs (skin of nose), histopathology

Leishmania tropica
Morales, N. M.; et al., 1972, J. Protozool., v. 19 (4), 667-672
Leishmania tropica, effects of ethidium bromide and several acridine dyes on kinetoplast DNA

Leishmania tropica, illus.
Muehlpfordt, H., 1975, Tropenmed. und Parasitol., v. 26 (4), 385-389
Leishmania donovani, L. tropica, L. braziliensis, comparative electron morphology of developing promastigote forms using cationized ferritin to determine labelling of negative charges on cell surfaces

Leishmania tropica
Mukkada, A. J.; et al., 1974, J. Protozool., v. 21 (2), 393-397
Leishmania tropica, role of exogenous glucose in growth media, delayed utilization

Leishmania tropica
Mukkada, A. J.; and Simon, M. W., 1977, Exper. Parasitol., v. 42 (1), 87-96
Leishmania tropica promastigotes, properties of active transport system responsible for uptake and accumulation of L-methionine

Leishmania tropica
Oelerich, S., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 296-304
Leishmania spp., Trypanosoma cruzi, Crithidia sp., Mycobacterium smegmatis, antigenic analyses and cross reactions, double gel-diffusion, complement-fixation, indirect hemagglutination

Leishmania tropica
Pieragostini, E.; et al., 1974, Atti Soc. Ital. Sc., v. 28, 854-857
Leishmania donovani (3 strains), Leishmania tropica, analysis of sterols, comparison; cholesterol possibly from culture medium

Leishmania tropica
Preston, P. M.; and Dumonde, D. C., 1976, Clin. and Exper. Immunol., v. 23 (1), 126-138
Leishmania tropica in CBA mice as experimental model of leishmaniasis in man: relationship of inoculum dose to size and duration of lesions, antibody production, and delayed hypersensitivity responses; infections manifest both during and after healing stages; immunization with sonicated promastigotes; lymphoid cells from immune mice conferred protection upon recipients

Leishmania tropica
Rabinowitz, H.; and Spira, D. T., 1973, J. Protozool., v. 20 (4), 533
Leishmania tropica-infected macrophages, suppression of phagocytic activity, increased stimulation of lysosomal activity, increased acid phosphatase activity

Leishmania tropica
Ranque, P.; Quilici, M.; and Debaene, A., 1972, Medecine Trop., v. 32 (4), 499-503
Arvicanthis niloticus, exper. infections with varying strains of Leishmania tropica, L. donovani and Leishmania sp., minimal skin infections with no secondary visceral invasion

Leishmania tropica
Rezai, H. R.; Ardehali, S.; and Gettner, S., 1975, Ann. Trop. Med. and Parasitol., v. 69 (1), 29-33
Leishmania enriettii, L. tropica, quantification of growth inhibition activity of normal sera from humans, dogs, rabbits, guinea pigs, rats, sheep, cats, mice, and chickens

Leishmania tropica, illus.
Rodriguez-Gonzalez, C.; and Garcia-Segura, L. M., 1975, Rev. Iber. Parasitol., v. 35 (3-4), 301-309
Leishmania tropica, electron microscopy of flagellum and sub-membranous fibers, suggestion that fibers can modify relative position of cell body and flagellum, evidence for coordination of movement

Leishmania tropica, illus.
Sandbank, M., 1977, Brit. J. Dermat., v. 96 (6), 653-657
human cutaneous leishmaniasis, presence of banded cross-striated material discovered in proximity to macrophages containing leishmania, possibly the structures are indication of active, accelerated collagen production

Leishmania tropica
Schaefer, F. W. III; Martin, E.; and Mukkada, A. J., 1974, J. Protozool., v. 21 (4), 592-596
Leishmania tropica promastigotes, glucose transport system

Leishmania tropica
Schaefer, F. W. III; and Mukkada, A. J., 1976, J. Protozool., v. 23 (3), 446-449
Leishmania tropica promastigotes, specificity of glucose transport system

Leishmania tropica
Schnur, L. F., 1976, Ann. Trop. Med. and Parasitol., v. 70 (3), 371-373
Leishmania tropica, attempted vaccination of Syrian hamsters using live vaccine at low concentrations

L[eishmania] tropica
Schnur, L. F., 1977, J. Protozool., v. 24 (2), Suppl., 5A-6A [Abstract]
Leptomonas, Crithidia, Herpetomonas, L[eishmania] tropica, L[eishmania] tropica major, L[eishmania] tropica minor, L[eishmania] donovani, excreted metabolic factors (EF) and homologous antisera, cross-reactions, no EF apparently produced by Trypanosoma cruzi, suggests closer relationship between insect flagellates and leishmanias than between these and trypanosomes like Trypanosoma cruzi

Leishmania tropica
Schnur, L. F.; and El-On, J., 1974, J. Protozool., v. 21 (3), 465-466 [Abstract]
Leishmania donovani, L. tropica, evidence of haptenic nature of leishmanial excreted factors

Leishmania tropica
Schnur, L. F.; and Zuckerman, A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 434-435 [Demonstration]
Leishmania spp. from Sudan, Ethiopia and Kenya, differentiation of leishmanial serotypes

Leishmania tropica
Schnur, L. F.; and Zuckerman, A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 273-294
Sudanese, Kenyan and Ethiopian leishmanial isolates from human visceral and cutaneous infections, various wild animals and sandflies, leishmanial excreted factor (EF) serotypes determined by gel diffusion, discussion of distribution and relationships

Leishmania tropica
Simon, M. W.; and Mukkada, A. J., 1977, Exper. Parasitol., v. 42 (1), 97-105
Leishmania tropica promastigotes, methionine uptake system shows broad specificity and is subject to regulation at the level of carrier activity

Leishmania tropica
Simon, M. W.; Rusnak, J. M.; and Mukkada, A. J., 1976, Exper. Parasitol., v. 39 (1), 51-58
Leishmania tropica promastigotes in vitro, sensitivity to toxic effects of bilirubin (loss of viability, decreased sugar and amino acid uptake, increased efflux of intracellular sugars, hexokinase activity, lowered respiration), results suggest irreversible damage to cell membrane, possible culture loss if bilirubin concentration of hemoglobin solution used is too high

Leishmania tropica
Slutzky, G. M.; and Greenblatt, C. L., 1977, FEBS Letters, v. 80 (2), 401-404
Leishmania tropica, isolation of immunologically active factor from cultures, contains carbohydrate segment produced by parasite and protein segment incorporated from growth medium

Leishmania tropica
Slutzky, G. M.; and Greenblatt, C. L., 1977, J. Protozool., v. 24 (4), 67A [Abstract]
Leishmania tropica, isolation and characterization of immunologically active carbohydrate-protein complex from cultures

Leishmania tropica
Stadler, B.; et al., 1976, Ztschr. Immunitaetsforsch., v. 152 (2), 117-118 [Abstract]
Leishmania tropica, internal labelling with ^{3}H-thymidine in order to study attachment to vs. phagocytosis by macrophages in vitro under different conditions (with and without antibodies, activation of macrophages, etc.)

Leishmania tropica
Trigari, G.; et al., 1974, Atti Soc. Ital. Sc. Vet., v. 28, 846-849
Leishmania donovani, Leishmania tropica, analysis of phospholipids, comparison

Leishmania tropica
Weintraub, J.; and Weinbaum, F. I., 1977, J. Immunol., v. 118 (6), 2288-2290
Leishmania tropica, BCG-treated mice, reduction in severity of cutaneous disease and significant decrease in mortality without evidence of visceralization when compared with non-treated controls

Leishmania tropica
Weissberger, H.; Spira, D. T.; and Zuckerman, A., 1973, J. Protozool., v. 20 (4), 534-535
Leishmania enriettii-infected guinea pigs, delayed hypersensitivity reactions to homologous and heterologous (L. tropica, L. donovani, L. tarentolae) antigens, skin test, blast transformation, macrophage migration inhibition

Leishmania tropica major
Alexander, J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 10 [Demonstration]
unusual arrangements of microtubules in flagellar axoneme of Leishmania enrietti and L. mexicana not present in L. tropica major, possible use as taxonomic marker for differentiation between New and Old World species

Leishmania tropica major
Alexander, J.; and Phillips, R. S., 1975, J. Protozool., v. 22 (3), 48A [Abstract]
Leishmania mexicana, immunity in mice, cross-protection between L. tropica major and L. mexicana

Leishmania tropica major
Ardehali, S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 266-267 [Demonstration]
Leishmania enriettii, L. tropica major, culture forms, possible species differentiation by enumeration of free moving promastigotes growing in presence of rabbit antisera, trials with homologous and heterologous antisera

Leishmania tropica major Yakimov and Schokhor, 1914
Bejarano, J. F. R., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 522
Leishmania tropica major, L. tropica minor, named in accordance with Rules of Zoological Nomenclature

Leishmania tropica major
Bray, R. S.; Ashford, R. W.; and Bray, M. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 345-348
as syn. of Leishmania major Yakimov, 1915 new rank

Leishmania tropica major
Chance, M. L.; and Gardener, P. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 9 [Demonstration]
Leishmania spp., morphological and biochemical differentiation into 4 significantly different groups within the genus

Leishmania tropica major
Chance, M. L.; Peters, W.; and Griffiths, H. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 24-25 [Demonstration]
Leishmania spp., differences in DNA buoyant density

Leishmania tropica major
Chance, M. L.; Peters, W.; and Schchory, L., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 307-316
Leishmania spp., buoyant density of total cell DNA determined by isopycnic centrifugation in CsCl, wide spectrum of values, results discussed in terms of taxonomy; isolated kinetoplast DNA, minicircle contour length showed no evidence of species specificity

Leishmania tropica major, illus.
Gardener, P. J., 1974, Ann. Trop. Med. and Parasitol., v. 68 (2), 167-176
Trypanosoma cruzi, Leishmania spp., amastigotes, comparative electron microscope study, pellicle-associated structures

Leishmania tropica major
Gardener, P. J.; Chance, M. L.; and Peters, W., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 317-325
Leishmania spp., variation in electrophoretic mobility of malate dehydrogenase using disc polyacrylamide-gel method, results correlated with DNA buoyant density determinations on same isolates, preliminary summary of biochemical taxonomy presented

Leishmania tropica major
Gardener, P. J.; Howells, R. E.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 23 [Demonstration]
Leishmania tropica major, L. mexicana amazonensis, L. chagasi, amastigotes, comparison revealed no distinct ultrastructural characteristics attributable to any of the strains studied

Leishmania tropica major (P isolate), illus.
Gardener, P. J.; Shchory, L.; and Chance, M. L., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 147-155
Leishmania species differentiation by morphometric comparisons of ultrastructural features of leishmanial amastigotes

Leishmania tropica major
Matossian-Rogers, A.; Lumsden, W. H. R.; and Dumonde, D. C., 1976, Immunology, v. 31 (1), 1-19
Leishmania enriettii, L. tropica major, L. aethiopica, L. mexicana amazonensis, numerical immunotaxonomy, differentiation according to reactivity and cross-reactivity in tests of parasite agglutination, indirect immunofluorescence, and passive cutaneous anaphylaxis

Leishmania tropica major, illus.
Mattock, N. M; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 349-357
Leishmania mexicana mexicana, L. tropica major, L. donovani amastigotes in dog sarcoma cells and in hamster peritoneal exudate cell line, techniques for studying sensitivity of parasites and hosts to drugs, concluded that tissue culture system is valid way of determining baseline sensitivity of a Leishmania to chemotherapy, illustrative data with amphotericin B and 7'-trifluormethyl-dihydrocinchonine dihydrochloride

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Leishmania spp. in tissue culture model system, variety of compounds used in treatment of parasitic or bacterial infections tested for activity, concluded that model may be of use in screening of drugs for leishmanicidal action

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parasitic protozoa stored frozen in liquid nitrogen, survival records

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Leptomonas, Crithidia, Herpetomonas, L[eishmania] tropica, L[eishmania] tropica major, L[eishmania] tropica minor, L[eishmania] donovani, excreted metabolic factors (EF) and homologous antisera, cross-reactions, no EF apparently produced by Trypanosoma cruzi, suggests closer relationship between insect flagellates and leishmanias than between these and trypanosomes like Trypanosoma cruzi

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Leishmania, relationship between clinical types and serotypes not as simple and direct as previously thought

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Syn.: Leishmania tropica urbis

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Wolf, R. E., 1976, J. Parasitol., v. 62 (2), 209-214
Leishmania tropica yotvata, Macaca mulatta as possible simian model of human disease, primary, secondary, and tertiary infection, clinical resistance, cellular- and humoral-immune responses, effects of antilymphocyte globulin therapy, results quantitatively and qualitatively different from those in humans

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Leishmaniasis (possibly Leishmania donovani)
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parasites indistinguishable from those causing cutaneous leishmaniasis in man found in Procavia habessinica and Heterohyrax brucei and in Phlebotomus longipes and Phlebotomus pedifer (Leishmania aethiopica), close association between hyrax and sandflies, man is incidental host: Ethiopia

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Leptomonas of insect origin, cross-resistance patterns of clones to a variety of medical and veterinary trypanocides and leishmanicides, potential as a model screening system

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human cutaneous leishmaniasis of lupoid form, diagnostic difficulties, case report, glucantime

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light, immunofluorescent and electron microscopic study of pathogenetic mechanism of glomerular lesions found in human leishmaniasis, immune complexes trapped within glomerular capillaries

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leishmaniasis, bibliography (1860-1974): Algeria

Leishmaniasis
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human visceral and cutaneous leishmaniasis, survey of 3 endemic areas for possible phlebotomine sandflies; Phlebotomus perniciosus thought to be vector in Monte Argentarion and P. perfiliewi the vector in the hilly inland territory of the Province of Grosseto, Tuscany, Italy

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human visceral leishmaniasis, value of immunoprecipitation and immunofluorescence in diagnosis, immigrants into France from endemic areas

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Trypanosoma cruzi, leishmaniasis, human, comparison of skin test reactions

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description of human cutaneous leishmaniasis imported into the Netherlands from Surinam, pathology, treatment with pentostam and lomidine

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"The statement that P[hlebotomus] papatasi is the vector of kala-azar in Malta is thus obviously wrong"

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review of forms of human leishmaniasis, pathology with emphasis on ocular lesions, case reports: Tchad

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human visceral leishmaniasis, 20 fatal cases, histological appearances of spleen and lymph nodes were suggestive of profound disturbance in cell-mediated immunity, depletion of small lymphocytes in thymus-dependent areas accompanied by abundance of parasite-containing histiocytes and hyperplasia of plasma cells

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kala-azar, increase in transmission in Western Tharaka after evacuation of Eastern Tharaka, Phlebotomus martini and P. vansomerenae as probable vectors: Meru district, Kenya

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Ctenolepisma nigra (mid gut, ventricular caeca): Kalyani, West Bengal, India

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Crithidia fasciculata, purification and properties of soluble, NAD-linked α-glycerophosphate dehydrogenase, similar preliminary studies with Leptomonas

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Leptomonas
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Leptomonas, need for minimal defined medium for drug studies

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Kottl, M. A.; et al., 1975, J. Protozool., v. 22 (3), 40A-41A [Abstract]
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as syn. of Malacozoomonas patellae (A. Porter, 1914) [n. comb.]

Leptomonas pessoai
Daggett, P. M.; and Decker, J. E., 1973, J. Protozool., v. 20 (4), 512
6 insect trypanosomatids, increased survival in mice with repeated passage associated with alteration in secretion

Leptomonas pessoai Galvao, Oliveira, Carvalho & Veiga, 1970
Roitman, I.; et al., 1976, J. Protozool., v. 23 (2), 291-293
"Leptomonas pessoai may be regarded as a thermophilic species of Herpetomonas. . .and we suggest that it be designated Herpetomonas samuelpessoai sp. n.", differentiation of original strain and clones from promastigote to opisthomastigote faster at high temperature and in defined medium

Leptomonas pessoai
Souza, M. do C.; et al., 1974, J. Protozool., v. 21 (4), 579-584
living culture forms of Leptomonas pessoai cross-protected mice against Trypanosoma cruzi challenge infection, circulating antibodies detected in immunized mice by immunodiffusion, passive haemagglutination, complement fixation, and antibody binding assay which cross-reacted with T. cruzi extracts, cellular immune response indicated by leucocyte migration inhibition

Leptomonas pessoai
Souza, M. do C. M.; and Roitman, I., 1971, Rev. Microbiol., S. Paulo, v. 2 (4), 187-189
Trypanosoma cruzi, high degree of protection in mice previously immunized with vaccine of Leptomonas pessoai (living suspension)

Leptomonas pessoai, illus.
de Souza, W., 1976, Ztschr. Parasitenk., v. 48 (3-4), 221-226
Leptomonas pessoai, carbohydrates in Golgi complex and cytoplasmic membranes, ultrastructure

Leptomonas pessoai, illus.
de Souza, W.; et al., 1976, Canad. J. Microbiol., v. 22 (2), 197-203
Leptomonas pessoai, "...this insect flagellate is a member of the genus Herpetomonas"; ultrastructure, electron microscopy of promastigotes

Leptomonas pessoai
de Souza, W.; Bunn, M. M.; and Angluster, J., 1976, J. Protozool., v. 23 (2), 329-333
Leptomonas pessoai, cell surface polysaccharides and/or glycoproteins demonstrated by agglutination with concanavalin A as well as cytochemical reactions

Leptomonas seymouri sp. n., illus.
Wallace, F. G., 1977, J. Protozool., v. 24 (4), 483-484
Dysdercus suturellus (free in midgut): Stuart, Florida

Leptotheca armatura Yoshino & Moser, 1974
Moser, M.; and Noble, E. R., 1976, J. Protozool., v. 23 (4), 490-492
measurements, rate of infection
Coryphaenoides acrolepis (gall bladder, bile ducts): off southern California
C. leptolepis (gall bladder): off northern California
C. pectoralis (gall bladder): off northern California

Leptotheca coelorhynchus Yoshino & Noble, 1973
Moser, M.; and Noble, E. R., 1976, J. Protozool., v. 23 (4), 490-492
measurements, rate of infection
Coelorinchus coelorhincus: off Ireland
Coryphaenoides armatus: off southern Australia
Ventrifossa nigrodorsalis: off Borneo
(gall bladder of all)

Leptotheca informis Auerbach, 1910
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides acrolepis
C. filifer
all from eastern Pacific

Leptotheca informis Auerbach 1910
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes chlorostictus: off southern California
S. goodei: off northern California
S. paucispinis: off northern California
S. rosaceus: off southern California
S. serranoides: off southern, central and northern California
(gallbladder of all)

Leptotheca informis Auerbach, 1910
Moser, M.; and Noble, E. R., 1976, J. Protozool., v. 23 (4), 490-492
measurements, rate of infection
Anoplopoma fimbria (gall bladder): off southern California
Coelorinchus scaphopsis (gall bladder): off Mexico
Coryphaenoides acrolepis (gall bladder, bile ducts): off northern Mexico; off southern, central, and northern California and Oregon
C. fernandezianus (gall bladder): off Chile
C. filifer (gall bladder): off Oregon and Canada
C. pectoralis (gall bladder): off northern California and Washington

Leptotheca longipes Auerbach 1910
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes serranoides (gall bladder): off central California

Leptotheca macrospora Auerbach 1909
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes serranoides (gall bladder): off southern and central California

Leptotheca minima Meglitsch, 1960
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides acrolepis
C. armatus
all from eastern Pacific

Leptotheca minima Meglitsch, 1960
Moser, M.; and Noble, E. R., 1976, J. Protozool., v. 23 (4), 490-492
measurements, rate of infection
Coryphaenoides acrolepis: off northern Mexico; off southern and northern California; off Oregon
C. armatus: off northern Mexico; off southern, central, and northern California; off Oregon
(gall bladder and bile ducts of all)

Leptotheca minuta sp. n., illus.
Moser, M.; and Noble, E. R., 1976, J. Protozool., v. 23 (4), 490-492
Coelorinchus occa: off Lesser Antilles
Nezumia condylura: off Japan
(gall bladder of all)

Leptotheca sebasta sp. n., illus.
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes serranoides (gall bladder): off southern California

Leucocytogregarina musculi (Porter, 1908) Sangiorgi, 1912
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Hepatozoon musculi

Leucocytozoid[ae sp.]
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Buccanodon leucotis (blood): Tanzania

Leucocytozoon Sambon, 1908
Bennett, G. F.; et al., 1975, J. Protozool., v. 22 (1), 24-30
extensive remarks on taxonomic and nomenclatural status of Leucocytozoon, classification (Haemosporina, Leucocytozoidae), emended generic diagnosis, proposal that authorship be credited to Sambon 1908 and that L. majoris (Laveran, 1902) be designated as type species (L. danilewskyi being a nomen nudum), rejection of recent synonymy of Akiba and Saurocytozoon with Leucocytozoon

Leucocytozoon
Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
3 spp. of Leucocytozoon, morphometric variation, "concluded that the morphometric characteristics were not valid criteria for species differentiation.", proposed synonymies

Leucocytozoon
Greiner, E. C.; et al., 1975, Canad. J. Zool., v. 53 (12), 1762-1787
avian hematozoa, prevalence with reference to distribution by geographic region, by host family, by vertical stratification of nesting sites and by feeding behavior of known vectors: North America north of Mexico
[Checklist includes 388 bird species and contains both published and unpublished records. For records from specific hosts, see entries in Supplement 22, Part 7, Hosts.]

Leucocytozoon Berestnev, 1904
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
checklist of species with hosts; synonymy

Leucocytozoon [sp.]
Andrews, S. E.; and Threlfall, W., 1975, Proc. Helminth. Soc. Washington, v. 42 (1), 24-28
Corvus brachyrhynchos: insular Newfoundland

Leucocytozoon [sp.]
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Ixobrychus minutus
Butorides striatus
Elanus caeruleus
Melierax gabar
Accipiter minullus
A. tachiro
A. badius
Francolinus sephaena
F. clappertoni
F. squamatus
Numida meleagris
Actophilornis africana
Streptopelia semitorquata
S. decipiens
S. senegalensis
Turtur tympanistria
T. afer
T. abyssinicus
Aplopelia larvata
Chrysococcyx caprius
Centropus monachus
C. superciliosus
Otus scops
Asio flammeus
Coracias abyssinica
Lybius leucomelas
Indicator indicator
I. minor
Jynx torquilla
Campethera nubica
Mesopicos goertae
Riparia riparia
R. paludicola
Anthus trivialis
A. cervinus
Pycnonotus barbatus
Chlorocichla flavicollis
Prionops plumata
Dryoscopus gambensis
Tchagra minuta
T. senegala
Laniarius aethiopicus
L. erythrogaster
L. funebris
Myrmecocichla semirufa
Cossypha semirufa
Turdus pelios
Turdoides leucopygius
Acrocephalus schoenobaenus
Sphenoeacus mentalis

Leucocytozoon [sp.].-- Continued.
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Hippolais pallida
Sylvia borin
S. atricapilla
Phylloscopus trochilus
P. collybita
Cisticola erythrops
C. galactotes
Prinia subflava
Sylvietta whytii
Melaenornis edolioides
Bradornis pallidus
Batis orientalis
Terpsiphone viridis
Parus leucomelas
Nectarinia olivacea
N. senegalensis
Emberiza poliopleura
Serinus mozambicus
S. citrinelloides
S. nigriceps
S. striolatus
S. tristriatus
Vidua paradisaea
Mandingoa nitidula
Pytelia phoenicoptera
Lagonosticta larvata
Ploceus baglafecht
P. cucullatus
P. superciliosus
P. ocularis
Malimbus rubriceps
Quelea erythrops
Euplectes afer
E. ardens
Passer griseus
P. eminibey
Petronia pyrgita
Lamprotornis chalybaeus
Oriolus oriolus
O. larvatus
all from Ethiopia

Leucocytozoon sp.
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Coracias caudata: Kenya
Dicrurius adsimilis: Tanzania
Anthropetes collaris: Kenya
Nectarinia venusta: Kenya

Leucocytozoon [sp.]
Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
survey, avian hematozoa, seasonal prevalence
Ispidina picta
Tympanistria tympanistria
Tchagra australis
Platysteira cyanea
Pycnonotus barbatus
Zosterops senegalensis
(blood of all): all from Uganda

Leucocytozoon [sp.]
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Bambusicola thoracica sonorivox (blood): Taiwan

Leucocytozoon [sp.]
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Liothrix luteus
Urocissa erythrorhyncha
(blood of all): all imported to U. S., origin unknown

Leucocytozoon [sp.], probably L. liothricis
Manwell, R. D.; and Sessler, G. J., 1973, J. Protozool., v. 20 (3), 363-364
Liothrix luteus (blood): imported from Southeast Asia

Leucocytozoon [sp.]
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Trachyphonus darnaudii (blood): Kenya
Pycnonotus barbatus dodsoni (blood): Kenya

Leucocytozoon [sp.], indistinguishable from L. monardi
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Sylvia communis (blood): Kenya

Leucocytozoon [sp.]
Thomas, S. E.; and Dobson, L. D., 1975, Onderstepoort J. Vet. Research, v. 42 (1), 67-68
Anas erythrorhyncha
Netta erythrophthalma
Francolinus swainsonii
Numida meleagris
Columba guinea
Passer domesticus
P. melanurus
(blood of all): all from vicinity of Onderstepoort, Republic of South Africa

Leucocytozoon or Akiba, unidentified sp., illus.
Walker, D.; and Garnham, P. C. C., 1972, Vet. Rec., v. 91 (3), 70, 71, 72 [Letter]
cysts in heart and gizzard of parakeets resembling megaloschizonts of Leucocytozoon or Akiba, description, clinical signs of disease, lesions, pathology, case report
Neophema pulchella: Surrey, England
Platycercus adelidae: Oxon., England
Alisterus scapularis: Cornwall, England
Polytelis anthopeplus: Glos., England
(heart of all)

Leucocytozoon akodoni (Carini & Maciel, 1915) Coatney, 1937
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Hepatozoon akodoni

Leucocytozoon anatis Wickware, 1915
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon simondi Mathis & Leger, 1910

Leucocytozoon andrewsi
Noblet, R.; Moore, H. S. IV; and Noblet, G. P., 1976, Poultry Science, v. 55 (1), 447-449
Gallus gallus (blood): South Carolina

Leucocytozoon anellobiae (Cleland and Johnson, 1911)
Bennett, G. F.; and Campbell, A. G., 1975,
Canad. J. Zool., v. 53 (6), 800-812
as syn. of Leucocytozoon fringillinarum
Woodcock, 1910

Leucocytozoon anseris Knuth and Magdeburg, 1922
Bennett, G. F.; and Campbell, A. G., 1975,
Canad. J. Zool., v. 53 (6), 800-812
as syn. of L. simondi Mathis and Leger, 1911

Leucocytozoon anseris Knuth & Magdeburg, 1922
Hsu, C. K.; Campbell, G. R.; and Levine, N. D.,
1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon simondi Mathis &
Leger, 1910

Leucocytozoon apiaster sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Merops apiaster]: north-east Azerbaidzhan

Leucocytozoon ardea
Bennett, G. F.; and Blancou, J., 1974, J.
Wildlife Dis., v. 10 (3), 239-240
Egretta dimorpha (blood): Madagascar

Leucocytozoon bacelari Tendeiro (1947)
Peirce, M. A.; and Cooper, J. E., 1977, Vet.
Rec., v. 100 (23), 493
as syn. of L. toddi

Leucocytozoon berestneffi Sambon, 1908
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Pica pica]: north-east Azerbaidzhan

Leucocytozoon bonasae
Eve, J. H.; and Davidson, W. R., 1976, J. Parasitol., v. 62 (1), 142-144
Bonasa umbellus (blood): Kentucky; Michigan;
West Virginia

Leucocytozoon bouffardi Leger and Blanchard, 1911
Bennett, G. F.; and Campbell, A. G., 1975,
Canad. J. Zool., v. 53 (6), 800-812
as syn. of Leucocytozoon fringillinarum
Woodcock, 1910

Leucocytozoon brimonti
Bennett, G. F.; and Herman, C. M., 1976, J.
Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Andropadus importunus: Tanzania
Pycnonotus barbatus: Zaire
P. xanthopygus: Tanzania

Leucocytozoon cambournaci Franca, 1912
Bennett, G. F.; and Campbell, A. G., 1975,
Canad. J. Zool., v. 53 (6), 800-812
as syn. of Leucocytozoon fringillinarum
Woodcock, 1910

Leucocytozoon cambournaci Franca, 1912
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Emberiza hortulana]: north-east Azerbaidzhan

Leucocytozoon canis James, 1905
Hsu, C. K.; Campbell, G. R.; and Levine, N. D.,
1973, J. Protozool., v. 20 (2), 195-203
as syn. of Hepatozoon canis

Leucocytozoon caulleryi, illus.
Fallis, A. M.; Jacobson, R. L.; and Raybould,
J. N., 1973, J. Protozool., v. 20 (3), 438-442
Guttera pucherani (blood): Amani, Tanzania
Numida mitrata (blood): vicinity of Handeni,
Tanzania

Leucocytozoon (Akiba) caulleryi Mathis & Leger, 1909
Hsu, C. K.; Campbell, G. R.; and Levine, N. D.,
1973, J. Protozool., v. 20 (2), 195-203
synonymy

Leucocytozoon centropi
Bennett, G. F.; and Blancou, J., 1974, J.
Wildlife Dis., v. 10 (3), 239-240
Centropus toulou (blood): Madagascar

Leucocytozoon coraciae de Mello & Alfonso, 1935,
emend. Bhatia 1938
Hsu, C. K.; Campbell, G. R.; and Levine, N. D.,
1973, J. Protozool., v. 20 (2), 195-203
synonymy

Leucocytozoon coraciae benghalensis de Mello &
Alfonso, 1935
Hsu, C. K.; Campbell, G. R.; and Levine, N. D.,
1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon coraciae de Mello
& Alfonso, 1935, emend. Bhatia 1938

Leucocytozoon danilewskyi
Bennett, G. F.; et al., 1975, J. Protozool.,
v. 22 (1), 24-30
extensive remarks on taxonomic and nomenclatural status of Leucocytozoon, classification (Haemosporina, Leucocytozoidae), emended generic diagnosis, proposal that authorship be credited to Sambon 1908 and that L. majoris (Laveran, 1902) be designated as type species (L. danilewskyi being a nomen nudum), rejection of recent synonymy of Akiba and Saurocytozoon with Leucocytozoon

Leucocytozoon danilewskyi (Ziemann, 1898) Berestnev, 1904
Hsu, C. K.; Campbell, G. R.; and Levine, N. D.,
1973, J. Protozool., v. 20 (2), 195-203
synonymy

Leucocytozoon danilewskii Berestnew, 1904
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Asio otus]
[Athene noctua]
all from north-east Azerbaidzhan

Leucocytozoon danilewskyi var. hirundinis Sergent
& Sergent, 1905
Hsu, C. K.; Campbell, G. R.; and Levine, N. D.,
1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon hirundinis Sergent &
Sergent, 1905, emend. [i.e., new rank]

Leucocytozoon dubreuili
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Catharus ustulatus
Turdus serranus
(blood of all): all from Colombia

Leucocytozoon dubreuili Mathis and Leger 1911, illus.
Bennett, G. F.; and Cameron, M. F., 1975, J. Parasitol., v. 61 (6), 1091-1095
Leucocytozoon fringillinarum, L. dubreuili, L. majoris, mixed infections, passeriform birds; concluded that species identification must be based on morphological criteria, and host specificity alone is not a valid diagnosis
Carpodacus purpureus: Newfoundland; Tantramar Marshes, New Brunswick
Melospiza georgiana: Newfoundland
Melospiza melodia: Newfoundland; Tantramar Marshes, New Brunswick
Pinicola enucleator: Newfoundland
Passerella iliaca: Newfoundland
Zonotrichia albicollis: Newfoundland; Tantramar Marshes, New Brunswick
Parus atricapillus: Newfoundland
Dendroica coronata: Newfoundland; Tantramar Marshes, New Brunswick
Dendroica petechia: Newfoundland; Tantramar Marshes, New Brunswick
Dendroica pinus: Newfoundland
Dendroica striata: Newfoundland
Seiurus novaboracensis: Newfoundland
Wilsonia pusilla: Newfoundland
Turdus migratorius: Newfoundland; Tantramar Marshes, New Brunswick
Bombycilla cedrorum: Tantramar Marshes, New Brunswick
Molothrus ater: Tantramar Marshes, New Brunswick
Geothlypis trichas: Tantramar Marshes, New Brunswick
Setophaga ruticilla: Tantramar Marshes, New Brunswick
Catharus guttatus: Tantramar Marshes, New Brunswick
Catharus ustulatus: Tantramar Marshes, New Brunswick

Leucocytozoon dubreuili Mathis and Leger, 1911, illus.
Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
3 spp. of Leucocytozoon, morphometric variation, "concluded that the morphometric characteristics were not valid criteria for species differentiation.", proposed synonymies
Catharus ustulatus: Newfoundland
Turdus migratorius: Newfoundland; New Brunswick

Leucocytozoon dubreuili
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Turdoides sp.: Zaire
Myrmecocichla aethiops: Kenya

Leucocytozoon dubreuili
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Turdus merula (blood): Bramley, Hampshire, Bramham, Yorkshire and Sussex, England

Leucocytozoon dubreuili Mathis & Leger, illus.
Wong, T. C.; and Desser, S. S., 1976, J. Protozool., v. 23 (1), 115-126
Leucocytozoon dubreuili, fine structure of oocyst transformation and sporozoites

Leucocytozoon dubreuili, illus.
Wong, T. C.; and Desser, S. S., 1977, J. Parasitol., v. 63 (4), 766-768
finding of a sporozoite in a gametocyte in an erythrocyte of an experimentally infected Turdus migratorius

Leucocytozoon dubreuili Mathis et Leger,1910
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Turdus merula]
[Monticola saxatilis]
all from north-east Azerbaidzhan

Leucocytozoon eurystomi
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Coracias spatulata (blood): Zaire

Leucocytozoon eurystomi Kerandel, 1913
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon francae Tendeiro, 1947 non L. francai Nikitin, 1927

Leucocytozoon felis-domesticae Patton, 1908
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Hepatozoon felisdomesticae

Leucocytozoon francae Tendeiro, 1947 non L. francai Nikitin, 1927
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon eurystomi Kerandel, 1913

Leucocytozoon franchini Franca, 1927
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon laverani Franchini, 1923 non L. laverani Franca, 1912

Leucocytozoon franchini Franca, 1927
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Garrulus glandarius]: north-east Azerbaidzhan

Leucocytozoon fringillinarum
Bennett, G. F.; and Blancou, J., 1974, J. Wildlife Dis., v. 10 (3), 239-240
Foudia madagascariensis (blood): Madagascar

Leucocytozoon fringillinarum
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Tangara ruficervix (blood): Colombia

Leucocytozoon fringillinarum Woodcock 1910, illus.
Bennett, G. F.; and Cameron, M. F., 1975, J. Parasitol., v. 61 (6), 1091-1095
Leucocytozoon fringillinarum, L. dubreuili, L. majoris, mixed infections, passeriform birds; concluded that species identification must be based on morphological criteria, and host specificity alone is not a valid diagnosis
Carpodacus purpureus: Newfoundland; Tantramar Marshes, New Brunswick
Melospiza georgiana: Newfoundland
M. lincolnii: Newfoundland
M. melodia: Newfoundland; Tantramar Marshes, New Brunswick
Pinicola enucleator: Newfoundland
Passerella iliaca: Newfoundland
Zonotrichia albicollis: Newfoundland; Tantramar Marshes, New Brunswick
Parus atricapillus: Newfoundland; Tantramar Marshes, New Brunswick
P. hudsonicus: Newfoundland
Dendroica coronata: Newfoundland; Tantramar Marshes, New Brunswick
D. petechia: Newfoundland; Tantramar Marshes, New Brunswick
D. pinus: Newfoundland
D. striata: Newfoundland; Tantramar Marshes, New Brunswick
Mniotilta varia: Newfoundland
Seiurus novaboracensis: Newfoundland
Wilsonia pusilla: Newfoundland
Dendroica pennsylvanica: Tantramar Marshes, New Brunswick
Molothrus ater: Tantramar Marshes, New Brunswick
Dumetella carolinensis: Tantramar Marshes, New Brunswick
Wilsonia canadensis: Tantramar Marshes, New Brunswick
Turdus migratorius: Newfoundland; Tantramar Marshes, New Brunswick
Catharus guttatus: Tantramar Marshes, New Brunswick
Bombycilla cedrorum: Tantramar Marshes, New Brunswick

Leucocytozoon fringillinarum Woodcock, 1910, illus.
Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
3 spp. of Leucocytozoon, morphometric variation, "concluded that the morphometric characteristics were not valid criteria for species differentiation.", proposed synonymies
Passerella iliaca: Newfoundland
Dendroica striata: Newfoundland
Empidonax minimus: Labrador; Newfoundland
Ploceus baglafecht: Tanzania

Leucocytozoon fringillinarum
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Emberiza cabanisi: Tanzania
Lagonosticta senegala: Zaire
Carduelis citrinelloides: Tanzania
Bradornis pumilus: Tanzania
Musophago violacea: Zaire
Anthropetes collaris: Tanzania
Cyanomitra olivacea: Tanzania
Nectarinia amethystina: Zaire
Oriolus auratus: Zaire
Hyphanturgis ocularis: Tanzania
Mandingoa nitidula: Tanzania

Leucocytozoon fringillinarum.-- Continued.
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Othypantes stuhlmanni: Tanzania
Passer iagoensis: Kenya
Ploceus baglafecht: Kenya; Tanzania
P. ocularia: Kenya
P. reichenowi: Kenya
P. spekei: Kenya
Cisticola robusta: Kenya

Leucocytozoon fringillinarum/majoris
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Serinus atrogullaris: Kenya
S. mozambicus: Zaire
Lanius collaris: Kenya; Zaire
Laniarius ferrugineus: Kenya
Nectarinia kilimensis: Kenya
Lamprocolius corruscus: Tanzania
Lamprotornis chalybaeus: Kenya

Leucocytozoon fringillinarum
Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
survey, avian hematozoa, seasonal prevalence
Ploceus aurantius
P. cucullatus
P. weynsi
(blood of all): all from Uganda

Leucocytozoon fringillinarum
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Sylvia borin (blood): Bramham, Yorkshire, England

Leucocytozoon fringillinarum?, illus.
Greiner, E. C., 1976, J. Parasitol., v. 62 (4), 545-547
Zoothera marginata (blood): Chiengmai, Thailand

Leucocytozoon fringillinarum
Noblet, R.; Moore, H. S. IV; and Noblet, G. P., 1976, Poultry Science, v. 55 (1), 447-449
Quiscalus quiscula (blood): South Carolina

Leucocytozoon funambuli Patton, 1906
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Hepatozoon funambuli

Leucocytozoon galli Ivanic, 1937
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
identification uncertain, not Leucocytozoon

Leucocytozoon gentili Leger, 1913
Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
as syn. of Leucocytozoon fringillinarum Woodcock, 1910

Leucocytozoon grusi
Forrester, D. J.; Bush, A. O.; and Williams, L. E., jr., 1975, J. Parasitol., v. 61 (3), 547-548
Grus canadensis pratensis (blood): Florida

Leucocytozoon hirundinis Sergent & Sergent, 1905, emend. [i.e., new rank]
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon danilewskyi var. hirundinis Sergent & Sergent, 1905

Leucocytozoon hirundinis Sergent et Sergent, 1905
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Hirundo rustica]: north-east Azerbaidzhan

Leucocytozoon laverani Franchini, 1923 non L. laverani Franca, 1912
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon franchini Franca, 1927

Leucocytozoon leitaoi Tendeiro, 1947
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon coraciae de Mello & Alfonso, 1935, emend. Bhatia 1938

Leucocytozoon leporis Patton, 1908
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Hepatozoon leporis

Leucocytozoon liothricis (Laveran & Marullaz, 1914) Coatney, 1937
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Haemamoeba liothricis Laveran & Marullaz, 1914

Leucocytozoon maccluri sp. n., illus.
Greiner, E. C., 1976, J. Parasitol., v. 62 (4), 545-547
Zoothera marginata (blood): Chiengmai, Thailand

Leucocytozoon majoris (Laveran, 1902)
Bennett, G. F.; et al., 1975, J. Protozool., v. 22 (1), 24-30
extensive remarks on taxonomic and nomenclatural status of Leucocytozoon, classification (Haemosporina, Leucocytozoidae), emended generic diagnosis, proposal that authorship be credited to Sambon 1908 and that L. majoris (Laveran, 1902) be designated as type species (L. danilewskyi being a nomen nudum), rejection of recent synonymy of Akiba and Saurocytozoon with Leucocytozoon

Leucocytozoon majoris (Laveran 1902), illus.
Bennett, G. F.; and Cameron, M. F., 1975, J. Parasitol., v. 61 (6), 1091-1095
Leucocytozoon fringillinarum, L. dubreuili, L. majoris, mixed infections, passeriform birds; concluded that species identification must be based on morphological criteria, and host specificity alone is not a valid diagnosis
Melospiza georgiana: Newfoundland
Melospiza lincolnii: Newfoundland
Melospiza melodia: Newfoundland; Tantramar Marshes, New Brunswick
Passerella iliaca: Newfoundland
Zonotrichia albicollis: Newfoundland; Tantramar Marshes, New Brunswick
Parus atricapillus: Newfoundland; Tantramar Marshes, New Brunswick
Parus hudsonicus: Newfoundland
Dendroica petechia: Newfoundland; Tantramar Marshes, New Brunswick
Dendroica striata: Newfoundland; Tantramar Marshes, New Brunswick
Mniotilta varia: Newfoundland
Seiurus novaboracensis: Newfoundland
Wilsonia pusilla: Newfoundland
Turdus migratorius: Newfoundland; Tantramar Marshes, New Brunswick
Dumetella carolinensis: Tantramar Marshes, New Brunswick
Dendroica coronata: Tantramar Marshes, New Brunswick
Dendroica pennsylvanica: Tantramar Marshes, New Brunswick
Dendroica petechia: Tantramar Marshes, New Brunswick
Geothlypis trichas: Tantramar Marshes, New Brunswick
Setophaga ruticilla: Tantramar Marshes, New Brunswick
Wilsonia canadensis: Tantramar Marshes, New Brunswick
Catharus ustulatus: Tantramar Marshes, New Brunswick

Leucocytozoon majoris/fringillinarum
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Serinus atrogullaris: Kenya
S. mozambicus: Zaire
Lanius collaris: Kenya; Zaire
Laniarius ferrugineus: Kenya
Nectarinia kilimensis: Kenya
Lamprocolius corruscus: Tanzania
Lamprotornis chalybaeus: Kenya

Leucocytozoon majoris
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Parus caeruleus (blood): Bramley, Hampshire and Bramham, Yorkshire, England

Leucocytozoon majoris?, illus.
Greiner, E. C., 1976, J. Parasitol., v. 62 (4), 545-547
Zoothera marginata (blood): Chiengmai, Thailand

Leucocytozoon majoris (Laveran, 1902) Coatney, 1937
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Haemoproteus majoris Laveran, 1902

Leucocytozoon majoris Coatney, 1937
  Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
    [Parus major]: north-east Azerbaidzhan

Leucocytozoon marchouxi
  Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
    (blood of all)
    Streptopelia lugens: Kenya
    S. semitorquata: Kenya
    Tympanistra tympanistra: Tanzania

Leucocytozoon marchouxi Mathis & Leger, 1910
  Greiner, E. C., 1975, J. Wildlife Dis., v. 11 (2), 150-156
    Zenaida macroura: Ames, Iowa

Leucocytozoon marchouxi Mathis & Leger, 1910
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    Syn.: Leucocytozoon turtur Ortega & Berenguer, 1946

Leucocytozoon marchouxi
  Stabler, R. M.; Kitzmiller, N. J.; and Braun, C. E., 1977, J. Wildlife Management, v. 41 (1), 128-130
    Columba fasciata (blood): Colorado; California; Mexico

Leucocytozoon marchouxi Mathis et Leger, 1910
  Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
    [Streptopelia turtur]: north-east Azerbaidzhan

Leucocytozoon mathisi Franca, 1912
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    Syn.: Leucocytozoon mathisi var. buteonis Coatney & Roudabush, 1937

Leucocytozoon mathisi Franca, 1912
  Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
    [Accipiter]: north-east Azerbaidzhan

Leucocytozoon mathisi var. buteonis Coatney & Roudabush, 1937
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    as syn. of Leucocytozoon mathisi Franca, 1912

Leucocytozoon monardi Rodhain, 1931
  Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
    as syn. of Leucocytozoon fringillinarum Woodcock, 1910

Leucocytozoon muris Balfour, 1906
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    as syn. of Hepatozoon muris

Leucocytozoon musajevi sp. n. [nomen nudum]
  Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
    [Coracias garrulus]: north-east Azerbaidzhan

Leucocytozoon musculi Porter, 1908
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    as syn. of Hepatozoon muris

Leucocytozoon neavei
  Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
    (blood of all)
    Guttera pucherani: Tanzania
    Numida meleagris: Kenya; Tanzania; Zaire

Leucocytozoon neavei, illus.
  Fallis, A. M.; Jacobson, R. L.; and Raybould, J. N., 1973, J. Protozool., v. 20 (3), 438-442
    sporogony and transmission
    Guttera pucherani (blood): Amani, Tanzania
    Numida mitrata (blood)(nat. and exper.): vicinity of Handeni, Tanzania
    Simulium impukane group (exper.)
    Simulium adersi (exper.)
    Simulium nyasalandicum (exper.)

Leucocytozoon neavei (Balfour, 1906) Wenyon, 1908
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    synonymy

Leucocytozoon numidae Kerandel, 1913
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    as syn. of Leucocytozoon neavei (Balfour, 1906) Wenyon, 1908

Leucocytozoon piroplasmoides Ducloux, 1908
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    not identifiable, Leishmania ?

Leucocytozoon ranarum Carini, 1907
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    as syn. of Haemogregarina ranarum

Leucocytozoon ratti Adie, 1906
  Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
    as syn. of Hepatozoon ratti

Leucocytozoon roubaudi Mathis and Leger, 1911
  Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
    as syn. of Leucocytozoon fringillinarum Woodcock, 1910

Leucocytozoon sabrazi [i.e. ?, sabrazesi]
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Francolinus sp.: Zaire
Gallus gallus: Tanzania

Leucocytozoon sabrazesi
Chang, C. H., 1975, J. Chinese Soc. Vet. Sc., v. 1 (1), 1-6
Leucocytozoon sabrazesi, white Leghorn roosters, absence of relapse in naturally and experimentally infected birds, possible factors: Taiwan

Leucocytozoon sabrazesi
Colley, F. C.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 84-85 [Demonstration]
chickens (peripheral blood): Gombak, Selangor, Malaysia

Leucocytozoon sabrazesi Mathis & Leger, 1910
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon schueffneri von Prowazek, 1912, in part

Leucocytozoon sabrazesi
Lee, Y. C.; and Chang, C. H., 1975, J. Chinese Soc. Vet. Sc., v. 1 (1), 20-23
Leucocytozoon sabrazesi, chickens, preventive and therapeutic effect of Diameton

Leucocytozoon sakharoffi Sambon, 1908
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Leucocytozoon zuccarellii Leger, 1913

Leucocytozoon sakharoffi
Noblet, R.; Moore, H. S. IV; and Noblet, G. P., 1976, Poultry Science, v. 55 (1), 447-449
Cyanocitta cristata cristata (blood): South Carolina

Leucocytozoon sakharoffi Sambon, 1908
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Corvus corone]
[Corvus corone cornix]
all from north-east Azerbaidzhan

Leucocytozoon salvelini Fantham, Porter & Richardson, 1942
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Haemogregarina salvelini

Leucocytozoon schoutedeni, illus.
Fallis, A. M.; Jacobson, R. L.; and Raybould, J. N., 1973, J. Protozool., v. 20 (3), 438-442
sporogony and transmission
chicken (nat. and exper.)(blood): vicinity of Amani, Tanzania
Simulium adersi: vicinity of Amani, Tanzania
Simulium impukane group: vicinity of Amani, Tanzania
Simulium vorax (exper.)
Simulium nyasalandicum (exper.)

Leucocytozoon schoutedeni, "resembling"
Fallis, A. M.; Jacobson, R. L.; and Raybould, J. N., 1973, J. Protozool., v. 20 (3), 438-442
Guttera pucherani (blood): Amani, Tanzania

Leucocytozoon schueffneri von Prowazek, 1912, in part
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon (Akiba) caulleryi Mathis & Leger, 1909

Leucocytozoon schüeffneri von Prowazek, 1912, in part
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon sabrazesi Mathis & Leger, 1910

Leucocytozoon seabrae Franca, 1912
Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
as syn. of Leucocytozoon fringillinarum Woodcock, 1910

Leucocytozoon simondi
Bennett, G. F.; et al., 1974, J. Wildlife Dis., v. 10 (4), 442-451
survey, prevalence of hematozoa in anatids, infection rate increases with host age, seasonal distribution
Anas rubripes
Anas platyrhynchos
Anas platyrhynchos x A. rubripes
Anas discors
Anas carolinensis
Aix sponsa
Lophodytes cucullatus
(blood of all): all from Massachusetts

Leucocytozoon simondi
Bennett, G. F.; et al., 1975, J. Wildlife Dis., v. 11 (2), 280-289
hematozoa of Anatidae, prevalence survey
Anas boschas
Anas rubripes rubripes
Anas platyrhynchos platyrhynchos
Anas platyrhynchos platyrhynchos x A. rubripes rubripes
Anas crecca carolinensis
Anas discors
Anas acuta
Aix sponsa
Aythya collaris
Glaucionetta clangula
Mareca americana
Branta canadensis
all from Maritime provinces of Canada

Leucocytozoon simondi Mathis and Leger, 1911, illus.
Bennett, G. F.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (6), 800-812
3 spp. of Leucocytozoon, morphometric variation, "concluded that the morphometric characteristics were not valid criteria for species differentiation.", proposed synonymies
Syn.: L. anseris Knuth and Magdborg, 1922
Anas rubripes: New Brunswick
A. platyrhynchos: Massachusetts
A. rubripes x A. platyrhynchos: Massachusetts
A. crecca: New Brunswick
Aix sponsa: Massachusetts
Chen caerulescens: Northwest Territories
domestic goose: New Brunswick

Leucocytozoon simondi, illus.
Commichau, C.; and Jonas, D., 1977, Zentralbl. Vet.-Med., Reihe B, v. 24 (8), 662-667
Leucocytozoon simondi in Cairina moschata (Herz, Lunge, Leber, Niere, Dunndarm), histopathology: Deutschland

Leucocytozoon simondi, illus.
Desser, S. S.; Fallis, A. M.; and Allison, F. R., 1976, Canad. J. Zool., v. 54 (5), 799-801
Leucocytozoon simondi, L. tawaki, nuclear changes preceding microgamete formation
Anas platyrhynchos: Algonquin Park, Ontario

Leucocytozoon simondi, illus.
Desser, S. S.; and Ryckman, A. K., 1976, Canad. J. Zool., v. 54 (5), 634-643
Leucocytozoon simondi, development in captive Branta canadensis maxima, B. c. interior, and Anser domesticus exposed to natural infection on shores of Lake Sasajewan, Algonquin Park, Ontario, some anemia but no other pathology, no megaloschizonts or elongate gametocytes seen, identity confirmed by experimental infections of goslings and ducklings with gosling-derived sporozoites

Leucocytozoon simondi
Herman, C. M.; and Bennett, G. F., 1976, Canad. J. Zool., v. 54 (7), 1038-1043
exposure of captive sentinel Anatidae to natural infection, seasonal transmission
Anas platyrhynchos
Anser anser
all from Nova Scotia-New Brunswick border area

Leucocytozoon simondi Mathis & Leger, 1910
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
synonymy

Leucocytozoon simondi, illus.
Sterling, C. R.; and Aikawa, M., 1973, J. Protozool., v. 20 (1), 81-92
Plasmodium gallinaceum, Haemoproteus columbae, Leucocytozoon simondi, gametocyte ultrastructure compared in detail

Leucocytozoon simondi, illus.
Trefiak, W. D.; and Desser, S. S., 1973, J. Protozool., v. 20 (1), 73-80
crystalloid inclusions in Leucocytozoon, Parahaemoproteus, and Plasmodium, lipid-protein nature, ultrastructural morphology

Leucocytozoon smithi
Barnett, B. D., 1977, World's Poultry Sc. J., v. 33 (2), 76-87
life cycle, vectors, transmission, pathology, prevention and control. review

Leucocytozoon smithi Laveran and Lucet, 1905
Forrester, D. J.; et al., 1974, J. Protozool., v. 21 (4), 494-497
prevalence in relation to host age, host sex, geographic distribution, and year
Meleagris gallopavo (blood): Florida

Leucocytozoon smithi (Laveran & Lucet, 1905)
Kissam, J. B.; Noblet, R.; and Garris, G. I., 1975, J. Med. Entom., v. 12 (3), 359-362
Leucocytozoon smithi, turkeys, control by large scale aerial application of Abate to streams against larval Simulium vectors, effective and economically feasible

Leucocytozoon smithi (Laveran & Lucet, 1905) Sambon, 1908
Noblet, R.; Adkins, T. R.; and Kissam, J. B., 1972, J. Med. Entom., v. 9 (6), 580
Leucocytozoon smithi, Simulium congareenarum as vector in early spring and S. slossonae as vector in remainder of season: South Carolina

Leucocytozoon smithi (Laveran & Lucet)
Noblet, R.; Kissam, J. B.; and Adkins, T. R., jr., 1975, J. Med. Entom., v. 12 (1), 111-114
Leucocytozoon smithi, transmission to sentinel turkeys by Simulium slossonae and S. congareenarum in 2 areas near turkey breeder farms, incidence and seasonal distribution: Coastal Plains near Sumter and the Sandhills near Pageland, South Carolina

Leucocytozoon smithi
Noblet, R.; and Moore, H. S. IV, 1975, J. Wildlife Dis., v. 11 (4), 516-518
Leucocytozoon smithi and Haemoproteus meleagridis in Meleagris gallopavo, wild turkeys as important reservoir hosts for Leucocytozoon smithi: South Carolina

Leucocytozoon smithi
Noblet, R.; Moore, H. S. IV; and Noblet, G. P., 1976, Poultry Science, v. 55 (1), 447-449
turkeys, wild (blood): South Carolina

Leucocytozoon tawaki n. sp., illus.
Fallis, A. M.; Bisset, S. A.; and Allison, F. R., 1976, N. Zealand J. Zool., v. 3 (1), 11-16
development in blackfly vectors
Austrosimulium australense: Jackson Head, South Island, New Zealand
A. dumbletoni: Jackson Head, South Island, New Zealand
A. ungulatum: Jackson Head, South Island, New Zealand
Eudyptes pachyrhynchus (blood): Kaikoura and Jackson Head, South Island, New Zealand

Leucocytozoon tawaki, illus.
Desser, S. S.; Fallis, A. M.; and Allison, F. R., 1976, Canad. J. Zool., v. 54 (5), 799-801
Leucocytozoon simondi, L. tawaki, nuclear changes preceding microgamete formation
Eudyptes pachyrhynchus: Jackson's Bay, New Zealand

Leucocytozoon toddi
Bennett, G. F.; and Blancou, J., 1974, J. Wildlife Dis., v. 10 (3), 239-240
Buteo brachypterus (blood): Madagascar

Leucocytozoon toddi
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Accipiter tachiro (blood): Tanzania

Leucocytozoon toddi
Peirce, M. A.; and Cooper, J. E., 1977, Vet. Rec., v. 100 (23), 493
Syn.: L. bacelari Tendeiro (1947)
Accipiter gentilis
A. nisus
Falco peregrinus
(blood of all): all from Great Britain

Leucocytozoon (Saurocytozoon) tupinambi (Lainson & Shaw, 1969) nov. comb.
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
Syn.: Saurocytozoon tupinambi Lainson & Shaw, 1969

Leucocytozoon turtur Ortega & Berenguer, 1946
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon marchouxi Mathis & Leger, 1910

Leucocytozoon vulgaris sp. n. [nomen nudum]
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Sturnus vulgaris]: north-east Azerbaidzhan

Leucocytozoon ziemanni
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Bubo africanus (blood): Zaire

Leucocytozoon ziemanni Laveran, 1902
Hsu, C. K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon danilewskyi (Ziemann, 1898) Berestnev, 1904

Leucocytozoon ziemanni (Laveran) (=L. danilewskyi)
Khan, R. A., 1975, J. Parasitol., v. 61 (3), 449-457
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Molineaux, L.; et al., 1976, Bull. World Health Organ., v. 54 (4), 379-389
human malarial control, impact of propoxur residual spraying on Anopheles vectors, analysis of variations between sprayed villages and effects of pre-spraying variables: West African savannah

Malaria
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differentiation between malarial and schistosomal pigment

Malaria
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common parasitic infections of man encountered in the United States, recommendations for treatment, review

Malaria
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human antimalarial programs, progress review for 1975, comparison with previous statistics (regional surveys, training, research, immunology, chemotherapy, epidemiology)

Malaria
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emerging medical problems associated with U.S. army troops, clinical management and prophylaxis: Vietnam

Malaria
Ohlrogge, R.; and Schlipkoeter, H. W., 1973, Med. Microbiol. and Immunol., v. 158 (4), 267-274
deviations in erythrogram before and after treatment of human malaria infections with resochin, no such changes in hookworm and other worm infections: Liberia; Togo

Malaria
O'Holohan, D. R.; Dondero, T. J., jr.; and Ponnampalam, J. T., 1973, Med. J. Malaysia, v. 27 (4), 310
fansidar excellent drug for treatment of chloroquine resistant malaria but use as prophylaxis is experimental and not recommended

Malaria
Palmer, A., 1976, Brit. J. Clin. Pract., v. 30 (1), 19-20
Plasmodium vivax, increased incidence of malaria with increased travel, implications if associated with pregnancy, need for diagnostic awareness: United Kingdom

Malaria
Pandeya, S. N., 1977, Science and Culture, v. 43 (12), 520-522
malaria, human, history of drug therapy, review

Malaria
Payet, M.; and Coulaud, J. P., 1971, Medecine Afrique Noire, v. 18 (spec. no.), 135-137
human malaria, cardiac complications, clinical review

Malaria
Peck, C. C.; Lewis, A. N.; and Joyce, B. E., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 141-146
Fansidar (pyrimethamine + sulfadoxine) administered once monthly as a malaria suppressant, time course of serum sufadoxine concentrations after monthly doses attests to pharmacokinetic rationale for such infrequent dosing

Malaria
Ponnampalam, J. T., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 285 [Letter]
human malaria, increased risk of exposure of individuals to bite of mosquito vectors during the durian (local fruit) season (June-Sept.) when collecting fruit by night or living in the orchards during that time: West Malaysia

Malaria
Powell, R. D., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 181-183
diagnosis established in clinically ill woman with symptoms of malaria but with persistently negative blood smears when parasites were detected in lupus erythematosus cell preparation: Iowa (had just returned from 3 month visit to Zaire)

Malaria
Rao, V. N., 1974, Progr. Drug Research, v. 18, 245-251
human malarias, review of control problems experienced by the National Malaria Eradication Programme of India

Malaria
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Plasmodium spp., possible immunization against sporozoite challenge, review, symposium presentation

Malaria
Rutter, N., 1977, Brit. Med. J. (6098), v. 2, 1335-1336
malaria infections in children born and living in non-malarious areas who have contracted infections upon visiting malarious areas with their immigrant parents who possibly have some natural immunity to malaria although their children do not, importance of diagnostic awareness: Nottingham, England

Malaria
Sinel'shchikov, V. A., 1973, Parazity Zhivot. i Rasten., Akad. Nauk Moldavsk. SSR (9), 230-238
epidemiological significance of faunistic complexes of malarial mosquitoes in landscape-geographic regions of Moldavia

Malaria
Snow, W. F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 449-450 [Letter]
human malaria, trials with an electronic mosquito vector repelling device proved to be unsuccessful: West Africa

Malaria
Sulzer, A. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 440-441 [Letter]
proposal for establishment of central facility to produce antigen for serologic tests for malarial antibody in order to advance research in malariaology and reduce numbers of simian species needed for antigen production

Malaria
Sulzer, A. J.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 55-58
malaria, human, diagnosis, indirect fluorescent antibody test using multi-species thick smear antigen containing equal proportions of Plasmodium vivax, P. falciparum, and P. brasilianum (latter serologically equivalent in IFA test to P. malariae), circumvents need for multiple testing with several antigen species

Malaria
Sulzer, A. J.; and Latorre, C. R., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 553 [Letter]
primate malarias, method for in vitro production of schizont antigens for use in indirect immunofluorescence tests for malarial antibodies

Malaria
Tesh, R. B.; and Papaevangelou, G., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 163-166
insecticide spraying for malaria control also resulted in reduced incidence of sandfly fever in Athens, Greece

Malaria
Trigg, P. I., 1976, Bull. World Health Organ., v. 53 (4), 399-406
Plasmodium spp., parasite cultivation in relation to chemotherapy of malaria with special application to chloroquine resistance, review of current aspects and new developments

Malaria
Umlas, J.; and Fallon, J. N., 1971, Am. J. Trop. Med. and Hyg., v. 20 (4), 527-529
simple technique for lysing thick films with saponin and staining with Wright or Wright-Giemsa stains for routine laboratory diagnosis of suspected human malaria

Malaria
Vogel, C. W.; and Koenigk, E., 1975, Tropenmed. u. Parasitol., v. 26 (3), 278-280
human malaria, spectrofluorometric and spectrodensitometric determination of chloroquine in plasma and urine of persons undergoing malaria therapy

Malaria
Voller, A., 1975, Symposia Brit. Soc. Parasitol., v. 13, 69-84
immunopathology of malaria, extensive review: immunosuppression and auto-immunity; tropical splenomegaly syndrome; nephrotic syndrome

Malaria
Voller, A.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 5 [Demonstration]
human malaria, application of enzyme-linked immune sorbent assay (E.L.I.S.A.) for diagnosis

Malaria
Voller, A.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 431-437
human tropical parasitic diseases, comparison of enzyme-immunoassay and radio-immunoassay in detection of antibodies, both assays sensitive and reproducible and gave comparable results

Malaria
Waddy, B. B., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 39-50
human parasitism and public health problems associated with building of new lakes and dams, review of several water projects with emphasis on Africa

Malaria
Ward, R. A., 1977, Proc. 15. Internat. Cong. Entom. (Washington, D.C., Aug. 19-27, 1976), 523-529
malaria, factors of human ecology that influence recent epidemiological changes

Malaria
Warren, K. S.; and Mahmoud, A. A. F., 1976, J. Infect. Dis., v. 133 (5), 596-601
prevention of malaria, schistosomiasis and other tropical and exotic diseases, advice to travellers

Malaria
Webber, R. H., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (3), 430-434
Wuchereria bancrofti, area survey for possible disease control as additional result of Malaria Eradication Programme since both infections transmitted by same mosquito vector: Choiseul and Shortland of Western Solomon Islands

Malaria
Webber, R. H., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 396-400
control measures carried out by the Malaria Eradication Programme against Anopheles vectors of human malaria also resulted in the natural decline in the prevalence of human Wuchereria bancrofti infections since both diseases are transmitted by the same vector in the Solomon Islands

Malaria
Weise, H. J., 1976, Munchen. Med. Wchnschr., v. 118 (35), 1061-1068
human exotic parasitic diseases, risks to tourists travelling in tropical countries, prophylactic and control measures, need for improved diagnosis

Malaria
Weiss, N.; and Degremont, A., 1976, Praxis, Bern, v. 65 (24), 742-744
human malaria, comparative evaluation of Plasmodium gallinaceum, P. falciparum and P. cynomolgi bastianellii as antigens for indirect immunofluorescence used for diagnosis

Malaria
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efficiency of urinary hydroxyproline index as indicator of nutritional status in mass surveys evaluated in presence of schistosomiasis, hookworm and malaria; in endemic malaria areas index probably of little value without prior evaluation of malarial status of all subjects

Malaria
White, G. B., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 278-301
malaria and filariasis in humans, extensive review of Anopheles gambiae complex vector mosquitoes, their morphologic identification and relationship to disease spread, possible control measures: Africa

Malaria
Wilson, R. J. M.; and Williams, K., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 15-20
European with suspected pyrimethamine-resistant malaria, serological study (fluorescent antibody test, C-reactive protein, precipitin test, immunoglobulin levels), precipitating antibodies demonstrated against two specific malarial antigens, differences from antibodies seen in sera from repeatedly infected Africans: The Gambia, West Africa

Malaria
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achievements in tropical medicine during past 25 years, control, prophylaxis, treatment

Malaria
Woodruff, A. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 313-328
mechanisms involved in anemia associated with infection (schistosomiasis, kala-azar, malaria, trypanosomiasis) and splenomegaly in tropics, complement activation leading to hemolysis and splenomegaly due to erythrophagocytosis, review

Malaria, probable Hepatocystes infection, illus.
Wu, C., 1967, Nettai Igaku (Trop. Med.), v. 9 (1), 65-69
Harpiocephalus harpia harpia
Murina puta
Hipposideros armiger terasensis
all from Taiwan

Malaria
Wyler, D. J., 1975, N. England J. Med., v. 293 (4), 203-204 [Letter]
latent malaria in Southeast Asian refugees, need for diagnostic awareness, screening as blood donors, reporting of acute cases: United States

Malaria
Yoeli, M., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 24-35
Plasmodium berghei yoelii in mice (exper.), promising model for study of cerebral malaria in man caused by P. falciparum (history, origin of virulent strain, pathology, enzyme differentiation, virulence expressed by mutation and genetic change)

Malaria
Zuckerman, A., 1977, Exper. Parasitol., v. 42 (2), 374-446
Plasmodium, immunology, extensive review: immunodiagnosis and seroepidemiology; immunopathology; antigenic analysis; host responses; immunoglobulins; cell-mediated reactions

Malpighamoeba (Malamoeba) locustae
Harry, O. G., 1974, J. Protozool., v. 21 (3), 450 [Abstract]
Malpighamoeba locustae, life cycle in Schistocerca gregaria

Malpighamoeba locustae
Harry, O. G., 1977, Parasitology, v. 75 (2), xxix [Abstract]
Malpighamoeba locustae, encapsulation in dorsal septum and other areas of locusts

Malpighamoeba locustae, illus.
Harry, O. G.; and Finlayson, L. H., 1976, Parasitology, v. 72 (2), 127-135
Malpighamoeba locustae in Schistocerca gregaria, life cycle, ultrastructure, mode of feeding (phagocytosis)

Malpighamoeba mellificae
Niemczuk, R.; and Sobieszczanska, B., 1976, Med. Wet., v. 32 (5), 296-297
Nosema apis, Leptomonas apis, Malpighamoeba mellificae, occurrence in digestive tract of adult bees, disturbances in metabolism, decrease in productivity: Lower Silesia

Mantonella
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Mantonella hammondi sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1976, J. Protozool., v. 23 (1), 57-63
prevalence
Kinosternon flavescens spooneri: Iowa

Marteilia Grizel, Comps, Bonami, Cousserans, Duthoit & Le Pennec, emend.
Perkins, F. O., 1976, J. Protozool., v. 23 (1), 64-74
Haplosporea
diagnosis

Marteilia sp., probably identical to M. sydneyi sp. n.
Perkins, F. O.; and Wolf, P. H., 1976, J. Parasitol., v. 62 (4), 528-538
Crassostrea echinata: eastern coast of Australia

Marteilia refringens Grizel, Comps, Bonami, Cousserans, Duthoit & Le Pennec, emend., illus.
Perkins, F. O., 1976, J. Protozool., v. 23 (1), 64-74
diagnosis; ultrastructure of sporulation establishes as member of Haplosporea
Ostrea edulis: Aber Wrach, France

Marteilia sydneyi sp. n., illus.
  Perkins, F. O.; and Wolf, P. H., 1976, J. Parasitol., v. 62 (4), 528-538
    fine structure, light and electron microscopy
    Crassostrea commercialis (hepatopancreas): Moreton Bay, Queensland, Australia

Mastigamoeba [sp.]
  Jadin, J. B.; and Willaert, E., 1976, J. Protozool., v. 23 (4), 12A [Abstract]
    isolation of free-living amoebae from nasal mucus of lepers, amoebae contain mycobacteria
    malade (mucus nasal): Kapolowe (Shaba-Zaire)

Mattesia dispora Naville, illus.
  Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
    life cycle
    Syn.: Coelogregarina ephestiae Ghelelovitch, 1948 sensu Weiser, 1954 non Ghelelovitch, 1948
    Ephestia kuehniella (fat body): Poland

Mattesia dispora Naville, 1930
  Zizka, Z., 1975, J. Protozool., v. 22 (3), 70A-71A [Abstract]
    Malamoeba locustae cysts, Mattesia dispora spores, scanning electron microscopy

Mattesia grandis
  Thompson, A. C.; and McLaughlin, R. E., 1977, J. Invert. Path., v. 30 (1), 108-109
    Mattesia grandis in Anthonomus grandis, comparison of lipids and fatty acids of parasite and fat body of host, direct competition for these compounds by the pathogen

Mattesia oryzaephili n. sp., illus.
  Ormieres, R.; Louis, C.; and Kuhl, G., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 547-556
    life cycle, pathology
    Oryzaephilus surinamensis (nat. and exper.) (tissu adipeux)

Mattesia oryzaephili Ormieres, Louis et Kuhl, illus.
  Ormieres, R.; Louis, C.; and Kuhl, G., 1972, Protistologica, v. 8 (3), 287-294
    Mattesia oryzaephili, first generation schizogony, ultrastructure

Maupasella Cepede, 1910
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Maupasellidae, Maupasellinae

Maupasella sp. Wichterman, 1939
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    as syn. of Maupasella pontodrili sp. nov.

Maupasella algeriana de Puytorac, 1954
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Helodrilus schneideri

Maupasella cepedei de Puytorac, 1954
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    synonymy
    Allolobophora terrestris
    Helodrilus chloroticus
    Eisenia foetida
    Allolobophora longa
    A. caliginosa
    Eiseniella tetraedra f. typica
    Dendrobaena illyrica v. hintzei
    Lumbricus rubellus

Maupasella cepedei f. castanei Lom, 1961
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Lumbricus castaneus

Maupasella criodrili Heidenreich, 1935, illus.
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Criodrilus lacuum
    C. ochridensis

Maupasella cylindri Katashima, 1952
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Pheretima communissima

Maupasella dichogasteri de Puytorac et Rakotoarivelo 1965, illus.
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Dichogaster inermis

Maupasella dilatata de Puytorac et Rakotoarivelo 1965
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Dichogaster inermis

Maupasella glossoscolecis de Puytorac, 1952
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Glossoscolex grandis

Maupasella herculei de Puytorac, 1954
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Lumbricus herculeus

Maupasella lubeti de Puytorac, 1952
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Glossoscolex grandis (?)

Maupasella mucronata (Cepede, 1910) de Puytorac, 1954
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    synonymy
    Eiseniella tetraedra

Maupasella multistriata de Puytorac, 1954
  de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
    Allolobophora gigas
    A. savigni (?)

Maupasella nova Bathia et Gulatti, 1927
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Pheretima posthuma
P. hawayana

Maupasella nova proparte Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Maupasella mucronata (Cepede, 1910) nov. comb. de Puytorac, 1954

Maupasella nova proparte, Meier, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Maupasella mucronata (Cepede, 1910) nov. comb. de Puytorac, 1954

Maupasella nova p. parte Rossolimo, 1926 [et auct.]
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Maupasella cepedei de Puytorac, 1954

Maupasella pheretimae Katashima, 1952
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Pheretima communissima

Maupasella pontodrili sp. nov.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Maupasella sp. Wichterman, 1939
Pontodrilus bermudensis

Maupasella vacuolata Lom, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Pheretima rodericensis
P. hawayana

Maupasellidae Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryoidea nv. super-fam.
includes: Maupasellinae; Acanthophryinae n. s. fam.

Maupasellinae Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Maupasellidae
includes: Maupasella; Buchneriella; Georgevitchiella

Menospora polyacantha, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Agrion puella
Lestes viridis

Mesnilella, Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Mesnilellinae

Mesnilella aculeata Georgevitch, 1943
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Georgevitchiella aculeata (Georgevitch, 1943) de Puytorac, 1957

Mesnilella beani Cheissin, 1930, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Lamprodrilus isoporus: Baikal

Mesnilella bispiculata Rossolimo, 1926, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Rhynchelmis brachicephala f. typica

Mesnilella bohemica Lom, 1959, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Nais sp.

Mesnilella cepedei Andre, 1911 [? pro parte]
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella clavata (Leidy, 1855) Cepede, 1910

Mesnilella cepedei Andre, 1911 [? pro parte]
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella paranaidos (Pierantoni 1909) Heidenrich, 1935

Mesnilella clavata (Leidy, 1855) Cepede, 1910, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Lumbriculus variegatus

Mesnilella depressa Rossolimo, 1926
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella depressa (Rossolimo, 1926)

Mesnilella ? elongata (Delphy) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya elongata (Delphy, 1922) de Puytorac, 1957

Mesnilella elongata (Delphy) pp. Meier, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya elongata (Delphy, 1922) de Puytorac, 1957

Mesnilella fastigata (Mobius, 1888) Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Enchytraeus albidus

Mesnilella hynnis Georgevitch, 1941
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubificidae

Mesnilella kochi Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya elongata (Delphy, 1922) de Puytorac, 1957

Mesnilella maritui Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella maritui (Cheissin 1930)

Mesnilella multispiculata Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya multispiculata (Cheissin, 1930)

Mesnilella paranaidos (Pierantoni, 1909) Heidenrich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Paranais litoralis

Mesnilella radiata, Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya radiata (Cheissin 1930) de Puytorac, 1957

Mesnilella rostrata Rossolimo, 1926, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Styloscolex: Baikal
Teleuscolex: Baikal
Lamprodrilus: Baikal
Agriodrilus vermivorus: Baikal
Mesenchytraeus bungei

Mesnilella secans part. Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Mesnilella spiculata (Warpachowsky, 1886) Cepede, 1930, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Enchytraeus buchholzi

Mesnilella stammeri Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Dero limosa

Mesnilella trispiculata Kijenskij, 1925, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Lumbriculus variegatus

Mesnilella variabilis Rossolimo 1926
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella variabilis (Rossolimo, 1926)

Mesnilella vermicularis Cheissin, 1928
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella vermicularis (Cheissin, 1928)

Mesnilellinae Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae
includes: Mesnilella

Metaracoelophrya de Puytorac et Dragesco, 1968
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Metaracoelophryinae n. sous-fm.

Metaracoelophrya intermedia de Puytorac et Dragesco, 1968, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Metaracoelophryinae n. sous-fm.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam.
includes: Metaracoelophrya; Paracoelophrya; Coelophrya; Dicoelophrya

Metaradiophrya Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Metaradiophryinae n. s. fam.

Metaradiophrya almae de Puytorac et Dragesco, 1968, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Metaradiophrya asymmetrica Dale Beers, 1938
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eisenia loonbergi

Metaradiophrya bifulta de Puytorac, 1954, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Helodrilus schneideri

Metaradiophrya chlorotica Williams, 1942
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Allolobophora chlorotica

Metaradiophrya eminoscolecis de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eminoscolex

Metaradiophrya euoplos de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma

Metaradiophrya falcifera (Stein, 1861) Heidenreich, 1935, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Allolobophora caliginosa
Octolasium lacteum
Lumbricus rubellus
L. terrestris

Metaradiophrya gardneri Rees, 1961
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eisenia foetida

Metaradiophrya gigas de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Allolobophora gigas
A. savignyi

Metaradiophrya heidenreichi de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Helodrilus schneideri

Metaradiophrya hovassei de Puytorac, 1954, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eisenia loonbergi
Allolobophora chlorotica

Metaradiophrya lituiformis (Georgevitch, 1943) de Puytorac, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Hoplitophrya lituiformis Georg., 1943
Criodrilus lacuum
C. ochridensis

Metaradiophrya lumbrici (Dujardin, 1841), illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Lumbricus rubellus
L. herculeus

Metaradiophrya lumbrici (Duj.) p. p. Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Metaradiophrya lumbrici (Duj.) pp. Heidenreich 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya varians de Puytorac, 1954

Metaradiophrya lumbrici p. p. Meier, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Metaradiophrya lumbrici Williams, 1942
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya varians de Puytorac, 1954

Metaradiophrya varians de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Eisenia foetida

Metaradiophryinae n. s. fam.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam.
includes: Metaradiophrya; Cheissinophrya

Metasicuophora gen.n.
Albaret, J. L., 1973, J. Protozool., v. 20 (1), 51-57
Plagiotomidae
tod: M. petteri sp. n.

Metasicuophora petteri gen.n., sp.n. (tod), illus.
Albaret, J. L., 1973, J. Protozool., v. 20 (1), 51-57
Xenopus fraseri (rectum): La Maboke, Republique Centrafricaine

Microsporida
Kurtti, T. J.; and Brooks, M. A., 1976, Invert. Tissue Culture, Applications Med., Biol., and Agric. (Kurstak and Maramorosch), 395-398
propagation of Microsporida, experimental infection in cell line of Heliothis zea

Microsporida, undetermined sp. no. 1
Colley, F. C.; and Ow-Yang, C. K., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (2), 270-271 [Demonstration]
survey of microsporidan hyperparasites of trematode larvae from Malaysian snails, morphometric comparisons
Echinostoma dingeri

Microsporida, undetermined sp. no. 2
Colley, F. C.; and Ow-Yang, C. K., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (2), 270-271 [Demonstration]
survey of microsporidan hyperparasites of trematode larvae from Malaysian snails, morphometric comparisons
Echinostoma dunni

Microsporida [sp.]
Kurtti, T. J.; and Brooks, M. A., 1977, J. Invert. Path., v. 29 (2), 126-132
Microsporida [sp.], growth, development, and fumagillin sensitivity in vitro in moth (Heliothis zea) cell culture
Malacosoma disstria: northern Minnesota

Microsporida sp.
Lai, P. F.; Colley, F. C.; and Lim, H. K., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 132-133 [Demonstration]
microsporidan found in tissue of a symbiotic annelid (Chaetogaster sp.) living between the shell and mantle of laboratory-reared Lymnaea rubiginosa

Microsporida sp., probably undescribed Nosema sp.
Lai, P. F.; Colley, F. C.; and Lim, H. K., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 132-133 [Demonstration]
Lymnaea rubiginosa (hepatopancreas, heart, preputium, gizzard, mantle): Peninsular Malaysia

Microsporidan, illus.
Carter, J. B., 1976, J. Invert. Path., v. 27 (3), 409-410
Tipula paludosa (nerve ganglia): hill pasture, northeast England

Microsporidan
Daborn, G. R., 1976, Canad. J. Zool., v. 54 (7), 1161-1164
Branchinecta gigas: Fleeinghorse Lake, Alberta

Microsporidia
Vavra, J., 1976, J. Protozool., v. 23 (4), 21A [Abstract]
microsporidia, occurrence of paramural bodies (scindosomes)

Microsporidia
Vavra, J., 1977, J. Protozool., v. 24 (2), Suppl., 17A [Abstract]
microsporidian mitotic apparatus

Microsporidia
Weiser, J., 1976, J. Invert. Path., v. 28 (1), 147-149
microsporidian spores, staining nuclei, modified technique

Microsporidia sp.
Arvy, L.; and Sowa, R., 1976, Ann. Parasitol., v. 51 (1), 111-120
Baetis rhodani: region de Cracovie, Pologne

Microsporidia [sp.], illus.
Basch, P. F., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 380-383
possible venereal transmission between individual snails (Indoplanorbis exustus) of same generation as well as between generations since spores were found in the spermatheca, lumen of penis, in egg mass jelly and in newly hatched snails: West Malaysia

Microsporidia [sp.]
Carter, J. B., 1976, J. Applied Ecol., v. 13 (1), 103-122
Tipula paludosa: north-east England

Microsporidia [sp.]
Garris, G. I.; and Noblet, R., 1975, J. Med. Entom., v. 12 (4), 481-482
Mermithidae and Microsporidia as possible natural control agents of Simuliidae
Simulium vittatum
S. verecundum
S. venustum
S. tuberosum
Simulium sp.
Prosimulium magnum
Prosimulium sp.
all from South Carolina

Microsporidian, n. sp.
Gasc, C.; et al., 1976, Acta Trop., v. 33 (2), 169-176
description, effect on host
Habrodesmus falx (region anterieure: musculature du tube digestif, le tissu adipeux, les muscles segmentaires)
Oxydesmus granulosus
all from Sud-Dahomey

Microsporidian, illus.
Hubert, M., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 495-499
description
Cylindroiulus teutonicus (tissu adipeux; surface externe des tubes de Malpighi): Bretagne

Microsporidian
Lainson, R.; et al., 1976, Acta Amazonica, v. 6 (4), 55-60
Psychodopygus complexus: Municipio of Aripuana, Mato Grosso State, Brazil

Microsporidian-like organism
Michelson, E. H., 1976, J. Parasitol., v. 62 (4), 648-649
Schistosoma mansoni
Biomphalaria havanensis
all from Haiti

Microsporidian, "appears to be Plistophora sp."
Ramalingam, S.; Dissanaike, A. S.; and Fernando, M. A., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 148 [Demonstration]
microsporidian in Culex sitiens interfered with development of attempted experimental Plasmodium gallinaceum infections in the mosquito: Rantau Panjang area, West Malaysia

Microsporidian, illus.
Talukdar, J. N., 1976, Indian J. Entom., v. 36 (3), 1974, 232-236
Antheraea assamensis: Assam

Microsporidians
Weiser, J., 1975, J. Protozool., v. 22 (3), 70A [Abstract]
microsporidians in simuliids (including Pleistophora debaisieuxi in Odagmia ornata; and Glugea), provocation of neoplastic xenoma

Microsporidium [sp.]
Lainson, R.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 381 [Demonstration]
morphology
Lutzomyia, anduzei group (midgut): Monte Dourado area of North Brazil

Miescheria
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis

Miescheria cruzi, Hasselmann, 1926, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovicanis n. spec.

Miescheria cruzi, Hasselmann, 1926, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovifelis n. spec.

Miescheria cruzi, Hasselmann, 1926, pro parte
Heydorn, A. O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovihominis n. spec.

Miescheria cruzi Hasselmann 1926 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Miescheria cruzi Hasselmann 1926 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hirsuta Moule 1888

Miescheria cruzi Hasselmann 1926 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hominis (Railliet and Lucet 1891) comb. n.

Miescheria cruzi Hasselmann, 1926 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora cruzi (Hasselmann, 1926) comb. nov.

Miescheria cruzi Hasselmann, 1926 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hirsuta (Moule, 1887) comb. nov.

Miescheria cruzi Hasselmann, 1926 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hominis (Railliet & Lucet, 1891) comb. nov.

Miescheria muris Blanchard 1885
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis muris (Blanchard 1885) Labbe 1899

Miescheria muris Blanchard, 1885
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora muris (Blanchard, 1885) comb. nov.

Miescheria utriculosa, Harz, 1886, pro parte
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
as syn. of Sarcocystis suihominis n. spec.

Mimophrya de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Radiophryinae nov. comb.

Mimophrya acuta de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eminoscolex

Mimophrya communissima (Katashima, 1951) de Puytorac, 1970
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Radiophrya communissima Katashima, 1951
Pheretima communissima

Mimophrya elegans de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Eminoscolex

Mimophrya ovata (Katashima, 1951) de Puytorac, 1970
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Radiophrya ovata Katashima, 1951
Pheretima herpeiensis

Minchinia sp., illus.
Kern, F. G., 1976, J. Protozool., v. 23 (4), 498-500
description, sporulation
Crassostrea gigas: Republic of Korea

(?) Minchinia or Urosporidium sp., illus.
Newman, M. W.; Johnson, C. A. III; and Pauley, G. B., 1976, J. Invert. Path., v. 27 (3), 311-315
presence of haplosporosomes in cytoplasm suggests species of Minchinia or Urosporidium
Callinectes sapidus: Chincoteague Bay, Virginia; Beaufort, North Carolina

Minchinia nelsoni
Douglass, W. R.; and Haskin, H. H., 1976, J. Invert. Path., v. 27 (3), 317-323
Minchinia nelsoni disease development in susceptible oysters, Crassostrea virginica, alterations in hemolymph protein, aspartate and alanine aminotransferases, and phosphohexose isomerase, host metabolic changes; possible humoral defense mechanisms

Minchinia nelsoni Haskin, Stauber & Mackin
Farley, C. A., 1975, J. Protozool., v. 22 (3), 418-427
Minchinia nelsoni in Crassostrea virginica, epizootiologic relationships concerning parasite life cycle, pathologic effects on host, and effects of physical factors: mortality, incidence, seasonal prevalence of progressive disease stages, gross and histopathology, salinity and temperature effects, differences in host susceptibility between native and introduced hosts: Chesapeake Bay, Maryland

Mixtophrya de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Mixtophryinae n. s. fam.

Mixtophrya catenula de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Dero vaga: Gabon

Mixtophryinae n. s. fam.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae
includes: Mixtophrya

Mobilina Kahl
Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
includes: Urceolariidae, Trichodinopsidae fam. n., Trichodinidae

Monocercomonadidae Kirby, emend.
Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
Trichomonadida Kirby, emend.
diagnosis; includes: Monocercomonadinae Kirby, 1944, emend.; Protrichomonadinae Honigberg & Kulova, 1969; Dientamoebinae Grasse, emend.; Chilomitinae Honigberg, 1963; Hypotrichomonadinae Honigberg, 1963

Monocercomonadinae Kirby, 1944, emend.
Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
Trichomonadida Kirby, emend.; Monocercomonadidae Kirby, emend.

Monocercomonas sp.
Lindmark, D. G.; and Mueller, M., 1973, J. Protozool., v. 20 (4), 500
Tritrichomonas foetus, Monocercomonas sp., subcellular distribution of flavins, results in agreement with suggestion of flavin-mediated electron transport

Monocercomonas sp.
Lindmark, D. G.; and Mueller, M., 1974, J. Protozool., v. 21 (2), 374-378
Monocercomonas sp., oxidoreductases and hydrolases, activity, subcellular distribution, biochemical cytology very similar to the more highly evolved trichomonad Tritrichomonas foetus

Monocercomonas colubrorum
Honigberg, B. M., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 469-550
immunology of trichomonads, review

Monocercomonas colubrorum
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Monocercomonas testudinus
Kurashvili, B. E., 1975, Izvest. Akad. Nauk Gruzinsk. SSR, s. Biol., v. 1 (4), 317-320
antagonistic and synergetic interrelationships between intestinal parasites

Monocercomonoides chakravartii n. sp., illus.
Krishnamurthy, R.; and Sultana, T., 1976, Current Sc., Bangalore, v. 45 (5), 184-186
Polyphaga indica (gut): Maharashtra, India

Monocercomonoides marathwadensis n. sp., illus.
Krishnamurthy, R.; and Sultana, T., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (3), 109-115
Periplaneta americana (hind gut): Aurangabad, Maharashtra, India

Monocercomonoides oryctesae n. sp., illus.
Krishnamurthy, R.; and Sultana, T., 1977, Arch. Protistenk., v. 119 (1-2), 121-126
Oryctes rhinoceros (gut): Aurangabad, Maharashtra, India

Monocercomonoides polyphagae n. sp., illus.
Krishnamurthy, R.; and Sultana, T., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (3), 109-115
Polyphaga indica (hind gut): Aurangabad, Maharashtra, India

Monocercomonoides qadrii Bhaskar Rao 1969, illus.
Krishnamurthy, R.; and Sultana, T., 1977, Arch. Protistenk., v. 119 (1-2), 121-126
description
Oryctes rhinoceros (gut): Aurangabad, Maharashtra, India

Monocystella Valkanov, 1935
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis
Syn.: Pseudolankesteria Ormieres, 1965

Monocystella arndti (Valkanov, 1931) Valkanov, 1935
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Lankesteria arndti Valkanov, 1931

Monocystella compacta (Geus, 1967) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Lankesteria compacta Geus, 1967

Monocystella cyclopori (Poisson, 1921) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Lankesteria cyclopori Poisson, 1921; Pseudolankesteria cyclopori (Poisson, 1921) Ormieres, 1965

Monocystella discocelidis (Mingazzini, 1893) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Ophioidina discocelidis Mingazzini, 1893

Monocystella leptoplanae (Bhatia & Setna, 1924) comb. nov.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Leidyana leptoplanae Bhatia & Setna, 1924

Monocystella neodendrocoeli (Georgevitch, 1951) de Puytorac and Grain, 1960
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    synonymy

Monocystella plagiostomae (Georgevitch, 1951) comb. nov.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syns.: Monocystis plagiostomae Georgevitch, 1951; Lankesteria plagiostomae (Georgevitch, 1951) Geus, 1967

Monocystella planariae (Mingazzini, 1893) comb. nov.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syns.: Pleurozyga planariae Mingazzini, 1893; Lankesteria planariae (Mingazzini, 1893) Labbe, 1899; ? Monocystis planariae (Schultze) Lankester, 1863; Pseudolankesteria planariae (Mingazzini, 1893) Ormieres, 1965

Monocystella spelaea Codreanu & Balcesco, 1971
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Monocystella swarczewskyi Codreanu & Balcesco, 1971
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Lankesteria sp. Swarczewsky, 1910

Monocystis [sp.]
  Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
    Vulpes vulpes (feces): Wales; Exmoor

Monocystis sp., illus.
  McGhee, R. B.; and Pursglove, S. R., jr., 1977, J. Parasitol., v. 63 (1), 159-160
    gregarine with close similarity to Monocystis sp.
    Philohela minor (liver): Canaan Valley, Tucker County, West Virginia

Monocystis amaroecii Giard, 1873 lapsus calami for M. amaroucii
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    "Monocystis amaroecii Giard, 1873. Lapsus calami for Monocystis amaroucii and Syn. of Lankesteria amaroucii (Giard, 1873) Labbe, 1899."

Monocystis amaroucii
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    "Monocystis amaroecii Giard, 1873. Lapsus calami for Monocystis amaroucii and Syn. of Lankesteria amaroucii (Giard, 1873) Labbe, 1899."

Monocystis ascidiae Lankester, 1872
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891

Monocystis chagasi Adler & Mayrink, 1961
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Ascocystis chagasi (Adler & Mayrink, 1961) Tuzet & Rioux, 1966

Monocystis clavellinae (von Kolliker, 1848) Lankester, 1863
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lankesteria clavellinae (von Kolliker, 1848) Ormieres, 1965

Monocystis clymenellae Porter, 1897
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Pterospora clymenellae (Porter, 1897) Dogiel, 1909

Monocystis copiliae Rose, 1933
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Paraophioidina copiliae (Rose, 1933) comb. nov.

Monocystis holothuriae (Schneider, 1858) Lankester, 1863
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Urospora holothuriae (Schneider, 1858) Tregouboff, 1918

Monocystis lacrima Vejdovsky, 1882
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Paraophioidina lacrima (Vejdovsky, 1882) comb. nov.

Monocystis lacryma Vejdovsky, 1882 of Labbe (1899) lapsus calami
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Paraophioidina lacrima (Vejdovsky, 1882) comb. nov.

Monocystis legeri Blanchard, 1902
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Ascocystis legeri (Blanchard, 1902) comb. nov.

Monocystis mackiei Shortt & Swaminath, 1927
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ascocystis mackiei (Shortt & Swaminath, 1927) Ormieres, 1965

Monocystis mitis Leidy, 1882
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Schaudinnella enchytraei (von Kolliker, 1848) Meier, 1956

Monocystis nemertis (von Kolliker, 1845) Lankester, 1863
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora nemertis (von Kolliker, 1845) Schneider, 1875

Monocystis neodendrocoeli Georgevitch, 1951
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella neodendrocoeli (Georgevitch, 1951) de Puytorac and Grain, 1960

Monocystis plagiostomae Georgevitch, 1951
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella plagiostomae (Georgevitch, 1951) comb. nov.

Monocystis planariae (Schultze) Lankester, 1863
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
? as syn. of Monocystella planariae (Mingazzini, 1893) comb. nov.

Monocystis putanea (?) [sic] (Lachmann, 1859) Lankester, 1863
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Paraophioidina putanea [sic] (Lachmann, 1859) comb. nov.

Monocystis sagittata (Leuckart, 1861) Lankester, 1863
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Ancora sagittata (Leuckart, 1861) Labbe, 1899

Monocystis sipunculi (von Kolliker, 1845) Lankester, 1863
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora sipunculi (von Kolliker, 1845) Leger, 1892

Monocystis spionis (von Kolliker, 1848) Lankester, 1863
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
"Syn. of Polyrhabdina spionis (von Kolliker, 1848) Mingazzini, 1891 pro parte and of Selenidium spionis (von Kolliker, 1848) Ray, 1930 pro parte."

Monocystis telepsavi Stuart, 1871
Levine, N. D., 1974, J. Protozool., v. 21 (1), 8-9
as syn. of Selenidium telepsavi (Stuart, 1871) comb. nov.

Monodontophrya Vejdovsky, 1892
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Intoshellinidae

Monodontophrya kijenskyi Cheissin, 1930, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Eumonodontophrya kijenskyi Heidenreich, 1935
Tubifex inflatus

Monodontophrya longissima Vejdovsky, 1892
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Rhynchelmis limosella

Monodontophrya prolifera (Clap. et Lachm., 1858) Schuster, 1912 p. p.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya prolifera (Clap. et Lachm., 1858), Rossolimo, 1926

Monodontophrya rhynchelmis, Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Monodontophrya longissima Vejdovsky, 1892

Monoductidae Ray and Chakravarty, 1933 emended
Ramachandran, P., 1976, Zool. Anz., Jena, v. 196 (1-2), 125-132
characters emended

Mrazekiella Kijenskij, 1925
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Radiophryinae nov. comb.

Mrazekiella costata Kijenskij, 1925
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Branchiura coccinea var. inaequalis

Mrazekiella debaisieuxi (Delphy, 1922) Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Syn.: Anoplophrya debaisieuxi, Delphy, 1922
Clitellio arenarius

Mrazekiella intermedia Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mrazekiella costata Kijenskij, 1925

Mrazekiella nucleoramiformis Kijensky, 1928
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mrazekiella costata Kijenskij, 1925

Myriosporides Hennere, 1966
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Grelliidae fam. nov.
diagnosis, includes: M. amphiglenae Hennere, 1966 (type species)

Myxidium [sp.], illus.
Newman, M. W., 1977, Chesapeake Science, v. 18 (2), 240-242
Anguilla rostrata (small intestine, gill, body surface): Wicomico River, Wicomico County, Maryland

Myxidium sp., illus.
Reichenbach-Klinke, H. H.; and Sturm, G., 1975, Fisch u. Umwelt (1), 51-57
description, incidence
eel (Kiemen)

Myxidium bouixi n. sp., illus.
Siau, Y., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 563-570
Synodontis ansorgii (gall bladder): Porto-Novo (Dahomey)

Myxidium calcariferi Chakravarty 1943
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes paucispinis
S. miniatus
(gallbladder of all): all from off southern California

Myxidium coryphaenoidium Noble, 1966
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides leptolepis
C. filifer
all from eastern Pacific

Myxidium coryphaenoidium Noble 1966, illus.
Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
Myxidium coryphaenoidium, presence of 2 spore forms interpreted as result of polymorphism and not as mixed infection of 2 species; abnormal spores superficially resemble other Myxidium spp. and other genera (Myxobolus, Zschokkella, Auerbachia)
Coryphaenoides ariommus (gallbladder): off Chile
C. armatus (gallbladder): off Costa Rica; northern Mexico
C. carminifer (gallbladder): off Ecuador
C. fernandezianus (gallbladder): off Chile
C. filifer (gallbladder): off British Columbia; Oregon
C. leptolepis (gallbladder, bile ducts): off northern California; Oregon; Washington
C. nasutus (gallbladder): off Japan
C. rupestris (gallbladder): off Norway
Coelorinchus fasciatus (gallbladder, bile ducts): off Patagonia
C. innotabilis (gallbladder): off New Zealand
Gadomus multifilis (gallbladder): off Borneo
Lionurus proximus (gallbladder, bile ducts): off Japan
Nezumia bariger (gallbladder): off central Mexico
N. stelgidolepis (gallbladder): off southern California
N. liolepis (gallbladder): off southern California

Myxidium giardi, illus.
Ghittino, P.; Smith, F. G.; and Glenn, J. S., 1974, Riv. Ital. Piscicolt. e Ittiopatol., v. 9 (1), 13-18
histopathology, spore characteristics compared with various Myxidium spp.
Anguilla rostrata (dermis): freshwater tributaries of Chesapeake Bay

Myxidium incurvatum Thelohan 1892
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes carnatus: off central California
S. ensifer: off southern California
S. paucispinis: off southern California
S. serranoides: off central California
(gallbladder of all)

Myxidium iwamotoi sp. n., illus.
Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
Coryphaenoides carapinus (gallbladder): off Delaware
Ventrifossa potronus (gallbladder): off Chile

Myxidium lieberkuehni Buetschli, illus.
Reichenbach-Klinke, H. H.; and Sturm, G., 1975, Fisch u. Umwelt (1), 51-57

Myxidium macrourium sp. n., illus.
Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
Bathygadus melanobranchus (gallbladder): off French Guiana

Myxidium melanocetum Noble 1966
Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
Coelorinchus chilensis (gallbladder): off Chile
Nezumia latirostratus (gallbladder): off Ecuador
N. loricatus (gallbladder, bile ducts): off Chile
Ventrifossa atherodon (gallbladder): South China Sea

Myxidium pearcyi sp. n., illus.
Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
Coelorinchus macrohir (gallbladder): off Japan
Coryphaenoides armatus (gallbladder, bile ducts): off northern Mexico; northern California
C. leptolepis (gallbladder, bile ducts): off northern California; Oregon; Washington
C. mexicanus (gallbladder): Gulf of Mexico

Myxidium pearcyi Moser, Noble & Lee, 1976
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides leptolepis: eastern Pacific

Myxidium serotinum Kudo & Sprague, 1940, illus.
Clark, J. G.; and Shoemaker, J. P., 1973, J. Protozool., v. 20 (3), 365-366
Myxidium serotinum redescribed from Eurycea bislineata (gall bladder), trophozoites but no spores in tadpoles, trophozoites with spores in mature salamanders, no Salienta or other Caudata found infected: West Virginia

Myxidium truttae Leger, 1930
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Salmo trutta: Loch Leven, Scotland

Myxobilatus ansorgii n. sp.
Siau, Y., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 563-570
[p. 565 given in error for: M. synodontis n. sp.]

Myxobilatus ohioensis (Herrick 1941) Davis 1944
Cone, D. K.; and Anderson, R. C., 1977, J. Parasitol., v. 63 (4), 657-666
Lepomis gibbosus (lumina of ureters and urinary bladder): Ryan Lake, Algonquin Park, Ontario

Myxobilatus synodontis n. sp., illus.
Siau, Y., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 563-570
[p. 565 given in error as M. ansorgii n. sp.]
Synodontis ansorgii (gills): Porto-Novo (Dahomey)

Myxobolus
Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
Myxidium coryphaenoidium, presence of 2 spore forms interpreted as result of polymorphism and not as mixed infection of 2 species; abnormal spores superficially resemble other Myxidium spp. and other genera (Myxobolus, Zschokkella, Auerbachia)

Myxobolus spp.
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Phoxinus phoxinus: Loch Leven, Scotland

Myxobolus sp.
Heckmann, R.; and Farley, D. G., 1973, J. Wildlife Dis., v. 9 (3), 221-224
Hesperoleucus symmetricus symmetricus (gills): foothill streams east of Fresno, California

Myxobolus sp., illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
brief description
Bathygadus furvescens (urinary bladder): off Celebes

Myxobolus sp., illus.
Overstreet, R. M.; and Howse, H. D., 1977, Ann. N. York Acad. Sc., v. 298, 427-462
helminths and protozoans of estuarine fishes, incidence and intensity; possible relationships with water pollutants; tumorous growths
Mugil cephalus (scales): Akko, Israel

Myxobolus [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Mugil cephalus (gills, gut wall, and muscles): Elat, Gulf of Aqaba, Red Sea

Myxobolus dechtiari sp. n., illus.
Cone, D. K.; and Anderson, R. C., 1977, J. Parasitol., v. 63 (4), 657-666
Lepomis gibbosus (among basal cells at distal ends of gill lamellae): Ryan Lake, Algonquin Park, Ontario

Myxobolus dentium Fantham, illus.
George, E.; Harrison, E. J.; and Hadley, W. F., 1977, Tr. Am. Fish. Soc., v. 106 (5), 476-480
Myxobolus dentium in Esox masquinongy (cysts in palatine and vomerine teeth), incidence of infection, age of host, seasonal distribution: upper Niagara River

Myxobolus dispar
Kurashvili, B. E., 1975, Izvest. Akad. Nauk Gruzinsk. SSR, s. Biol., v. 1 (4), 317-320
antagonistic and synergetic interrelationships between intestinal parasites

Myxobolus ellipsoides bramaeformis (Dogel' & Akhmerov, 1960)
Aisa, E., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 517-524
Myxobolus ellipsoides bramaeformis, lesions in swimbladder of Tinca tinca: Lake Trasimeno

Myxobolus ellipsoides bramaeformis, illus.
Aisa, E., 1972, Parassitologia, v. 14 (2-3), 219-225
Myxobolus ellipsoides bramaeformis, infection levels similar in central and littoral portions of lake; infection of tench, Tinca tinca, localized mainly on gills, occasionally in swimbladder, rarely diffuse; slight pathological effects, no mortality; limnological data giving no ecological clue to distribution: lake Trasimeno

Myxobolus ellipsoides bramaeformis Dogel' e Akhmerov
Gattaponi, P., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 512-517
Tinca tinca: Lake Trasimeno

Myxobolus exiguus
Siau, Y., 1977, J. Protozool., v. 24 (4), 64A [Abstract]
Myxobolus exiguus, analysis of sulfur and calcium in spore

Myxobolus irinae sp. n., illus.
Daniiarov, M. R., 1975, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 18 (1), 67-68
Varicorhinus heratensis stendachneri
Barbus capito conocephalus
(kidney of all): all from Kafirnigan river, Tadzhik SSR

Myxobolus iucundus n. sp., illus.
Hine, P. M., 1977, J. Roy. Soc. N. Zealand, v. 7 (2), 157-161
Galaxias maculatus (musculature): Taumutu, Lake Ellesmere, South Island, New Zealand

Myxobolus koi Kudo, 1920
Hensley, G. H.; and Nahhas, F. M., 1975, Calif. Fish and Game, v. 61 (4), 201-208
Cyprinus carpio (gills): Sacramento-San Joaquin Delta, California

Myxobolus lairdi sp. n., illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Coryphaenoides rupestris (eye, brain): off Norway

Myxobolus lintoni, illus.
Overstreet, R. M.; and Howse, H. D., 1977, Ann. N. York Acad. Sc., v. 298, 427-462
helminths and protozoans of estuarine fishes, incidence and intensity; possible relationships with water pollutants; tumorous growth
Cyprinodon variegatus: Davis Bayou, Mississippi

Myxobolus magnaspherus sp. n., illus.
Cone, D. K.; and Anderson, R. C., 1977, J. Parasitol., v. 63 (4), 657-666
Lepomis gibbosus (parietal peritoneum of the kidney): Ryan Lake, Algonquin Park, Ontario

Myxobolus mexicanus Yoshino & Noble 1973
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Coelorinchus scaphopsis (kidney): Gulf of California
Coryphaenoides armatus (kidney): off northern California
Nezumia aequalis (kidney): Gulf of Mexico
Ventrifossa atherodon (gall bladder): off Hawaii

Myxobolus neurobius Schuberg and Schroder, 1905
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Salmo trutta: Loch Leven, Scotland

Myxobolus osburni Herrick 1936, illus.
Cone, D. K.; and Anderson, R. C., 1977, J. Parasitol., v. 63 (4), 657-666
description
Lepomis gibbosus (pancreatic tissue): Ryan Lake, Algonquin Park, Ontario

Myxobolus pfeifferi (Thelohan, 1895)
Willemse, J. J., 1968, Bull. Zool. Mus. Univ. Amsterdam, v. 1 (8), 83-87
Misgurnus fossilis (gonads): Doornenburg (Arnhem)

Myxobolus tadzhikistanica sp. n., illus.
Daniiarov, M. R., 1975, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 18 (1), 67-68
Schizothorax intermedius
Varicorhinus heratensis stendachneri
(spleen and kidney of all): all from Kafirnigan river; Chilu Chor Chashma spring, Tadzhik SSR

Myxobolus uvuliferis sp. n., illus.
Cone, D. K.; and Anderson, R. C., 1977, J. Parasitol., v. 63 (4), 657-666
Lepomis gibbosus (within fibrous tissue capsule surrounding metacercariae of Uvulifer amboplitis): Ryan Lake, Algonquin Park, Ontario

Myxobolus varicorhinii sp. nov., illus.
Dzhalilov, U. D.; and Daniiarov, M. R., 1975, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 18 (2), 68-69
Varicorhinus heratensis stendachneri (kidney and spleen)
Barbus capito conocephalus (skin)
all from Kafirnigan river basin and Chilu-Chor-Chasma spring, Shaartuzsk region, Tadzhik SSR

Myxoproteus abyssus Yoshino & Moser, 1974
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (4), 253-255
brief description
Bathygadus antrodes (urinary bladder): off Japan (east coast)
Coryphaenoides ariommus (gall bladder, urinary bladder, kidney): off Chile
C. armatus (urinary bladder): off northern California
C. cinereus (kidney): Bering Sea
C. fernandezianus (gall bladder): off Chile
C. filifer (urinary bladder): off Oregon
C. longifilis (urinary bladder): Bering Sea
C. acrolepis (gall bladder): off southern California; British Columbia
Coelorinchus occa (urinary bladder, kidney): off Lesser Antilles
Nezumia loricatus (kidney): off Galapagos Islands
Ventrifossa macropogon (urinary bladder): off Venezuela (Caribbean)

Myxoproteus californicus Yoshino & Noble, 1973
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides acrolepis: eastern Pacific

Myxoproteus californicus Yoshino & Noble, 1973
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (4), 253-255
brief description
Coryphaenoides acrolepis (urinary bladder, kidney): off northern Mexico (west coast); southern California; Oregon
C. pectoralis (urinary bladder, kidney): off northern California

Myxoproteus hubbsi new species, illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (4), 253-255
Coelorinchus coelorhincus carminatus (gall bladder): off Surinam
C. occa (gall bladder): off Lesser Antilles

Myxoproteus inexpectatus Schulman, 1966
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (4), 253-255
brief description
Hymenocephalus nascens (urinary bladder, kidney): off Borneo

Myxoproteus rosenblatti new species, illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (4), 253-255
Coryphaenoides acrolepis (gall bladder): off Oregon
C. ariommus (urinary bladder, kidney): off Chile
C. armatus (urinary bladder, kidney): off Costa Rica (west coast)
C. filifer (gall bladder): off Canada (west coast)

Myxosoma cerebralis
Daniels, S. B.; Herman, R. L.; and Burke, C. N., 1976, J. Protozool., v. 23 (3), 402-410
unidentified protozoon in Salmo gairdneri (epithelium) exposed for as little as 1 hour to water known to contain infective stages of Myxosoma cerebralis, fine structure and possible life cycle stages, relationship to M. cerebralis remains unresolved

Myxosoma cerebralis
Halliday, M. M., 1976, J. Fish Biol., v. 9 (4), 339-357
Myxosoma cerebralis (whirling disease in salmonids), geographic distribution, host range, life cycle, development and transmission, pathology, diagnosis, treatment and control, extensive review

Myxosoma cerebralis (Hofer, 1903)
Hewitt, G. C.; and Little, R. W., 1972, N. Zealand J. Marine and Freshwater Research, v. 6 (1-2), 1-10
Myxosoma cerebralis, life cycle, pathology, disease control
Salmo gairdneri: Otago Acclimatisation Society Hatchery at Waitati near Dunedin, New Zealand

Myxosoma cerebralis
Hoffman, G. L., 1975, J. Wildlife Dis., v. 11 (4), 505-507
Myxosoma cerebralis, disinfection of contaminated water with ultraviolet radiation at 2 dose levels, marked reduction in incidence and severity of whirling disease in Salmo gairdneri

Myxosoma cerebralis
Hoffman, G. L.; and Markiw, M. E., 1977, J. Fish Biol., v. 10 (2), 181-183
Myxosoma cerebralis, control of whirling disease, determination of minimum heat necessary to kill spores as demonstrated presumptively by staining with methylene blue

Myxosoma cerebralis
Hoffman, G. L.; and O'Grodnick, J. J., 1977, J. Fish Biol., v. 10 (2), 175-179
Myxosoma cerebralis, attempts to disinfect contaminated mud by air drying and applying hydrated lime, and contaminated water and mud with chlorine

Myxosoma cerebralis, illus.
Lunger, P. D.; et al., 1975, J. Parasitol., v. 61 (3), 476-480
Myxosoma cerebralis, mature spores, fine structure, thin-section electron microscopy

Myxosoma cerebralis
Munday, B. L., 1976, Austral. Vet. Practitioner, v. 6 (1), 46-50
brief review of protozoan and other infectious diseases. farmed trout: Australia

Myxosoma cerebralis
O'Grodnick, J. J., 1975, J. Wildlife Dis., v. 11 (1), 54-57
Myxosoma cerebralis, spore concentration in infected rainbow trout using continuous plankton centrifuge, effective diagnostic aid

Myxosoma (=Lentospora) cerebralis (Hofer) Plehn, 1905, illus.
Riechenbach-Klinke, H. H., 1975, Fisch u. Umwelt (1), 45-50
symptoms, transmission, distribution, control, therapy, review

Myxosoma cerebralis
Taylor, R. E. L.; and Haber, M. H., 1974, J. Wildlife Dis., v. 10 (4), 347-351
Myxosoma cerebralis-infected external opercular cysts in Salmo clarki henshawii raised in Myxosoma-contaminated water, cyst rupture could provide means of disease transmission from live naturally infected fish

Myxosoma cerebralis, illus.
Wolf, K.; and Markiw, M. E., 1976, J. Protozool., v. 23 (3), 425-427
Myxosoma cerebralis, in vitro sporulation, first report of culture of a myxosporidan

Myxosoma dahomeyensis n. sp., illus.
Siau, Y., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 563-570
Synodontis ansorgii (ovary): Porto-Novo (Dahomey)

Myxosoma dujardini Thelohan, illus.
Reichenbach-Klinke, H. H.; and Sturm, G., 1975, Fisch u. Umwelt (1), 51-57

Myxosoma funduli Kudo 1918
Knight, S. A.; Janovy, J., jr.; and Current, W. L., 1977, J. Parasitol., v. 63 (5), 897-902
Myxosoma funduli on Fundulus kansae (gills), summer epizootiology: parasite population overdispersed within host population, demographic characteristics of infected fish subpopulation virtually identical to those of whole fish population, distribution on individual gill bars, proportion of unilateral vs. bilateral infections, pre-existing infection does not preclude new infection: South Platte River, Nebraska

Myxosoma mulleri (Bond 1939)
Lee, R. L. G., 1977, Lond. Naturalist (1976) (56), 57-70
Perca fluviatilis (gills): Serpentine lake, Hyde Park and Kensington Gardens, central London

Myxosporida
Moser, M., 1977, Internat. J. Parasitol., v. 7 (5), 389-391
myxosporidan spores, size determined by parasite's development in host tissues vs. organ cavities, shape determined by presence of physiologically and behaviorally suitable host fish, constancy of spore size and shape a result of little selective pressure to change

Myxosporidian
McKenzie, R. A.; and Hall, W. T. K., 1976, Austral. Vet. J., v. 52 (5), 230-231
Strigeidae [sp.], mixed infection with myxosporidian spores and fungi, skin ulcers in Mugil cephalus, pathological findings: south-east Queensland

Naegleria
Cerva, L., 1973, J. Protozool., v. 20 (4), 535
Naegleria in vitro, sensitivity to brilliant and malachite green equal to amphotericin B

Naegleria, illus.
Cerva, L., 1975, Ang. Parasitol., v. 16 (3), 12 pp., (Merkbl. Ang. Parasitenk. u. Schadlingsbekampf.)
Naegleria, primary meningoencephalitis

Naegleria
Diamond, L. S.; and Mattern, C. F. T., 1976, Advances Virus Research, v. 20, 87-112
protozoal viruses, review

Naegleria
Dolensek, E. P.; Bihn, J. P.; and Napolitano, R. L., 1976, J. Protozool., v. 23 (2), 19A [Abstract]
Entamoeba invadens, Naegleria, and Hartmanella isolated from exhibited reptiles

Naegleria
Dunnebacke, T. H.; and Schuster, F. L., 1977, Am. J. Trop. Med. and Hyg., v. 26 (3), 412-421
Naegleria spp., pathogenic and non-pathogenic strains, partial isolation and characterization of cytopathogenic material indicates that the material is protein or has a protein component

Naegleria
Jadin, J. B.; and Willaert, E., 1972, Protistologica, v. 8 (4), 505-508
presence of limax group amoebae in swimming pools and drinking water system, possible control methods: Anvers

Naegleria
Kasprzak, W.; Mazur, T.; and Hadas, E., 1975, J. Protozool., v. 22 (3), 64A [Abstract]
Naegleria, strains pathogenic for mice isolated from fresh water lakes polluted thermally with effluents from industrial installation: [Poland ?]

Naegleria
Kramar, J.; and Cerva, L., 1975, J. Protozool., v. 22 (3), 65A
Naegleria, differentiation of pathogenic from nonpathogenic strains by indirect fluorescent antibody test

Naegleria
Martinez, A. J.; Markowitz, S. M.; and Duma, R. J., 1975, J. Infect. Dis., v. 131 (6), 692-699
electronmicroscopic and clinicopathologic features of pneumonia and encephalitis caused by Acanthamoeba castellanii and Acanthamoeba polyphaga in exper. mice, comparisons with infections caused by Naegleria

Naegleria
Visvesvara, G. S.; and Healy, G. R., 1973, J. Protozool., v. 20 (4), 522
pathogenic vs. free-living Naegleria, comparison by gel-diffusion and immunoelectrophoresis reactions

Naegleria sp., illus.
Kingston, N.; and Taylor, P. C., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 227-229
Physa gyrina (feces): Bear Lake, Wyoming

Naegleria spp.
Taylor, P. W., 1977, J. Parasitol., v. 63 (2), 232-237
isolation and experimental infection of free-living amebae in freshwater fishes, only Acanthamoeba polyphaga elicited pathological lesions
Tilapia nilotica (gill): Auburn University Experimental Station

Naegleria sp.
Willaert, E.; Jadin, J. B.; and Le Ray, D., 1972, Protistologica, v. 8 (4), 497-504
Naegleria, immunoelectrophoretic analysis of water soluble proteins, comparison of pathogenic and nonpathogenic strains, confirms existence of two separate species and gives evidence of homogeneity of pathogenic strains from different geographic areas, no antigenic relationship with Hartmannella castellanii or Entamoeba histolytica, study of antigenically intermediate strain between Naegleria gruberi and pathogenic Naegleria sp.

Naegleria aerobia, illus.
Maitra, S. C.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 56-60
Naegleria aerobia, ultrastructural differences in parasite grown in culture and from infected mouse brain

Naegleria aerobia, illus.
Maitra, S. C.; et al., 1976, Internat. J. Parasitol., v. 6 (6), 489-493
Naegleria aerobia, Hartmannella culbertsoni, comparison of ultrastructural characteristics in brain of mice with exper. primary amoebic meningoencephalitis, possible relationship of structure to pathogenicity

Naegleria aerobia Singh and Das
Visvesvara, G. S.; and Callaway, C. S., 1974, J. Protozool., v. 21 (2), 239-250
"N. fowleri by Carter, N. aerobia by Singh & Das and N. invades by Chang. . .all of these isolates belong to one species. . .we consider the pathogenic Naegleria species as N. fowleri, which takes precedence over the other names because it is the oldest available. . ."

Naegleria fowleri
Adams, A. C.; John, D. T.; and Bradley, S. G., 1976, Infect. and Immun., v. 13 (5), 1387-1391
Naegleria fowleri, mice (exper.), course of disease dependent upon infecting dose, no significant modification of resistance by bacterial endotoxin or other test substances, high resistance to challenge in mice that survived primary infection

Naegleria fowleri
Anderson, K. F., 1973, Med. J. Australia, v. 1 (18), 897-900
Naegleria fowleri, differential diagnosis and treatment of human meningitis

Naegleria fowleri
Cerva, L., 1975, J. Protozool., v. 22 (3), 60A [Abstract]
Naegleria fowleri, N. gruberi, axenic growth in liquid media in relation to temperature of incubation, concentration of culture medium, and size of inoculum, results show N. fowleri may produce dense populations under conditions which could occur in water which may become dangerous source of infection for human primary amebic meningoencephalitis

Naegleria fowleri
Cerva, L., 1977, J. Protozool., v. 24 (4), 50A [Abstract]
Naegleria fowleri and N. gruberi, growth characteristics in axenic culture compared

Naegleria fowleri, illus.
Cursons, R. T. M.; and Brown, T. J., 1976, N. Zealand J. Marine and Freshwater Research, v. 10 (2), 245-262
pathogenic free-living amebae, identification, classification, agents of primary amebic meningo-encephalitis

Naegleria fowleri
De Jonckheere, J., 1977, Applied and Environment. Microbiol., v. 33 (4), 751-757
Naegleria fowleri, use of an axenic medium for differentiation between pathogenic and nonpathogenic isolates, data obtained on 102 Naegleria strains during extensive ecological study gave further evidence that thermal polluted waters are the main origin of N. fowleri in the environment

Naegleria fowleri
De Jonckheere, J.; and Van de Voorde, H., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 10-15
Naegleria fowleri, evidence of proliferation of pathogenic strains in thermally polluted, biologically healthy water; isolates from discharges of thermal polluting factories highly virulent in mice (exper.), more amoeba present in summer than in winter: Belgium

Naegleria fowleri, illus.
De Jonckheere, J.; and Van de Voorde, H., 1977, J. Protozool., v. 24 (2), 304-309
Naegleria, 6 strains compared in structure, indirect fluorescent antibody test, effect of temperature on growth rate, effect of amphotericin B and fumagillin, growth and cytopathic effect upon Vero cell cultures, and pathogenicity for mice, results demonstrate occurrence of nonpathogenic variants of N. fowleri

Naegleria fowleri Carter, 1970
Diffley, P.; Skeels, M. R.; and Sogandares-Bernal, F., 1976, Ztschr. Parasitenk., v. 49 (2), 133-137
Naegleria fowleri, guinea pigs infected subcutaneously, tested with antigen derived from trophozoites, delayed hypersensitivity reaction; differences in immunocompetence of guinea pigs infected subcutaneously or intranasally discussed

Naegleria fowleri
Duma, R. J.; and Finley, R., 1976, Antimicrob. Agents and Chemotherapy, v. 10 (2), 370-376
Naegleria fowleri, Acanthamoeba castellanii, A. polyphaga, in vitro susceptibility to variety of therapeutic agents, results suggest that amphotericin B is most effective agent against Naegleria but few agents are effective against Acanthamoeba

Naegleria fowleri
Dunnebacke, T. H.; and Schuster, F. L., 1974, J. Protozool., v. 21 (2), 327-329
presence of cytopathogenic agent that is filterable and passageable only in strains of Naegleria (free-living gruberi as well as pathogenic fowleri) not in Acanthamoeba (pathogenic or free-living)

Naegleria fowleri
Dunnebacke, T. H.; and Schuster, F. L., 1977, Am. J. Trop. Med. and Hyg., v. 26 (3), 412-421
Naegleria spp., pathogenic and non-pathogenic strains, partial isolation and characterization of cytopathogenic material indicates that the material is protein or has a protein component

Naegleria fowleri, illus.
Feldman, M. R., 1977, Exper. Parasitol., v. 41 (2), 290-306
Naegleria fowleri, cytochemical assays using acid phosphatase and 3,3'-diaminobenzidine reveal extent to which lysosomes and heme proteins contribute to cytoarchitecture, data suggest that pathogen's rapid invasive behavior is result of availability of erythrocytes (primary source of nutriment) in host inflammatory reaction and combined mechanisms of enhanced extracellular destruction with subsequent phagocytosis

Naegleria fowleri
Griffin, J. L., 1973, J. Protozool., v. 20 (4), 497-498
Naegleria fowleri, environmental sampling, differential culture and preliminary identification

Naegleria fowleri
Hadas, E.; Kasprzak, W.; and Mazur, T., 1977, Tropenmed. u. Parasitol., v. 28 (1), 35-43
Naegleria fowleri, N. gruberi and strains of Hartmannella spp., taxonomic differentiation by means of disc electrophoresis of protein patterns

Naegleria fowleri
Jadin, J. B.; Hermanne, J.; and Willaert, E., 1972, Medecine et Malad. Infect., v. 2 (5), 205-210
Naegleria fowleri, human meningoencephalitis, case reports, diagnosis using spinal fluid inoculation into mouse brain and nasal instillation, need for prophylactic measures in addition to amphotericin B treatment

Naegleria fowleri
John, D. T.; and Martinez, A. J., 1975, J. Protozool., v. 22 (3), 39A [Abstract]
Naegleria fowleri, experimental primary amebic meningoencephalitis in mice inoculated intravenously

Naegleria fowleri
John, D. T.; Weik, R. R.; and Adams, A. C., 1977, Infect. and Immun., v. 16 (3), 817-820
mice, immunization with Naegleria fowleri or N. gruberi, protection against otherwise lethal N. fowleri challenge

Naegleria fowleri, illus.
Josephson, S. L.; Weik, R. R.; and John, D. T., 1977, Am. J. Trop. Med. and Hyg., v. 26 (5, part 1), 856-858
Naegleria gruberi, N. fowleri, concanavalin A-induced agglutination of N. gruberi but not N. fowleri indicating differences in polysaccharide structure of cell membranes, possible application to differentiating species and probing membrane properties associated with virulence

Naegleria fowleri
Karr, S. L., jr.; Wong, M. M.; and Brummer, M., 1977, J. Electr. Micr., v. 26 (2), 176 [Abstract]
Naegleria fowleri in monkey kidney cell cultures, inoculation of immune monkey serum prevented destruction of cells by amebae, light and scanning electron microscopy

Naegleria fowleri
Lastovica, A. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 286-287 [Letter]
Naegleria amoebae, ultrastructural differentiation of pathogenic and non-pathogenic agents

Naegleria fowleri, illus.
Lastovica, A. J., 1976, Ztschr. Parasitenk., v. 50 (3), 245-250
Naegleria fowleri, electron microscopy of 2 types of microfilaments, their significance in amoeboid motion

Naegleria fowleri
Lyons, T. B. III.; and Kapur, R., 1977, Applied and Environment. Microbiol., v. 33 (3), 551-555
limax amoebae in public swimming pools, concentration, correlations, and public health significance, preliminary tests for the human pathogen Naegleria fowleri were inconclusive: Albany, Schenectady, and Rensselaer counties, New York

Naegleria fowleri, illus.
Martinez, A. J.; and Rodriquez-Jurado, P., 1974, Patologia, v. 12 (4), 191-203
Naegleria fowleri as cause of primary amoebic meningoencephalitis in humans, clinical aspects, importance of diagnostic differentiation from Entamoeba histolytica

Naegleria fowleri
Molet, B.; et al., 1976, Ann. Parasitol., v. 51 (4), 401-406
no pathogenic strain of Naegleria fowleri found among 75 strains of free-living amoeba isolated from public drinking water supplies, swimming pools, and official swimming ponds in Strasbourg

Naegleria fowleri, illus.
Nicoll, A. M., 1973, N. Zealand Med. J. (496), v. 78, 108-112
Naegleria fowleri in man with resulting meningoencephalitis, case report, history of swimming in natural thermal pool, fatal illness: New Zealand

Naegleria fowleri, illus.
Schuster, F. L., 1975, J. Protozool., v. 22 (3), 352-359
Naegleria spp., comparison of cyst ultrastructure

Naegleria fowleri
Schuster, F. L.; and Dunnebacke, T. H., 1976, J. Protozool., v. 23 (2), 7A [Abstract]
Naegleria spp. lysates causing cytopathic effects in vitro

Naegleria fowleri, illus.
Schuster, F. L.; and Dunnebacke, T. H., 1977, J. Protozool., v. 24 (4), 489-497
Naegleria fowleri, mice, ultrastructural observations of experimental meningoencephalitis, intranuclear inclusions in amebae and host cells

Naegleria fowleri
Schuster, F. L.; and Rechthand, E., 1975, Antimicrob. Agents and Chemotherapy, v. 8 (5), 591-605
Naegleria fowleri, N. gruberi, amphotericin B, in vitro effects on growth, viability, and ultrastructure

Naegleria fowleri
Simione, F. P., jr.; and Daggett, P. M., 1976, J. Parasitol., v. 62 (1), 49
Naegleria fowleri, N. gruberi, successful freeze preservation of pathogenic and nonpathogenic Naegleria species

Naegleria fowleri
Stevens, A. R.; and Stein, S., 1977, J. Parasitol., v. 63 (1), 151-152
Acanthamoeba, Naegleria, pathogenic and nonpathogenic species (strains), susceptibility to lectin-induced agglutination

Naegleria fowleri, illus.
Visvesvara, G. S.; and Callaway, C. S., 1974, J. Protozool., v. 21 (2), 239-250
Naegleria fowleri, pathogenesis in mice and in monkey kidney cell cultures, observations show that amebas invade and destroy brain tissue by active phagocytosis
"N. fowleri by Carter, N. aerobia by Singh & Das and N. invades by Chang. . .all of these isolates belong to one species. . .we consider the pathogenic Naegleria species as N. fowleri, which takes precedence over the other names because it is the oldest available. . ."

Naegleria fowleri
Visvesvara, G. S.; Callaway, C. S.; and Healy, G. R., 1973, J. Protozool., v. 20 (4), 498
Naegleria fowleri, mechanism of pathogenesis in mouse brain and in monkey kidney cell cultures, light and electron microscopy, observations clearly show that amoebae invade and destroy brain tissue by active phagocytosis

Naegleria fowleri
Visvesvara, G. S.; and Healy, G. R., 1974, J. Protozool., v. 21 (3), 438 [Abstract]
Naegleria gruberi of Shumaker et al., 1971 identified as pathogenic strain of N. fowleri, first report of isolation from human nasal passages (7-year-old boy who previously swam in fresh-water lake in Virginia)

Naegleria fowleri
Visvesvara, G. S.; Healy, G. R.; and Jones, D. B., 1975, J. Protozool., v. 22 (3), 27A [Abstract]
Naegleria fowleri, 2 pathogenic strains, in vitro effect of amphotericin B methyl ester hydrochloride

Naegleria fowleri
Weik, R. R.; and John, D. T., 1977, J. Parasitol., v. 63 (1), 150-151
Naegleria fowleri, quantitation and cell size, electronic particle counting as highly sensitive, reproducible, and convenient method

Naegleria fowleri
Weik, R. R.; and John, D. T., 1977, J. Parasitol., v. 63 (5), 868-871
Naegleria fowleri, agitated axenic mass cultivation, growth kinetics, medium changes during growth, data suggest predominantly aerobic metabolism and utilization of non-carbohydrate sources as carbon and energy sources

Naegleria fowleri
Willaert, E. J. P.; De Jonckheere, J. F.; and Oktem, N., 1977, J. Parasitol., v. 63 (2), 389-390
Naegleria fowleri, immunoelectrophoretic identification of environmental pathogenic isolate from Belgium

Naegleria fowleri
Willaert, E.; and Stevens, A. R., 1976, Lancet, London (7988), v. 2, 741
Naegleria fowleri, isolation from artificially heated water (thermal-discharge water of electric power stations): Florida

Naegleria fowleri
Wong, M. M.; Karr, S. L., jr.; and Chow, C. K., J. Parasitol., v. 63 (5), 872-878
Naegleria fowleri maintained in vitro, changes in virulence

Naegleria gruberi
Cerva, L., 1975, J. Protozool., v. 22 (3), 60A [Abstract]
Naegleria fowleri, N. gruberi, axenic growth in liquid media in relation to temperature of incubation, concentration of culture medium, and size of inoculum, results show N. fowleri may produce dense populations under conditions which could occur in water which may become dangerous source of infection for human primary amebic meningoencephalitis

Naegleria gruberi, illus.
Cursons, R. T. M.; and Brown, T. J., 1976, N. Zealand J. Marine and Freshwater Research, v. 10 (2), 245-262
pathogenic free-living amebae, identification, classification, agents of primary amebic meningo-encephalitis

Naegleria gruberi
Hadas, E.; Kasprzak, W.; and Mazur, T., 1977, Tropenmed. u. Parasitol., v. 28 (1), 35-43
Naegleria fowleri, N. gruberi and strains of Hartmannella spp., taxonomic differentiation by means of disc electrophoresis of protein patterns

Naegleria gruberi
Jadin, J. B.; and Willaert, E., 1972, Protistologica, v. 8 (1), 95-100
Naegleria gruberi, isolation from 3 human cases of primary amoebic meningo-encephalitis, characteristics different from Naegleria isolated in swimming pools or in nature, theory about process by which pathogenicity is acquired, suggestions for control in swimming pools; literature review of cases of primary amoebic meningo-encephalitis: Anvers, Belgique

Naegleria gruberi
Lastovica, A. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 286-287 [Letter]
Naegleria amoebae, ultrastructural differentiation of pathogenic and non-pathogenic agents

Naegleria gruberi
Nagington, J.; and Richards, J. E., 1976, J. Clin. Path., v. 29 (7), 648-651
in vitro testing of 24 compounds (9 found promising) for possible cysticidal activity against Acanthamoeba polyphaga and A. castellanii (2 strains), comparison with results with Naegleria gruberi

Naegleria gruberi
O'Dell, D. S.; et al., 1976, Biochem. Soc. Tr., v. 4 (1), 124-125
Naegleria gruberi, amoeboid vs. flagellate locomotory phenotypes, cell surface receptors, reactions with antibodies, lectins, and cationized ferritin

Naegleria gruberi, illus.
Schuster, F. L., 1975, J. Protozool., v. 22 (3), 352-359
Naegleria spp., comparison of cyst ultrastructure

Naegleria gruberi
Schuster, F. L.; and Dunnebacke, T. H., 1976, J. Protozool., v. 23 (2), 7A [Abstract]
Naegleria spp. lysates causing cytopathic effects in vitro

Naegleria gruberi
Schuster, F. L.; and Rechthand, E., 1975, Antimicrob. Agents and Chemotherapy, v. 8 (5), 591-605
Naegleria fowleri, N. gruberi, amphotericin B, in vitro effects on growth, viability, and ultrastructure

Naegleria gruberi
Simione, F. P., jr.; and Daggett, P. M., 1976, J. Parasitol., v. 62 (1), 49
Naegleria fowleri, N. gruberi, successful freeze preservation of pathogenic and nonpathogenic Naegleria species

Naegleria gruberi (Schardinger)
Visvesvara, G. S.; and Balamuth, W., 1975, J. Protozool., v. 22 (2), 245-256
Acanthamoeba, Naegleria, Hartmannella, comparative studies on free-living and pathogenic amebae: cyst structure; nutrition; protein composition; immunology; cell free plaques and other cytopathic effects; phospholipase liberation; sensitivity to amphotericin B

Naegleria gruberi
Visvesvara, G. S.; and Healy, G. R., 1974, J. Protozool., v. 21 (3), 438 [Abstract]
Naegleria gruberi of Shumaker et al., 1971 identified as pathogenic strain of N. fowleri, first report of isolation from human nasal passages (7-year-old boy who previously swam in fresh-water lake in Virginia)

Naegleria invades
Visvesvara, G. S.; and Callaway, C. S., 1974, J. Protozool., v. 21 (2), 239-250
"N. fowleri by Carter, N. aerobia by Singh & Das and N. invades by Chang. . .all of these isolates belong to one species. . .we consider the pathogenic Naegleria species as N. fowleri, which takes precedence over the other names because it is the oldest available. . ."

Naegleria jadini, illus.
Schuster, F. L., 1975, J. Protozool., v. 22 (3), 352-359
Naegleria spp., comparison of cyst ultrastructure

Naegleria jadini
Schuster, F. L.; and Dunnebacke, T. H., 1976, J. Protozool., v. 23 (2), 7A [Abstract]
Naegleria spp. lysates causing cytopathic effects in vitro

Nellocystis Gates, 1933
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Aikinetocystidae
diagnosis; checklist of species

Nematocystis magna Schmidt, illus.
MacMillan, W. G., 1973, J. Protozool., v. 20 (2), 267-274
Nematocystis magna trophozoites, conformational changes in cortical region during peristaltic movements

Nematocystis magna Schmidt
MacMillan, W. G., 1974, J. Protozool., v. 21 (3), 451 [Abstract]
Nematocystis magna, types of movement, ultrastructural and cytochemical analysis

Nematocystis magna Schmidt
MacMillan, W. G., 1975, J. Protozool., v. 22 (3), 42A [Abstract]
nature of cell junction between Nematocystis magna and an epithelial cell of its host Lumbricus terrestris

Nematocystis magna Schmidt
MacMillan, W. G.; and Norton, M. Y., 1975, J. Protozool., v. 22 (3), 53A-54A [Abstract]
Nematocystis magna, technique for determination of surface area

Nematodomonas [n. gen.]
Nicoli, R. M.; and Penaud, A., 1971, Bull. Soc. Zool. France, v. 96 (1), 15-17
"nous proposons pour les Trypanosomatides monoxeniques des Nematodes le genre Nematodomonas avec pour espece type Nematodomonas goodeyi nom. nov. designant le flagelle parfaitement decrit mais non denomme par T. Goodey et M. J. Triffitt, 1927, saprobie du tube digestif du nematode Diplogaster longicauda . . ."

Nematodomonas n. gen.
Nicoli, R. M.; Penaud, A.; and Timon-David, P., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 405-415
Trypanosomatida
tod: N. goodeyi (R. M. Nicoli, 1965) [n. comb.]

Nematodomonas goodeyi nom. nov. (tod) [On the basis of information available in this article, this appears to be a new species; however, on the basis of Nicoli; Penaud; and Timon-David, [1972], it appears to be a new combination.]
Nicoli, R. M.; and Penaud, A., 1971, Bull. Soc. Zool. France, v. 96 (1), 15-17
"nous proposons pour les Trypanosomatides monoxeniques des Nematodes le genre Nematodomonas avec pour espece type Nematodomonas goodeyi nom. nov. designant le flagelle parfaitement decrit mais non denomme par T. Goodey et M. J. Triffitt, 1927, saprobie du tube digestif du nematode Diplogaster longicauda . . ."

Nematodomonas goodeyi (R. M. Nicoli, 1965) [n. comb.] (tod), illus.
Nicoli, R. M.; Penaud, A.; and Timon-David, P., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 405-415
Syns.: Leptomonas goodeyi R. M. Nicoli, 1965; innomine H. E. Ziegler, ?1895; innomine T. Goodey et M. J. Triffitt, 1927; innomine R. Ph. Dollfus, 1946

Nephridiophaga
Narasimhamurti, C. C.; and Kalavati, C., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (1), 20-25
genus should be retained

Nosema
Wille, H., 1976, Schweiz. Bienen-Ztg., v. 99 (10), 527-535
Nosema, bees, higher incidence correlated with decreased longevity, particularly in early months of the year, statistical analysis

Nosema
Zherebkin, M. V., 1976, Apiacta, v. 11 (1), 5-9
Nosema disease, resistance in honey bees, relationship of the enzyme chimozine to peritrophic membrane in midgut and protection against parasite

Nosema n. sp., spores
Daborn, G. R., 1976, Canad. J. Zool., v. 54 (7), 1161-1164
Nosema n. sp. in Branchinecta gigas and B. mackini, brief epizootics (May-June, 1970, 1971, 1972), host mortality, no effect on host egg production; possibly caused by or resulting in brief (1970, 1971) infection of Daphnia similis: Fleeinghorse Lake, Alberta

Nosema [sp.]
Desportes, I.; Ginsburger Vogel, T.; and Zerbib, C., 1976, J. Protozool., v. 23 (4), 8A-9A [Abstract]
Orchestia gammarellus (ovaire)

Nosema sp., illus.
Helms, T. J.; and Wedberg, J. L., 1976, J. Invert. Path., v. 28 (3), 383-384
Nosema-infected Loxagrotis albicosta (midgut epithelium), Bacillus thuringiensis treatment had beneficial not debilitative effect

Nosema [sp.]
Jakowska, S., 1964, An. 2. Cong. Latino-Am. Zool., v. 1, 265-273
Lophius americanus (celulas nervosas): Aquario de New York

Nosema sp.
Maddox, J. V., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 99-104
persistence of Microsporida, resistance of spores to environmental conditions, transmission and dispersal, host range and infectivity, effect of temperature on development
Acrosternum hilare

Nosema sp., illus.
Narayanan, K.; and Subramaniam, T. R., 1976, Current Sc., Bangalore, v. 45 (6), 223
Plusia peponis (fat body): Coimbatore, India

Nosema sp.
Nordin, G. L., 1976, J. Kansas Entom. Soc., v. 49 (1), 32-40
Malacosoma americanum eggs, decreased percentages of hatch and survival among Nosema-infected colonies, predisposal of infected larvae to other biotic and abiotic mortality factors: Kentucky

Nosema sp. of Margileth et al., 1973
Sprague, V., 1974, Tr. Am. Micr. Soc., v. 93 (3), 400-402
as syn. of Nosema connori n. sp.

Nosema sp.
Subrahmanyam, M., 1974, Indian J. Marine Sc., v. 3 (2), 182-183
Metapenaeus monoceros: Pulicat Lake, Adyar estuary, and Ennur estuary

Nosema sp., illus.
Walker, M. H.; and Hinsch, G. W., 1975, J. Parasitol., v. 61 (6), 1074-1080
Nosema sp., mature spore, structure, cytochemistry, changes in infected cells
Libinia dubia (reproductive tract): Bear Cut, Biscayne Bay, Florida

Nosema sp., illus.
Watanabe, H., 1976, J. Invert. Path., v. 28 (3), 321-328
Nosema sp. of Spodoptera litura, morphology, life cycle and development, host range, pathogenicity, comparisons with other Nosema spp.
Spodoptera litura: laboratory colony, Nisso Institute for Life Science, Oiso, Kanagawa, Japan
Pieris rapae crucivora (exper.)
Hyphantria cunea (exper.)
Chilo suppressalis (exper.)
Agrotis ipsilon (exper.)
Adoxophyes orana (exper.)

Nosema sp.
Weiser, J.; and Hostounsky, Z., 1974, J. Protozool., v. 21 (3), 459 [Abstract]
Colorado potato beetle (epithelium of the gut) (exper.)
Gastroidea viridula

Nosema acridophagus
Henry, J. E.; and Oma, E. A., 1974, Acrida, v. 3 (4), 223-231
Nosema locustae, N. acridophagus, N. cuneatum, virulence in Melanoplus bivittatus; N. locustae probably most useful for biological control

Nosema algerae, illus.
Undeen, A. H., 1975, J. Protozool., v. 22 (1), 107-110
Nosema algerae, growth in pig kidney cell cultures, early developmental stages described

Nosema algerae
Undeen, A. H., 1976, J. Invert. Path., v. 27 (3), 343-347
in vivo germination of Nosema algerae in mosquitoes, role in host susceptibility (Anopheles stephensi and A. albimanus most susceptible; A. quadrimaculatus and Culex pipiens relatively unsusceptible; A. atroparvus almost completely refractory), differences in midgut pH and passage rate through midgut not accountable for differences in spore germination percentages

Nosema algerae, illus.
Undeen, A. H.; and Alger, N. E., 1976, Exper. Parasitol., v. 40 (1), 86-88
Nosema algerae, white mice, spores subcutaneously injected into ears, tail, and feet, infections were transient and localized at infection sites, spore germination tests in blood plasma indicate it unlikely that spores injected by infected mosquito bite would result in infection

Nosema algerae
Undeen, A. H.; and Alger, N. E., 1977, Science Biol. J., v. 3 (1), 259-262
Nosema algerae in white mice (exper.), antibody detected by indirect fluorescent antibody test and by slide spore agglutination test, possible usefulness in safety evaluation phase of prospective microsporida biological control agents to determine mammalian exposure

Nosema algerae, illus.
Van Essen, F. W.; and Anthony, D. W., 1976, J. Invert. Path., v. 28 (1), 77-85
only 1 of 9 predators found susceptible after feeding on infected Anopheles quadrimaculatus larvae
Notonecta undulata (gut, muscle, fat body, Malpighian tubules, tracheal epithelium, testes, brain, hypodermis, ommatidia) (exper.)

Nosema anophelis Kudo, 1924
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania anophelis (Kudo 1924) [n. comb.]

Nosema apis Zander
dos Anjos, L. S.; and da Silva, R. M. B., 1973, Bol. Indust. Animal, Sao Paulo, v. 30 (1), 191-194
Apis mellifera (intestines): Pindamonhangala, Sao Paulo State

Nosema apis Zander
Clinch, P. G., 1976, N. Zealand J. Exper. Agric., v. 4 (1), 125-126
Nosema apis, rapid method of examining honey bees individually for spores

Nosema apis Zander, illus.
Hryniewiecka-Szyfter, Z.; and Banaszak, J., 1975, Bull. Soc. Amis Sc. et Let. Poznan, v. 15, s. D, 83-86
Nosema apis, histological investigation, mid-gut of infected bees (exper.), individual and mass feeding

Nosema apis
Lehnert, T., 1977, J. Apicult. Research, v. 16 (3), 163-164
Nosema apis in Apis mellifera, fumagillin, no detectable spores in queen mailing cages; need for entire colony of bees to be treated to prevent reinfection

Nosema apis, illus.
Liu, T. P., 1976, Apiacta, v. 11 (1), 10-13
Nosema apis, Fumidil B, ultrastructural alteration in spore membrane; effect on lipids of infected midgut epithelial cells of honey bees

Nosema apis
Niemczuk, R.; and Sobieszczanska, B., 1976, Med. Wet., v. 32 (5), 296-297
Nosema apis, Leptomonas apis, Malpighamoeba mellificae, occurrence in digestive tract of adult bees, disturbances in metabolism, decrease in productivity: Lower Silesia

Nosema apis
Rinderer, T. E.; and Elliott, K. D., 1977, J. Invert. Path., v. 30 (1), 110-111
Nosema apis, reduction in hoarding behavior of infected Apis mellifera

Nosema blissi sp. n., illus.
Liu, H. J.; and McEwen, F. L., 1977, J. Invert. Path., v. 29 (2), 141-146
Blissus leucopterus hirtus (Malpighian tubules): Guelph, Ontario

Nosema bombycis Naegeli
Skladzien-Bielatowicz, D., 1974, Roczniki Nauk. Zootech., v. 1, 17-23
Nosema bombycis, silkworm parasite, chloramine to kill spores, poor results

Nosema bombycis Naegeli, illus.
Takizawa, H.; Vivier, E.; and Petitprez, A., 1975, J. Protozool., v. 22 (3), 359-368
Nosema bombycis, cytochemistry, presence of nucleic acids, polysaccharides, and acid phosphatases demonstrated, localization in various stages of development from schizont to spore

Nosema campoletidis Brooks and Cranford, 1972, illus.
Brooks, W. M., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 105-111
Nosema campoletidis, N. heliothidis, N. cardiochilis, insect host-insect parasite-protozoan pathogen interrelationships, transovarian transmission of protozoans by insect parasites and hyperparasites of pest insects

Nosema cardiochilis Brooks and Cranford, 1972
Brooks, W. M., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 105-111
Nosema campoletidis, N. heliothidis, N. cardiochilis, insect host-insect parasite-protozoan pathogen interrelationships, transovarian transmission of protozoans by insect parasites and hyperparasites of pest insects

Nosema chagasi (Torres) 1927
Weiser, J., 1975, J. Protozool., v. 22 (3), 70A [Abstract]
microsporidians pathogenic to man

Nosema chapmani Kellen et al., 1967
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania periculosa (Kellen and Wills 1962)

Nosema connori n. sp.
Sprague, V., 1974, Tr. Am. Micr. Soc., v. 93 (3), 400-402
human, infant
Syn.: Nosema sp. of Margileth et al., 1973

Nosema connori
Weiser, J., 1975, J. Protozool., v. 22 (3), 70A [Abstract]
microsporidians pathogenic to man

Nosema costelytrae n. sp., illus.
Hall, I. M.; Oliver, E. H. A.; and Given, B. B., 1977, N. Zealand J. Zool., v. 4 (1), 1-3
Costelytra zealandica (fat body): Maihihi, Takapau and Te Awamutu, North Island, and Methven and Rakaia, South Island, New Zealand

Nosema cuneatum Henry
Henry, J. E.; and Oma, E. A., 1974, Acrida, v. 3 (4), 223-231
Nosema locustae, N. acridophagus, N. cuneatum, virulence in Melanoplus bivittatus; N. locustae probably most useful for biological control

Nosema cuniculi
Cox, J. C.; and Gallichio, H. A., 1977, Research Vet. Sc., v. 22 (1), 50-52
Nosema cuniculi, rabbits, serological diagnosis, indirect immunofluorescence test, compared with histopathological methods, no cross-reactivity between N. cuniculi and Toxoplasma gondii

Nosema cuniculi, illus.
Cox, J. C.; Walden, N. B.; and Nairn, R. C., 1972, Research Vet. Sc., v. 13 (6), 595-597
Nosema cuniculi, rabbits (urine, kidneys), clinical aspects and mortality, presumptive diagnosis using immunofluorescence technique: laboratory colony

Nosema cuniculi
Moffatt, R. E.; and Schiefer, B., 1973, Lab. Animal Sc., v. 23 (2), 282-284
Encephalitozoon cuniculi (Nosema cuniculi) causing typical encephalitic and nephritic lesions in apparently healthy guinea pigs in a closed laboratory colony, implications for experimental use of these animals

Nosema cuniculi
Mohn, S. F.; and Ødegaard, O. A., 1977, Acta Vet. Scand., v. 18 (2), 290-292
Nosema cuniculi, blue fox (Alopex lagopus), diagnosis by indirect fluorescent antibody test

Nosema cuniculi, illus.
Nordstoga, K.; and Westbye, K., 1976, Acta Path. et Microbiol. Scand., Sect. A., Path., v. 84A (3), 291-296
pathologic findings in polyarteritis nodosa associated with Nosema cuniculi in blue foxes, possible relationship to immunological disturbance

Nosema cuniculi, illus.
Somvanshi, R.; et al., 1977, Current Sc., Bangalore, v. 46 (8), 272 [Letter]
white laboratory mouse (brain): India

Nosema (Glugea) disstriae, illus.
Sohi, S. S.; and Wilson, G. G., 1976, Canad. J. Zool., v. 54 (3), 336-342
Nosema disstriae, cell lines developed from hemocytes and ovarian tissues of naturally infected Malacosoma disstria larvae, spores from hemocyte cultures infectious to host larvae, possible use in large-scale production of insect pathogens; ovarian cultures disappeared after several passages

Nosema disstriae
Wilson, G. G., 1977, Canad. Entom., v. 109 (7), 1021-1022
Nosema disstriae, Pleistophora schubergi, possible biological control agents of Malacosoma disstria (exper.), host survival

Nosema disstriae
Wilson, G. G.; and Sohi, S. S., 1977, Canad. J. Zool., v. 55 (4), 713-717
Nosema disstriae-infected and non-infected Malacosoma disstria hemocytes, continuous cultures, temperature effects on growth and viability

Nosema eurytremae (Canning and Basch, 1968) Canning, 1972, illus.
Canning, E. U.; Lai Peng Foon; and Lie Kian Joe, 1974, J. Protozool., v. 21 (1), 19-25
Fasciola hepatica (exper.)
Echinoparyphium dunni: Kuala Pilah, West Malaysia
Echinostoma hystricosum: kampung Gedok, West Malaysia
E. malayanum: kampung Gedok and kampung Ayer Kuning, West Malaysia

Nosema eurytremae Canning 1972, illus.
Colley, F. C.; Lim, H. K.; and Lie, K. J., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 142-143
Nosema eurytremae found in tissue of Bradybaena similaris (epicardium, pericardium) as well as hyperinfecting Postharmostomum gallinum, first evidence that Nosema life cycle can be completed in snail tissue

Nosema eurytremae
Higby, G. C.; and Canning, E. U., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 16 [Demonstration]
attempted use of Nosema eurytremae as biological control measure against larval stage of Fasciola hepatica

Nosema eurytremae
Higby, G. C.; Canning, E. U.; and Pilley, B. M., 1977, J. Protozool., v. 24 (4), 43A [Abstract]
Nosema eurytremae, cultivation through complete life cycle in vertebrate and invertebrate cell lines

Nosema eurytremae
Lai, P. F.; Ow-Yang, C. K.; and Lie, K. J., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (2), 277 [Demonstration]
Nosema eurytremae infections in Lymnaea rubiginosa (within tissues), comparison of infection rate in trematode free snails and those infected also with Echinostoma audyi

Nosema eurytremae
Lim, H. K.; et al., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 133 [Demonstration]
Nosema eurytremae, hyperparasite of Malaysian snails (Indoplanorbis exustus) also transmissible to several trematode species in Biomphalaria glabrata (exper.)

Nosema fumiferanae (Thom.)
Harvey, G. T.; and Gaudet, P. M., 1977, Canad. Entom., v. 109 (7), 987-993
Nosema fumiferanae in Choristoneura fumiferana in laboratory culture, benomyl, MBC phosphate, both not very effective in controlling microsporidian levels but had detrimental effects on budworm development and survival (reduced percentage of fertile matings and egg eclosions; male budworms more drug sensitive than females)

Nosema fumiferanae
Wilson, G. G., 1976, Canad. Entom., v. 108 (4), 383-386
Nosema fumiferanae, method of mass producing spores in Choristoneura fumiferana

Nosema gastroidei
Hostounsky, Z.; and Kodys, F., 1975, J. Protozool., v. 22 (3), 62A-63A [Abstract]
Nosema gastroidei causing mortality in Leptinotarsa decemlineata (exper.)

Nosema gastroideae
Weiser, J.; and Hostounsky, Z., 1974, J. Protozool., v. 21 (3), 459 [Abstract]
Colorado potato beetle (muscles, fat body, Malphigian tubules, muscularis of intestine) (exper.)
Gastroidea polygoni

Nosema gastroideae
Zizka, Z.; and Hostounsky, Z., 1977, J. Protozool., v. 24 (4), 54A-55A [Abstract]
Nosema gastroideae spores, polar filament and germ

Nosema gigantica sp. nov., illus.
Canning, E. U.; and Madhavi, R., 1977, Parasitology, v. 75 (3), 293-300
prevalence, hatching of spores, pathogenicity, possible mode of transmission
Allocreadium fasciatusi (parenchymatous tissue): Waltair, Andra Pradesh, India

Nosema heliothidis Lutz and Splendor, illus.
Brooks, W. M., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 105-111
Nosema campoletidis, N. heliothidis, N. cardiochilis, insect host-insect parasite-protozoan pathogen interrelationships, transovarian transmission of protozoans by insect parasites and hyperparasites of pest insects

Nosema heliothidis
Sikorowski, P. P.; and Lashomb, J. H., 1977, J. Invert. Path., v. 30 (1), 95-96
Nosema heliothidis spores isolated from Heliothis zea, loss of infectivity after exposure to sunlight

Nosema heterosporum
Kucera, M., 1976, Acta Entom. Bohemoslov., v. 73 (6), 367-372
Nosema heterosporum-infected Bararthra brassicae larvae, host enzymes, purification by isoelectric focusing, Michaelis constants determined, host metabolism

Nosema kingi
Armstrong, E., 1976, J. Invert. Path., v. 27 (3), 363-366
Nosema kingi, Drosophila willistoni (exper.), Fumidil B less deleterious to flies than benomyl, drug efficacy

Nosema kingi
Armstrong, E., 1976, Ztschr. Parasitenk., v. 48 (3-4), 275-280
Nosema kingi, longevity of Drosophila willistoni host, varied diets and age of host at time of infection, host susceptibility at various ages

Nosema kingi
Armstrong, E., 1976, Ztschr. Parasitenk., v. 50 (2), 161-165
Nosema kingi, fecal and transovarial transmission in Drosophila willistoni, infectivity to other hosts
Drosophila sturtevanti
D. melanogaster
D. persimilis
D. pseudoobscura
D. virilis
D. simulans
D. ananassae
D. nebulosa
(all exper.)

Nosema locustae Canning
Henry, J. E., 1972, Acrida, v. 1 (2), 111-120
Nosema locustae, epizootiology in grasshoppers, seasonal incidence, possible biological control agent
Melanoplus bivittatus
Melanoplus sanguinipes sanguinipes
Oedaleonotus enigma
Melanoplus f. femurrubrum
Melanoplus f. foedus
Pseudopomala brachyptera
Chorthippus curtipennis
Dissosteira carolina
Camnula pellucida
Aulocara elliotti
Amphitornus coloradus
Arphia p. pseudonietana
all from near Fairfield, Camas County, Idaho

Nosema locustae Canning
Henry, J. E.; and Oma, E. A., 1974, Acrida, v. 3 (4), 223-231
Nosema locustae, N. acridophagus, N. cuneatum, virulence in Melanoplus bivittatus; N. locustae probably most useful for biological control

Nosema necatrix Kramer, 1965
Fowler, J. L.; and Reeves, E. L., 1974, J. Protozool., v. 21 (4), 538-542
microsporidan isolate previously considered to represent mixed infection of Nosema necatrix and Thelohania diazoma determined to be one species with spore dimorphism, electrophoretic protein profile of two spore types, temperature effects on ratio of mono- to octospores, recommended that name Nosema necatrix be retained for this isolate
Trichoplusia ni (exper.)
Heliothis zea (exper.)

Nosema necatrix
Maddox, J. V., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 99-104
persistence of Microsporida, resistance of spores to environmental conditions, transmission and dispersal, host range and infectivity, effect of temperature on development
Pseudaletia unipuncta

Nosema necatrix
Maddox, J. V., 1977, Misc. Publication Entom. Soc. Am., v. 10 (3), 3-18
Nosema necatrix, N. trichoplusiae, viability of spores, effects of sunlight, temperature, water or humidity, substrate or chemicals; potential stability as biological control agents against insect pests

Nosema necatrix
Nordin, G. L., 1976, J. Invert. Path., v. 27 (3), 397-398
surface application technique for administering Nosema necatrix spores to third-instar Pseudaletia unipuncta larvae using bioassay containers

Nosema necatrix
Pilley, B. M., 1976, J. Invert. Path., v. 27 (3), 349-350
preservation of Nosema necatrix infective spores in Spodoptera exempta by lyophilization

Nosema parkeri sp. n., illus.
Krinsky, W. L., 1977, J. Protozool., v. 24 (1), 52-56
schizogonic and sporogonic stages described, transmission is transovarial and transstadial, no pathologic effect on tick host, failure to infect mice
Ornithodoros parkeri (nat. and exper.) (hemolymph, hypodermis, connective tissue, muscles, salivary glands, Malpighian tubules, midgut, central nerve mass, ovary, testes): laboratory colony, Rocky Mountain Laboratory, Montana
Dermacentor andersoni (exper.)

Nosema plodiae
Hostounsky, Z., 1977, J. Protozool., v. 24 (2), Suppl., 15A [Abstract]
spores, rate of germination after peroral administration to starved Mamestra brassicae V instar caterpillars

Nosema plodiae
Maddox, J. V., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 99-104
persistence of Microsporida, resistance of spores to environmental conditions, transmission and dispersal, host range and infectivity, effect of temperature on development
Plodia interpunctella

Nosema plodiae, illus.
Weiser, J., 1976, J. Invert. Path., v. 28 (1), 147-149
microsporidian spores, staining nuclei, modified technique
Mamestra brassicae (exper.)

Nosema polygrammae
Weiser, J.; and Hostounsky, Z., 1974, J. Protozool., v. 21 (3), 459 [Abstract]
Colorado potato beetle (epithelium of the gut) (exper.)
Polygramma undecimlineata

Nosema ptyctimae n. sp., illus.
Purrini, K.; and Baeumler, W., 1976, Anz. Schadlingsk., v. 49 (11), 169-171
Rhysotritia ardua (fat body, nephrocytes): Fichtenwald

Nosema pulvis
Vivares, C. P.; Bouix, G.; and Manier, J. F., 1977, J. Protozool., v. 24 (1), 83-94
Carcinus mediterraneus: etangs cotiers de la region du Languedoc mediterraneen

Nosema pulvis Perez, 1905, illus.
Vivares, C. P.; Loubes, C.; and Bouix, G., 1976, Ann. Parasitol., v. 51 (1), 1-14
Thelohania maenadis, Nosema pulvis, cytochemistry
Carcinus mediterraneus: divers etangs littoraux languedociens et catalans

Nosema pyrausta
Windels, M. B.; Chiang, H. C.; and Furgala, B., 1976, J. Invert. Path., v. 27 (2), 239-242
Nosema pyrausta, infected pupa and adult Ostrinia nubilalis, adverse effect on host longevity, oviposition, fecundity and fertility

Nosema stegomyiae Fox and Weiser (in part), 1959 [et auct.]
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania anophelis (Kudo 1924) [n. comb.]

Nosema stricklandi Jirovec, 1943, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Nosema strigeoideae, illus.
Palmieri, J. R.; Cali, A.; and Heckmann, R. A., 1976, J. Parasitol., v. 62 (2), 325-326
Diplostomum spathaceum in Lymnaea auricularia, experimental biological control by protozoan hyperparasite (Nosema strigeoideae)

Nosema takapauensis n. sp., illus.
Hall, I. M.; Oliver, E. H. A.; and Given, B. B., 1976, N. Zealand J. Zool., v. 3 (3), 257-260
schizogony, sporogony, spores
Costelytra zealandica (fat tissue): Takapau, southern Hawkes Bay, New Zealand

Nosema telengae sp. n., illus.
Simchuk, P. A., 1977, Zool. Zhurnal, v. 56 (1), 10-16
Laspeyresia pyrivora (tissues and organs): Bachoi and Pugoi, Moldaviia

Nosema transitellae sp. n., illus.
Kellen, W. R.; Hoffmann, D. F.; and Collier, S. S., 1977, J. Invert. Pathol., v. 29 (3), 289-296
life cycle, pathogenicity, ultrastructure, host range
Paramyelois transitella (fat, silk and salivary glands, midgut, muscle, and nerve tissue) (nat. and exper.): hosts reared on contaminated trash almonds in laboratory (these in-hull nuts were collected near Ballico, Madera County, California)
Tribolium castaneum (exper.)
Cadra cautella (exper.)
C. figulilella (exper.)
Ephestia elutella (exper.)
Galleria mellonella (exper.)

Nosema trichoplusiae
Maddox, J. V., 1977, Misc. Publication Entom. Soc. Am., v. 10 (3), 3-18
Nosema necatrix, N. trichoplusiae, viability of spores, effects of sunlight, temperature, water or humidity, substrate or chemicals; potential stability as biological control agents against insect pests

Nosema vasicola sp. n., illus.
Canning, E. U.; Lai Peng Foon; and Lie Kian Joe, 1974, J. Protozool., v. 21 (1), 19-25
Echinostoma hystricosum: rice fields near kampung Gedok, State of Negri Sembilan, West Malaysia

Nosema vivieri
Devauchelle, G.; and Vinckier, D., 1976, Ztschr. Parasitenk., v. 48 (3-4), 297-298 [Abstract]
Nosema vivieri in gregarines of nemerteans; bacteria in gregarines of Lithobia, earthworms and flourworms

Nosema vivieri Vinckier, Devauchelle & Prensier, 1970
Vinckier, D., 1975, J. Protozool., v. 22 (2), 170-184
as syn. of Nosemoides vivieri (Vinckier, Devauchelle & Prensier, 1970) comb. nov.

Nosematosis
Dade, A. W.; et al., 1977, J. Am. Vet. Med. Ass., v. 171 (9), 885-886
Baylisascaris spp. in Myocaster coypus (brain), complicated by nosematosis, case history, clinical and pathologic findings: city zoo, Grand Rapids, Michigan

Nosematosis
Madatov, R. I.; and Mershchiev, V. M., 1973, Vestnik Sel'skokhoz. Nauki Kazakhstana (11), 75-78
nosematosis, bees, chemotherapy, sulfapyridazine + chlortetracyclin most effective

Nosemoides gen. n.
Vinckier, D., 1975, J. Protozool., v. 22 (2), 170-184
"microsporidie"
mt: N. vivieri (Vinckier, Devauchelle & Prensier, 1970) comb. nov.

Nosemoides vivieri (Vinckier, Devauchelle & Prensier, 1970) comb. nov. (mt), illus.
Vinckier, D., 1975, J. Protozool., v. 22 (2), 170-184
Nosemoides vivieri comb. nov., life cycle, transformation of sporoblasts, organitogenesis of spore
Syn.: Nosema vivieri Vinckier, Devauchelle & Prensier, 1970
Lecudina linei: littoral boulonnais, Station de Biologie maritime, Wimereux

Nuttallia
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Nuttallia sp.
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
Tchitrea nigriceps
Anthropetes collaris
Cyanomitra olivacea
Amaurestes fringiloides
Ploceus baglafecht
Andropadus importunus
Pycnonotus xanthopygus
(blood of all): all from Tanzania

Nuttallia balearicae sp. n., illus.
Peirce, M. A., 1973, J. Protozool., v. 20 (5), 543-546
Balearica pavonina pavonina
B. pavonina gibbericeps
(blood of all): all captive in collections in Surrey, England

Nuttallia danii
Hadani, A.; et al., 1977, J. Protozool., v. 24 (4), 65A [Abstract]
Nuttallia danii, gerbils, relatively short span of time between tick (Hyalomma excavatum) transmission of parasite and its appearance in the blood makes the existence of exo-erythrocytic forms highly improbable

Nuttallia danii (Tsur, Hadani & Pipano, 1960)
Hadani, A.; Kauffman, B.; and Barnett, S. F., 1975, J. Protozool., v. 22 (3), 71A [Abstract]
Nuttallia danii, development in salivary glands of Hyalomma excavatum

Nuttallia danii (Tsur, Hadani & Pipano, 1960)
Hadani, A.; and Theodor, O., 1973, J. Protozool., v. 20 (4), 532
factors affecting the rate of infection of Hyalomma excavatum nymphs with Nuttallia danii: host splenectomy, stage of disease, route of transmission (blood vs. tick-borne)

Nuttallia meri
Gunders, A. E., 1974, J. Protozool., v. 21 (3), 464 [Abstract]
failure of Ornithodoros erraticus to transmit Nuttallia meri transovarially

Nuttallia meri
Gunders, A. E., 1974, J. Protozool., v. 21 (3), 464 [Abstract]
Nuttallia meri, transmission through bites of Ornithodoros erraticus up to 225 days after field collection of the ticks, ability to transmit on more than one occasion

Nuttallia meri
Gunders, A. E., 1974, J. Protozool., v. 21 (3), 464-465 [Abstract]
Nuttallia meri, transmission by laboratory-infected Ornithodoros erraticus, results suggest that only a proportion of clean ticks ever become infected after exposure

Nuttallia meri, illus.
Gunders, A. E., 1977, Experientia, v. 33 (7), 892-893
Nuttallia meri, infective forms first appear as spindle-shaped sporozoites in and around salivary glands of Ornithodoros erraticus 30 days after ticks had fed on infected blood (Psammomys obesus or hamsters)

Nuttallia meri
Gunders, A. E.; and Hadani, A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 536-538
Nuttallia meri, transmission to Psammomys obesus by field-collected Ornithodoros erraticus (both by intraperitoneal injection of triturated ticks and by bite), believed to be first recorded transmission of mammalian piroplasm by argasid tick: Jordan Valley

Nuttallia meri Gunders, 1971
Gunders, A. E.; and Hadani, A., 1974, Nature (5438), v. 247, 225-226
Nuttallia meri, demonstration of transmisson to Psammomys obesus by bite of field-collected Ornithodoros erraticus

Nuttallia rustica Jakunin et Krivkova, 1971, illus.
Subkhanov, M., 1975, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 18 (7), 52-53
Hirundo rustica (erythrocytes): foot of Khodzhi-Kazien, on right bank of river Vakhsh, Tadzhikistan

Nuttallia shortti
Peirce, M. A., 1973, J. Protozool., v. 20 (5), 543-546
"On the available evidence. . .regarded as valid avian piroplasm. . ."

Nycteria
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Nycteria congolensis
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Nycteria medusiformis
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Nyctotherus ampullariarum Cordero, 1928
Gascon, A., 1975, Rev. Biol. Uruguay, v. 3 (2), 111-125
as syn. of Parasicuophora ampullariarum (Cordero, 1928) n. comb.

Nyctotherus cordiformis
Jeserich, G., 1973, Mikrokosmos, v. 62 (4), 107-108
Opalina ranarum, Balantidium entozoon, Nyctotherus cordiformis, comparative fine structure morphology

Nyctotherus cordiformis
Lank, D. R., jr., 1971, Proc. Indiana Acad. Sc., v. 81 (2), 359-364
Rana catesbeiana: Indiana

Nyctotherus macropharyngeus Bezzenberger, 1904
Albaret, J. L., 1973, J. Protozool., v. 20 (1), 51-57
as syn. of Sicuophora macropharyngea (Bezzenberger, 1904) comb. nov.

Ochridanus Georgevitch, 1941
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Anthonyellinae n. s. fam.

Ochridanus kozarovi de Puytorac, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Limnodrilus hoffmeisteri

Ochridanus ocellatus (Georgevitch, 1941) de Puytorac, 1959, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubifex ochridanus f. typica

Octomitus Prowazek
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Diplomonadina emend.

Octomitus muris
Matthiesen, T.; Kunstyr, I.; and Tuch, K., 1976, Ztschr. Versuchstierk., v. 18 (3), 113-120
as syn. of Hexamita muris

Octosporea sp. "probably"
Lushbaugh, W. B.; Rowton, E. D; and McGhee, R. B., 1975, J. Protozool., v. 22 (3), 10A-11A [Abstract]
Drosophila melanogaster (intestine)

Octosporea adipovora n. sp.
Codreanu, R.; and Codreanu-Balcescu, D., 1976, J. Protozool., v. 23 (4), 7A [Abstract]
Antheraea pernyi (tissu adipeux)

Octosporea effeminans
Bulnheim, H. P., 1977, Biol. Zentralbl., v. 96 (1), 61-78
Thelohania herediteria and Octosporea effeminans in Gammarus d. duebeni (ovary) (nat. and exper.), feminized hosts, androgenic gland development inhibited, partially suppressed endocrine function: Gebiet der Elbmundung (Graben des Deichvorlandes bei Muggendorf)

Octosporea intestinalis n. sp. [nomen nudum]
Codreanu, R.; and Codreanu-Balcescu, D., 1975, J. Protozool., v. 22 (3), 77A-78A [Abstract]
Rhithrogena semicolorata (mesenteron antérieur): environs de la Station Zoologique de Sinaia, Carpates Meridionales, Roumanie

Octosporea muscaedomesticae Kramer
McLaughlin, R. E., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 95-98
entomopathogenic protozoa as control agents of insect pests, methods of application, economic factors, further research needs, review

Octosporea muscaedomesticae
Maddox, J. V., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 99-104
persistence of Microsporida, resistance of spores to environmental conditions, transmission and dispersal, host range and infectivity, effect of temperature on development Phormia regina

Octosporea muscaedomesticae Flu, 1911, illus.
Ormieres, R.; et al., 1976, J. Protozool., v. 23 (2), 320-328
Octosporea muscaedomesticae, various life cycle stages, ultrastructure
Ceratitis capitata (intestin) (nat. and exper.): laboratoires de l'INRA, Antibes, France

Octosporea muscaedomesticae
Teetor, G. E.; and Kramer, J. P., 1976, J. Invert. Path., v. 28 (1), 37-41
Octosporea muscaedomesticae, quantitative pathogenicity in Phormia regina, dosage effect study with reference to reproductive cycle of pathogen, relationship between number of spores ingested and subsequent infection rate

Octosporea muscaedomesticae
Teetor, G. E.; and Kramer, J. P., 1977, J. Invert. Path., v. 30 (3), 348-353
Octosporea muscaedomesticae, effect of ultraviolet radiation on naked spores, on various stages within host (Phormia regina), and on spores in host feces (survival, infectivity, development), uv-protective function ascribed to components provided by host's tissues and feces

Octosporea simulii Debaisieux, 1926, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Octosporella
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Octosporella sanguinolenti sp. n., illus.
Ovezmukhammedov, A., 1975, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (4), 87-88
Agama sanguinolenta (intestine): Turkmenistan

Oligochaetocystis pachydrili (Lankester, 1863) Meier, 1956
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
synonymy

Oodinioides vastator Reichenbach-Klinke
Herkner, H., 1975, Fisch u. Umwelt (1), 139-146
parasites as cause of disease in aquarium fish, review

Oodinium ocellatum, illus.
Herkner, H., 1975, Fisch u. Umwelt (1), 139-146
parasites as cause of disease in aquarium fish, review

Oodinium pillularis, illus.
Herkner, H., 1975, Fisch u. Umwelt (1), 139-146
parasites as cause of disease in aquarium fish, review

'Oodinium' pillularis
Lom, J., 1977, J. Protozool., v. 24 (2), Suppl., 16A [Abstract]
'Oodinium' pillularis trophonts, mode of attachment to gill surface of Puntius conchonius

Opalina
Mignot, J. P.; and Brugerolle, G., 1974, J. Protozool., v. 21 (5), 649-658
Opalina, Cepedea, ultrastructure of 2 different types of cysts, cortical morphogenesis

Opalina Intosh, 1872
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtamesnilella taupodrili (Kijenskij, 1925) de Puytorac

Opalina armata Stein, 1854
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Opalina filum Claparede, 1860
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Delphyella filum (Claparede, 1860) de Puytorac, 1969

Opalina inermis, Stein, 1859
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Acanthophrya inermis (Stein, 1859) Heidenreich, 1935

Opalina lumbrici Dujardin, 1841
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Metaradiophrya lumbrici (Dujardin, 1841)

Opalina lumbriculi Buchholz, 1863
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Opalina naidos part. Lankester, 1870
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Opalina naidos Schmidt, 1846
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya naidos (Schmidt, 1846) Heidenreich, 1935

Opalina pachydrili Clap., 1861
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya pachydrili (Claparede, 1861) Heidenreich, 1935

Opalina prolifera Clap et Lachm., 1858
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya prolifera (Clap. et Lachm., 1858) Rossolimo, 1926

Opalina ranarum
Jeserich, G., 1973, Mikrokosmos, v. 62 (4), 107-108
Opalina ranarum, Balantidium entozoon, Nyctotherus cordiformis, comparative fine structure morphology

Opalina secans Stein, 1859
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Opalina spiculata Warpachowsky, 1886
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mesnilella spiculata (Warpachowsky, 1886) Cepede, 1930

Opalinata
Sandon, H., 1976, Tr. Am. Micr. Soc., v. 95 (3), 357-366
opalinids with special reference to Protoopalina, problems in specific identification, speculations about evolution

Ophioidina Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Ophioidina bonelliae (Frenzel, 1885) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Gregarina bonelliae Frenzel, 1885

Ophioidina discocelidis Mingazzini, 1893
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella discocelidis (Mingazzini, 1893) comb. nov.

Ophioidina elongata Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lecudina elongata (Mingazzini, 1891) Kamm, 1922

Ophioidina haeckelii Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Paraophioidina haeckeli (Mingazzini, 1891) comb. nov.

Ophioidina heterocephala Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lecudina heterocephala (Mingazzini, 1891) Kamm, 1922

Ormieresia [gen. n.]
Vivares, C. P.; Bouix, G.; and Manier, J. F., 1976, J. Protozool., v. 23 (4), 21A-22A [Abstract]

Ormieresia gen. n.
Vivares, C. P.; Bouix, G.; and Manier, J. F., 1977, J. Protozool., v. 24 (1), 83-94
mt: Ormieresia carcini sp. n.

Ormieresia carcini [sp. n.]
  Vivares, C. P.; Bouix, G.; and Manier, J. F., 1976, J. Protozool., v. 23 (4), 21A-22A [Abstract]
    Ormieresia carcini, life cycle, ultrastructure
    Carcinus mediterraneus

Ormieresia carcini sp. n. (mt), illus.
  Vivares, C. P.; Bouix, G.; and Manier, J. F., 1977, J. Protozool., v. 24 (1), 83-94
    life cycle, ultrastructure
    Carcinus mediterraneus (musculature): Sete, Etang de Thau (Herault, France)

Pachyporospora retorta
Vivares, C. P., 1976, Ztschr. Parasitenk., v. 48 (3-4), 298 [Abstract]
Pachyporospora retorta, nucleus containing crystalloids composed of thousands of particles, possibly of viral origin
Macropipus vernalis (intestine): Tunis gulf

Pachysoma Mingazzini, 1891
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora Schneider, 1875

Pachysoma sipunculi (von Kolliker, 1845) Mingazzini, 1891
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
as syn. of Urospora sipunculi (von Kolliker, 1845) Leger, 1892

Paracoelophrya de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Metaracoelophryinae n. sous-fm.

Paracoelophrya almae de Puytorac, 1969
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma sp.

Paracoelophrya intermedia de Puytorac, 1969, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Alma emini

Paradinium caulleryi n. sp., illus.
Chatton, E.; and Soyer, M. O., 1973, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 15 (1), 27-60 [Posthumous article of Chatton, 1883-1947]
internal cycle, morphological development, systematic study shows probable relationship to mycetozoarians
Oncoea media: Cap Bear, Banyuls

Paradinium mesnili n. sp., illus.
Chatton, E.; and Soyer, M. O., 1973, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 15 (1), 27-60 [Posthumous article of Chatton, 1883-1947]
internal cycle, morphological development, systematic study shows probable relationship to mycetozoarians
Oncoea conifera: Cap Bear, Banyuls

Paradinium poucheti Chatton, illus.
Chatton, E.; and Soyer, M. O., 1973, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 15 (1), 27-60 [Posthumous article of Chatton, 1883-1947]
Paradinium poucheti, external cycle, morphological development, systematic study shows probable relationship to mycetozoarians
Acartia clausi: Banyuls-sur-Mer

Paradistigma Wita
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (1), 59-62
key to species parasitizing copepods, includes: Paradistigma ergasili; P. triangulatum; P. hyperparasiticum

Paradistigma triangulatum Wita
Michajlow, W.; and Wita, I., 1976, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 24 (2), 113-116
Ergasilus sieboldi and its hyperparasite Paradistigma triangulatum, survival in water contaminated with Vapam at various concentrations

Paragonospora Lang, 1954
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Urosporidae
checklist of species

Parahaemoproteus fringillae, illus.
Trefiak, W. D.; and Desser, S. S., 1973, J. Protozool., v. 20 (1), 73-80
crystalloid inclusions in Leucocytozoon, Parahaemoproteus, and Plasmodium, lipid-protein nature, ultrastructural morphology

Parahaemoproteus velans, illus.
Trefiak, W. D.; and Desser, S. S., 1973, J. Protozool., v. 20 (1), 73-80
crystalloid inclusions in Leucocytozoon, Parahaemoproteus, and Plasmodium, lipid-protein nature, ultrastructural morphology

Parahistomonas wenrichi
Fine, P. E. M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 2-3 [Demonstration]
Parahistomonas wenrichi, in vitro maintenance at -79C

Parahistomonas wenrichi (Lund, 1963) Honigberg and Kuldova, 1969
Fine, P.E.M., 1975, J. Helminth., v. 49 (4), 229-243
Heterakis gallinarum in chickens with concomitant Parahistomonas wenrichi infections, worm fertility (egg production), worm mortality, inter-caecal distribution and migration tendencies of worms with reference to probability of pairing of males and females

Parahistomonas wenrichi
Lund, E. E.; and Chute, A. M., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 73-76
larval Heterakis gallinarum, reciprocal transfer between Phasianus colchicus and Coturnix coturnix japonica, influence of early development in incompatible host on subsequent development; effect on liberation of Histomonas meleagridis and Parahistomonas wenrichi and transmission of H. meleagridis

Parahistomonas wenrichi
Lund, E. E.; and Chute, A. M., 1975, J. Protozool., v. 22 (3), 27A-28A [Abstract]
Parahistomonas wenrichi, factors in transmission by Heterakis gallinarum

Parahistomonas wenrichi
Madsen, H., 1976, Norwegian J. Zool., v. 24 (4), 460 [Abstract]
as syn. of Histomonas meleagridis (Smith, 1895) Tyzzer, 1920

Paramoeba perniciosa, illus.
Johnson, P. T., 1977, J. Invert. Pathol., v. 29 (3), 308-320
Paramoeba perniciosa in Callinectes sapidus, seasonal and geographic distribution, symptoms and diagnosis, tissue location, morphology, and replication, pathology, host defense reactions, laboratory transmission
Callinectes sapidus (connective tissues and hemal spaces of various organs, circulating blood) (nat. and exper.): Chincoteague Bay, Virginia; Long Island Sound, Connecticut; Sandy Hook Bay, New Jersey

Paramoeba perniciosa
Sawyer, T. K., 1976, Tr. Am. Micr. Soc., v. 95 (2), 271 [Abstract]
as cause of phagocytic nodules in crustaceans
Cancer irroratus
Homarus americanus
all from Sandy Hook Bay, New Jersey and/or New York Bight apex

Paranaplasma
Kreier, J. P.; and Ristic, M., 1973, Proc. 6. National Anaplasmosis Conf. (Las Vegas, Nevada, March 19-20, 1973), 24-28
Rickettsiales, Anaplasmataceae

Paranaplasma caudatum
Gallo, C.; and Balbo, S. M., 1976, Riv. Zootecn. e Vet. (3), 229-245
classification, morphology, epizootiology, transmission, vectors, pathogenicity, immunity, diagnosis, prophylaxis, review

Paranaplasma caudata, illus.
Simpson, C. F., 1977, Am. J. Vet. Research, v. 38 (10), 1509-1513
Paranaplasma caudata, calves, morphologic alteration of paranaplasma bodies following treatment with oxytetracycline; blood and liver biopsy specimens

Paraophioidina gen. n.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
tod: Paraophioidina haeckeli (Mingazzini, 1891) comb. nov.

Paraophioidina apsteini (Theodorides & Desportes, 1972) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Ganymedes apsteini Theodorides & Desportes, 1972

Paraophioidina apsteini (Theodorides & Desportes, 1972) Levine, 1977
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Ganymedes apsteini Theodorides and Desportes, 1972

Paraophioidina copiliae (Rose, 1933) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Monocystis copiliae Rose, 1933

Paraophioidina haeckeli (Mingazzini, 1891) comb. nov. (tod)
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Ophioidina haeckelii Mingazzini, 1891; Ganymedes haeckeli (Mingazzini, 1893) Theodorides & Desportes, 1972

Paraophioidina haeckeli (Mingazzini, 1893) Levine, 1977
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Ganymedes haeckeli (Mingazzini, 1893) Theodorides and Desportes, 1972

Paraophioidina lacrima (Vejdovsky, 1882) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Monocystis lacrima Vejdovsky, 1882; M. lacryma Vejdovsky, 1882 of Labbe (1899) lapsus calami

Paraophioidina oaklandi (Jones, 1968) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Ganymedes oaklandi Jones, 1968

Paraophioidina oaklandi (Jones, 1968) Levine, 1977
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Ganymedes oaklandi Jones, 1968

Paraophioidina pisae (Leger & Duboscq, 1911) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Porospora pisae Leger & Duboscq, 1911

Paraophioidina portuni (Frenzel, 1885) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Gregarina portuni Frenzel, 1885

Paraophioidina putanea [sic] (Lachmann, 1859) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Zygocystis putanea [sic] Lachmann, 1859; Monocystis putanea (?) [sic] (Lachmann, 1859) Lankester, 1863

Paraophioidina vibiliae (Theodorides & Desportes, 1972) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Ganymedes vibiliae Theodorides & Desportes, 1972

Paraophioidina vibiliae (Theodorides and Desportes, 1972) Levine, 1977
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Syn.: Ganymedes vibiliae Theodorides and Desportes, 1972

Parasicuophora
Albaret, J. L., 1973, J. Protozool., v. 20 (1), 51-57
definition

Parasicuophora ampullariarum (Cordero, 1928) n. comb., illus.
Gascon, A., 1975, Rev. Biol. Uruguay, v. 3 (2), 111-125
redescription
Syn.: Nyctotherus ampullariarum Cordero, 1928
Pomacea canaliculata (tubo digestivo): lago del Parque Rodo, ciudad de Montevideo

Parasicuophora corderoi n. sp., illus.
Gascon, A., 1975, Rev. Biol. Uruguay, v. 3 (2), 111-125
Pomacea canaliculata (tubo digestivo): lago del Parque Rodo, ciudad de Montevideo

Parasicuophora mantellae, illus.
Albaret, J. L., 1973, J. Protozool., v, 20 (1), 51-57

Parasicuophora xavierae sp.n., illus.
Albaret, J. L., 1973, J. Protozool., v. 20 (1), 51-57
Petropedetes natator (rectum): Mont Nimba, Liberia

Parastasia Michajłow
Michajłow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (1), 59-62
key to species parasitizing copepods, includes: Parastasia norvegica; P. fennica; P. parvicula; P. hanoiensis; P. helvetica; P. coelomae; P. cyclopis; P. sophiensis; P. bulgarica

Parathelohania Codreanu 1966
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
redescription

Parathelohania africanus sp. n., illus.
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
Anopheles gambiae (adipose tissue, oenocytes, ovaries): Nigeria

Parathelohania anomala (Sen 1941)
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
Syn.: Thelohania anomala Sen, 1941
Anopheles ramsayi (oenocytes, in area of adipose tissue): India

Parathelohania anophelis (Kudo 1924) [n. comb.], illus.
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
synonymy, description, reexamination
Anopheles quadrimaculatus (adipose tissue, oenocytes, ovaries): Georgia, New York, Louisiana, Florida, USA

Parathelohania illinoisensis (Kudo 1921), illus.
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
snyonymy, description, reexamination
Anopheles punctipennis (adipose tissue, oenocytes, ovaries): Illinois, Louisiana, Connecticut, USA; Quebec, Canada

Parathelohania indica (Kudo 1929)
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept.Agric., 1-26
synonymy
Anopheles hyrcanus (adipose tissue): India

Parathelohania legeri (Hesse 1904)
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
synonymy, description, reexamination
Anopheles maculipennis (adipose tissue, oenocytes, ovaries): France; Italy; Czechoslovakia

Parathelohania obesa (Kudo 1924), illus.
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
synonymy, description, reexamination
Anopheles crucians
A. quadrimaculatus
all from U. S. A.

Parathelohania obscura (Kudo 1929)
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
synonymy
Anopheles varuna: India

Parathelohania octolagenella sp. n., illus.
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
Anopheles pretoriensis (adipose tissue, oenocytes, ovaries): Nigeria

Parathelohania periculosa (Kellen and Wills 1962)
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
synonymy
Anopheles franciscanus (oenocytes, adipose tissue): California, U. S. A.

Paratrichodina
Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Trichodinidae

Pentatrichomonas gallinarum
Kulda, J.; Suchankova, E.; and Svoboda, S., 1974, Acta Vet. Brno, v. 43 (1), 53-64
as syn. of Tetratrichomonas gallinarum (Martin & Robertson 1911) Honigberg 1963

Pentatrichomonas hominis
Giboda, M.; Hoffman, F.; and Rimar, P., 1971, Ceskoslov. Pediat., v. 26 (2), 88-89
intestinal protozoal infections in children aged 1-15 years, clinical trials with metronidazole, results and parasites eliminated varied with age and parasite: Kosice, Czechoslovakia

Pentatrichomonas hominis
Honigberg, B. M., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 469-550
immunology of trichomonads, review

Perezia [sp.]
Jakowska, S., 1964, An. 2. Cong. Latino-Am. Zool., v. 1, 265-273
Lophius americanus (celulas nervosas): Aquario de New York

Perezia helminthorum
Colley, F. C.; Lie, K. J.; and Cheong Weng Hooi, 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 411 [Demonstration]
Aedes togoi
A. albopictus
A. aegypti
Anopheles balabacensis
(all exper.)

Perezia helminthorum
Colley, F. C.; Lie, K. J.; and Zaman, V., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 411 [Demonstration]
Perezia helminthorum, electron microscopy of spore

Perezia helminthorum
Colley, F. C.; and Ow-Yang, C. K., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (2), 270-271 [Demonstration]
survey of microsporidan hyperparasites of trematode larvae from Malaysian snails, morphometric comparisons
Trematod[a spp.]

Perezia helminthorum
Lie, K. J.; and Colley, F., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 410-411 [Demonstration]
Perezia helminthorum recovered from rediae of Echinostoma malayanum, stages in life cycle

P[erezia] helminthorum
Lie, K. J.; Kwo, E. H.; and Ow-Yang, C. K., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (2), 237-243
Schistosoma spindale, studies in biological control by trematode antagonism with Echinostoma malayanum, P[erezia] helminthorum infection of trematode larvae leading to suppression of cercarial production and reduction in vector snail population due to parasitic castration and high mortality of infected snails

Peritrichia sp. Calkins, 1933
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Esox lucius: Loch Leven, Scotland

Phytomonas davidi
Postell, F. J.; and McGhee, R. B., 1975, J. Protozool., v. 22 (3), 10A [Abstract]
transmission by hemipteron vector, ultrastructure of promastigotes
Pachybrachius bilobata scutellatus (salivary glands): Columbia County, Florida

Phytomonas elmassiani
Ayala, S. C.; de Quintero, O. B.; and Barreto, P., 1975, Rev. Biol. Trop., v. 23 (1), 5-15
Phytomonas elmassiani, naturally and experimentally infected Oncopeltus cingulifer and O. unifasciatellus, transmission of P. elmassiani to laboratory raised laticiferous plants (Asclepias curassavica, A. fruticosa): Colombia; Costa Rica

Phytomonas elmassiani, illus
Cosgrove, W. B., 1973, J. Protozool., v. 20 (2), 191-194
Trypanosomatidae, history and present status of knowledge of kinetoplasts, presidential address

Pirhemocyton [sp.], illus.
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses
Leptodactylus pentadactylus: Panama
Leptodactylus sp.: Venezuela
Sceloporus occidentalis: California, U.S.A.
Gehyra australis (blood): New South Wales, Australia
Egernia striolata (blood): New South Wales, Australia
Gehyra variegata (blood): New South Wales, Australia

Pirhemocyton [sp.]
de Sousa, M. A.; de Biasi, P.; and Pessoa, S. B., 1973, J. Protozool., v. 20 (4), 530
Bothrops alternatus (blood): Brazil

Pirhemocyton chelonarum n. sp., illus.
Acholonu, A. D., 1974, J. Protozool., v. 21 (5), 659-664
Chelydra serpentina serpentina
Pseudemys scripta elegans
P. floridana hoyi
Terrapene carolina carolina
T. carolina triunguis
(blood of all): all from southeastern Louisiana

Piroplasms
Van Peenen, P. F. D.; et al., 1977, J. Protozool., v. 24 (2), 310-312
piroplasms, probably geographic strain of Babesia microti [but those from bandicoot rats are also referred to as B. bandicootia]; occurrence and appearance; transtadial infection of Ixodes granulatus; infectivity for laboratory hosts; effect of syringe passage, splenectomy, and irradiation; pathology; cross-infection experiments; lack of susceptibility of bandicoot rats and spiny rats to Gray strain of B. microti
Bandicota indica nemorivaga (nat. and ? exper.): Taiwan
Rattus coxinga (nat. and exper.): Taiwan
R. norvegicus (exper.)
Mesocricetus auratus (exper.)
Meriones unguiculatus (exper.)
Bandicota indica setifera (exper.)
Ixodes granulatus: Taiwan

Piroplasma-like parasite
Svahn, K., 1976, Norwegian J. Zool., v. 24 (4), 460-461 [Abstract]
Lacerta agilis (erythrocytes): Bornholm, Denmark

Piroplasmosis
Dorchies, Ph., 1976, Rev. Med. Vet., Toulouse, v. 127 (11), 1523-1528
equine piroplasmosis, latent infection, problems of international competition and importation of horses, importation restrictions as control measure, possible alternative measures, discussed in relation to situation at Montreal Olympic Games

Piroplasmosis
Guilhon, J.; and Morel, P., 1976, Bull. Acad. Vet. France, v. 49 (1), 157-159
piroplasmosis or babesiosis, human: l'ouest de la France (Sarthe; Calvados)

Piroplasmosis
Iwata, A.; et al., 1976, Nippon Zyuisi-Kai Zassi (J. Japan. Vet. Med. Ass.), v. 29 (12), 673-675
piroplasmosis, Holstein cows, time interval between virulent blood inoculation and acquisition of immunity

Piroplasmosis
Ohshima, K.; et al., 1976, J. Fac. Agric., Iwate Univ., v. 13 (2), 161-176
viral disease in cattle previously or concurrently having fascioliasis, dictyocaulosis, and/or piroplasmosis, histopathological study of mucosal lesions, differential diagnosis: Japan

Plagiotoma lumbrici
Albaret, J. L., 1973, J. Protozool., v. 20 (2), 217
Plagiotoma lumbrici, somatic ciliature, morphogenesis

Plasmodium
Allison, A. C.; and Clark, I. A., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, Pt. 2), 216-222
specific and non-specific immunity to haemoprotozoa with emphasis on malaria and Babesia infections, workshop report: inherited differences in erythrocytes which influence susceptibility; specific responses of B- and T-lymphocytes to parasite antigens; macrophage activation

Plasmodium
Bannister, L. H., 1977, Symposia Brit. Soc. Parasitol., v. 15, 27-55
Plasmodium, invasion of red cells, symposium presentation: process of invasion (structure and formation of merozoites; release of merozoites from schizont; extracellular transit; adhesion to new host cell; invasion (theories of red cell deformation, removal of cell coat, passage of merozoite into parasitophorous vacuole, comparison with invasion in other genera of coccidians, recognition of red cell by merozoites); transformation of merozoites into trophic parasite); immunological aspects of invasion

Plasmodium
Capron, A.; et al., 1977, Ann. Immunol., v. 128C (1-2), 541-556
impairment of immune response in parasitic infections characterized by high prevalence of autoantibodies and by immunosuppression, review discussing malaria, trypanosomiasis, trichinosis, and schistosomiasis, with some original material on the last

Plasmodium
Clark, I. A.; Wills, E. J.; and Richmond, J. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 12 [Demonstration]
intra-erythrocytic death of Babesia and Plasmodium as host acquires immunity, possibly caused by soluble mediator liberated by host cells

Plasmodium
Cohen, S., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 283-286
malarial immunity, differences in immune mechanisms induced by malarial infection vs. merozoite vaccination, symposium presentation

Plasmodium
Cohen, S.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, Pt. 2), 223-232
malaria, role of humoral and cell-mediated mechanisms in specific acquired immunity to erythrocytic stage, current status of merozoite vaccination, workshop report

Plasmodium
Cohen, S.; Butcher, G. A.; and Mitchell, G.H., 1977, Advances Exper. Med. and Biol., v. 93, 89-112
immunization against erythrocytic forms of malaria parasites, review: malaria life cycle; innate and acquired resistance to malaria (specific malarial antibody, protective malarial antibody, synergistic action of malarial antibody and cells, role of specific cell-mediated immunity in malaria, isolation and properties of merozoites); vaccination against erythrocytic forms of malaria (vaccination and challenge using undefined and defined variants of Plasmodium knowlesi, adjuvant requirement for successful vaccination)

Plasmodium
Diamond, L. S.; and Mattern, C. F. T., 1976, Advances Virus Research, v. 20, 87-112
protozoal viruses, review

Plasmodium
Garnham, P. C. C., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 767-791
primate malaria, immunology, review

Plasmodium
Garnham, P. C. C., 1973, J. Protozool., v. 20 (1), 37-42
second roundtable discussion of taxonomic problems relating to malaria parasites

Plasmodium
Greiner, E. C.; et al., 1975, Canad. J. Zool., v. 53 (12), 1762-1787
avian hematozoa, prevalence with reference to distribution by geographic region, by host family, by vertical stratification of nesting sites and by feeding behavior of known vectors: North America north of Mexico
[Checklist includes 388 bird species and contains both published and unpublished records. For records from specific hosts, see entries in Supplement 22, Part 7, Hosts.]

Plasmodium
Guerra, F., 1977, J. Trop. Med. and Hyg., v. 80 (7), 135-140
Plasmodium, introduction of cinchona in the treatment of human malaria, historical review

Plasmodium
Houba, V., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, Pt. 2), 233-239
immunopathology mechanisms in protozoal infections, workshop report: malaria (nephropathies, splenomegaly); African trypanosomiasis

Plasmodium
Irvin, A. D.; and Young, E. R., 1977, Nature, London (5627), v. 269, 407-409
Babesia spp., in vitro test for screening drug activity which exploits finding that Babesia parasites readily incorporate hypoxanthine in an in vitro culture system, may also be used to detect drug resistance patterns and have possible application in primary screening of antimalarial drugs

Plasmodium
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium
Leeflang, P.; et al., 1976, Internat. J. Parasitol., v. 6 (2), 159-161
survey of 173 males using indirect fluorescent antibody technique with Babesia bovis, B. bigemina, and B. ratti as antigens, babesial antibodies detected in 54%, no correlation found with mode of living, contact with livestock, incidence of malaria parasitaemias or malarial antibodies, no Babesia organisms found in blood smears or by animal subinoculation, source of antibodies unknown but subclinical infection not likely: Nigeria

Plasmodium
Lepes, T., 1976, Bol. Ofic. San Panamer., v. 80 (3), 229-234
brief review of malaria research in the world

Plasmodium
Le Viguelloux, J.; and Barabe, P., 1974, Medecine Trop., v. 34 (5), 653-660
possible mechanisms of immunity in human Plasmodium and application to epidemiologic studies, review

Plasmodium
McGhee, R. B., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 331-369
immunity to avian malaria, review

Plasmodium
Mauel, J.; and Behin, R., 1974, Transplant. Rev., v. 19, 121-146
cell-mediated and humoral immunity to protozoan infections (leishmaniasis, malaria, trypanosomiasis), review

Plasmodium
Miller, L. H., 1977, Advances Exper. Med. and Biol., v. 93, 113-120
malaria, critique of merozoite and sporozoite vaccines

Plasmodium
Miller, L. H.; and Carter, R., 1976, Exper. Parasitol., v. 40 (1), 132-146
mechanisms of innate resistance in malaria, extensive review

Plasmodium
Nussenzweig, R. S., 1977, Advances Exper. Med. and Biol., v. 93, 75-87
immunoprophylaxis of malaria, sporozoite-induced immunity, review: sporozoite-induced protective immunity in rodent malaria, characteristics and possible mechanisms; simian malaria immunization (Plasmodium cynomolgi-rhesus monkey system); immunization of man against sporozoite-induced malaria

Plasmodium
Ristic, M.; and Smith, R. D., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 41-63
zoonoses caused by hemoprotozoa, review: trypanosomiasis, malaria, babesiosis

Plasmodium
Sinden, R. E.; Canning E. U.; and Toye, P. J., 1977, J. Protozool., v. 24 (4), 39A [Abstract]
Plasmodium, nuclear organization during gametogenesis

Plasmodium
Trigg, P. I.; and Gutteridge, W. E., 1977, Symposia Brit. Soc. Parasitol., v. 15, 57-81
parasitic Protozoa, morphological, biochemical, and physiological changes occurring during life cycles, symposium presentation

Plasmodium
Voller, A., 1975, Symposia Brit. Soc. Parasitol., v. 13, 69-84
immunopathology of malaria, extensive review: immunosuppression and auto-immunity; tropical splenomegaly syndrome; nephrotic syndrome

Plasmodium
  Walliker, D., 1976, Symposia Brit. Soc. Parasitol., v. 14, 25-44
    Plasmodium, genetic techniques (hybridization and cloning; genetic markers--enzyme polymorphism, drug-resistance, etc.), genetic recombination experiments, genetic factors influencing host-parasite relationships (strain-specific immunity; virulence), symposium presentation

Plasmodium
  Wan, Y. P.; Porter, T. H.; and Folkers, K., 1974, Proc. National Acad. Sc., v. 71 (3), 952-956
    inhibition of electron transfer mechanisms of Plasmodium by quinones, possible use as curative and prophylactic antimalarials, laboratory trials with experimental animals

Plasmodium
  Zuckerman, A., 1977, Exper. Parasitol., v. 42 (2), 374-446
    Plasmodium, immunology, extensive review: immunodiagnosis and seroepidemiology; immunopathology; antigenic analysis; host responses; immunoglobulins; cell-mediated reactions

Plasmodium [sp.}
  Andrews, S. E.; and Threlfall, W., 1975, Proc. Helminth. Soc. Washington, v. 42 (1), 24-28
    Corvus brachyrhynchos: insular Newfoundland

Plasmodium sp.
  Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
    Ixobrychus minutus
    Anthus similis
    Pycnonotus barbatus
    Laniarius funebris
    Turdus pelios
    Sylvia communis
    S. curruca
    Sylvietta brachyura
    Ploceus velatus
    Quelea erythrops
    Q. quelea
    Euplectes ardens
    Passer griseus
    Creatophora cinerea
    Oriolus larvatus
    all from Ethiopia

Plasmodium spp.
  Bannister, L. H., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 275-276
    Plasmodium spp., structural aspects relevant to, and prospects of, vaccination against human malarias, symposium presentation

Plasmodium sp., illus.
  Barnes, H. J., 1974, Vet. Rec., v. 95 (10), 218-219
    Plasmodium sp., turkeys (blood) (nat. and exper.), clinical symptoms: Zaria area, northern Nigeria

Plasmodium spp.
  Bennett, G. F.; et al., 1975, J. Wildlife Dis., v. 11 (2), 280-289
    hematozoa of Anatidae, prevalence survey
    Anas rubripes rubripes
    Anas platyrhynchos platyrhynchos
    Anas crecca carolinensis
    Anas discors
    Anas acuta
    Aix sponsa
    Aythya collaris
    Glaucionetta clangula
    Mareca americana
    Branta canadensis
    all from Maritime provinces of Canada

Plasmodium [sp.]
  Bennett, G. F.; and Blancou, J., 1974, J. Wildlife Dis., v. 10 (3), 239-240
    Saxicola torquata (blood): Madagascar

Plasmodium [sp.]
  Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
    Columbigalina talpacoti
    Psarocolius angustifrons
    (blood of all): all from Colombia

Plasmodium [sp.]
  Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
    Anas platyrhynchus (blood): Tanzania

Plasmodium [sp.] or Haemoproteus [sp.]
  Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
    (blood of all)
    Dioptrornis fischeri
    Phylloscapys trochilus
    all from Kenya

Plasmodium [sp.]
  Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
    survey, avian hematozoa, seasonal prevalence
    Ispidina picta
    Riparia cincta
    Tchagra australis
    Platysteira cyanea
    Cyanomitra olivacea
    Ploceus nigricollis
    Pycnonotus barbatus
    Ciccaba woodfordii
    Camaroptera brevicaudata
    Sylvia borin
    Trichastoma fulvescens
    Turdus olivaceus
    (blood of all): all from Uganda

Plasmodium sp.
  Bishop, C. A.; and Threlfall, W., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 25-35
    Somateria mollissima (blood): insular Newfoundland and/or southern Labrador

Plasmodium sp.
  Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
    Parus major (blood): Great Britain

Plasmodium [sp.]
  Choudhury, A.; and Misra, K. K., 1973, J. Protozool., v. 20 (4), 514
    Lanius schach collurio (blood): India

Plasmodium sp., characteristic of P. juxtanucleare
  Colley, F. C.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 84-85 [Demonstration]
    chickens (peripheral blood): Gombak, Selangor, Malaysia

Plasmodium [sp.]
  Crewe, S. M., 1974, J. Protozool., v. 21 (3), 449 [Abstract]
    species of malaria with morphological characters resembling those of Plasmodium circumflexum, quotidian periodicity of schizogony
    Streptopelia senegalensis: Nigeria
    Ploceus nigerrimus: Nigeria
    Ploceus cucullatus: Nigeria
    Serinus mozambicus (exper.)
    Serinus canarius (exper.)

Plasmodium [sp.]
  Crewe, S. M., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 477-478
    Aedomyia as possible vector of avian malaria
    Aedomyia africana (nat. and exper.): Nigeria
    Culex poicilipes: Nigeria
    Aedomyia furfurea (exper.)

Plasmodium sp.
  Das, S. K.; Strauss, J. M.; and Chee, C. T., 1966, Med. J. Malaya, v. 20 (4), 323
    Plasmodium inui-like infection presumably producing fatal illness in Indian leaf monkey (Presbytis johnii): National Zoo, Malaysia (natural habitat in Southern India)

Plasmodium sp.
  Forrester, D. J.; et al., 1974, J. Protozool., v. 21 (4), 494-497
    Meleagris gallopavo (blood): Florida

Plasmodium sp.
  Kocan, R. M.; and Perry, M. C., 1976, J. Wildlife Dis., v. 12 (1), 30-33
    Plasmodium sp. associated with high mortality in captive wild-trapped Aythya valisineria: Chesapeake Bay

Plasmodium sp., probably P. relictum
  Lewis, J. C.; Carpenter, J. W.; and Morrison, J. A., 1975, J. Wildlife Dis., v. 11 (4), 537-539
    Toxostoma rufum (blood): near Fairview, Major County, Oklahoma

Plasmodium sp.
  Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
    Butorides striatus amurensis
    Anas c. crecca
    Accipiter virgatus affinis
    Anborophila crudiigularis
    Bambusicola thoracica sonorivox
    Phasianus colchicus formosanum
    Turnix suscitator rostrata
    Gallinula chloropus indica
    Amaurornis phoenicurus chinensis
    Rostratula b. benghalensis
    Charadrius dominicus fulvus
    Streptopelia orientalis orii
    Otus bakkamoena glabripes
    O. spilocephalus hambroecki
    Corvus macrorhynchus colonorum
    Pomatorhinus ruficollis musicus
    Paradoxornis webbianus bulomachus
    Spizixos semitorques cinereicapillus
    Turdus c. chrysolaus
    Hypothymis azurea obholseri
    Lanius cristatus lucionensis
    L. schach formosae
    Zosterops japonica simplex
    Lonchura malacca formosana
    Passer montanus taiwanensis
    Emberiza spodocephala subsp.
    (blood of all): all from Taiwan

Plasmodium sp.
  Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
    Cyanocorax affinis
    Turdus fulviventris
    Euplectes a. afra
    E. orix franciscinana
    Ploceus ocularis
    Sporopipes frontalis
    Lonchura/Euodice cantans
    (blood of all): all imported to U. S., origin unknown

Plasmodium (Novyella) sp.
  Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
    Ramphastos vitellinus ariel
    Cyanocorax affinis
    Ramphocoelus carbo
    Uraeginthus angolensis
    (blood of all): all imported to U. S., origin unknown

Plasmodium [sp.], incertae sedis, illus.
  Nandi, N. C.; and Mandal, A. K., 1976, Indian J. Animal Health, v. 15 (2), 161-163
    description
    Hipposideros lankadiva (blood): forest near Khutimari, north-west of Gayerkata, Jalpaiguri District, West Bengal

Plasmodium sp.
  Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
    Terpsiphone paradisi: Orissa, India

Plasmodium sp.
  Noblet, R.; Moore, H. S. IV; and Noblet, G. P., 1976, Poultry Science, v. 55 (1), 447-449
    Quiscalus quiscula (blood): South Carolina

Plasmodium [sp.], resembled P. (Novyella) vaughani
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Coturnix delegorguei (blood): Kenya

Plasmodium spp.
Richards, W. H. G., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 279-280
Plasmodium spp., possible immunization against sporozoite challenge, review, symposium presentation

Plasmodium [sp.]
Thomas, S. E.; and Dobson, L. D., 1975, Onderstepoort J. Vet. Research, v. 42 (1), 67-68
Numida meleagris (blood): vicinity of Onderstepoort, Republic of South Africa

Plasmodium spp.
Trigg, P. I., 1976, Bull. World Health Organ., v. 53 (4), 399-406
Plasmodium spp., parasite cultivation in relation to chemotherapy of malaria with special application to chloroquine resistance, review of current aspects and new developments

Plasmodium sp.
Warren, M.; Mason, J.; and Hobbs, J., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 545-546
natural infections of Anopheles albimanus with Plasmodium in small malaria focus suggest that A. albimanus meets criteria as effective malaria vector: El Salvador

Plasmodia spp.
Wing, E. J.; and Remington, J. S., 1977, West. J. Med., San Francisco, v. 126 (1), 14-31
role of cell-mediated immunity (and enhancement by levamisole) in host defense against human parasitic diseases, extensive review

Plasmodium [sp.]
Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
Peromyscus maniculatus (blood): Chaco Canyon National Monument, San Juan County, New Mexico

Plasmodium spp.
Worms, M. J., 1972, Zool. J. Linn. Soc., London, v. 51, Suppl. 1, 53-67
microfilariae, Plasmodium spp., trypanosomes, rhythmic behavior, significance in relation to transmission, review

Plasmodium [sp.], illus.
Zaman, V.; and Visuvalingam, N., 1968, Med. J. Malaya, v. 22 (3), 228
Hylobates klossi

Plasmodium achiotense
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Basiliscus basiliscus: Panama

Plasmodium aegyptensis n. sp., illus.
Abd-El-Aziz, G. A.; Landau, I.; and Miltgen, F., 1975, Ann. Parasitol., v. 50 (4), 419-424
Arvicanthis niloticus (sang): region d'Assiut sur la rive Ouest du Nil, a 375 km au Sud du Caire, Haute-Egypte

Plasmodium anasum
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Anas clypeata (blood): Taiwan

Plasmodium atheruri
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium aurulentum
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis limifrons: Panama
Thecadactylus rapicaudus: Panama

Plasmodium balli
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis humilis: Costa Rica
A. limifrons: Costa Rica; Panama
A. lemurinus: Costa Rica; Belize
A. lionotus: Costa Rica; Panama
A. poecilopus: Panama

Plasmodium basilisci
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Basiliscus vittatus: Honduras
B. basiliscus: Panama
Iguana iguana: Honduras

Plasmodium beltrani, illus.
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Sceloporus teapensis: Belize

Plasmodium berghei
Alger, N. E.; and Harant, J., 1976, Exper. Parasitol., v. 40 (2), 261-268
Plasmodium berghei, mice, vaccination, heat-treated sporozoites administered iv; attempted adoptive transfer of immunity; preliminary sporozoite fractionation studies

Plasmodium berghei
Alger, N. E.; and Harant, J., 1976, Exper. Parasitol., v. 40 (2), 269-272
Plasmodium berghei, mice immunized by repeated ip injections of normal mosquito salivary glands or heads were protected from ip sporozoite challenge but not from iv sporozoite challenge, suggested that hypersensitive Type 1 reaction may explain part of this protection

Plasmodium berghei
Alger, N. E.; and Harant, J., 1976, Exper. Parasitol., v. 40 (2), 273-280
Plasmodium berghei, mice, concluded that hypersensitivity may possibly be at least partly responsible for protection by injections of 70 mosquito salivary glands but that sporozoite immunity is not primarily due to hypersensitivity

Plasmodium berghei
Alger, N. E.; and Harant, J., 1976, Science Biol. J., v. 2 (3), 89-103
Plasmodium berghei, mice, vaccination, comparison of x-irradiated, heat-treated, or formalin-treated sporozoites administered intravenously, intraperitoneally, intracutaneously, or intramuscularly, some groups given BCG, sodium alginate, or Freund's complete adjuvant

Plasmodium berghei
Alger, N. E.; and Harant, J., 1976, Science Biol. J., v. 2 (3), 104-110
Plasmodium berghei, mice, vaccination, frozen-thawed sporozoites given IP or IV, comparison with lyophilized, heated, and formalin-treated sporozoites

Plasmodium berghei
Areekul, S.; et al., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (4), 492-498
Plasmodium berghei, erythrocyte adenosine triphosphate levels in infected mice (exper.) compared with levels in normal mice, levels in infected mice found to be considerably higher

P[lasmodium] berghei
Areekul, S.; et al., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (3), 452 [Demonstration]
P[lasmodium] spp., erythrocyte adenosine triphosphate content compared in controls and infected man, monkeys and mice

Plasmodium berghei
Barnwell, J. W.; and Desowitz, R. S., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 429-433
Plasmodium berghei, mice (exper.), identification of indicators of impending parasitic crisis (penultimate stage)

Plasmodium berghei
Barr, M. L.; et al., 1977, Cellular Immunol., v. 33 (2), 447-451
Plasmodium berghei, transfer of immunity to mice by RNA from spleens and lymph nodes of immune rats

Plasmodium berghei
Beaudoin, R. L.,; et al., 1976, Exper. Parasitol., v. 39 (3), 438-443
Plasmodium berghei berghei ANKA strain, mice, immunization with irradiated sporozoites protected against challenge with sporozoites but not with erythrocytic stages, immunization of mice with extracts of Anopheles stephensi thorax failed to protect them, mosquito antigens apparently not responsible for protective immunity

Plasmodium berghei
Beaudoin, R. L.; et al., 1977, Exper. Parasitol., v. 42 (1), 1-5
Plasmodium berghei, mice, immunization against ANKA strain using unaltered sporozoite as antigen, suppressive doses of chloroquine throughout immunization period with curative courses of primaquine prior to challenge, all mice survived sporozoite challenge but succumbed to challenge with infected erythrocytes

Plasmodium berghei
Belehu, A.; Poulter, L. W.; and Turk, J. L., 1976, Infect. and Immun., v. 14 (2), 457-462
hamsters, effect of Plasmodium berghei or P. yoelii malaria on concurrent Leishmania enriettii infection depended on relative timing of two infections

Plasmodium berghei
Black, R. H.; and Ray, A. P., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 131-139
Plasmodium berghei-infected mice (exper.), potentiation of therapeutic effects of proguanil and pyrimethamine by dapsone

Plasmodium berghei
Bliznakov, E. G.; and Tao, S. J., 1976, J. Interdisciplinary Cycle Research, v. 7 (3), 215-222
Plasmodium berghei, mice, significant periodic fluctuations in mortality confirmed by application of statistical periodic analysis, 5-year study

Plasmodium berghei
Boonpucknavig, S.; et al., 1976, J. Nat. Research Council Thailand, v. 8 (1), 51-66
Plasmodium berghei, mice, immune complex nephritis, clinical, histopathological and immunofluorescent studies

Plasmodium berghei
Booth, M.; et al., 1977, J. Protozool., v. 24 (4), 41A [Abstract]
Plasmodium berghei, aspartate transcarbamylase, characterization

Plasmodium berghei
Bosworth, A. B.; Schneider, I.; and Freier, J. E., 1975, J. Parasitol., v. 61 (4), 769-772
Plasmodium berghei, mass isolation of Anopheles stephensi salivary glands infected with malarial sporozoites

Plasmodium berghei
Brambilla, E.; Galli, L.; and Caprotti, M., 1974, Riv. Parassitol., Roma, v. 35 (4), 237-244
Plasmodium berghei in Mus musculus subject to thermal stress or whole-body irradiation, lower dose of chloroquine is therapeutically effective

Plasmodium berghei
Brown, K. N., 1977, Advances Exper. Med. and Biol., v. 93, 5-25
antigenic variation in malaria, review: Plasmodium knowlesi, schizont-infected cell agglutination test, protective variant-specific antibodies, induction of antigenic variation, antigenic variation and the cell cycle, antigenic variation and red cell penetration, variant-specific antibody synthesis and protective immunity; Plasmodium berghei, lymphocyte subpopulations and immunity transcending antigenic variation; gametocytes, antigenic variation, and protection; antigenic variation and immunization against malaria

Plasmodium berghei
Brown, K. N.; Jarra, W.; and Hills, L. A., 1976, Infect. and Immun., v. 14 (4), 858-871
Plasmodium berghei, rats, development of protective T cell activity

Plasmodium berghei
Bucci, A.; et al., 1974, Riv. Parassitol., Roma, v. 35 (4), 245-252
Plasmodium berghei incubated in vitro with sera of immune rats (recovered from infection or immunized), inhibition of subsequent infectivity, implications for role of serum antibody in protection

Plasmodium berghei
Buengener, W., 1974, Tropenmed. u. Parasitol., v. 25 (3), 309-312
Plasmodium berghei, P. vinckei, P. chabaudi, mice, rats, allopurinol stimulated multiplication of plasmodia and infections caused death more rapidly or in more animals, this effect could result from hypoxanthine concentration being limiting factor for parasite multiplication, in these parasite-host combinations host-purine-dependent parasites would have a selective advantage over non-dependent ones

Plasmodium berghei
Buengener, W., 1975, Tropenmed. u. Parasitol., v. 26 (3), 285-290
Plasmodium berghei-infected mice (exper.), added infection of Trypanosoma musculi results in rapid multiplication of trypanosome parasites with distribution into multiple body organs and tissues, infection course

Plasmodium berghei
Buengener, W., 1975, Tropenmed. u. Parasitol., v. 26 (3), 291-294
Trypanosoma musculi strain passaged several years in mice (exper.), infection course and development in pregnant mice, Plasmodium berghei-infected mice, or normal mice given either heavy or weak infections

Plasmodium berghei
Cabrera, E. J.; Alger, N. E.; and Silverman, P. H., 1973, J. Protozool., v. 20 (3), 449-452
Plasmodium berghei, rats, adoptive immunity transferred by 2 x 10^7 or 2 x 10^8 but not 2 x 10^6 immune spleen cells, spleen cells kept at 47 C for 45 min were no longer able to transfer protection; capacity to transfer adoptive immunity not found in spleen cells from unexposed adult rats capable of age immunity, but found in spleen cells from rats that had suffered very transient parasitemia

Plasmodium berghei
Cantrell, W.; and Elko, E. E., 1976, Exper. Parasitol., v. 40 (2), 281-285
Wistar and Sprague-Dawley strains of rat differ in their ability to maintain phagocytic hyperactivity toward colloidal carbon during Plasmodium berghei infection

Plasmodium berghei
Carroll, F. I.; et al., 1976, J. Med. Chem., v. 19 (9), 1111-1119
Plasmodium berghei-infected mice, Plasmodium cynomolgi-infected rhesus monkeys, laboratory trials with reduced 8-aminoquinoline analogues

Plasmodium berghei
Cenedella, R. J.; and Saxe, L. H., 1971, Am. J. Trop. Med. and Hyg., v. 20 (4), 530-534
attempted evaluation of extent to which DDS inhibits glucose utilization by parasite in Plasmodium berghei infected mice, partial reversal of antimalarial activity of drug by induced hyperglycemia in host

Plasmodium berghei
Chen, D. H.; Tigelaar, R. E.; and Weinbaum, F. I., 1977, J. Immunol., v. 118 (4), 1322-1327
Plasmodium berghei, immunization of T and B cell-deficient mice with x-irradiated sporozoites, results demonstrate preeminent role for T cells in induction of protective immunity against sporozoite infection

P[lasmodium] berghei
Chen, E. H.; et al., 1977, J. Med. Chem., v. 20 (8), 1107-1109
P[lasmodium] berghei in mice, P. cynomolgi in monkeys, 5-phenoxy derivatives of primaquine are more active and less toxic than primaquine in antimalarial screening trials

Plasmodium berghei, illus.
Cochrane, A. H.; et al., 1976, J. Immunol., v. 116 (3), 859-867
Plasmodium berghei, P. cynomolgi, antibody-induced ultrastructural changes of sporozoites, surface coat formation

Plasmodium berghei
Coleman, R. M.; et al., 1975, Immunology, v. 29 (1), 49-54
Plasmodium berghei, mice, cell-mediated cytotoxic activity against erythrocytes from malaria-infected animals demonstrated in vitro, splenic macrophages and nylon-purified spleen cells are implicated, antibody found to enhance cell-mediated lysis

Plasmodium berghei
Coleman, R. M.; et al., 1976, J. Parasitol., v. 62 (1), 138-140
Plasmodium berghei, transitory but heightened rate of destruction of normal transfused erythrocytes in infected rats

Plasmodium berghei
Coleman, R. M.; Bruce, A.; and Rencricca, N. J., 1976, J. Parasitol., v. 62 (1), 137-138
Plasmodium berghei, inhibition of macrophage migration in vitro may be an analog of macrophage disappearance reaction in vivo, mice

Plasmodium berghei
Das, B. P.; and Boykin, D. W., 1977, J. Med. Chem., v. 20 (4), 531-536
Trypanosoma rhodesiense in mice, successful synthesis and antiprotozoal activity of 2,5-bis(4-guanylphenyl)furans in laboratory trials; little activity of these compounds against Plasmodium berghei in mice

Plasmodium berghei
Das, B. P.; and Boykin, D. W., 1977, J. Med. Chem., v. 20 (9), 1219-1221
2,5-bis(4-guanylphenyl)thiophenes and -pyrroles, significant antiprotozoal activity against Trypanosoma rhodesiense but little activity against Plasmodium berghei in mice (exper.), laboratory synthesis and comparison trials with stilbamidine, hydroxystilbamidine and pentamidine

Plasmodium berghei
Dave, D.; Gutteridge, W. E.; and Richards, W. H. G., 1976, Parasitology, v. 73 (2), xvii [Abstract]
Plasmodium spp., conversion of dihydroorotate to orotate, activities were irreversible, required O_2, and were inhibited by CN^- and Antimycin A, no involvement of reduced pteridines or pyridine nucleotides detected, this area of metabolism may be important site for selective drug action

Plasmodium berghei
Davies, E. E.; and Howells, R. E., 1976, Ztschr. Parasitenk., v. 48 (3-4), 297 [Abstract]
Plasmodium berghei and associated infections in Anopheles stephensi mid-gut: rickettsial-like particles, no concomitant infection; virus-like particles within morphologically abnormal oocysts and similar particles within mid-gut epithelial cells; virus-like particles in mid-gut epithelial cells but not in oocysts

Plasmodium berghei, illus.
Davies, E. E.; Howells, R. E.; and Peters, W., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 461-464
Plasmodium berghei oocysts after exposure to primaquine, no changes in morphology of mitochondria (as opposed to erythrocytic stages), possible reasons for insensitivity of oocyst to primaquine

Plasmodium berghei
Davies, E. E.; Warhurst, D. C.; and Peters, W., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 4-5 [Demonstration]
Plasmodium berghei, ultrastructural changes after treatment with quinine or WR 122,455

Plasmodium berghei
Desowitz, R. S., 1965, Med. J. Malaya, v. 20 (1), 52-53
oxygen consumption of Plasmodium berghei in presence of normal or immune rat serum and chloroquine, possible usefulness in drug screening and determination of drug-serum clearance rates

Plasmodium berghei
Desowitz, R. S., 1971, Science (3988), v. 172, 1151-1152
Plasmodium berghei, immunization (with non-living antigen) of young white rats born of immune mothers, significantly higher level of immunity than unvaccinated littermates or vaccinated rats born of normal nonimmune mothers

Plasmodium berghei
Desowitz, R. S., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 238-244
Plasmodium berghei, rats, factors influencing maternally-transmitted protective immunity: effect of duration of maternal infection and of rechallenge; immunogenic activity of non-viable antigen in inducing and maintaining maternally-transmitted immunity

Plasmodium berghei
Desowitz, R. S.; and Barnwell, J. W., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 475-476
Plasmodium berghei very young forms, deep vascular sequestration in heart and kidney of white rat equal to or greater than that in bone marrow, lung, liver, and spleen; hitherto unrecorded site of schizont concentration in lung

Plasmodium berghei
Diggs, C. L.; and Osler, A. G., 1975, J. Immunol., v. 114 (4), 1243-1247
Plasmodium berghei, rats, evidence suggests that schizonts and/or merozoites are targets of protective antibody action

Plasmodium berghei
Dimopoullos, G. T.; and Finerty, J. F., 1976, Am. J. Vet. Research, v. 37 (6), 693-695
Anaplasma marginale, Plasmodium lophurae, P. berghei, cross reactivity in passive haemagglutination tests

Plasmodium berghei
Djudovic, P.; et al., 1976, Pharmazie, v. 31 (12), 845-849
quinoline-3-, 6-methoxyquinoline-3-, and quinoxaline-2-derivatives, synthesis, activity against drug-sensitive and resistant strains of Plasmodium spp. in white mice and canaries, comparison with known antimalarials

Plasmodium berghei
Dokow, S.; et al., 1974, J. Protozool., v. 21 (3), 463 [Abstract]
Plasmodium berghei-infected rats, splenocyte population, change in balance between formation of immunocompetent and erythropoietic cells

Plasmodium berghei
Druilhe, P.; and Monjour, L., 1975, Compt. Rend. Soc. Biol., Paris, v. 169 (4), 1089-1095
human malaria, serodiagnosis, gel diffusion test using Plasmodium falciparum and P. berghei antigens prepared by different methods, compared with indirect immunofluoresence

Plasmodium berghei
Eaton, J. W.; et al., 1976, Nature (5588), v. 264, 758-760
Plasmodium berghei, mice, vitamin E-deficiency moderates course of infection, supports idea that oxidant sensitivity of host erythrocytes may limit severity of malaria

Plasmodium berghei
Eckman, J. R.; et al., 1977, J. Lab. and Clin. Med., v. 90 (4), 767-770
Plasmodium berghei in chloroquine-resistant and chloroquine-sensitive mice (exper.), studies confirm that malaria pigment is composed of precipitated host cell hemoglobin and suggest that drug resistance is accompanied by basic alteration in parasite-mediated hemoglobin catabolism

Plasmodium berghei, illus.
Einheber, A.; Palmer, D. M.; and Aikawa, M., 1976, Exper. Parasitol., v. 40 (1), 52-61
Plasmodium berghei, chloroquine-induced pigment clumping, inhibition and reversal by quinine, WR 33,063, WR 171,669, WR 30,090, and WR 142,490, possible application in assay for oral bioavailability of such candidate antimalarials

Plasmodium berghei, illus.
Eling, W. M. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (3), 267-268
Plasmodium berghei-infected mice (exper.), centrifugation technique for separation of parasitized erythrocytes from infected mouse blood

Plasmodium berghei
Eling, W.; and Jerusalem, C., 1977, Tropenmed. u. Parasitol., v. 28 (2), 158-174
Plasmodium berghei infected mice (exper.), immunization studies using sulfathiazole-treated drinking water and comparison with para-aminobenzoic acid deficient diet for control of parasitic proliferation during sensitization period; balance between suppressive effect of drug and survival of parasites in treated host is important for induction of immunity

Plasmodium berghei
Eling, W.; and Jerusalem, C., 1977, Tropenmed. u. Parasitol., v. 28 (3), 293-301
Plasmodium berghei, Swiss and C3H/StZ mice (exper.), evaluation of conditions that may affect immunizing capacity of inoculum; actual immunizing capacity depends on magnitude and time of initiation of sulfathiazole treatment after inoculation as well as conditions of storage, medium, and temperature; conditions leading to immunity are apparently strain specific

Plasmodium berghei
Ewers, W. H., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 89 [Demonstration]
experimental infection in splenectomized fruit bat (Dobsonia moluccensis), infection of red blood cells found 19 days after inoculation; no infections developed in unsplenectomized bats

Plasmodium berghei, illus.
Fajardo, L. F., 1973, Nature (5405), v. 243, 298-299
Plasmodium falciparum, P. vivax, human, ultrastructural changes in blood platelets; P. berghei, mice (exper.), presence of parasites in platelets

Plasmodium berghei
Ferone, R., 1973, J. Protozool., v. 20 (3), 459-464
Plasmodium berghei, synthesis of dihydropteroate and dihydrofolate, some properties of enzymes involved, enzymic synthesis of dihydropteroate inhibited by several sulfonamides and diaminodiphenylsulfone

Plasmodium berghei
Fitch, C. D., 1977, Life Sc., v. 20 (7), 1281-1284
Plasmodium berghei, chloroquine-resistant and -sensitive parasites have identical infective potencies, indicates that chloroquine-resistant parasites select and preferentially infect immature erythrocytes rather than lacking the capability of infecting mature erythrocytes

Plasmodium berghei
Fitch, C. D., 1977, Life Sc., v. 21 (10), 1511-1514
chloroquine-resistant Plasmodium berghei found to be as susceptible to chloroquine as chloroquine-susceptible P. berghei in vitro, indicates that chloroquine resistance is due to inadequate exposure of the intracellular parasites to the drug

Plasmodium berghei
Fitch, C. D.; Chevli, R.; and Gonzalez, Y., 1975, J. Pharmacol. and Exper. Therap., v. 195 (3), 389-396
Plasmodium berghei and P. vinckei chloroquine-resistant and -susceptible strains, P. berghei yoelii spontaneously resistant to chloroquine, variations of substrate-stimulated chloroquine accumulation

Plasmodium berghei
Fitch, C. D.; Gonzalez, Y.; and Chevli, R., 1975, J. Pharmacol. and Exper. Therap., v. 195 (3), 397-403
Plasmodium berghei chloroquine-resistant and -susceptible, amodiaquin accumulation by infected mouse erythrocytes, comparison with chloroquine accumulation

Plasmodium berghei
Fletcher, K. A.; Canning, M. V.; and Theakston, R. D. G., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 125-130
Plasmodium berghei-infected mice, rats and hamsters (all exper.) and P. knowlesi-infected monkeys (exper.), absence of glucose-6-phosphate activity of "parasite origin" and presence of 6-phosphogluconate dehydrogenases of "parasite origin" in erythrocytes of infected animals, possible relationships in erythrocyte metabolism and parasite growth

Plasmodium berghei
Foley, D. A.; and Vanderberg, J. P., 1977, Exper. Parasitol., v. 43 (1), 69-81
Plasmodium berghei, successful transmission to rats, mice, and hamsters using liver from sporozoite-infected rats without concomitant blood infections, concluded that infections were due to immature exoerythrocytic schizonts (EES), other life cycle stages rigorously excluded as cause of infectivity, immunity to malaria in sporozoite-immune mice does not extend to EES

Plasmodium berghei
Foley, D. A.; and Vanderberg, J. P., 1977, Experientia, v. 33 (7), 911-912
Plasmodium berghei, rats, mice, simple method for transmitting infection by inoculation of immature exoerythrocytic schizonts, possibility that infection has been induced by other life cycle stages has been experimentally excluded, potential applications of this technique

Plasmodium berghei
Frankenburg, S.; et al., 1977, Exper. Parasitol., v. 43 (2), 362-369
Plasmodium berghei, rats, appropriate balance between immunocompetent and erythropoietic cells in spleen is one of critical factors in determining fate of infected animal

Plasmodium berghei
Frankenburg, S.; and Greenblatt, C. L., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
Plasmodium berghei, mice with primary vs. secondary infections, cellular changes in bone marrow

Plasmodium berghei
Gesinski, R. M.; and Napoletano, T. S., 1976, Science Biol. J., v. 2 (3), 84-88
Plasmodium berghei-infected mice, increased oxygen consumption by bone marrow cells, decreased oxygen consumption in presence of antibodies

Plasmodium berghei
Gesinski, R. M.; Napoletano, T. S.; and Algaier, R. J., 1976, Science Biol. J., v. 2 (6), 244-250
Plasmodium berghei, mice, volume changes and osmotic fragility of normal and infected erythrocytes, non-infected erythrocytes from infected animals lyse earlier than erythrocytes from malaria-free animals, malaria-infected erythrocytes were least susceptible to volume changes and lysis

Plasmodium berghei
Golenser, J.; et al., 1974, J. Protozool., v. 21 (3), 464 [Abstract]
Plasmodium berghei-infected rats, dynamics of response to sheep erythrocytes

Plasmodium berghei
Golenser, J.; et al., 1976, Ztschr. Parasitenk., v. 50 (1), 95-98
Plasmodium berghei, spleen cells from previously infected rats and mice, lymphocyte transformation test with plasmodial antigen, higher parasitemia correlated with higher in vitro stimulation of lymphocytes

Plasmodium berghei
Golenser, J.; et al., 1977, Clin. and Exper. Immunol., v. 29 (1), 43-51
Plasmodium berghei, rats immunized with sporozoites or infected blood, indirect fluorescent antibody tests, crossreactivity using as antigen sporozoites, exoerythrocytic forms, or blood schizonts, protection or lack of protection against challenge with sporozoites or infected blood

Plasmodium berghei
Golenser, J.; et al., 1977, Trop. and Geogr. Med., v. 29 (2), 204-205 [Abstract]
Plasmodium berghei-infected rats (exper.), estimation of density of exo-erythrocytic (EEF) forms at different stages of infection and comparison with normal controls indicates that blood stage interferes with the development of EEF in liver parenchymal cells of infected animals

P[lasmodium] berghei
Golenser, J.; et al., 1977, Trop. and Geogr. Med., v. 29 (3), 318 [Abstract]
P[lasmodium] berghei, detection of antibodies against sporozoites, parasitized erythrocytes and exoerythrocytic forms using the indirect fluorescent antibody test

Plasmodium berghei
Golenser, J.; Shmuel, Z.; and Spira, D. T., 1975, J. Protozool., v. 22 (3), 71A [Abstract]
Plasmodium berghei-infected rats injected with sheep erythrocytes, numbers of plaque-forming cells in spleen and popliteal lymph nodes during course of infection

Plasmodium berghei
Golenser, J.; Spira, D. T.; and Shmuel, Z., 1976, Parasitology, v. 73 (2), xiii [Abstract]
rats, mutual influence of infection with Plasmodium berghei and Nippostrongylus brasiliensis

Plasmodium berghei
Golenser, J.; Spira, D. T.; and Zuckerman, A., 1973, J. Protozool., v. 20 (4), 531-532
Plasmodium berghei, splenocytes from infected rats, spontaneous incorporation of ^{3}H-thymidine in vitro, unresponsiveness to phytohemagglutinin stimulation

Plasmodium berghei
Golenser, J.; Verhave, J. P.; and Meuwissen, J. H. E. T., 1977, Parasitology, v. 75 (2), xxxi-xxxii [Abstract]
Plasmodium berghei, specificity of antibody responses to different life-cycle stages, results indicate different antigenic determinants but also certain common antigens

Plasmodium berghei
Gorenflot, A.; et al., 1976, Ann. Pharm. Franc., v. 34 (5-6), 199-209
Plasmodium berghei, mice, morphology of infected erythrocytes, optical and scanning electron microscopy, proposed hypothetical cycle explaining erythrocyte polymorphism

Plasmodium berghei
Gravely, S. M.; Hamburger, J.; and Kreier, J. P., 1976, Infect. and Immun., v. 14 (1), 178-183
Plasmodium berghei, young vs. adult rats, T and B cell population changes in relation to mechanism of age-related immunity

Plasmodium berghei
Gravely, S. M.; and Kreier, J. P., 1974, Tropenmed. u. Parasitol., v. 25 (2), 198-206
Babesia microti, removal from infected hamster erythrocytes by continuous-flow ultrasonication, freed parasites collected by differential centrifugation, effectiveness of this system of lysis evaluated by electron microscopy and parasites evaluated serologically by complement-fixation testing, cross-reactions with sera from Plasmodium berghei-infected hamsters may be due to erythrocyte contamination

Plasmodium berghei
Gravely, S. M.; and Kreier, J. P., 1976, Infect. and Immun., v. 14 (1), 184-190
Plasmodium berghei, rats, adoptive transfer experiments with T and B lymphocytes alone or in combination, results argue strongly for immunity transferrable by antibody-producing cells and not by T cells acting in classical cell-mediated immune responses

Plasmodium berghei
Gun, F.; and Greenblatt, C. L., 1973, J. Protozool., v. 20 (4), 532
Plasmodium berghei, P. gallinaceum, cultivation in cell hybrids formed by fusing infected red blood cells with HeLa cells, chick fibroblasts, baby hamster kidney cells and macrophages

Plasmodium berghei, illus.
Hamburger, J.; and Kreier, J. P., 1976, Exper. Parasitol., v. 40 (2), 158-169
Plasmodium berghei free parasites (but not parasites in erythroctes) become coated with antibodies after incubation in recovered rat serum (as demonstrated by fluorescent antibody technique), this immune serum did not protect mice against inoculation of free parasites but did protect rats partially or completely, phagocytes ingested parasites more readily in prescence of immune vs. normal serum

Plasmodium berghei
Hamburger, J.; and Zuckerman, A., 1976, Exper. Parasitol., v. 39 (3), 460-478
Plasmodium berghei, soluble extract, fractionation by preparative disc electrophoresis, subsequent analytical disc electrophoresis, detection of precipitinogens by immunoelectrophoresis or by double immunodiffusion in agar gel, induction of precipitins in rabbits, physicochemical properties of soluble components, antigenic contaminants of host blood origin

Plasmodium berghei
Hamburger, J.; and Zuckerman, A., 1976, Exper. Parasitol., v. 39 (3), 479-495
Plasmodium berghei, soluble extract, separation into 12 fractions by preparative disc electrophoresis, employment of fractions to seek precipitins in hyperimmune rat serum and in the vaccination of rats

Plasmodium berghei
Hansen, B. D.; and Pappas, P. W., 1977, Ohio J. Sc., v. 77 (4), 189-191
Plasmodium berghei, changes in metabolic rate of mice with acute infections

Plasmodium berghei
Hockmeyer, W. T.; et al., 1976, Abst. Ann. Meet. Am. Soc. Microbiol., 86
Plasmodium berghei, mice, T-cell dependency of sporozoite antigens in inducing protection against subsequent sporozoite challenge

Plasmodium berghei

Holbrook, T. W.; Spitalny, G. L.; and Palczuk, N. C., 1976, J. Parasitol., v. 62 (5), 670-675

mice immunized with Plasmodium fallax exoerythrocytic stages, stimulation of resistance to sporozoite-induced P. berghei malaria

Plasmodium berghei, illus.

Homewood, C. A.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 283-287

Plasmodium berghei, purification and some properties of malarial pigment

P[lasmodium] berghei

Homewood, C. A.; and Atkinson, E. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 26-27 [Demonstration]

P[lasmodium] berghei: carbohydrate metabolism; dependence of chloroquine-induced pigment clumping on composition of culture medium

Plasmodium berghei

Homewood, C. A.; and Howells, R. E., 1973, J. Protozool., v. 20 (4), 526-527

Plasmodium berghei, cyclical changes in carbohydrate metabolism, gain and loss of enzymes of citric acid cycle follows change of host

Plasmodium berghei

Homewood, C. A.; and Neame, K. D., 1976, Ann. Trop. Med. and Parasitol., v. 70 (2), 249-251

Plasmodium berghei, cellulose powder successful for removal of white cells from malaria-infected blood in order to facilitate parasite study

Plasmodium berghei, illus.

Howells, R. E.; and Davies, E. E., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 451-459

Plasmodium berghei oocysts, sequence of post-meiotic nuclear divisions, nuclear changes may be incorporated into general pattern of oocyst differentiation or cytokinesis

Plasmodium berghei, illus.

Howells, R. E.; and Maxwell, L., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 279-283

Plasmodium berghei, mitochondrial changes during life cycle, electrophoretic demonstration that malarial oocyst possesses isoenzyme variants of NAD- and NADP-dependent isocitrate dehydrogenases distinguishable from those of midgut of Anopheles stephensi

Plasmodium berghei

Howells, R. E.; and Maxwell, L., 1973, Ann. Trop. Med. and Parasitol. v. 67 (3), 285-300

Plasmodium berghei, chloroquine-resistant and -sensitive strains, intraerythrocytic forms do not possess NAD- or NADP-dependent isocitrate dehydrogenase, elevation of levels of NADP-isocitrate dehydrogenase in reticulocytes, possible relationship of this reticulocyte stimulation to chloroquine resistance

Plasmodium berghei

Howells, R. E.; and Maxwell, L. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 19 [Demonstration]

Plasmodium berghei, isocitrate dehydrogenase activity in relation to mitochondrial changes during life-cycle; enhancement of enzyme activity greater in reticulocytes infected by chloroquine-resistant vs. -sensitive strain

P[lasmodium] berghei

Hussain, Q. Z.; et al., 1976, Indian J. Med. Research, v. 64 (12), 1836-1840

Plasmodium berghei in rats, successful immunization with liver antigen

Plasmodium berghei

Ilan, J.; Pierce, D. R.; and Miller, F. W., 1977, Proc. Nat. Acad. Sc., v. 74 (8), 3386-3390

Plasmodium berghei, 9-β-D-arabinofuranosyladenine markedly depresses total protein synthesis and induces pronounced changes in spectrum of proteins synthesized (indicating changes in commitment for gene expression)

Plasmodium berghei

Jacobs, R. L.; and Koontz, L. C., 1976, Exper. Parasitol., v. 40 (1), 116-123

Plasmodium berghei, mice, induction of resistance to clindamycin and minocycline, cross-resistance patterns and other characteristics of resistant strains

Plasmodium berghei

Jacobson, R. L.; Zuckerman, A.; and Greenblatt, C. L., 1977, J. Protozool., v. 24 (4), 67A [Abstract]

Plasmodium berghei, rats, adoptive transfer of immunity in allogeneic neonates with spleen cells from hyperimmunized outbred female rats, effect mitigated when young rats are weaned on immune milk

Plasmodium berghei, illus.

Jadin, J.; Timperman, G.; and De Ruysser, F., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 235-237

pyrimethamine-resistant Plasmodium berghei virulent strains, variations in virulence and biological behavior after cryopreservation

Plasmodium berghei, illus.

Jakstys, B. P.; et al., 1974, J. Protozool., v. 21 (2), 344-348

Plasmodium berghei, ultrastructural analysis of sporozoite antigens prepared by freeze-thawing vs. heat inactivation

P[lasmodium] berghei

Jaroonvesma [sic], N.; and Maegraith, B. G., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 588 [Demonstration]

P[lasmodium] berghei, comparison of changes in color of liver and kidney of mice and rats (both exper.) infected with chloroquine sensitive and resistant strains

P[lasmodium] berghei
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P[lasmodium] berghei, platelets and platelet aggregation in rats and mice (both exper.) infected with chloroquine sensitive and resistant strains of parasites

Plasmodium berghei
Jarra, W.; et al., 1977, J. Protozool., v. 24 (4), 40A [Abstract]
Plasmodium berghei, in vitro maintenance, multiplication and incorporation of radiolabelled hypoxanthine and isoleucine, possible application to malaria immunology

Plasmodium berghei
Jarra, W.; Hills, L. A.; and Brown, K. N., 1976, Parasitology, v. 73 (2), xii [Abstract]
Plasmodium berghei, rats, protection conferred by adoptive transfer of unfractionated spleen cells and T cells alone from donors at various stages of infection, protection conferred by T cells increased with duration of infection in donors, additional presence of B cells in transferred lymphocyte populations enhanced their protective capacity over T cells alone

Plasmodium berghei
van der Kaay, H. J.; Boorsma, L.; and Noordink, J. P. W., 1977, Trop. and Geogr. Med., v. 29 (3), 317 [Abstract]
Plasmodium berghei, labelling of parasite with adenine-^{14}C in midgut sections of vector mosquitoes Anopheles atroparvus

P[lasmodium] berghei
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P[lasmodium] berghei, drug-sensitive and -resistant strains, action of minocycline and/or chloroquine

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Plasmodium berghei drug sensitive and resistant strains, action of minocycline, in combination with chloroquine the interaction was no more than additive

Plasmodium berghei
Kasper, L. H.; and Alger, N. E., 1973, J. Protozool., v. 20 (3), 445-449
Plasmodium berghei, mice, adoptive transfer of immunity, effect of number of cells transferred, source of cells (spleen vs. lymph node), and age of host from which they are derived

Plasmodium berghei
Kitchen, A. G.; and Di Luzio, N. R., 1971, J. Reticuloendothel. Soc., v. 9 (3), 237-247
Plasmodium berghei, rats, functional state of reticuloendothelial system during course of infection, evaluation in vitro of serum recognition factor activity and hepatic phagocytosis

Plasmodium berghei
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pharmacological action and efficacy of combination sulfamoxole and trimethoprim against Plasmodium gallinaceum in chickens and Plasmodium berghei in mice, laboratory trials

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Plasmodium berghei, mice, albumin enhances protection of intramuscularly injected irradiated sporozoites probably by increasing motility but intravenously injected sporozoites still offer more protection

Plasmodium berghei
Kreier, J. P., 1976, Vet. Parasitol., v. 2 (1), 121-142
Plasmodium berghei, rats, mechanisms for implementation of immune response by humoral antibody and phagocytes, changes in cellular systems upon which development of immune response is dependent, review

Plasmodium berghei, illus.
Kreier, J. P.; et al., 1976, Tropenmed. u. Parasitol., v. 27 (1), 82-88
Plasmodium berghei, characteristics of selected population of small blood stage parasites obtained by differential centrifugation of population of parasites freed by continuous flow sonication

Plasmodium berghei
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Trypanosoma cruzi, mice, exacerbating effect of acute infection by Plasmodium berghei superimposed on course of chronic Chagas' disease

Plasmodium berghei
Krettli, A.; Chen, D. H.; and Nussenzweig, R. S., 1973, J. Protozool., v. 20 (5), 662-665
Plasmodium berghei, P. cynomolgi, sporozoites isolated by density-gradient centrifugation, infectivity, ability to induce protective immunity and formation of antisporozoite antibodies, in vitro reactivity in circum-sporozoite precipitation reaction

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Krettli, A. U.; and Nussenzweig, R., 1974, Cellular Immunol., v. 13 (3), 440-446
Plasmodium berghei, mice, depletion of T and B lymphocytes during infection, probable causal relationship with malaria-induced immunosuppression

Plasmodium berghei
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Plasmodium berghei, mice, level of immune complex release activity from surface of lymphocytes, alternative complement pathway function. C3 levels in serum

Plasmodium berghei
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Plasmodium spp., laboratory trials in mice and monkeys using primaquine, 4-methylprimaquine and their 4-substituted analogues showed suppressive antimalarial activity

Plasmodium berghei
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Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium berghei
Loose, L. D.; and Di Luzio, N. R., 1976, Am. J. Trop. Med. and Hyg., v. 25 (2), 221-228
Plasmodium berghei, mice, temporal relationship between reticuloendothelial system phagocytic alterations and antibody responses, implications for mechanism of malaria-induced immunosuppression

Plasmodium berghei
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Plasmodium berghei-infected mice, IgG and IgM present on circulating erythrocytes (both parasitized and nonparasitized), this Ig could be part of immune complexes nonspecifically bound to cell surface or could constitute autoantibodies against reticulocytes or antibodies against parasite antigens present on cell membrane

Plasmodium berghei
McAlister, R. O., 1977, J. Parasitol., v. 63 (3), 455-463
Plasmodium berghei, time-dependent loss of invasive ability of merozoites in vitro

Plasmodium berghei
McAlister, R. O.; and Gordon, D. M., 1976, J. Parasitol., v. 62 (5), 664-669
Plasmodium berghei- or P. yoelii-parasitized mouse erythrocytes, differential fractionation by ultracentrifugation using preformed isodensity gradients of discontinuous type (Stractan II as gradient material), produces significant fractionation of various erythrocytic stages, yields cell suspensions both morphologically and physiologically intact

Plasmodium berghei
McAlister, R. O.; and Gordon, D. M., 1977, J. Parasitol., v. 63 (3), 448-454
Plasmodium berghei, evaluation of 5 methods for artificial removal of merozoites from infected mouse erythrocytes, free merozoites found to be noninvasive in vitro irrespective of isolation method, infective potential in vivo of these "free" parasite preparations could be accounted for solely on basis of contamination by intact parasitized cells

Plasmodium berghei
McBride, J. S.; Micklem, H. S.; and Ure, J. M., 1977, Immunology, v. 32 (5), 635-644
Plasmodium yoelii yoelii, P. berghei, mice, depression of antibody response to type III pneumococcal polysaccharide

Plasmodium berghei
MacCallum, D. K., 1969, J. Reticuloendothel. Soc., v. 6 (3), 232-252
Plasmodium berghei-infected Mesocricetus auratus, sequential changes demonstrated by hepatic system of macrophages during acute infection and after treatment, evidence indicates that macrophages containing endogenous pigment formed by disease process react in identical fashion to macrophages containing exogenous particulate matter (carbon)

Plasmodium berghei
MacCallum, D. K., 1969, J. Reticuloendothel. Soc., v. 6 (3), 253-270
Plasmodium berghei-infected Mesocricetus auratus, macrophage interactions with pulmonary vasculature during acute infection and after drug therapy

Plasmodium berghei
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successful laboratory trials and synthesis of several 1,2,4-triazines against Plasmodium berghei-infected mice and Plasmodium gallinaceum-infected chicks

Plasmodium berghei
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curative therapeutic action of 6 compounds of methylthio- and methylsulfonyl-substituted 9-phenanthrenemethanols against Plasmodium berghei in mice

Plasmodium berghei
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Babesia rodhaini, hypothymic BALB/c.nu/nu mice are more resistant than intact BALB/c. nu/+ mice, proportion of nu/nu survivors is not affected by administration of amicarbalide whereas majority of nu/+ mice develop long-lasting resistance to infection, data suggest that T cell dependent activities are involved in acceleration of parasitaemia and in development of drug-assisted resistance; nu/nu mice are not more resistant than nu/+ mice to Plasmodium berghei

Plasmodium berghei
Miyata, A., 1973, Nettai Igaku (Trop. Med.), v. 15 (4), 204-213
Entamoeba histolytica, Toxoplasma gondii, Plasmodium berghei, Trypanosoma gambiense, preservation by freezing, evaluation of precooling and cryo-protective solutions, length of survival; T. gondii (Beverley strain) unable to survive freezing

Plasmodium berghei
Momen, H., 1976, Parasitology, v. 73 (2), xvi [Abstract]
Plasmodium spp., Babesia spp., Anthemosoma garnhami, carbohydrate metabolism: enzyme activity, glucose catabolism

Plasmodium berghei
Moore, G. A.; and Boothroyd, B., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 489
Plasmodium berghei, direct resolution of lattice planes of malarial pigment, results imply pigment is synthesized as excretory product and is not merely residue of haemoglobin digestion

Plasmodium berghei
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Schistosoma mansoni, Plasmodium berghei, comparison of structures of pigment

Plasmodium berghei
Natarajan, P. N.; Wan, A. S.; and Zaman, V., 1974, Planta Med., v. 25 (3), 258-260
Plasmodium berghei in mice, Entamoeba invadens in monophasic medium, poor results in laboratory trials with gentianine and 3-hydroxymethyl-4-hydroxyethyl-5-vinyl-pyridine

Plasmodium berghei
Neame, K. D.; Homewood, C. A.; and Momen, H., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 8 [Demonstration]
Plasmodium berghei, Babesia rodhaini, effect of intraerythrocytic parasites on the permeability of red-cell membrane

Plasmodium berghei
Newland, B. G.; and John, D. T., 1977, Virginia J. Sc., v. 28 (2), 65 [Abstract]
Plasmodium berghei, mice, pretreatment with bacterial endotoxin and lipid A (endotoxin-pretreated mice demonstrated increased resistance; lipid A had little effect)

Plasmodium berghei
Norby, S. W.; and Alger, N. E., 1977, Science Biol. J., v. 3 (1), 271-279
Plasmodium berghei, mice, successful transfer of adoptive immunity by intramuscular injection of immune spleen cells

Plasmodium berghei
Orsay, C. P.; and Heath, J. E., 1973, Nature (5429), v. 246, 162-163
Plasmodium berghei, mice, direct febrile response to injection of infected erythrocytes in pre-optic area

Plasmodium berghei
Parashar, A.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 474-480
Plasmodium berghei, mice, passive transfer experiments confirm that protection against infection involves both humoral and cellular immune reactions and response to 'processed antigen' produced by sensitized cells, possibly macrophages

Plasmodium berghei
Parker, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 387-390
Babesia argentina, direct counting technique for estimating high parasitaemias, also suitable for B. bigemina and Plasmodium berghei but not for B. rodhaini or Anaplasma centrale

Plasmodium berghei
Paul, K.; and Blanton, C. D., jr., 1976, J. Pharm. Sc., v. 65 (10), 1527-1530
5-aryloxy-6-methoxy-8-aminoquinolines synthesized and screened for potential antimalarial activity, poor results in comparison trials with primaquine phosphate

Plasmodium berghei
Peters, W., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 155-167
Plasmodium berghei, different drug-sensitive and -resistant strains, blood schizontocidal action of 3 newer sulphonamides selected for low antibacterial activity, alone or potentiated with pyrimethamine or proguanil

Plasmodium berghei
Peters, W.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 407-418
Plasmodium berghei, chloroquine sensitive and resistant strains, or mixed infections with P. yoelii nigeriensis, efficacy and mode of action of mefloquine in mice (exper.), evidence of potent blood schizontocide with effects enhanced by addition of pyrimethamine, sulfaphenazole or primaquine

Plasmodium berghei-like
Peters, W.; and Gregory, K. G., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 133-141
Plasmodium berghei-like parasite, N67 strain, primary resistance to mepacrine

Plasmodium berghei
Peters, W.; and Porter, M., 1976, Ann. Trop. Med. and Parasitol., v. 70 (3), 271-281
Plasmodium berghei, drug trials with mice comparing activity of WR 122,455 against drug resistant strains with activity of common antimalarials

Plasmodium berghei
Peters, W.; Portus, J. H.; and Robinson, B. L., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 143-154
Plasmodium berghei, effect of chloroquine on development of resistance to sulphaphenazole, effect of other sulphonamides on sulphaphenazole-resistant line, effect of chloroquine on development of resistance to pyrimethamine, effect of chloroquine-pyrimethamine combination against chloroquine resistant strain first attacked with chloroquine alone

Plasmodium berghei
Peters, W.; Portus, J. H.; and Robinson, B. L., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 155-171
Plasmodium berghei, use of drug-sensitive strain and several drug-resistant lines derived from it for evaluation of a selection of old and new compounds, value as screening system for blood schizontocidal activity

Plasmodium berghei
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Plasmodium berghei, mice (exper.), studies on development of resistance to mefloquine and possible means of slowing down resistance process by mixing mefloquine with pyrimethamine, sulfaphenazole or primaquine; recommendation that mefloquine be used only to treat chloroquine resistant P. falciparum

Plasmodium berghei
Phillips, R. S., 1976, Parasitology, v. 73 (2), xii [Abstract]
Plasmodium berghei, mice, immunization with irradiated infected red cells, Bordetella pertussis and B.C.G. as potentiators of protection induced

Plasmodium berghei
Playfair, J. H. L.; de Souza, J. B.; and Cottrell, B. J., 1977, Immunology, v. 32 (5), 681-687
Plasmodium spp., Babesia microti, mouse helper T-cell response to parasites measured by using them as carriers for a standard hapten, results show extensive cross-reaction between the 4 parasites as carriers apparently unrelated to known serological cross-reactions but are against the idea that helper T-cells are exclusively responsible for resistance

Plasmodium berghei
Playfair, J. H. L.; de Souza, J. B.; and Cottrell, B. J., 1977, Immunology, v. 33 (4), 507-515
Plasmodium yoelii, P. vinckei, P. berghei, mice, regime of killed homologous vaccine plus Bordetella pertussis adjuvant, differences between species in effectiveness of protection, some cross-protection but largely species-specific, passive transfer of immunity to P. yoelii by serum or spleen cells

Plasmodium berghei
Poels, L. G.; et al., 1977, Exper. Parasitol., v. 42 (1), 182-193
Plasmodium berghei, active immunization of chloroquine-protected mice, immunofluorescence and immunoperoxidase studies, transfer of malaria-immunized spleen cells and/or serum, priming with immune spleen cells, evidence for selective release of protective antigens during course of infection

Plasmodium berghei
Poels, L. G.; et al., 1977, Exper. Parasitol., v. 43 (1), 255-267
Plasmodium berghei, formation of immune complexes and their role in nephropathies in infected mice in comparison with chloroquine-cured and hyperimmune mice, comparative study on nude mouse model, immune complexes deposited in renal glomeruli of acutely infected and hyperimmune mice but not in glomeruli of infected nude mice, pathological ultrastructural alterations found in glomeruli of first two groups

Plasmodium berghei
Poels, L. G.; and van Niekerk, C. C., 1977, Exper. Parasitol., v. 42 (1), 235-247
Plasmodium berghei-infected mice, depression of immune responsiveness to sheep erythrocytes, tolerance induced during period of severe immunosuppression, responsiveness restored with chloroquine cure, elevated levels of nonspecific antibodies produced during immunosuppression, possible relationship between hyperimmunoglobulinemia and immunosuppression

Plasmodium berghei
Porter, M.; and Peters, W., 1976, Ann. Trop. Med. and Parasitol., v. 70 (3), 259-270
Plasmodium berghei, drug-sensitive strains successfully suppressed by WR 122,455, laboratory trials in mice comparing action with that of chloroquine and quinine

Plasmodium berghei
Prunescu, P.; and Prunescu, C., 1975, Arch. Roumaines Path. Exper. et Microbiol., v. 34 (4), 341-348
Plasmodium berghei in mice and hamsters (exper.), pathologic changes in spleen during course of infection

Plasmodium berghei
Rees, R. W. A.; et al., 1976, J. Med. Chem., v. 19 (5), 723-725
13 analogs of pyrimethamine tested in Plasmodium berghei-infected mice, the more potent drugs were then tested in Plasmodium gallinaceum-infected chicks, promising activity

Plasmodium berghei
Reid, V. E.; and Friedkin, M., 1973, Molec. Pharm., v. 9 (1), 74-80
Plasmodium berghei-infected mouse erythrocytes, increase of thymidylate synthetase, kinetic and physical properties, suggests possible target for chemotherapeutic attack

Plasmodium berghei
Reisen, W. K.; and Hillis, T. C., 1975, J. Parasitol., v. 61 (5), 937-940
Plasmodium berghei, failure to protect mice with footpad injections of killed parasites incorporated in complete Freund's adjuvant, possible explanations for failure to immunize

Plasmodium berghei
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significant antimalarial activity in benzo-[h]quinoline-4-methanols tested in laboratory trials with Plasmodium berghei-infected mice, some phototoxicity also present

Plasmodium berghei
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Plasmodium berghei
Richards, W. H. G.; and Williams, S. G., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 179-190
Plasmodium berghei, in vitro evaluation of drug activities using whole blood vs. leucocyte-free blood-dilution cultures and ^{3}H-leucine uptake, comparison with P. knowlesi

Plasmodium berghei
Richards, W. H. G.; and Williams, S. G., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 249-250
technique for removal of leucocytes from malaria infected blood

Plasmodium berghei
Rosales-Ronquillo, M. C.; and Silverman, P. H., 1973, J. Protozool., v. 20 (4), 512
Plasmodium berghei, mode of locomotion of ookinete cultivated in vitro

Plasmodium berghei, illus.
Rosales-Sharp, M. C.; and Silverman, P. H., 1976, Invert. Tissue Culture, Applications Med., Biol., and Agric. (Kurstak and Maramorosch), 77-85
Plasmodium berghei, cultivation of sporogonic forms in vector and non-vector cell substrate, review

Plasmodium berghei
Saul, K. W.; and Kreier, J. P., 1977, Tropenmed. u. Parasitol., v. 28 (3), 302-318
Plasmodium berghei-infected rats (exper.), immunization with antigens of a sonically freed preparation of erythrocytic parasites rich in merozoites, evaluation in rats of 3 age groups and of vaccine with and without adjuvants, freeze-thawed freed parasites did not lose antigenicity when stored up to 2 weeks

Plasmodium berghei, illus.
Seed, T. M.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 525-535
Plasmodium gallinaceum, P. berghei, intra- and extracellular parasites, general topography and surface ultrastructure, scanning and transmission electron microscopy

Plasmodium berghei, illus.
Seed, T. M.; et al., 1976, Exper. Parasitol., v. 40 (3), 380-390
Plasmodium berghei (intraerythrocytic and ultrasonically liberated), effect of osmotic stress on cell ultrastructure and parasite infectivity, comparison with mouse host erythrocytes, results suggest these parasites have osmotic regulatory capacities at least comparable to host cells

Plasmodium berghei, illus.
Seed, T. M.; Aikawa, M.; and Sterling, C. R., 1973, J. Protozool., v. 20 (5), 603-605
Plasmodium berghei, limiting membranes of malaria parasites vs. those of host red cells, differential staining when exposed to positively charged iron colloid, cytochemical method to assay degree of host cell membrane contamination of "erythrocyte-free" malaria parasite preparations

Plasmodium berghei, illus.
Seed, T. M.; and Kreier, J. P., 1976, Infect. and Immun., v. 14 (6), 1339-1347
Plasmodium berghei, surface charge and lectin-binding capacity of isolated malaria parasites vs. host erythrocytes compared by chromatographic, electrophoretic, and cytochemical methods

Plasmodium berghei
Seitz, H. M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 88-90
Plasmodium vinckei, P. berghei, counter-current immunoelectrophoresis used to detect malarial antigens and antibodies in rat and mouse sera, more sensitive than double diffusion methods

Plasmodium berghei
Seitz, H. M., 1975, Tropenmed. u. Parasitol., v. 26 (4), 417-425
Plasmodium berghei, strain K 173 in isogenic mouse strains (exper.), infection course, immunization by intermittent suppression of parasite multiplication by maintaining mice on milk diets of varying lengths, F1-hybrids most resistant and immunization attempts most successful with this strain

Plasmodium berghei
Seitz, H. M., 1976, Tropenmed. u. Parasitol., v. 27 (1), 33-43
description of antigens and antibodies demonstrated during course of Plasmodium berghei infections in mice, comparison of animals with lethal infections and those maintained on milk-diet suppressed infections

Plasmodium berghei
Seitz, H. M., 1976, Tropenmed. u. Parasitol., v. 27 (2), 197-201
Plasmodium berghei infections in neonatally thymectomized mice showed same infection patterns as normal infected mice; cell transfer experiments were unsuccessful in transferring immunity from immune to non-immune animals by transfer of lymphoid cells but acquired immunity destroyed by irradiation could be restored by injection of immune spleen and lymphnode cells

Plasmodium berghei
Seitz, H. M.; and Kaltenbach, H., 1971, Tr. Roy. Soc. Trop. Med. and Hyg., v. 65 (5), 682 [Letter]
Plasmodium berghei, stain techniques for estimating level of parasitemia in mice (exper.) using cresyl fast violet and Wright's stain

Plasmodium berghei
Shapiro, M.; Espinal-Tejada, C.; and Nussenzweig, R. S., 1975, J. Parasitol., v. 61 (6), 1105-1106
Plasmodium berghei, P. cynomolgi var. bastianelli, evaluation of method of Rosales-Ronquillo et al. for in vitro ookinete development, failure to yield adequate numbers of ookinetes

Plasmodium berghei
Shetty, R. V.; Wetter, W. P.; and Blanton, C. D., jr., 1977, J. Med. Chem., v. 20 (10), 1349-1351
Plasmodium berghei in mice (exper.) and P. cynomolgi in rhesus monkeys (exper.), synthesis of 2-benzyloxy and 2 benzylthio analogues of primaquine, laboratory trials showed significant schizonticidal antimalarial activity and decreased toxicity

Plasmodium berghei
Shmuel, Z.; Golenser, J.; and Spira, D. T., 1975, J. Protozool., v. 22 (3), 73A [Abstract]
rats infected with Trypanosoma lewisi at different stages in course of Plasmodium berghei infection, influence on parasitemia of each

Plasmodium berghei
Sinden, R. E.; and Canning, E. U., 1973, J. Protozool., v. 20 (4), 528
Plasmodium berghei, nuclear development, cytogenetics

Plasmodium berghei
Speer, C. A.; Rosales-Ronquillo, M. C.; and Silverman, P. H., 1973, J. Protozool., v. 20 (4), 512-513
scanning electron microscopy of Plasmodium berghei-infected primary mosquito cell cultures

Plasmodium berghei, illus.
Speer, C. A.; Silverman, P. H.; and Schiewe, S. G., 1976, J. Parasitol., v. 62 (5), 657-663
Plasmodium berghei, erythrocytic stages, cultivation in primary cultures of rodent bone marrow cells, unsuccessful with P. vinckei vinckei and P. knowlesi

Plasmodium berghei, illus.
Speer, C. A.; Silverman, P. H.; and Schiewe, S. G., 1976, Ztschr. Parasitenk., v. 50 (3), 237-244
Plasmodium berghei, P. vinckei vinckei, P. coatneyi, P. knowlesi, attempts to cultivate erythrocytic stages in various cell lines, success only with P. berghei in mouse Leydig cell testicular tumor cultures

Plasmodium berghei
Spira, D. T.; Golenser, J.; and Gery, I., 1976, Clin. and Exper. Immunol., v. 24 (1), 139-145
Plasmodium berghei, reactivity of lymphoid macrophage populations of malarious rat spleen to non-specific mitogens

Plasmodium berghei
Spitalny, G. L.; et al., 1977, Exper. Parasitol., v. 42 (1), 73-81
Plasmodium berghei, mice, effect of T cell deprivation on sporozoite immunization, eliminated or reduced capacity to develop protection, sporozoite-neutralizing activity, or circumsporozoite antibodies, capacity fully restored by giving thymocytes prior to immunization, data demonstrate T cell dependence of sporozoite-induced immunity

Plasmodium berghei
Spitalny, G. L.; Rivera-Ortiz, C. I.; and Nussenzweig, R. S., 1976, Exper. Parasitol., v. 40 (2), 179-188
Plasmodium berghei, mice, effect of splenectomy before and after immunization on development and manifestation of sporozoite-induced immunity, monitoring of protective immunity and production of antisporozoite antibodies (circumsporozoite precipitate and sporozoite neutralization activity), effect of passive transfer of hyperimmune sera

Plasmodium berghei
Sweeney, A. W.; and Saint-Yves, I. F. M., 1974, Ann. Trop. Med. and Parasitol., v. 68 (1), 1-3
Plasmodium berghei, Anopheles annulipes refractory to ANKA strain and almost completely refractory to NK 65 strain, Anopheles amictus hilli refractory to NK 65 and susceptible to ANKA strain but not a good vector

Plasmodium berghei
Tanabe, K.; et al., 1977, Exper. Parasitol., v. 43 (1), 143-152
Plasmodium berghei, mice, suppressed response of antibody-forming cells

Plasmodium berghei, illus.
Terzakis, J. A.; Vanderberg, J. P.; and Hutter, R. M., 1974, J. Protozool., v. 21 (2), 251-253
pre-erythrocytic Plasmodium berghei mitochondria, ultrastructure, demonstration of cristae, technical and interpretive pitfalls

Plasmodium berghei, illus.
Terzakis, J. A.; Vanderberg, J. P.; and Weiss, M. M., 1976, J. Parasitol., v. 62 (3), 366-371
ultrastructural appearance of viruslike particles in several Plasmodium spp. at different times in sporogony

Plasmodium berghei, illus.
Theakston, R. D. G.; and Fletcher, K. A., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 441-450
Plasmodium berghei, preference for young mature erythrocytes as opposed to more mature cells, dynamics of hemolysis, isotopic tracer techniques

Plasmodium berghei
Theakston, R. D. G.; and Fletcher, K. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 16-17 [Demonstration]
Plasmodium spp., 6-phosphogluconate dehydrogenase in infected erythrocytes, activity present in parasite cytoplasm as well as in host cell

Plasmodium berghei
Tosta, C. E.; and Filho, N., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 531 [Letter]
Plasmodium berghei, factor(s) in normal adult rat serum not related to malarial antibodies that causes decrease in infectivity of parasite

Plasmodium berghei
Tsukamoto, M., 1974, Nettai Igaku (Trop. Med.), v. 16 (2), 55-69
Plasmodium berghei, diagnostic differentiation of enzymes by polyacrylamide gel electrophoresis

Plasmodium berghei, illus.
Vanderberg, J. P., 1974, J. Protozool., v. 21 (4), 527-537
Plasmodium spp. sporozoites, motility, locomotion, movement, effects of several factors (parasite species, oocyst vs. salivary gland sporozoites, presence of albumin or globulins, temperature), relationship to infectivity, immunogenicity (circumsporozoite precipitate reaction), and secretory activity

Plasmodium berghei
Vanderberg, J. P., 1977, Exper. Parasitol., v. 42 (1), 169-181
Plasmodium berghei, quantitation of sporozoites injected by Anopheles stephensi feeding on rats and mice

Plasmodium berghei
Van Dyke, K., 1977, Exper. Parasitol., v. 42 (2), 274-281
Plasmodium berghei, nucleotides from erythrocyte-free parasites, isolation and high pressure liquid chromatographic analysis

Plasmodium berghei
Vray, B., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (3), 255-256 [Letter]
Plasmodium berghei-infected mice (exper.), possible relationships between hepatosplenic and blood rates of para-aminobenzoic acid and parasite development, diet-deficiency study

Plasmodium berghei
Warhurst, D. C., 1973, J. Protozool., v. 20 (4), 529
Plasmodium berghei N67, P. berghei yoelii 17X, pigmented strains naturally resistant to chloroquine, surprisingly shown to possess high-affinity chloroquine receptors present in sensitive strains, presence of receptors apparently linked with production of haemozoin which will itself concentrate chloroquine, suggested that chloroquine interferes with sequestration of haemin as haemozoin

Plasmodium berghei
Warhurst, D. C.; Homewood, C. A.; and Baggaley, V. C., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 265-281
Plasmodium berghei, erythrocytic forms in vitro, technique for study of autophagic vacuole formation (pigment clumping), effect of chloroquine alone or with various cytotoxic drugs

P[lasmodium] berghei
Warhurst, D. C.; Robinson, B. L.; and Peters, W., 1974, J. Protozool., v. 21 (3), 453 [Abstract]
P[lasmodium] berghei, erythromycin, antimalarial action against blood stages confirmed and causal prophylactic activity detected, potentiation and chloroquine examined in chloroquine-sensitive and -resistant strains, erythromycin stearate more active than the base when given orally

Plasmodium berghei
Warhurst, D. C.; Robinson, B. L.; and Peters, W., 1976, Ann. Trop. Med. and Parasitol., v. 70 (3), 253-258
Plasmodium berghei, mice, erythromycin potentiation of action of chloroquine on chloroquine-resistant strains, comparison of oral and subcutaneous routes and erythromycin base and stearate

Plasmodium berghei, illus.
Warhurst, D. C.; and Thomas, S. C., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 417-420
Plasmodium berghei, P. falciparum, localization of mepacrine by fluorescence microscopy

Plasmodium berghei
Warhurst, D. C.; and Thomas, S. C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 7-8 [Demonstration]
Plasmodium berghei in vitro, localization of mepacrine in merozoites

Plasmodium berghei
Warhurst, D. C.; and Thomas, S. C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 428 [Demonstration]
Plasmodium berghei, chloroquine treatment in vitro causes digestive vesicles containing hemozoin to clump together, competitive inhibition of this clumping by quinine and oligomycin, possibility that clumping site is a membrane ATP-ase used by the intraerythrocytic parasite

Plasmodium berghei
Wedderbun, N.; and Dracott, B. N., 1977, Clin. and Exper. Immunol., v. 28 (1), 130-137
mice with acute and chronic malaria, suppression of immune response to type III pneumococcal polysaccharide

Plasmodium berghei
Weiss, M. L., 1976, Exper. Parasitol., v. 40 (1), 103-111
Plasmodium berghei, mouse strain noninfective but highly immunogenic for Meriones unguiculatus was adapted to M. unguiculatus through serial passage of infected blood, antigenic changes during adaptation, loss of infectivity for mice, different antigens apparently responsible for immunogenicity vs. infectivity, vaccination led to production of some protective antibody but also to blocking and enhancing antibody

Plasmodium berghei
Weiss, M. M.; and Vanderberg, J. P., 1974, J. Protozool., v. 21 (3), 423 [Abstract]
Plasmodium berghei, P. cynomolgi, isolation and concentration of ookinetes from Anopheles stephensi midguts

Plasmodium berghei, illus.
Weiss, M. M.; and Vanderberg, J. P., 1976, J. Protozool., v. 23 (4), 547-551
Plasmodium berghei ookinetes, technic for isolation and concentration of large numbers from Anopheles stephensi midgut, evaluation of physiologic state of these stages upon isolation

Plasmodium berghei
Weiss, M. M.; and Vanderberg, J. P., 1977, J. Parasitol., v. 63 (5), 933-934
Plasmodium berghei, in vitro formation of ookinetes

Plasmodium berghei
Weissberger, H.; Spira, D. T.; and Golenser, J., 1977, J. Protozool., v. 24 (4), 68A [Abstract]
Plasmodium berghei, rats convalescent from infection, lymphoblast transformation by viable and nonviable antigens

Plasmodium berghei
Wellde, B. T.; et al., 1973, Isotopes and Radiation Parasitol. III, 187-192
Plasmodium berghei, mice, immunizing effect of irradiated parasitized RBC's, effects of various factors on development of resistance (splenectomy before and after immunization; lysis in French pressure cell or by freeze thawing of irradiated parasitized cells; increasing doses of radiation applied to parasitized cells; gluteraldehyde fixation of irradiated parasitized cells)

Plasmodium berghei
Wells, R. A.; and Diggs, C. L., 1976, J. Parasitol., v. 62 (4), 638-639
protective activity of sera from mice immunized with irradiated Plasmodium berghei-infected erythrocytes

Plasmodium berghei
Wells, R. A.; Diggs, C. L.; and Phillips, S. M., 1977, J. Immunol., v. 118 (2), 472-477
Plasmodium berghei, mice, general immunosuppression induced by cyclophosphamide alone and specific immunosuppression induced by cyclophosphamide in combination with antigen

Plasmodium berghei
Whitelaw, A.; Miller, J. F. A. P.; and Mitchell, G. F., 1977, Cellular Immunol., v. 32 (1), 216-222
Plasmodium berghei, delayed-type hypersensitivity response to blood cells from infected mice can be elicited in sensitized mice

Plasmodium berghei
Williams, A. I. O.; Rosen, F. S.; and Hoff, R., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 179-185
Plasmodium berghei, mice, role of complement components in susceptibility to infection

Plasmodium berghei
Williams, S. G.; and Fanimo, O., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 301-309
Plasmodium berghei, intracellular pH of rat erythrocytes parasitized with chloroquine-sensitive vs. resistant strains, no evidence to support hypothesis that intracellular pH of drug resistant strains is higher than that for sensitive strains

Plasmodium berghei
Williams, S. G.; and Richards, W. H. G., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 169-178
Plasmodium berghei, techniques for preparation and culture of leucocyte-free blood-dilution cultures

Plasmodium berghei
Wyler, D. J.; and Gallin, J. I., 1977, J. Immunol., v. 118 (2), 478-484
Plasmodium spp. in mice and monkeys, spleen-derived mononuclear cell chemotactic factor as a possible mechanism for splenic macrophage accumulation

Plasmodium berghei, illus.
Yoeli, M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 302-305
Plasmodium berghei NK65 strain, slow maturing primary exo-erythrocytic schizonts in laboratory bred Thamnomys surdaster, differentiation of this "slowed down" growth rhythm from that of "chronic" forms of P. b. yoelii

Plasmodium berghei, illus.
Yoeli, M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 306-310
Plasmodium berghei NK65 strain from highlands of Katanga, experimental approach shows only single generation of exo-erythrocytic schizonts formed in Thamnomys surdaster (exper.)

Plasmodium berghei
Zuckerman, A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 793-829
rodent malaria, immunology, review

Plasmodium berghei
Zuckerman, A.; et al., 1977, J. Protozool., v. 24 (2), Suppl., 20A [Abstract]
Plasmodium berghei-immune donor rats treated with cyclophosphamide, residual spleen cell population retained protection-inducing potential, cyclophosphamide mainly destroyed B-cells but also reduced relative number of T-cells

Plasmodium berghei
Zuckerman, A.; and Jacobson, R. L., 1975, J. Protozool., v. 22 (3), 73A-74A [Abstract]
Plasmodium berghei, rats, adoptive transfer of protection by spleen cells from recovered rats (untreated and pretreated with cyclophosphamide)

Plasmodium berghei
Zuckerman, A.; and Jacobson, R. L., 1976, Internat. J. Parasitol., v. 6 (2), 103-106
Plasmodium berghei, rats, transfer of normal or immune spleen cells induces accelerated fluorescent antibody response, only immune cells induce protection against challenge; pre-treatment of donors with cyclophosphamide depletes spleens but affects neither antibody response nor protection-inducing potential in recipients

Plasmodium berghei
Zuckerman, A.; Spira, D. T.; and Golenser, J., 1977, J. Protozool., v. 24 (2), Suppl., 20A [Abstract]
Plasmodium berghei-immune donor rats treated with cyclophosphamide, residual spleen cell population retained protection-inducing potential, cyclophosphamide mainly destroyed B-cells but also reduced relative number of T-cells

Plasmodium berghei anka
Moyou, R.; Forestier, M. F.; and Pautrizel, R., 1977, J. Protozool., v. 24 (4), 62A [Abstract]
Plasmodium berghei berghei, P. b. anka, mice maintained at 35°C. vs. 20-22°C., differences in development of infection and reinfection

Plasmodium berghei berghei
Al-Mashhadani, H. M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 267-268 [Demonstration]
Plasmodium berghei berghei, selection of Anopheles gambiae sp. A vectors for susceptibility and refractoriness to malaria parasites

Plasmodium berghei berghei
Boonpucknavig, S.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 410-415
Plasmodium berghei berghei, chronological development in tissue of infected mice described by means of immunofluorescent and histological study, change in morphological appearance of immunofluorescent-stained parasites from early to late stage of infection

Plasmodium berghei berghei
Boonpucknavig, S.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (6), 116-119
serum soluble malaria antigen probably responsible for soluble immune complex causing glomerulonephritis in Plasmodium berghei berghei-infected mice

Plasmodium berghei berghei
Carter, R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 274 [Demonstration]
Plasmodium berghei subspp., electrophoretic variation in glutamate dehydrogenase

Plasmodium berghei berghei
Chance, M. L.; and Warhurst, D. C., 1973, J. Protozool., v. 20 (4), 524
Plasmodium berghei berghei, P. berghei yoelii, P. chabaudi, P. vinckei, DNA relationships in the subgenus Vinckeia

Plasmodium berghei berghei
Davies, E. E.; and Howells, R. E., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 20 [Demonstration]
Plasmodium berghei berghei oocysts, uptake of ^{3}H-adenosine but not ^{3}H-thymidine

Plasmodium berghei berghei, illus.
Davies, E. E.; Warhurst, D. C; and Peters, W., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 147-153
Plasmodium berghei berghei in mice, fine structural changes following treatment with quinine or WR 122,455, relationship of changes to drug mode of action

Plasmodium berghei berghei
Djudovic, P.; et al., 1976, Pharmazie, v. 31 (12), 845-849
quinoline-3-, 6-methoxyquinoline-3-, and quinoxaline-2-derivatives, synthesis, activity against drug-sensitive and resistant strains of Plasmodium spp. in white mice and canaries, comparison with known antimalarials

P[lasmodium] b. berghei
Gillet, J.; and Herman, F., 1974, J. Protozool., v. 21 (3), 471 [Abstract]
P[lasmodium] b. berghei, mice, experimental congenital infection

Plasmodium berghei berghei
Golenser, J.; Spira, D. T.; and Zuckerman, A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 251-258
Plasmodium berghei berghei in rats (exper.) injected with hyperimmune serum (HIS), site, mode and time of action of neutralizing antiplasmodial antibody, degree of suppression of parasitemia proportional to dose of HIS and mortality rate inversely proportional

Plasmodium berghei berghei
Hamburger, J.; and Kreier, J. P., 1976, Tropenmed. u. Parasitol., v. 27 (3), 385-390
Plasmodium berghei berghei in immune serum, changes in dilution demonstrate interaction between protective antibodies and malaria parasites, probable involvement of low avidity antibodies that make malaria parasite-protective antibody interaction readily reversible

Plasmodium berghei berghei
Hargreaves, B. J.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 289-299
Plasmodium berghei yoelii, mice, protective immunity induced by mild parasite strains against virulent line, effect of increased inocula and time course of infection, further studies testing cross-protection against P. b. berghei and P. v. vinckei

Plasmodium berghei berghei
Jayawardena, A. N.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 426 [Demonstration]
Plasmodium berghei yoelii-infected intact and T-cell-deprived mice, P. b. berghei-infected mice, responses of spleen cells to phytohaemagglutinin (PHA) and bacterial lipopolysaccharide (LPS) investigated to define changes in functional activity of T- and B-cell populations and relationship to immunodepression

Plasmodium berghei berghei
Jayawardena, A. N.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 427 [Demonstration]
Plasmodium berghei yoelii, P. b. berghei in CBA mice, relationship between T-cell activity and pathology of parasitic infection

Plasmodium berghei berghei Vincke and Lips, 1948
Killick-Kendrick, R., 1971, Symposia Brit. Soc. Parasitol., v. 9, 39-64
murine malaria parasites: collection of strains from rodents; collection of strains from mosquitoes; establishment of strains in the laboratory; maintenance by cyclical transmission, extensive review

Plasmodium berghei berghei Vincke and Lips, 1948
Killick-Kendrick, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 261-277
Plasmodium berghei nigeriensis subsp. nov., life cycle, sporogony, effect of high and low temperatures on sporogony, infectivity of sporozoites, primary exoerythrocytic schizogony, blood stages; zoogeography; comparison with P. b. berghei, P. b. yoelii, and P. b. killicki

Plasmodium berghei berghei
Merkli, B.; and Peters, W., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 473-474
Plasmodium berghei berghei, selection for primaquine resistance, comparison of two different methods

Plasmodium berghei berghei
Moyou, R.; Forestier, M. F.; and Pautrizel, R., 1977, J. Protozool., v. 24 (4), 62A [Abstract]
Plasmodium berghei berghei, P. b. anka, mice maintained at 35°C. vs. 20-22°C., differences in development of infection and reinfection

Plasmodium berghei berghei
Poels, L. G., 1977, Exper. Parasitol., v. 41 (1), 83-88
Plasmodium berghei berghei-infected mice, polyribosome profiles in spleen, results suggest spleen not involved in synthesis of antibodies against parasitic antigen during course of primary lethal infection with P. berghei

P[lasmodium] berghei berghei
Porter, M.; and Peters, W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 17 [Demonstration]
P[lasmodium] berghei berghei, two lines, development of resistance to WR 122,455, cross-resistance to quinine and possibly of one line also to chloroquine

Plasmodium berghei berghei, illus.
Schrevel, J.; Asfaux-Foucher, G.; and Bafort, J. M., 1977, J. Ultrastructure Research, v. 59 (3), 332-350
Plasmodium b. berghei in Anopheles stephensi (midgut), ultrastructural study of sporogony, multiple nuclear divisions

Plasmodium berghei berghei
Suzuki, M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 20 [Demonstration]
Plasmodium berghei berghei, P. yoelii nigeriensis, lung pathology in mice (exper.)

Plasmodium berghei berghei
Trigg, P. I.; and McColm, A. A., 1976, Parasitology, v. 73 (2), xxiii-xxxiv [Abstract]
Plasmodium berghei berghei, erythrocytic stages, in vitro cultivation

Plasmodium berghei berghei
Wedderburn, N.; Turk, J. L.; and Hutt, M. S. R., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 468-470
chronic malarial infections, mice, histologic changes in spleen and liver, depression of IgG plaque-forming cell response, chronically infected mice as laboratory model for tropical splenomegaly

Plasmodium berghei killicki
Carter, R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 274 [Demonstration]
Plasmodium berghei subspp., electrophoretic variation in glutamate dehydrogenase

Plasmodium berghei killicki Landau, Michel and Adam, 1968
Killick-Kendrick, R., 1971, Symposia Brit. Soc. Parasitol., v. 9, 39-64
murine malaria parasites: collection of strains from rodents; collection of strains from mosquitoes; establishment of strains in the laboratory; maintenance by cyclical transmission, extensive review

Plasmodium berghei killicki Landau, Michel and Adam, 1968
  Killick-Kendrick, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 261-277
    Plasmodium berghei nigeriensis subsp. nov., life cycle, sporogony, effect of high and low temperatures on sporogony, infectivity of sporozoites, primary exoerythrocytic schizogony, blood stages; zoogeography; comparison with P. b. berghei, P. b. yoelii, and P. b. killicki

Plasmodium berghei nigeriensis subsp. nov., illus.
  Killick-Kendrick, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 261-277
    Plasmodium berghei nigeriensis subsp. nov., life cycle, sporogony, effect of high and low temperatures on sporogony, infectivity of sporozoites, primary exoerythrocytic schizogony, blood stages; zoogeography; comparison with P. b. berghei, P. b. yoelii, and P. b. killicki
    Thamnomys rutilans: near Ilobi, Western State, Nigeria
    white mice (exper.)
    white rats (exper.)
    Grammomys surdaster (exper.)
    Praomys natalensis (exper.)

    Anopheles stephensi (exper.)

Plasmodium berghei nigeriensis
  Carter, R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 274 [Demonstration]
    Plasmodium berghei subspp., electrophoretic variation in glutamate dehydrogenase

Plasmodium berghei nigeriensis, illus.
  Davies, E. E., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 283-290
    Plasmodium berghei nigeriensis, early ookinete stage and its transformation into oocyst, ultrastructural studies

Plasmodium berghei nigeriensis, illus.
  Davies, E. E., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 379-384
    Plasmodium berghei nigeriensis sporogonic stages, acid phosphatase and beta-glucuronidase not detected within parasite itself but clearly seen within mosquito epithelial cells, aryl sulphatase detected within oocysts

Plasmodium berghei nigeriensis
  Davies, E. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 4 [Demonstration]
    Plasmodium berghei nigeriensis, host and parasite enzymes, detection within sporogonic stages

P[lasmodium] b[erghei] nigeriensis
  Davies, E. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 5 [Demonstration]
    P[lasmodium] b[erghei] nigeriensis, ultrastructure of ookinete

Plasmodium berghei nigeriensis (P. yoelii nigeriensis)
  Hill, J., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 421-427
    Plasmodium berghei nigeriensis, screening compounds for causal prophylactic activity, 2 methods for testing compounds against tissue stages (one for compounds with little or no residual effect and one for those with prolonged residual effect), 9 compounds examined

Plasmodium berghei nigeriensis, illus.
  Sinden, R. E.; and Garnham, P. C. C., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (5), 631-637
    Plasmodium spp., sporozoite ultrastructure within oocyst and salivary glands, incidence of micropores and possible relationship to infectivity, sporozoite morphogenesis, abnormalities in development

Plasmodium berghei yoelii
  Barker, L. R., 1971, Tr. Roy. Soc. Trop. Med. and Hyg., v. 65 (5), 586-590
    Plasmodium berghei yoelii, mice, primary infection, antibody synthesis and protection persisted at least 17 months, cross-protection against virulent isolate of parent strain but not against P. berghei berghei, not possible to detect persisting antigen or persisting infectious organisms in immune mice

Plasmodium berghei yoelii
  Bomford, R.; and Wedderburn, N., 1973, Nature (5398), v. 242, 471-473
    Plasmodium berghei yoelii infection potentiates induction of lymphomas in mice by Moloney leukaemia virus, effect accompanied by reduction in detectable levels of circulating neutralizing antibody to the virus and in particular by absence of IgG neutralizing antibody

Plasmodium berghei yoelii
  Carter, R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 274 [Demonstration]
    Plasmodium berghei subspp., electrophoretic variation in glutamate dehydrogenase

Plasmodium berghei yoelii
  Chance, M. L.; and Warhurst, D. C., 1973, J. Protozool., v. 20 (4), 524
    Plasmodium berghei berghei, P. berghei yoelii, P. chabaudi, P. vinckei, DNA relationships in the subgenus Vinckeia

Plasmodium berghei yoelii
  Clark, I. A.; Allison, A. C.; and Cox, F. E., 1976, Nature, London (5541), v. 259, 309-311
    Babesia, Plasmodium, protection of mice by previous infection with BCG, possible mechanism

Plasmodium berghei yoelii
  Crandall, R. B.; Crandall, C. A.; and Muth, B., 1976, J. Parasitol., v. 62 (2), 321-323
    acute Plasmodium berghei yoelii infection in mice, comparison of delayed hypersensitivity response mediated through spleen vs. that mediated through peripheral lymph nodes

Plasmodium berghei yoelii
Dallas, A. B. C., 1976, J. Trop. Med. and Hyg., v. 79 (8), 182-188
Trypanosoma brucei rhodesiense, Plasmodium berghei yoelii, mice, mutual potentiation of concurrent infections, possibly caused by immunosuppression

Plasmodium berghei yoelii
Finerty, J. F.; and Krehl, E. P., 1976, Infect. and Immun., v. 14 (4), 1103-1105
Plasmodium berghei yoelii, mice, protection from lethal infection by pretreatment with cyclophosphamide, development of resistance preceded by increased hypersensitivity demonstrated by delayed footpad swelling technique

Plasmodium berghei yoelii
Finerty, J. F.; and Krehl, E. P., 1977, Am. J. Trop. Med. and Hyg., v. 26 (3), 377-381
Plasmodium berghei yoelii, reactions in mice immunized with malarial antigen show that cell-mediated immunity depends on route of immunization and type of antigen

P[lasmodium] berghei yoelii
Fink, E., 1976, Ztschr. Parasitenk., v. 50 (2), 183-184
P[lasmodium] berghei yoelii, P. cathemerium, schizogony and sporogony stages, metabolites and antimetabolites

Plasmodium berghei yoelii
Fink, E.; et al., 1973, Prog. Med., Paris, v. 101 (7), 281 [Abstract]
Plasmodium berghei yoelii, mice (exper.), establishment of mean causal prophylactic dose of selected antimalarial and experimental compounds

Plasmodium berghei yoelii
Fitch, C. D.; Chevli, R.; and Gonzalez, Y., 1975, J. Pharmacol. and Exper. Therap., v. 195 (3), 389-396
Plasmodium berghei and P. vinckei chloroquine-resistant and -susceptible strains, P. berghei yoelii spontaneously resistant to chloroquine, variations of substrate-stimulated chloroquine accumulation

Plasmodium berghei yoelii
George, C. R. P.; Parbtani, A.; and Cameron, J. S., 1976, J. Path., v. 120 (4), 235-249
Plasmodium berghei yoelii in mouse model, nephropathy and immunologic responses before and after antimalarial, immunosuppressive and anticoagulant therapy

Plasmodium berghei yoelii
Greenwood, B. M.; and Greenwood, A. M., 1971, Tr. Roy. Soc. Trop. Med. and Hyg., v. 65 (5), 581-585
Plasmodium berghei yoelii-infected NZB and B/W hybrid mice, adult mice more susceptible to infection than young mice of same strains, probably due to defective cell-mediated immunity in adults

Plasmodium berghei yoelii
Hargreaves, B. J.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 289-299
Plasmodium berghei yoelii, mice, protective immunity induced by mild parasite strains against virulent line, effect of increased inocula and time course of infection, further studies testing cross-protection against P. b. berghei and P. v. vinckei

Plasmodium berghei yoelii
Jayawardena, A. N.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 426 [Demonstration]
Plasmodium berghei yoelii-infected intact and T-cell-deprived mice, P. b. berghei-infected mice, responses of spleen cells to phytohaemagglutinin (PHA) and bacterial lipopolysaccharide (LPS) investigated to define changes in functional activity to T- and B-cell populations and relationship to immunodepression

Plasmodium berghei yoelii
Jayawardena, A. N.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 427 [Demonstration]
Plasmodium berghei yoelii, P. b. berghei in CBA mice, relationship between T-cell activity and pathology of parasitic infection

Plasmodium berghei yoelii, illus.
Jayawardena, A. N.; et al., 1977, Immunology, v. 32 (6), 849-859
Plasmodium berghei yoelii (P. yoelii), CBA mice, immunological response, general characteristics and effects of T-cell deprivation and reconstitution with thymus grafts

Plasmodium berghei yoelii Landau and Killick-Kendrick, 1966
Killick-Kendrick, R., 1971, Symposia Brit. Soc. Parasitol., v. 9, 39-64
murine malaria parasites: collection of strains from rodents; collection of strains from mosquitoes; establishment of strains in the laboratory; maintenance by cyclical transmission, extensive review

Plasmodium berghei yoelii Landau and Killick-Kendrick, 1966
Killick-Kendrick, R., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 261-277
Plasmodium berghei nigeriensis subsp. nov., life cycle, sporogony, effect of high and low temperatures on sporogony, infectivity of sporozoites, primary exoerythrocytic schizogony, blood stages; zoogeography; comparison with P. b. berghei, P. b. yoelii, and P. b. killicki

Plasmodium berghei yoelii
Lewinsohn, R., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 51-56
Schistosoma mansoni and Plasmodium berghei yoelii in mice (exper.), dynamics of anemia resulting from mixed infections

Plasmodium berghei yoelii
Michel, J. C., 1975, Compt. Rend. Acad. Sc., Paris, v. 281, s. D, Sc. Nat. (17), 1281-1283
Plasmodium berghei yoelii, non-specific resistance in mice increased by injection of bacterial phospholipid extract

Plasmodium berghei yoelii
Roberts, D. W.; et al., 1977, Infect. and Immun., v. 16 (3), 821-826
Plasmodium berghei yoelii produced fatal infection in nude mice or in mice made B cell deficient by treatment with anti-μ-chain serum, malaria recrudesced in Nu/Nu mice after drug termination of acute disease, recrudescence prevented by thymic grafting or treatment with hyperimmune serum, data suggest that crucial role of thymus is to provide helper function in production of protective antibody

Plasmodium berghei yoelii
Warhurst, D. C., 1973, J. Protozool., v. 20 (4), 529
Plasmodium berghei N67, P. berghei yoelii 17X, pigmented strains naturally resistant to chloroquine, surprisingly shown to possess high-affinity chloroquine receptors present in sensitive strains, presence of receptors apparently linked with production of haemozoin which will itself concentrate chloroquine, suggested that chloroquine interferes with sequestration of haemin as haemozoin

Plasmodium berghei yoelii
Warren, H. S.; and Weidanz, W. P., 1976, European J. Immunol., v. 6 (11), 816-819
in vitro response of spleen cells from Plasmodium berghei yoelii-infected mice to horse erythrocytes is depressed, concluded that a splenic adherent cell (probably the macrophage) is functionally defective as an accessory cell

Plasmodium berghei yoelii
Wedderburn, N.; Turk, J. L.; and Hutt, M. S. R., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 468-470
chronic malarial infections, mice, histologic changes in spleen and liver, depression of IgG plaque-forming cell response, chronically infected mice as laboratory model for tropical splenomegaly

Plasmodium berghei yoelii
Weinbaum, F. I.; Evans, C. B.; and Tigelaar, R. E., 1976, J. Immunol., v. 116 (5), 1280-1283
description of in vitro T cell-dependent proliferative response of immune BALB/c mouse spleen cells to Plasmodium berghei yoelii-infected syngeneic RBCs and to saline-soluble-extract prepared from schizonts

Plasmodium berghei yoelii
Weinbaum, F. I.; Evans, C. B.; and Tigelaar, R. E., 1976, J. Immunol., v. 117 (5), pt. 2, 1999-2005
Plasmodium berghei yoelii, course of infection in T cell and B cell deficient mice, results establish requirement for presence of both T cells and B cells for effective resistance

Plasmodium berghei yoelii, illus.
Weise, R.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 271-278
Plasmodium berghei yoelii, mice, ultrastructure of kidneys, glomerular changes at different stages of disease, findings add to evidence in favour of transient immune complex glomerulonephritis

Plasmodium berghei yoelii
Yoeli, M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 302-305
Plasmodium berghei NK65 strain, slow maturing primary exo-erythrocytic schizonts in laboratory bred Thamnomys surdaster, differentiation of this "slowed down" growth rhythm from that of "chronic" forms of P. b. yoelii

Plasmodium berghei yoelii, illus.
Yoeli, M., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 24-35
Plasmodium berghei yoelii in mice (exper.), promising model for study of cerebral malaria in man caused by P. falciparum (history, origin of virulent strain, pathology, enzyme differentiation, virulence expressed by mutation and genetic change)

Plasmodium berghei yoelii
Yoeli, M.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 173-178
Plasmodium berghei yoelii strain 17X, sudden increase in virulence

Plasmodium berghei yoelii
Yoeli, M.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 4-5 [Demonstration]
Plasmodium berghei yoelii, virulent mutated strain, fulminating fatal cerebral infection in mice (exper.)

Plasmodium (Vinckeia) booliati sp. nov., illus.
Sandosham, A. A.; Yap, L. F.; and Omar, I., 1965, Med. J. Malaya, v. 20 (1), 3-7
Petaurista petaurista melanotus (heart blood): Bukit Lagong Forest Reserve, near Kuala Lumpur, State of Selangor, Malaysia

Plasmodium brasilianum
Bidwell, D.; and Voller, A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 6 [Demonstration]
Plasmodium spp., counter immunoelectrophoresis studies on development of precipitating antibodies

Plasmodium brasilianum
Deane, L. de M., 1976, Scient. Publication (317), Pan Am. Health Organ., 144-163
Plasmodium brasilianum, P. simium, monkeys, epidemiology: Brazil

Plasmodium brasilianum
Houba, V.; et al., 1976, Clin. Immunol. and Immunopathol., v. 6 (1), 1-12
Plasmodium brasilianum, P. malariae, P. falciparum, humans, Aotus monkeys, increased binding of malarial antibodies with sera, faster disappearance of this antibody from circulation, and its increased deposition in renal glomeruli, serum complement levels during progress of infection, important role of P. malariae in pathogenesis of chronic progressive nephropathies occurring in malarial areas

Plasmodium brasilianum, illus.
Hutt, M. S. R.; and Voller, A., 1976, J. Trop. Med. and Hyg., v. 79 (5), 106-110
histopathology of hepatic changes in exper. Aotus monkeys and possible role of liver in immunologic response to malaria

Plasmodium brasilianum
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium brasilianum
Mathews, H. M.; and Dilworth, D. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (2), 351-352
Plasmodium brasilianum, use as antigen in indirect hemagglutination test for diagnosing human P. falciparum, P. vivax, P. ovale, P. malariae

Plasmodium brasilianum
Sulzer, A. J.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 55-58
malaria, human, diagnosis, indirect fluorescent antibody test using multi-species thick smear antigen containing equal proportions of Plasmodium vivax, P. falciparum, and P. brasilianum (latter serologically equivalent in IFA test to P. malariae), circumvents need for multiple testing with several antigen species

Plasmodium brasilianum
Thomas, V.; and Dissanaike, A. S., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 602-606
Plasmodium spp., detection of malaria endemicity among Orang Asli aborigines using Plasmodium falciparum and P. brasilianum antigens and indirect fluorescent antibody test (IFA), age dependent increase in number of positive results, IFA valuable as adjunct to blood slide examination especially when parasites are at very low levels: Malaysia

Plasmodium brasilianum
Weise, M.; and Voller, A., 1975, Tropenmed. und Parasitol., v. 26 (4), 390-394
Plasmodium brasilianum, proteinuria and other pathologic changes in urinary tract of Aotus trivirgatus (exper.), possible use of Aotus-P. brasilianum system to study renal immunopathology of human malaria

Plasmodium bubalis
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium cathemerium
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Cisticola natalensis: Ethiopia

Plasmodium cathemerium
Djudovic, P.; et al., 1976, Pharmazie, v. 31 (12), 845-849
quinoline-3-, 6-methoxyquinoline-3-, and quinoxaline-2-derivatives, synthesis, activity against drug-sensitive and resistant strains of Plasmodium spp. in white mice and canaries, comparison with known antimalarials

P[lasmodium] cathemerium
Fink, E., 1976, Ztschr. Parasitenk., v. 50 (2), 183-184
P[lasmodium] berghei yoelii, P. cathemerium, schizogony and sporogony stages, metabolites and antimetabolites

Plasmodium cathemerium
Maier, W. A., 1976, Ztschr. Parasitenk., v. 50 (2), 192
Plasmodium cathemerium-infected Culex pipiens fatigans, amino acid uptake

Plasmodium cathemerium
Maier, W. A., 1977, Ztschr. Parasitenk., v. 51 (2), 199-211
Plasmodium cathemerium-infected and normal Culex pipiens fatigans, labeling of tissues after feeding on tritiated amino acids; intensive uptake of essential amino acids; nearly no uptake of non-essential amino acids

Plasmodium cathemerium-relictum
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Anas c. crecca
Lophura swinhoei
Capella gallinago
Streptopelia chinensis formosae
Ketupa flavipes
Apus affinis kuntzi
Hirundo daurica formosae
Pomatorhinus ruficollis musicus
Turdus c. chrysolaus
T. pallidus
Hypothymis azurea obholseri
Muscicapa rufilata
Anthus hodgsoni yunnanensis
Motacilla alba leucopsis
Lanius cristatus lucionensis
L. schach formosae
(blood of all): all from Taiwan

Plasmodium cathemerium
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Thraupis virens (blood): imported to U.S., origin unknown

Plasmodium cathemerium, illus.
Senaud, J.; Chobotar, B.; and Scholtyseck, E., 1976, Tropenmed. u. Parasitol., v. 27 (2), 145-159
comparative fine structure morphology of Plasmodium cathemerium, Eimeria ferrisi, E. stiedai, Besnoitia jellisoni, and Frenkelia sp. micropores, discussion of micropore function in parasite nutrition

Plasmodium cathemerium Hartman, 1927
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Muscicapa striata]
[Pica pica]
all from north-east Azerbaidzhan

Plasmodium cathemerium muris
Djudovic, P.; et al., 1976, Pharmazie, v. 31 (12), 845-849
quinoline-3-, 6-methoxyquinoline-3-, and quinoxaline-2-derivatives, synthesis, activity against drug-sensitive and resistant strains of Plasmodium spp. in white mice and canaries, comparison with known antimalarials

Plasmodium chabaudi
Buengener, W., 1974, Tropenmed. u. Parasitol., v. 25 (3), 309-312
Plasmodium berghei, P. vinckei, P. chabaudi, mice, rats, allopurinol stimulated multiplication of plasmodia and infections caused death more rapidly or in more animals, this effect could result from hypoxanthine concentration being limiting factor for parasite multiplication, in these parasite-host combinations host-purine-dependent parasites would have a selective advantage over non-dependent ones

Plasmodium chabaudi
Carter, R.; and Walliker, D., 1974, J. Protozool., v. 21 (3), 449 [Abstract]
Thamnomys rutilans (blood): Central African Republic

Plasmodium (Vinckeia) chabaudi Landau, 1965, illus.
Carter, R.; and Walliker, D., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 187-196
redescription of blood stages, sporogony, and enzyme forms; "From the results reported here it is now clear from the account in her paper (1965) that Landau was describing blood forms of both P. chabaudi and P. v. petteri."
Thamnomys rutilans (mature erythrocytes): near Bangui, Central African Republic

Plasmodium chabaudi
Chance, M. L.; and Warhurst, D. C., 1973, J. Protozool., v. 20 (4), 524
Plasmodium berghei berghei, P. berghei yoelii, P. chabaudi, P. vinckei, DNA relationships in the subgenus Vinckeia

Plasmodium chabaudi
Clark, I. A.; Cox, F. E. G.; and Allison, A. C., 1977, Parasitology, v. 74 (1), 9-18
mice pre-treated with killed Corynebacterium parvum, completely resistant to infection with Babesia microti or B. rodhaini, protected from death caused by Plasmodium vinckei or P. chabaudi infection, no antibody detected, suggested that non-specific soluble mediator may play important role in protection observed

Plasmodium chabaudi, illus.
Herman, R., 1977, Exper. Parasitol., v. 42 (1), 211-220
Plasmodium chabaudi, physical interaction in vitro between splenic lymphocytes from immune mice and syngeneic peritoneal macrophages which had phagocytized and processed infected red cells, specific antigen-mediated binding of sensitized lymphocytes to macrophage membranes was demonstrated, interaction possible expression of role for T cells in immunity in rodent malaria

Plasmodium chabaudi
Koenigk, E.; and Mirtsch, S., 1976, Ztschr. Parasitenk., v. 50 (2), 184-185
Plasmodium chabaudi-infected mouse erythrocytes, fine structure of erythrocyte membranes

Plasmodium chabaudi
Koenigk, E.; and Mirtsch, S., 1977, Tropenmed. u. Parasitol., v. 28 (1), 17-22
Plasmodium chabaudi, characterization of membrane preparations from infected and uninfected erythrocytes of mice by specific activities of membrane-associated enzymes and by patterns of proteins and glycoproteins using gel electrophoresis

Plasmodium chabaudi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium chabaudi
Lukow, I.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 500-504
Plasmodium chabaudi, adenosine monophosphate salvage synthesis

Plasmodium chabaudi
McDonald, V.; and Phillips, R. S., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 286 [Demonstration]
Plasmodium chabaudi, mice, adoptive transfer of immunity using B-cell enriched (T-cell depleted) and B-cell depleted (T-cell enriched) spleen cell populations

Plasmodium chabaudi
McDonald, V.; and Phillips, R. S., 1977, J. Protozool., v. 24 (2), Suppl., 4A [Abstract]
Plasmodium chabaudi, mice, transfer of immune spleen cells and immune serum, protective activity was potentiated in irradiated hosts

Plasmodium chabaudi
McLeod, R. A. F.; and Brown, A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 286 [Demonstration]
Plasmodium chabaudi, intravenous injection of unadapted strain into splenectomized rats, development of high number of gametocytes

Plasmodium chabaudi
Mazen, L.; Gull, K.; and Gutteridge, W. E., 1975, J. Protozool., v. 22 (3), 54A [Abstract]
Plasmodium chabaudi-infected erythrocytes examined using freeze fracture technique, nature of host-parasite interface

Plasmodium chabaudi
Musoke, A. J.; and Cox, H. W., 1977, J. Parasitol., v. 63 (3), 464-470
Plasmodium chabaudi, adaptation to rat host, immune responses in rats and in mice to rat strain, resistance to challenge with homologous and heterologous strains and to Babesia rodhaini, elaboration of soluble serum antigen in mice infected with rat strain

Plasmodium chabaudi
Musoke, A. J.; Cox, H. W.; and Williams, J. F., 1977, J. Parasitol., v. 63 (6), 1081-1088
Plasmodium chabaudi, rats, antigens and antibodies found associated with anemia, splenomegaly, and glomerulonephritis, suggested that soluble complexes of parasite antigen and antibody may have been causal in this syndrome

Plasmodium chabaudi
Phillips, R. S.; and McDonald, V., 1977, Parasitology, v. 75 (2), xxxiii-xxxiv [Abstract]
Plasmodium chabaudi in irradiated mice, depression of early primary parasitemia, nonspecific resistance

Plasmodium chabaudi
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Plasmodium chabaudi
Rosario, V. E., 1976, Nature (5561), v. 261, 585-586
Plasmodium chabaudi, genetic basis of chloroquine resistance investigated in crosses between lines differing additionally in pyrimethamine-susceptibility

Plasmodium chabaudi
do Rosario, V. E., 1977, J. Protozool., v. 24 (2), Suppl., 4A [Abstract]
Plasmodium chabaudi, genetics of chloroquine resistance

Plasmodium chabaudi
Rosario, V. E.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 287 [Demonstration]
Plasmodium chabaudi capacity of drug-resistant forms to compete with drug-sensitive forms in absence of drug pressure, pyrimethamine-resistant mutants are at disadvantage compared to sensitive forms but chloroquine-resistant mutants appear to outgrow sensitive forms

P[lasmodium] chabaudi
do Rosario, V. E.; Walliker, D.; and Beale, G. H., 1977, J. Protozool., v. 24 (2), Suppl., 4A [Abstract]
P[lasmodium] chabaudi, competition between chloroquine-resistant and chloroquine-sensitive lines in absence of drug pressure, apparent selective advantage of resistant over sensitive forms

Plasmodium chabaudi
Schmidt, G.; Walter, R. D.; and Koenigk, E., 1974, Tropenmed. u. Parasitol., v. 25 (3), 301-308
adenosine kinase from normal mouse erythrocytes and from Plasmodium chabaudi, partial purification and characterization, comparison

Plasmodium chabaudi
Walter, R. D.; and Koenigk, E., 1974, Tropenmed. u. Parasitol., v. 25 (2), 227-235
Plasmodium chabaudi, hypoxanthine-guanine phosphoribosyltransferase and adenine phosphoribosyltransferase, purification and characterization

Plasmodium chabaudi adami subsp. nov., illus.
Carter, R.; and Walliker, D., [1977], Ann. Parasitol., v. 51 (6), 1976, 637-646
description of the blood-forms, sporogony and enzyme forms
laboratory mice (exper.)
Grammomys surdaster (exper.)
Anopheles stephensi (exper.)
Thamnomys rutilans: near the village of N'ganga Lingolo, 15 km west of Brazzaville, Congo

Plasmodium chabaudi chabaudi
Carter, R.; and Walliker, D., [1977], Ann. Parasitol., v. 51 (6), 1976, 637-646
sporozoite length and enzyme forms
Thamnomys rutilans: Central African Republic

Plasmodium circumflexum
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Gallinago gallinago
Motacilla flava
Laniarius aethiopicus
Cisticola erythrops
all from Ethiopia

Plasmodium circumflexum
Bennett, G. F.; et al., 1974, J. Wildlife Dis., v. 10 (4), 442-451
survey, prevalence of hematozoa in anatids, infection rate increases with host age, seasonal distribution
Anas rubripes
Anas platyrhynchos
Anas platyrhynchos x A. rubripes
Anas discors
Anas carolinensis
Aix sponsa
Lophodytes cucullatus
(blood of all): all from Massachusetts

Plasmodium circumflexum
Crewe, S. M., 1974, J. Protozool., v. 21 (3), 449 [Abstract]
species of malaria with morphological characters resembling those of Plasmodium circumflexum, quotidian periodicity of schizogony
Streptopelia senegalensis: Nigeria
Ploceus nigerrimus: Nigeria
P. cucullatus: Nigeria
Serinus mozambicus (exper.)
S. canarius (exper.)

Plasmodium circumflexum
Crewe, S. M., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 477-478
Aedomyia as possible vector of avian malaria
Streptopelia senegalensis: Nigeria

Plasmodium circumflexum
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Turdoides somervillei
Cinnyricin leucogaster
Lamprotornis chalybaeus
Thraupis virens
(blood of all): all imported to U. S., origin unknown

Plasmodium circumflexum Kikuth
Meyer, C. L.; and Bennett, G. F., 1976, Canad. J. Zool., v. 54 (2), 133-141
Plasmodium circumflexum, P. polare, sporogony
Mansonia perturbans (exper.)
Culiseta morsitans (exper.)
Anas platyrhynchos (exper.)
Anser cygnoides (exper.)
Aix sponsa (blood): New Brunswick

Plasmodium (Giovannolaia) circumflexum
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Philomachus pugnax (blood): Kenya

Plasmodium clelandi sp.n., illus.
Manawadu, B. R., 1972, J. Protozool, v. 19 (4), 587-589
Varanus cepedianus (erythrocytes): Kal-Aru, Mannar District, Ceylon

Plasmodium cnemidophori Carini, 1941, illus.
Telford, S. R., jr., 1973, J. Protozool., v. 20 (2), 203-207
Ameiva ameiva praesignis (proerythrocytes, erythrocytes, erythroblast): Panama Province and Canal Zone, Republic of Panama

Plasmodium cnemidophori
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Ameiva ameiva: Panama

Plasmodium coatneyi
Areekul, S.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 408 [Demonstration]
Plasmodium coatneyi, turnover rate of ^{131}I-fibrinogen compared in normal and infected monkeys

Plasmodium coatneyi
Areekul, S.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 455-461
Plasmodium coatneyi, metabolism of ^{131}I-labelled fibrinogen in infected rhesus monkeys (exper.) and comparison with normal controls

Plasmodium coatneyi
Areekul, S.; Kanakakorn, K.; and Kasemsuthi, R., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (2), 261-262
Plasmodium coatneyi in Macaca mulatta (exper.), mechanism of intravascular haemolysis occurring in infected monkeys

Plasmodium coatneyi
Areekul, S.; Kanakakorn, K.; and Kasemsuth, R., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 584 [Demonstration]
Plasmodium coatneyi, estimates of fraction of intravascular haemolysis taking place in a continuing process of haemolysis during course of infection in Macaca mulatta (exper.)

Plasmodium coatneyi
Glew, R. H.; and Levin, J., 1975, Proc. Soc. Exper. Biol. and Med., v. 148 (2), 508-510
Plasmodium falciparum, P. vivax in humans (exper.), P. coatneyi in Macaca mulatta (exper.), unable to demonstrate the presence of endotoxins in blood of either humans or monkeys infected with malaria in studies using the Limulus amebocyte lysate test

Plasmodium coatneyi
Kasemsuthi, R.; et al., 1970, Southeast Asian J. Trop. Med. and Pub. Health, v. 1 (4), 558-559 [Demonstration]
Plasmodium coatneyi, fibrinogen metabolism compared in infected and control monkeys

Plasmodium coatneyi
Kilejian, A., 1975, J. Protozool., v. 22 (3), 40A [Abstract]
Plasmodium coatneyi, P. lophurae, DNA properties

Plasmodium coatneyi, illus.
Kilejian, A.; Abati, A.; and Trager, W., 1977, Exper. Parasitol., v. 42 (1), 157-164
Plasmodium falciparum, Plasmodium coatneyi, knob-like protrusions on infected monkey erythrocyte membranes are antigenically different from adjacent areas devoid of knobs

Plasmodium coatneyi
Speer, C. A.; Silverman, P. H.; and Schiewe, S. G., 1976, Ztschr. Parasitenk., v. 50 (3), 237-244
Plasmodium berghei, P. vinckei vinckei, P. coatneyi, P. knowlesi, attempts to cultivate erythrocytic stages in various cell lines, success only with P. berghei in mouse Leydig cell testicular tumor cultures

Plasmodium colombiense sp. n., illus.
Ayala, S. C.; and Spain, J. L., 1976, J. Parasitol., v. 62 (2), 177-189
host blood pictures, parasitemia, parasite structure and structural variance, infection states, host populations structure, epidemiology
Anolis auratus (erythrocytes): western Columbia (Cauca River valley basin)

Plasmodium cynomolgi
Bennett, G. F.; Warren, M.; and Cheong, W. H., 1964, Med. J. Malaya, v. 19 (1), 32
strain variations of Plasmodium cynomolgi identified by sporogonic cycles

Plasmodium cynomolgi
Carroll, F. I.; et al., 1976, J. Med. Chem., v. 19 (9), 1111-1119
Plasmodium berghei-infected mice, Plasmodium cynomolgi-infected rhesus monkeys, laboratory trials with reduced 8-aminoquinoline analogues

Plasmodium cynomolgi (Langur strain)
Carter, R.; and Voller, A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 14-15 [Demonstration]
Plasmodium spp., survey of electrophoretic forms of 4 enzymes, variation between species, subspecies, and strains

Plasmodium cynomolgi
Chen, D. H.; Nussenzweig, R. S.; and Collins, W. E., 1976, J. Parasitol., v. 62 (4), 636-637
rat antisera against sporozoites of 6 primate Plasmodium spp. reacted in circum-sporozoite precipitation tests only with sporozoites of homologous species, geographically different strains of same species cross-reacted intensely however

P[lasmodium] cynomolgi
Chen, E. H.; et al., 1977, J. Med. Chem., v. 20 (8), 1107-1109
P[lasmodium] berghei in mice, P. cynomolgi in monkeys, 5-phenoxy derivatives of primaquine are more active and less toxic than primaquine in antimalarial screening trials

Plasmodium cynomolgi
Cheong, W. H.; et al., 1965, Med. J. Malaya, v. 20 (1), 52
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Plasmodium falciparum, Santa Lucia strain isolated from El Salvador, sensitive to commonly used antimalarial drugs, established in Aotus trivirgatus griseimembra (exper.), successful transmission from monkey to monkey both by transfusion of infected blood and through bites of infected Anopheles freeborni, A. maculatus, and A. albimanus (all exper.), description of course of infection, higher parasitemia and mortality in animals previously exposed to Plasmodium vivax, parasitemia in animals exposed to P. vivax and P. cynomolgi lower than those exposed to P. vivax only

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M. nemestrina
M. speciosa
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saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Ameiva ameiva: Panama

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clinical trials using trimethoprim in combination with sulfalene in treatment of drug resistant Plasmodium falciparum, results not as satisfactory as presently used regimens except in patients having repeated recrudescences after previous standard therapy

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Plasmodium falciparum, variations in electrophoretic forms of the enzymes glucose phosphate isomerase (GPI), lactate dehydrogenase (LDH) and 6-phosphogluconate dehydrogenase (6GPD) in isolates collected from various areas of Africa

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human Plasmodium falciparum, sensitivity of Thai strain of malaria tested against quinine alone and in combinations with tetracycline and pyrimethamine

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Cianci, P.; et al., 1972, Mil. Med., v. 137 (3), 113-114
Plasmodium falciparum, war casualties, stress of surgical repair of wounds and anesthesia as factor in development of clinical malaria

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sporozoite-induced human malaria (exper.), suppressive activity of mefloquine, more effective against Plasmodium falciparum than against P. vivax

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Plasmodium falciparum
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Plasmodium falciparum, Santa Lucia strain, comparative infectivity to different anophelines, infection rates highest if fed on Aotus trivirgatus between 19th and 25th day of patent gametocytemia
Anopheles freeborni (exper.)
A. maculatus (exper.)
A. balabacensis balabacensis (exper.)
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Plasmodium falciparum
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[Plasmodium] falciparum
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falciparum malaria, warning on use of tetracycline

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suggested technique for identification of scanty numbers of malarial parasites in human blood

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malarial antibodies compared in New Guineans with tropical splenomegaly syndrome and normal controls; results of study cannot implicate any one species of Plasmodium in production of this pathological condition

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Plasmodium falciparum
Dennis, D. T.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (3), 241-245
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Plasmodium falciparum in young girl, strain with resistance to chloroquine, quinacrine, pyrimethamine and chlorguanide, radical cure obtained with pyrimethamine and sulfadiazine in combination; delayed primary attack with Plasmodium vivax: Maryland (had traveled and engaged in field work in remote area of eastern Colombia)

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Plasmodium falciparum, lymphocyte sub-population changes in children with acute malaria, transient increase in K cell activity suggests that K cells have role in protective response to infection

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Plasmodium falciparum
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human falciparum malaria, clinical trials comparing quinine and chloroquine regimens (both drugs given with associated pyrimethamine and sulfa) in U.S. troops in Vietnam and volunteers in the United States; combinations using quinine most successful

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Plasmodium falciparum, probable chloroquine resistant malaria in woman who acquired infection while travelling in Central Java despite taking weekly prophylactic chloroquine-primaquine medication: Indonesia

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P[lasmodium] falciparum
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Plasmodium falciparum
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Kilejian, A.; Abati, A.; and Trager, W., 1977, Exper. Parasitol., v. 42 (1), 157-164
Plasmodium falciparum, Plasmodium coatneyi, knob-like protrusions on infected monkey erythrocyte membranes are antigenically different from adjacent areas devoid of knobs

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clinical review and management of 2 human cases of falciparum malaria presenting with psychiatric symptoms: Canada

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Plasmodium falciparum, suppression of malaria in Aotus trivirgatus fed exclusively on milk diet, indicates that P. falciparum is dependent on exogenous p-aminobenzoic acid supply for normal growth, supports view that dietary factors are involved in infant resistance to malaria

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Plasmodium falciparum, human, marked reduction in incidence of cold haemagglutinin

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Plasmodium falciparum, hospital trials with Gambian children testing BRL 50216 alone or in combinations with sulphadimethoxine or sulphafurazole (acting as potentiators); some response also with P. malariae; sporontocidal effect only when BRL 50216 given in presence of potentiators

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humans: Municipio of Aripuana, Mato Grosso State, Brazil

Plasmodium falciparum, illus.
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Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

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Plasmodium falciparum, possible mechanism for protection against malaria by sickle-cell trait, involves increased buffering by haemoglobin of fatty acids produced within erythrocyte by parasite, may prevent or retard release of merozoites and associated intravascular haemolysis

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acute renal insufficiency as complication of Plasmodium falciparum malaria, clinical management, diagnosis, case reports, pathologic findings, therapy

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survey of 173 males using indirect fluorescent antibody technique with Babesia bovis, B. bigemina, and B. ratti as antigens, babesial antibodies detected in 54%, no correlation found with mode of living, contact with livestock, incidence of malaria parasitaemias or malarial antibodies, no Babesia organisms found in blood smears or by animal subinoculation, source of antibodies unknown but subclinical infection not likely: Nigeria

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Plasmodium falciparum, P. knowlesi, acid protease activity in parasites and in ghosts of their respective host red cells, possible value of protease inhibitors in inhibiting growth of malarial parasites

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Plasmodium falciparum, P. vivax, suppression of malaria with monthly administration of combined sulphadoxine and pyrimethamine, compared with control groups on placebo and on weekly chloroquine, no serious side effects observed

Plasmodium falciparum, illus.
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Plasmodium falciparum in humans, physiopathology of hemolysis associated with pernicious parasitic infection

P[lasmodium] falciparum
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Plasmodium falciparum antigen slides made from in vitro culture for use in indirect immunofluorescence test

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parasitic infestation, preschool children, malnutrition and impaired immune response, brief review comment: Nigeria

P[lasmodium] falciparum
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malaria, prevalence survey of highland natives using enlargement of spleen as sign of infection, considerations for control: around Bokondini (Jayawijaya), Irian Jaya, Indonesian New Guinea

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Plasmodium spp., homologous and heterologous malaria pathogens used as antigens to determine malaria antibodies by direct immunofluorescence, pre- and post-treatment with chloroquine compared

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Mannweiler, E.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (1), 23-25
Plasmodium falciparum, P. vivax, strain related differences in immunodiagnosis, antigen variations as cause of antibody titer differences

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Plasmodium falciparum resulting in acute pulmonary insufficiency in African woman, successful treatment with intermittent positive pressure ventilation: Salisbury, Rhodesia

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malaria field studies, 1971-1973, incidence, seasonal occurrence, densities in regard to host age and sex: coastal area of El Salvador

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Plasmodium brasilianum, use as antigen in indirect hemagglutination test for diagnosing human P. falciparum, P. vivax, P. ovale, P. malariae

Plasmodium falciparum
Mathews, H. M.; Fried, J. A.; and Kagan, I. G., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 417-422
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Plasmodium falciparum, P. malariae, infants and young children, prevalence of malaria antibody evaluated using indirect hemagglutination test with P. falciparum antigen and filter paper blood specimens, slight decline in 6- to 8-month-old children with no demonstrable parasitemia but those older than 10 months had similar antibody levels regardless of presence or absence of demonstrated parasites in blood smears: Ivory Coast

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Plasmodium falciparum, identification of suspected exophilic vector, Anopheles aruni?, exper. laboratory infection from human volunteer: northeastern Transvaal, South Africa

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human malarias, current prospects and problems in developing a vaccine for malaria immunization

Plasmodium falciparum
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Plasmodium falciparum, P. knowlesi, susceptibility of human erythrocytes lacking various blood group antigens to invasion, differential effect of enzyme treatment of human erythrocytes on invasion, evidence for difference in erythrocyte surface receptors for these two parasites

Plasmodium falciparum
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erythrocyte receptors for malaria merozoites: Duffy blood group system and invasion by Plasmodium knowlesi and P. vivax, erythrocyte requirements for invasion by P. falciparum, workshop report

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Plasmodium falciparum, techniques for in vitro cultivation, immune IgG-mediated inhibition of development in vitro

Plasmodium falciparum
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Plasmodium falciparum, successful vaccination of Aotus trivirgatus griseimembra (exper.) with erythrocytic merozoites, immunity specific for falciparum malaria, technique for isolating merozoites from infected human blood

Plasmodium falciparum
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Plasmodium falciparum, rapid development of gametocytes in culture

Plasmodium falciparum
Mitra, P.; et al., 1976, Med. and Surg., Bombay, v. 16 (7), 28-29
case reports of Plasmodium falciparum in 2 young boys, clinical symptoms of cerebral malaria and bronchopneumonia, medical management: India

Plasmodium falciparum
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phytohaemagglutinin-induced lymphocyte transformation in leucocyte cultures from malarious, malnourished, and control Gambian children, some depression of response to low doses of PHA in children with malaria

Plasmodium falciparum
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Gambian children with acute Plasmodium falciparum malaria or with acute protein-energy malnutrition, phytohaemagglutin-induced lymphocyte transformation, depressed responses of purified lymphocytes in autologous plasma paralleled results with whole blood cultures

Plasmodium falciparum
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presence or absence of anti-Plasmodium falciparum and/or anti-P. malariae antibodies in patients with different types of glomerulo-nephritis: Senegal; Cameroun

Plasmodium falciparum
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Plasmodium falciparum cerebral malaria in children, possible relationships between malarial infection and nutritional state; use of heparin as adjunct to malarial therapy shows promising results in small clinical trial: Indonesia

Plasmodium falciparum
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Plasmodium spp. in humans, epidemiologic survey to determine prevalence rate in rural areas of the Philippines

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P[lasmodium] falciparum
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human malaria, statistical survey of cases imported into Japan from 1972-1974

Plasmodium falciparum
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Plasmodium falciparum, immunity to induced malaria, workshop report

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human antimalarial programs, progress review for 1975, comparison with previous statistics (regional surveys, training, research, immunology, chemotherapy, epidemiology)

Plasmodium falciparum
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Plasmodium cynomolgi, P. falciparum, production of antibodies against sporozoites in rats, no cross-reaction between two species

Plasmodium falciparum
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deviations in erythrogram before and after treatment of human malaria infections with resochin, no such changes in hookworm and other worm infections: Liberia; Togo

Plasmodium falciparum
O'Holohan, D. R., 1975, Med. J. Malaysia, v. 29 (3), 157-162
statistical survey of individual physician's practice for current trends in human malarial infections, review of presenting clinical symptoms: Seremban, West Malaysia

Plasmodium falciparum
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Plasmodium vivax, P. falciparum, human, analysis of clinical and laboratory findings of 1000 cases, need for diagnostic awareness and repeated blood smears in endemic areas: Malaysia

Plasmodium falciparum
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Plasmodium falciparum, humans, increasing evidence of chloroquine resistant strains in Africa

Plasmodium falciparum
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increasing incidence of Schistosoma mansoni in Gezira area of Sudan, clinical findings in inhabitants, frequent hepatosplenomegaly attributed to schistosomiasis and possibly hyperendemic malaria

Plasmodium falciparum
Otieno, L. H.; et al., 1971, Trop. and Geogr. Med., v. 23 (4), 369-375
human Plasmodium spp., dynamics of epidemiologic survey in highly endemic malarious area, data show decreasing incidence in recent years: north-eastern Tanzania

Plasmodium falciparum
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Plasmodium falciparum, human, Rieckmann in vitro field test confirmed sensitivity to chloroquine in agreement with in vivo tests: Ethiopia

Plasmodium falciparum
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multiple articles focusing on human malaria in Papua New Guinea (epidemiology, public health, control programs, vectors, diagnosis, toxicity of antimalarials, statistics of surveys)

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epidemiologic survey in South Sulawesi for human malarias and filariasis conducted 8 and 22 months after arrival of transmigrants from Central Java showed decreased incidence of malaria and increased filariasis, discussion of associated problems and effects on populace: Indonesia

P[lasmodium] falciparum
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P[lasmodium] falciparum, host resistance to infection probably related to genetics and acquired alterations in red blood cells

Plasmodium falciparum
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Plasmodium falciparum in vitro, preferential invasion of young red cells but no difference in rate of development in red cells of differing ages, foetal haemoglobin (Hb F) has no direct effect on rate of invasion of red cells but does cause retardation of parasite growth and development and may therefore offer some degree of protection, this may be possible mechanism for maintenance of β thalassaemia polymorphism since there is a retardation of rate of decline of Hb F production in infants heterozygous for β thalassaemia

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impact of control measures on malaria transmission and general mortality, extensive survey, results show relationship between efficacy of antivectorial measures (fenithrothion spraying) and decrease in general mortality and infant mortality in Kenya

Plasmodium falciparum
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Plasmodium falciparum, P. vivax, chloroquine resistant strains, field trials comparing efficacy of pyrimethamine combined with either sulfadoxine or diformyldapsone: Thailand

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Plasmodium falciparum, P. vivax, chemosuppressive field trials testing efficacy of diformyldapsone alone and in combination with pyrimethamine, comparisons with dapsone and pyrimethamine: Thailand

Plasmodium falciparum
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Plasmodium berghei, mice (exper.), studies on development of resistance to mefloquine and possible means of slowing down resistance process by mixing mefloquine with pyrimethamine, sulfaphenazole or primaquine; recommendation that mefloquine be used only to treat chloroquine resistant P. falciparum

Plasmodium falciparum
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Plasmodium vivax, P. falciparum, human, centrifuged vacutainer of venous blood procedure of choice in quantitative comparisons of methods of thin blood smear preparations for diagnosis

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Plasmodium falciparum, in vitro penetration of human red cells

Plasmodium falciparum
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Plasmodium falciparum, differentiation of gametocytes in vitro

Plasmodium falciparum
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"semi-immune" humans with acute attacks of Plasmodium falciparum, comparison drug trials using antemal and nivaquine, biochemical reactions of drug agents on trophozoites

P[lasmodium] falciparum
Ponnampalam, J. T., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 285 [Letter]
human malaria, increased risk of exposure of individuals to bite of mosquito vectors during the durian (local fruit) season (June-Sept.) when collecting fruit by night or living in the orchards during that time: West Malaysia

Plasmodium falciparum
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human malaria prophylaxis, controlled study comparing efficacy of dapsone combined with pyrimethamine (Maloprim) to that of chloroquine for use in long-term chemosuppression of infection: Malaysia

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Plasmodium falciparum
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changes in distribution and determinants of malaria that occurred with the development process in the highlands of Papua New Guinea between 1930 and 1970

Plasmodium] falciparum
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Plasmodium falciparum, P. vivax, humans, jaundice and hepatomegaly associated with attacks of primary uncomplicated malaria

lasmodium falciparum
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unsuccessful attempts to infect Macaca philippinensis philippinensis for use as laboratory host

Plasmodium falciparum
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Plasmodium falciparum, P. vivax, humans, evaluation of blood schizonticidal and radical curative activities of sulfamonomethoxine alone or combined with pyrimethamine in strains resistant and sensitive to chloroquine

Plasmodium falciparum
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Plasmodium falciparum, human, possible relationships between complement and decreased hemoglobin in parasitemia

Plasmodium falciparum
Reid, H. A.; and Nkrumah, F. K., 1972, Lancet, London (7744), v. 1, 218-221
Plasmodium falciparum, high levels of fibrin-degradation products in serum of patients with cerebral malaria, possible relationships to pathogenic processes of falciparum malaria

Plasmodium falciparum
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Plasmodium falciparum
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technique for removal of leucocytes from malaria infected blood

Plasmodium falciparum
Richards, W. H. G.; and Williams, S. G., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 135-140
Plasmodium falciparum in Aotus trivirgatus, treatment with chloroquine or 1-amidino-3-(3-chloro-4-cyanophenyl) urea, comparison of assessment in vivo with an in vitro test of protein-synthesizing activity (incorporation of radioactive leucine), in vitro system should prove useful in drug metabolism studies and in experiments using large or expensive animals

Plasmodium falciparum
Ridley, D. S.; and Hedge, E. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 522-525
microfilariae of various spp., immunofluorescent reactions involving sheath, cuticle, and cytoplasm, relevance to immuno-evasive mechanisms: (1) microfilariae failed to adsorb non-specific immunoglobulins in contrast to other helminth larvae and non-blood protozoa; (2) sheath of Wuchereria bancrofti and Loa loa adsorbed specific A and B blood group antigens; (3) low titer reaction between microfilarial cytoplasm (L. loa and W. bancrofti) and host serum

Plasmodium falciparum
Rieckmann, K. H.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (3), 258-259 [Letter]
Plasmodium falciparum, sporozoite induced immunity in human using infected irradiated mosquitoes

Plasmodium falciparum
Rinehart, J.; Arnold, J.; and Canfield, C. J., 1976, Am. J. Trop. Med. and Hyg., v. 25 (6), 769-774
Plasmodium falciparum, human, WR 122,455, WR 171,669, oral tolerance, toxicity, antimalarial efficacy

Plasmodium falciparum
Ringelhann, B.; et al., 1976, Am. J. Hum. Genet., v. 28 (3), 270-279
possible relationships between hemoglobin types and human malarial infection rate, parasite species, parasite density, host age and sex; correlations with transplacental and passive immunity

Plasmodium falciparum
Robinson, M. J.; and Eykyn, S., 1977, Brit. Med. J. (6103), v. 2, 1667 [Letter]
childhood malaria, increasing incidence with increased travel to and from endemic areas, diagnostic problem in children: United Kingdom

Plasmodium falciparum
Rosenberg, R.; and Maheswary, N. P., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 533 [Letter]
Plasmodium falciparum in humans, chloroquine resistant strain discovered in Bangladesh

Plasmodium falciparum
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development in Panamanian subspecies of howler monkey for possible laboratory host Aloutta villosa trabeata (exper.)

Plasmodium falciparum
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efficacies of potential antimalaria drugs (WR-142,490; WR-171,669; WR-172,435; WR-154,904; WR-180,409) using the owl monkey/Plasmodium falciparum model system

Plasmodium falciparum
Roux, J.; Picq, J. J.; and Marcadet, Y., 1974, Medecine Trop., v. 34 (2), 145-155
Plasmodium falciparum, application of indirect fluorescent antibody reaction with homologous antigen to epidemiologic and chemoprophylactic studies in human endemic areas: Upper Volta

P[lasmodium] falciparum
Roy, R. G.; et al., 1976, Indian J. Med. Research, v. 64 (10), 1451-1455
Plasmodium vivax, P. falciparum, humans, nirmalaria (indigenous compound of 4 herbal ingredients) unsuccessful in trials as inhalation therapy: India

Plasmodium falciparum
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malaria infections in children born and living in non-malarious areas who have contracted infections upon visiting malarious areas with their immigrant parents who possibly have some natural immunity to malaria although their children do not, importance of diagnostic awareness: Nottingham, England

Plasmodium falciparum
Ryan, B. P., 1975, Med. J. Australia, v. 1 (4), 101-103
clinical trials using combined sulfadoxine and pyrimethamine to treat human Plasmodium falciparum: Vietnam

Plasmodium falciparum
Rytel, M. W.; Rose, H. D.; and Stewart, R. D., 1973, Proc. Soc. Exper. Biol. and Med., v. 144 (1), 122-123
Trypanosoma cruzi, Plasmodium falciparum, P. vivax, humans, lack of circulating humoral interferon in serum of persons with acute illness

Plasmodium falciparum, illus.
Sauer, O., 1974, Immun. u. Infekt., v. 2 (4), 189-193
Plasmodium falciparum, children, 2 case reports, pathogenesis, epidemiology, treatment, need for diagnostic awareness with increased travel into tropical areas: Germany

P[lasmodium] falciparum
Schelp, F. P.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (3), 319-322
human falciparum malaria, effect of parasitism untreated for 2-4 days or 5-10 days on the host serum protein pattern, hematocrit levels, SGOT activity, and blood creatinine; measurement of these parameters may give evidence of duration and severity of infection

Plasmodium falciparum
Schmidt, L. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 446-474
Plasmodium falciparum and Plasmodium vivax in Aotus trivirgatus as model systems for chemotherapeutic studies, handling of animals, course of infections and response to standard antimalarial drugs compared with that in man, use of model in assessing activities of new agents and drug resistance

Plasmodium falciparum
Schmidt, L. H.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (5, part 1), 837-849
Plasmodium cynomolgi in Macaca mulatta, P. vivax and P. falciparum in Aotus trivirgatus griseimembra (both exper.), therapeutic potentiation with combinations of pyrimethamine and sulfonamides, good response in pyrimethamine susceptible and resistant strains except for Malayan Camp and Vietnam Smith strains of P. falciparum

Plasmodium falciparum
Segal, H. E.; et al., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (4), 499-503
Plasmodium falciparum, humans, assessment of gastrointestinal function and quinine absorption, findings suggest minimal distortion of absorptive function of little clinical importance

Plasmodium falciparum
Segal, H. E.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 139-142
Plasmodium falciparum in humans, clinical trials comparing efficacy of diaminodiphenylsulphone + pyrimethamine and sulfadoxine + pyrimethamine: Northeast Thailand

Plasmodium falciparum
Seigneurin, D., 1976, Nouv. Presse Med., v. 5 (38), 2552 [Letter]
human Plasmodium falciparum, presence of intra-erythrocytic parasites in blood resulted in error in platelet counting

Plasmodium falciparum
Shanmugam, C. A. K.; et al., 1976, Indian J. Med. Research, v. 64 (9), 1314-1317
Plasmodium falciparum, P. vivax, humans treated with chloroquine and/or primaquine, follow-up for chloroquine sensitivity status: Tamil Nadu State, India

Plasmodium falciparum
Shaw, P. K.; Brodsky, R. E.; and Schultz, M. G., 1976, J. Infect. Dis., v. 133 (1), 95-101
malaria surveillance, statistics for 1974, increasing incidence with increasing international travel: United States

Plasmodium falciparum, illus.
Siddiqui, W. A., 1977, Science (4301), v. 197, 388-389
Plasmodium falciparum, successful immunization of Aotus trivirgatus

Plasmodium falciparum
Siddiqui, W. A.; and Richmond-Crum, S. M., 1977, J. Parasitol., v. 63 (3), 583-584
Plasmodium falciparum, in vitro cultivation, fatty acid-free bovine albumin as replacement for plasma

Plasmodium falciparum, illus.
Sinden, R. E.; and Smalley, M. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 344-345
Plasmodium falciparum gametocytes, phagocytosis by leucocytes in vivo (in blood meal within Anopheles gambiae midgut) and in vitro, much higher percentage ingested in vitro

Plasmodium falciparum
Sinden, R. E.; and Smalley, M. E., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 381-382 [Demonstration]
Plasmodium falciparum, ingestion of gametocytes of Plasmodium by leucocytes in the bloodmeal of mosquitoes, comparison with incidence of ingestion of activated gametocytes in vitro

Plasmodium falciparum
Smalley, M. E., 1976, Nature (5583), v. 264, 271-272
Plasmodium falciparum, gametocytogenesis in vitro

Plasmodium falciparum
Smalley, M. E., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 526-529
Plasmodium falciparum, asexual erythrocytic parasites destroyed by chloroquine but mature gametocytes are not, the drug clumped pigment of developing gametocytes with only immature gametocytes in the final stages of development surviving

Plasmodium falciparum
Smalley, M. E.; and Sinden, R. E., 1977, Parasitology, v. 74 (1), 1-8
Plasmodium falciparum, gametocytes are both long-lived and show persistent infectivity to mosquitoes, they can stimulate antibody production but immune response appears to play no part in their elimination

Plasmodium falciparum
Smitskamp, H.; and Wolthuis, F. H., 1976, Nederl. Tijdschr. Geneesk., v. 120 (35), 1503-1505
human Plasmodium falciparum, recommendations for medical management of severe infection using intravenous quinine, improved microcirculation and prevention of edema: Rotterdam, Netherlands

Plasmodium falciparum
Srichaikul, T.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 36-38
Plasmodium falciparum, man, blood histamine changes and correlation with severity of complications occurring during infection, possible release of histamine through activation of complement system and immune destruction of platelets

Plasmodium falciparum
Srichaikul, T.; Siriasawakul, T.; and Poshyachinda, M., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 244-246
Plasmodium falciparum in humans, defect in hemoglobin synthesis during infection, effect on normoblastic development in vitro, possible role of complement in depression of erythropoiesis

Plasmodium falciparum
Sroczynski, J., 1977, Polski Tygod. Lekar., v. 32 (17), 637-638
Plasmodium falciparum, relatively mild course of infection in persons with sickle cell anemia

P[lasmodium] falciparum
Sucharit, P.; et al., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 145-146 [Demonstration]
P[lasmodium] falciparum in humans with pernicious manifestations, attempted correlation of clinical features and laboratory evidence of intravascular coagulation

[Plasmodium] falciparum
Sucharit, P.; et al., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 33-39
[Plasmodium] falciparum, studies show that coagulation and fibrinolysis develop only in persons who have cerebral manifestations of malaria or have high parasitemia

Plasmodium falciparum
Sucharit, P.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 401-405
Plasmodium falciparum, evaluation of relationships between chloroquine concentrations in in vitro cultures which inhibit maturation of asexual erythrocytic forms of parasite strains found in Thailand and the in vivo response of 3 parasite strains

Plasmodium falciparum
Sulzer, A. J.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 55-58
malaria, human, diagnosis, indirect fluorescent antibody test using multi-species thick smear antigen containing equal proportions of Plasmodium vivax, P. falciparum, and P. brasilianum (latter serologically equivalent in IFA test to P. malariae), circumvents need for multiple testing with several antigen species

Plasmodium falciparum
Sulzer, A. J.; and Latorre, C. R., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 553 [Letter]
primate malarias, method for in vitro production of schizont antigens for use in indirect immunofluorescence tests for malarial antibodies

Plasmodium falciparum
Szarfman, A.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 19-24
evaluation of specificity of EVI factor observed in Trypanosoma cruzi cases using indirect fluorescent antibody test on sera from patients with other parasitic diseases, immunofluorescence of anti-skeletal muscle antibody from leishmaniasis

P[lasmodium] falciparum
Thaver, S.; and Draper, C. C., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 8 [Demonstration]
P[lasmodium] falciparum, P. vivax and T[rypanosoma] cruzi antibodies tested for reliability of epidemiologic field survey collection methods, use of chromatography paper for collecting capillary blood recommended

Plasmodium falciparum
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Plasmodium falciparum-infected human erythrocytes, glucose-6-phosphate and 6-phosphogluconate dehydrogenase activity

Plasmodium falciparum
Thomas, V.; and Dissanaike, A. S., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 602-606
Plasmodium spp., detection of malaria endemicity among Orang Asli aborigines using Plasmodium falciparum and P. brasilianum antigens and indirect fluorescent antibody test (IFA), age dependent increase in number of positive results, IFA valuable as adjunct to blood slide examination especially when parasites are at very low levels: Malaysia

Plasmodium falciparum
Thompson, R. G.; et al., 1977, Brit. Med. J. (6074), v. 1, 1468 [Letter]
human malaria, increasing number of infections seen in travelers returning from endemic areas, suggested prophylactic measures: United Kingdom

Plasmodium falciparum
Tin, F.; and Nyunt Hlaing, 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (1), 104-112
Plasmodium falciparum, humans, proposed new treatment regimens for recently discovered infections resistant to standard doses of chloroquine, pyrimethamine, and proguanil: Burma

P[lasmodium] falciparum
Topley, E.; Knight, R.; and Woodruff, A. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 51-54
malaria patients, direct antiglobulin test and immunoconglutinin titres, possible significance of results in understanding mechanism of anemia

Plasmodium falciparum
Trager, W.; and Jensen, J. B., 1976, Science (4254), v. 193, 673-675
Plasmodium falciparum, continuous culture in human erythrocytes incubated at 38°C in RPMI 1640 medium with human serum under an atmosphere with 7% CO_2 and low oxygen

Plasmodium falciparum
Truong-Minh-Ky, D.; et al., 1976, Medecine Afrique Noire, v. 23 (4), 215-223
Plasmodium malariae, Plasmodium falciparum, survey of village infants for presence of malarial antibodies: Abidjan

Plasmodium falciparum
Tsukamoto, M.; et al., 1975, Nettai Igaku (Trop. Med.), v. 17 (1), 13-26
Plasmodium spp. in humans, field trials of biochemical analysis of blood and urine, value comparisons before and after antimalarial therapy: Philippines

Plasmodium falciparum
Valera, C. V.; and Shute, G. T., 1976, Bull. World Health Organ., v. 53 (4), 391-398
Plasmodium falciparum in humans, preliminary evaluation of distribution of chloroquine resistant strains and local incidence using the Rieckmann in vitro technique: Manila, the Philippines

Plasmodium falciparum
Vanderberg, J. P., 1974, J. Protozool., v. 21 (4), 527-537
Plasmodium spp. sporozoites, motility, locomotion, movement, effects of several factors (parasite species, oocyst vs. salivary gland sporozoites, presence of albumin or globulins, temperature), relationship to infectivity, immunogenicity (circumsporozoite precipitate reaction), and secretory activity

Plasmodium falciparum
Verdrager, J.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (3), 58-66
chloroquine resistant Plasmodium falciparum reported in East Kalimantan, Island of Borneo, Indonesia, correlation of resistant malaria with area of dispersion of A[nopheles] balabacensis vector mosquitoes

Plasmodium falciparum
Voller, A.; Bartlett, A.; and Bidwell, D. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 98-106
application of enzyme immunoassay to diagnosis of human parasitic diseases, technique for microplate enzyme-linked immunosorbent assay

Plasmodium falciparum, illus.
Warhurst, D. C.; and Thomas, S. C., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 417-420
Plasmodium berghei, P. falciparum, localization of mepacrine by fluorescence microscopy

Plasmodium falciparum
Warren, K. S.; and Mahmoud, A. A. F., 1976, J. Infect. Dis., v. 133 (5), 596-601
prevention of malaria, schistosomiasis and other tropical and exotic diseases, advice to travellers

Plasmodium falciparum
Warren, M.; et al., 1976, Am. J. Trop. Med. and Hyg., v. 25 (1), 20-25
Plasmodium falciparum, human, localized epidemics, serologic assessment with indirect fluorescent antibody method provides valuable information but must be interpreted in association with other known epidemiologic factors: El Salvador; Panama

Plasmodium falciparum
Warren, M.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 607-611
differences in susceptibility to Plasmodium vivax and P. falciparum of inbred morphologic phenotypes of Anopheles albimanus vector mosquitoes, significance in epidemiologic surveys and development of vector control measures: El Salvador

[Plasmodium] falciparum
Wartofsky, L.; et al., 1977, J. Clin. Endocrinol. and Metab., v. 44 (1), 85-90
acute falciparum malaria, nature of thyroidal suppression, integrity of pituitary response to thyrotropin-releasing hormone and alterations in serum T_3 and reverse T_3

Plasmodium falciparum
Webber, R. H., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 396-400
control measures carried out by the Malaria Eradication Programme against Anopheles vectors of human malaria also resulted in the natural decline in the prevalence of human Wuchereria bancrofti infections since both diseases are transmitted by the same vector in the Solomon Islands

Plasmodium falciparum
Weiss, N.; and Degremont, A., 1976, Praxis, Bern, v. 65 (24), 742-744
human malaria, comparative evaluation of Plasmodium gallinaceum, P. falciparum and P. cynomolgi bastianellii as antigens for indirect immunofluorescence used for diagnosis

Plasmodium falciparum
Welch, S. G.; McGregor, I. A.; and Williams, K., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 295-296
survey of 2 rural Gambian villages showed total absence of Duffy blood group antigens and absence of Plasmodium vivax infection although P. falciparum, P. malariae and P. ovale were present; findings consistant with theory that absence of Duffy phenotypes constitutes basis of innate resistance to P. vivax

Plasmodium falciparum
Wilkinson, R. N.; Colwell, E. J.; and Neoypatimanond [sic], S., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 148-149 [Letter]
Plasmodium falciparum, gametocytes not sterilized by sulphamethoxazole-trimethoprim treatment, this treatment regimen should be supplemented by effective gametocytocidal or sporontocidal drug to interrupt transmission

Plasmodium falciparum
Wilkinson, R. N.; Noeypatimanondh [sic], S.; and Gould, D. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 306-307
Plasmodium falciparum, malaria patients before and after chloroquine treatment, infectivity for Anopheles balabacensis and A. minimus, possible relationship to drug resistance: Thailand

Plasmodium falciparum, illus.
Williams, A. I. O., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (5), 621-630
Plasmodium falciparum, preparation and use of malarial placenta antigen for immunodiffusion studies: Ibadan, Western State of Nigeria

Plasmodium falciparum
Williamson, J.; and Cover, B., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 78-87
Plasmodium falciparum, application of differential sucrose gradient centrifugation to separation and purification of human erythrocytic stages and gametocytes and of nitrogen cavitation for host-cell-free parasite preparation, gel diffusion analyses showed that highly purified infected cell preparations retained precipitinogenic spectrum of original crude preparation, possible use in vaccine preparation

Plasmodium falciparum
Wilson, R. J. M.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 308-312
Plasmodium falciparum, human, precipitating antibody response to malarial S-antigens, age distribution, other factors affecting production and detection of such antibodies: The Gambia

Plasmodium falciparum
Wilson, R. J. M.; McGregor, I. A.; and Hall, P. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 460-467
Plasmodium falciparum, persistence and recurrence of S-antigens in human serum following antimalarial treatment, correlation with density of parasitemia and antigen titer observed before treatment

Plasmodium falciparum
Wilson, R. J. M.; McGregor, I. A.; and Williams, K., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 453-459
Plasmodium falciparum, prevalence of malarial S-antigens in human serum or plasma, relationship of antigen presence to season and infection intensity

Plasmodium falciparum
Wilson, R. J. M.; and Pasvol, G., 1977, J. Protozool., v. 24 (2), Suppl., 4A-5A [Abstract]
Plasmodium falciparum, distribution and growth compared in human red blood cells containing adult vs. fetal hemoglobins, results suggest that young metabolically active cells may be preferentially invaded but that parasite growth was impaired in fetal cells, this may be an important protective mechanism for both fetus and newborn

Plasmodium falciparum
Wilson, R. J. M.; and Phillips, R. S., 1976, Nature, London (5573), v. 263, 132-134
Plasmodium falciparum in Gambian children, in vitro screening of sera for anti-Plasmodium falciparum activity raised questions about antigenic diversity and duration of inhibitory antibody response, method devised to test known homologous or heterologous combinations of parasitised cells and sera (parasite multiplication and uptake of ^{3}H-isoleucine in serum obtained before and after chloroquine treatment)

Plasmodium falciparum
Wilson, R. J. M.; and Phillips, R. S., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 279-280 [Demonstration]
Plasmodium falciparum, multiplication in vitro of cryopreserved parasites as a method to test inhibitory antibody responses to wild populations of parasites of potentially disparate antigenicity

Plasmodium falciparum
Wilson, R. J. M.; and Williams, K., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 15-20
European with suspected pyrimethamine-resistant malaria, serological study (flurorescent antibody test, C-reactive protein, precipitin test, immunoglobulin levels), precipitating antibodies demonstrated against two specific malarial antigens, differences from antibodies seen in sera from repeatedly infected Africans: The Gambia, West Africa

Plasmodium falciparum
Wyler, D. J., 1976, Clin. and Exper. Immunol., v. 23 (3), 471-476
Plasmodium falciparum, human (30 children, 3 adults), peripheral lymphocyte subpopulations altered by infection, decrease in % and concentration of T cells, increase in % but not concentration of B cells, increase in % and concentration of 'null' cells, effects rapidly reversible after antimalarial treatment, presumably represent sequestration of T cells in spleen or other organs: The Gambia, West Africa

Plasmodium falciparum
Wyler, D. J.; and Brown, J., 1977, Clin. and Exper. Immunol., v. 29 (3), 401-407
Plasmodium falciparum, malaria antigen-specific T-cell responsiveness not abrogated during infection, need for further work to identify basis of immunosuppression in malarial infection; possibility of acquiring sensitized T cells without experiencing clinically apparent infections

Plasmodium falciparum
Wyler, D. J.; and Gallin, J. I., 1977, J. Immunol., v. 118 (2), 478-484
Plasmodium spp. in mice and monkeys, spleen-derived mononuclear cell chemotactic factor as a possible mechanism for splenic macrophage accumulation

Plasmodium falciparum
Yoeli, M., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 24-35
Plasmodium berghei yoelii in mice (exper.), promising model for study of cerebral malaria in man caused by P. falciparum (history, origin of virulent strain, pathology, enzyme differentiation, virulence expressed by mutation and genetic change)

Plasmodium falciparum
Young, M. D.; Baerg, D. C.; and Rossan, R. N., 1976, Lab. Animal Sc., v. 26 (6, pt. 2), 1131-1137
Plasmodium falciparum, P. vivax, induced malarias in Aotus monkeys, transmission to other New World monkeys and man, use as biological model, literature review

Plasmodium] falciparum
Ziessman, H. A., 1976, J. Nuclear Med., v. 17 (9), 794-796
increased lung uptake of sulfur colloid during liver scan in human and experimental malaria due to enhanced activity of phagocytic cells of the reticuloendothelial system of liver, spleen, and lung

Plasmodium falciparum
Zuidema, P. J., 1971, Nederl. Tijdschr. Geneesk., v. 115 (29), 1209-1212
man, case history of mixed infections of Entamoeba histolytica, Necator americanus, Plasmodium falciparum and Schistosoma mansoni, medical management: Netherlands (native of Surinam)

Plasmodium fallax
Holbrook, T. W.; Spitalny, G. L.; and Palczuk, N. C., 1976, J. Parasitol., v. 62 (5), 670-675
mice immunized with Plasmodium fallax exoerythrocytic stages, stimulation of resistance to sporozoite-induced P. berghei malaria

Plasmodium fallax
Huchzermeyer, F. W., 1976, Rhodesian Vet. J., v. 7 (3), 52-57
turkeys (blood): Lower Gwelo, Rhodesia

Plasmodium fieldi, illus.
Aikawa, M.; Hsieh, C. L.; and Miller, L. H., 1977, J. Parasitol., v. 63 (1), 152-154
Plasmodium simiovale- and P. fieldi-infected Macaca mulatta, ultrastructural alterations in cell membrane of erythrocytes, caveola-vesicle complexes likely correspond to Schüffner's dots, electron dense excrescences at protruded tip of infected erythrocytes

Plasmodium fieldi
Carter, R.; and Voller, A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 14-15 [Demonstration]
Plasmodium spp., survey of electrophoretic forms of 4 enzymes, variation between species, subspecies, and strains

Plasmodium fieldi
Chen, D. H.; Nussenzweig, R. S.; and Collins, W. E., 1976, J. Parasitol., v. 62 (4), 636-637
rat antisera against sporozoites of 6 primate Plasmodium spp. reacted in circum-sporozoite precipitation tests only with sporozoites of homologous species, geographically different strains of same species cross-reacted intensely however

P[lasmodium] fieldi
zum Felde, I.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (4), 477-484
P[lasmodium] falciparum, P. vivax, P. ovale, P. malariae, human, indirect immunoflourescence test with same 4 Plasmodium spp. plus P. fieldi as antigen, species-specific and cross-reactions, effect of therapy, implications for immunodiagnosis

Plasmodium fieldi
Mannweiler, E.; et al., 1976, Munchen. Med. Wchnschr., v. 118 (36), 1139-1144
Plasmodium spp., homologous and heterologous malaria pathogens used as antigens to determine malaria antibodies by direct immunofluorescence, pre- and post-treatment with chloroquine compared

Plasmodium fieldi
Warren, M.; et al., 1964, Med. J. Malaya, v. 19 (1), 31
morphological variations, Macaca spp.

Plasmodium floridense
Jordan, H. B., 1975, J. Protozool., v. 22 (2), 241-244
Plasmodium floridense, differential course of infection in Anolis carolinensis vs. Sceloporus undulatus, duration and intensity of parasitemia, % frequency of merozoite means during periods of acute rise and decline, natural and acquired immunity: Echols and Clinch counties, Georgia

Plasmodium floridense
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis humilis: Costa Rica
A. limifrons: Costa Rica; Panama
A. lemurinus: Belize
A. lionotus: Costa Rica
A. tropidonotus: Honduras
A. biporcatus: Panama
A. frenatus: Panama
A. pentaprion: Panama

Plasmodium foleyi Buck et al., 1952, illus.
Garnham, P. C. C.; and Uilenberg, G., 1975, Ann. Parasitol., v. 50 (4), 409-418
redescription
Syn.: P. lemuris Huff & Hoogstraal, 1963

Plasmodium foleyi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium formosanum Manwell
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Anborophila crudiigularis (blood): Taiwan

Plasmodium fragile
Collins, W. E.; et al., 1975, J. Parasitol., v. 61 (4), 718-721
successful infection of Anopheles balabacensis balabacensis by feeding on splenectomized Macaca mulatta, transmission to Macaca mulatta via bite of Anopheles balabacensis balabacensis or via inoculation of infected salivary glands; comparative infectivity to Anopheles freeborni, Anopheles maculatus, Anopheles quadrimaculatus

Plasmodium fragile, illus.
Collins, W. E.; et al., 1977, J. Parasitol., v. 63 (2), 226-231
Plasmodium fragile, exoerythrocytic stages in Macaca mulatta (liver), description

Plasmodium fragile, illus.
Fremount, H. N.; and Miller, L. H., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 1-8
organ distribution and ultrastructure of Plasmodium fragile-infected erythrocytes adherent to vascular endothelium

Plasmodium fragile
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium fragile (Dissanaike, Nelson and Garnham, 1965)
Nelson, P., 1971, Tr. Roy. Soc. Trop. Med. and Hyg., v. 65 (5), 695-696 [Letter]
Anopheles elegans established as vector of simian malaria in Ceylon
rhesus monkey (exper.)

Plasmodium gallinaceum
Atkinson, E. R.; and Granchelli, F. E., 1976, J. Pharm. Sc., v. 65 (4), 618-620
4-aminobenzothiazoles ineffective in laboratory trials with Plasmodium gallinaceum in chicks

Plasmodium gallinaceum
Bisseru, B.; and Lim, K. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 412 [Demonstration]
Corvus splendens protegatus (blood): Klang, Selangor, Malaysia

Plasmodium gallinaceum
Carter, R.; and Chen, D. H., 1976, Nature, London (5572), v. 263, 57-60
vaccination of chickens with Plasmodium gallinaceum gametes reduced infectivity of malarious chickens for Aedes aegypti at least 99.9% below control levels

Plasmodium gallinaceum
Carter, R.; and Nijhout, M. M., 1977, Science (4276), v. 195, 407-409
Plasmodium gallinaceum, control of gamete formation (exflagellation) in vitro solely by change in pH in blood as it moves from environment of circulation to that of atmosphere, the pH rise being mediated by fall in carbon dioxide tension as blood equilibrates with atmosphere

Plasmodium gallinaceum
Dave, D.; Gutteridge, W. E.; and Richards, W. H. G., 1976, Parasitology, v. 73 (2), xvii [Abstract]
Plasmodium spp., conversion of dihydroorotate to orotate, activities were irreversible, required O_2, and were inhibited by CN^- and Antimycin A, no involvement of reduced pteridines or pyridine nucleotides detected, this area of metabolism may be important site for selective drug action

Plasmodium gallinaceum
Dojmi Di Delupis, G.; Palmieri, C.; and Piccione, G., 1972, Parassitologia, v. 14 (2-3), 305-308
Plasmodium gallinaceum, maintenance in lines of Anopheles stephensi selected for low or high receptivity, development of oocysts in various generations of both lines

Plasmodium gallinaceum
Fernando, M. A.; and Dissanaike, A. S., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 144 [Demonstration]
Gallus gallus spadiceus: Tasek Berah, State of Pahang
Aedes aegypti (exper.)

Plasmodium gallinaceum, Brumpt, 1935, illus.
Fernando, M. A.; and Dissanaike, A. S., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 25-32
Plasmodium gallinaceum and P. juxtanucleare, developmental and morphologic comparisons, pleomorphism of gametocytes and erythrocytic schizonts of P. gallinaceum
Gallus gallus spadiceus (blood): Peninsular Malaysia

Plasmodium gallinaceum
Ferone, R., 1973, J. Protozool., v. 20 (3), 459-464
Plasmodium spp., enzymic synthesis of dihydropteroate and dihydrofolate

Plasmodium gallinaceum
Ferris, D. H.; Beamer, P. D.; and Stutz, D. R., 1973, Avian Dis., v. 17 (1), 12-23
Plasmodium gallinaceum, response to exper. infection in dysgammaglobulinemic chickens compared to infected sham bursectomized and intact birds: parasitemia, histopathology; results indicate that resistance to initial infection is related to bursa-mediated defenses and humoral factors have a role in protection against malaria

Plasmodium gallinaceum
Gun, F.; and Greenblatt, C. L., 1973, J. Protozool., v. 20 (4), 532
Plasmodium berghei, P. gallinaceum, cultivation in cell hybrids formed by fusing infected red blood cells with HeLa cells, chick fibroblasts, baby hamster kidney cells and macrophages

Plasmodium gallinaceum
Gwadz, R. W., 1976, Science (4258), v. 193, 1150-1151
Plasmodium gallinaceum, reduction or elimination of gametocyte infectivity and oocyst development in Aedes aegypti by immunizing chickens on which mosquitoes feed with formalin or x-ray treated infected red blood cells, protection apparently related to immobilization of microgametes in mosquito gut and associated with IgG fraction of serum

Plasmodium gallinaceum
Kohlmann, F. W.; and Sous, H., 1976, Arzneimittel-Forsch., v. 26 (4a), 618-620
pharmacological action and efficacy of combination sulfamoxole and trimethoprim against Plasmodium gallinaceum in chickens and Plasmodium berghei in mice, laboratory trials

Plasmodium gallinaceum, illus.
Lushbaugh, W. B.; McGhee, R. B.; and Singh, S. D., 1976, J. Protozool., v. 23 (1), 127-134
Plasmodium gallinaceum, erythrocytic stages in embryonic and neonate chicks, abnormal morphology and development associated with adaptation to immature host

Plasmodium gallinaceum
McGhee, R. B., 1976, Exper. Parasitol., v. 39 (1), 88-94
Plasmodium gallinaceum-infected immunoincompetent chicken embryos, changes in blood picture in response to injection of serum from hyperimmunized chickens, results suggest definite role of immunity in anemia accompanying malaria, failure to clarify question of autoimmunity

Plasmodium gallinaceum
McGhee, R. B.; and Singh, S. D., 1974, J. Protozool., v. 21 (3), 422-423 [Abstract]
Plasmodium gallinaceum, adaptation sufficient to allow continuous maintenance in chicken embryos, selection toward virulence

Plasmodium gallinaceum
McGhee, R. B.; Singh, S. D.; and Lushbaugh, W. B., 1975, J. Protozool., v. 22 (3), 18A [Abstract]
Plasmodium gallinaceum in chicken embryos of different ages, differences in development and reproduction may be due to different hemoglobin composition

Plasmodium gallinaceum
McGhee, R. B.; Singh, S. D.; and Lushbaugh, W. B., 1977, Exper. Parasitol., v. 43 (1), 220-230
Plasmodium gallinaceum, changing virulence patterns during adaptation from neonate chick to chicken embryos

Plasmodium gallinaceum
McGhee, R. B.; Singh, S. D.; and Weathersby, A. B., 1977, Exper. Parasitol., v. 43 (1), 231-238
Plasmodium gallinaceum, chicks, vaccination with embryo-adapted strain, completely effective as protection against erythrocytic stages

Plasmodium gallinaceum
Mandarini-Galli, L.; Brambilla, E.; and Sacchi, L., 1974, Riv. Parassitol., Roma, v. 35 (4), 253-260
Plasmodium gallinaceum, chicks, course of infection (parasitemia, symptomatology, mortality) varies with ambient temperature

Plasmodium gallinaceum
March, L. C.; et al., 1976, J. Med. Chem., v. 19 (6), 845-848
successful laboratory trials and synthesis of several 1,2,4-triazines against Plasmodium berghei-infected mice and Plasmodium gallinaceum-infected chicks

Plasmodium gallinaceum
Prior, R. B.; and Kreier, J. P., 1977, Ohio J. Sc., v. 77 (2), 91-96
Plasmodium gallinaceum, release from chicken erythrocytes by ultrasound, energy frequency, intensity, and exposure time necessary for erythrocyte breakdown

Plasmodium gallinaceum
Ramalingam, S.; Dissanaike, A. S.; and Fernando, M. A., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 148 [Demonstration]
microsporidian in Culex sitiens interfered with development of attempted experimental Plasmodium gallinaceum infections in the mosquito: Rantau Panjang area, West Malaysia

Plasmodium gallinaceum
Rank, R. G.; and Weidanz, W. P., 1976, Proc. Soc. Exper. Biol. and Med., v. 151 (2), 257-259
Plasmodium gallinaceum, fatal fulminating parasitemias in agammaglobulinemic chickens, B-cell deficient chickens rescued from primary infection by chloroquine therapy resisted challenge infection, data suggest that premunition to malaria in chickens is B-cell independent

Plasmodium gallinaceum
Rees, R. W. A.; et al., 1976, J. Med. Chem., v. 19 (5), 723-725
13 analogs of pyrimethamine tested in Plasmodium berghei-infected mice, the more potent drugs were then tested in Plasmodium gallinaceum-infected chicks, promising activity

Plasmodium gallinaceum
Richards, W. H. G.; and Latter, V. S., 1977, Parasitology, v. 75 (2), xxix-xxx [Abstract]
Plasmodium gallinaceum, chicks, immunization

Plasmodium gallinaceum
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Plasmodium gallinaceum, illus.
Seed, T. M.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 525-535
Plasmodium gallinaceum, P. berghei, intra- and extracellular parasites, general topography and surface ultrastructure, scanning and transmission electron microscopy

Plasmodium gallinaceum
Sinden, R. E.; and Garnham, P. C. C., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (5), 631-637
Plasmodium spp., sporozoite ultrastructure within oocyst and salivary glands, incidence of micropores and possible relationship to infectivity, sporozoite morphogenesis, abnormalities in development

Plasmodium gallinaceum
Skippon, J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 3 [Demonstration]
domestic chickens (blood): vicinity of Kota Kinabalu, Borneo

Plasmodium gallinaceum
Soni, J. L.; and Cox, H. W., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 423-430
Plasmodium gallinaceum, chickens, soluble complexes of serum antigen and its antibody may be mediator of acute anemia, serologic identity of serum antigen from malarious chickens and from Babesia rodhaini-infected rats and its distinction from parasite antigen suggest that it might be an autoantigenic macroglobulin

Plasmodium gallinaceum
Soni, J. L.; and Cox, H. W., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 431-438
Plasmodium gallinaceum, chickens, complexes of serum antigen and its antibody may cause glomerulonephritis associated with acute avian malaria

Plasmodium gallinaceum
Soni, J. L.; and Cox, H. W., 1976, Indian J. Med. Research, v. 64 (8), 1177-1184
immune complex associated with malarious nephritis of Plasmodium gallinaceum-infected chicken, isolation and description of antigen component of complex

Plasmodium gallinaceum, illus.
Sterling, C. R.; and Aikawa, M., 1973, J. Protozool., v. 20 (1), 81-92
Plasmodium gallinaceum, Haemoproteus columbae, Leucocytozoon simondi, gametocyte ultrastructure compared in detail

Plasmodium gallinaceum, illus.
Terzakis, J. A.; Vanderberg, J. P.; and Weiss, M. M., 1976, J. Parasitol., v. 62 (3), 366-371
ultrastructural appearance of viruslike particles in several Plasmodium spp. at different times in sporogony

Plasmodium gallinaceum
Theakston, R. D. G.; and Fletcher, K. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 16-17 [Demonstration]
Plasmodium spp., 6-phosphogluconate dehydrogenase in infected erythrocytes, activity present in parasite cytoplasm as well as in host cell

Plasmodium gallinaceum, illus.
Trefiak, W. D.; and Desser, S. S., 1973, J. Protozool., v. 20 (1), 73-80
crystalloid inclusions in Leucocytozoon, Parahaemoproteus, and Plasmodium, lipid-protein nature, ultrastructural morphology

Plasmodium gallinaceum
Tsuneda, K., 1974, Nettai Igaku (Trop. Med.), v. 16 (1), 11-20
Plasmodium gallinaceum, asexual forms and gametocytes, cyclic changes in numbers in blood of infected fowls, effect of chloroquine treatment, exflagellation of gametocytes (as indication of infectivity for mosquito vectors) demonstrated as long as mature gametocytes could be found in blood

Plasmodium gallinaceum
Weidanz, W. P.; and Rank, R. G., 1975, Proc. Soc. Exper. Biol. and Med., v. 148 (3), 725-728
Plasmodium gallinaceum, chickens (exper.), immunosuppressive effects of infection

Plasmodium gallinaceum
Weiss, N.; and Degremont, A., 1976, Praxis, Bern, v. 65 (24), 742-744
human malaria, comparative evaluation of Plasmodium gallinaceum, P. falciparum and P. cynomolgi bastianellii as antigens for indirect immunofluorescence used for diagnosis

Plasmodium gallinaceum
Yamaguchi, E.; and Nakabayashi, T., 1976, Nettai Igaku (Trop. Med.), v. 18 (1), 45-48
Plasmodium gallinaceum, influence of temperature and carbon dioxide on exflagellation in vitro

Plasmodium garnhami
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Upupa epops: Ethiopia

Plasmodium garnhami Guindy, Hoogstraal et Mohammed, 1965
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Coracias garrulus]: north-east Azerbaidzhan

Plasmodium girardi, illus.
Garnham, P. C. C.; and Uilenberg, G., 1975, Ann. Parasitol., v. 50 (4), 409-418
redescription
Lemur macaco macaco: Madagascar
L. macaco fulvus: Madagascar
L. mongoz mongoz (exper.)

Plasmodium girardi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium gonatodi
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Gonatodes albogularis: Panama

Plasmodium gonderi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium hegneri
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Anas c. crecca (blood): Taiwan

Plasmodium (Huffia) hermani sp. n., illus.
Telford, S. R., jr.; and Forrester, D. J., 1975, J. Protozool., v. 22 (3), 324-328
pathology, strong immune response of infected birds
Meleagris gallopavo (erythrocytes, erythroblasts, bone marrow, spleen, liver, lung, brain): Palmdale, Glades County, and Lochloosa Wildlife Management Area, Alachua County, Florida
Colinus virginianus (exper.)
Anser anser (exper., transient)
Serinus canaria (exper., transient)

Plasmodium hexamerium
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Emberiza spodocephala subsp. (blood): Taiwan

Plasmodium huffi Muniz, Soares & Battista pro parte
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
as syn. of P. nucleophilum toucani Manwell & Sessler 1971

Plasmodium hylobati
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium inui
Carter, R.; and Voller, A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 14-15 [Demonstration]
Plasmodium spp., survey of electrophoretic forms of 4 enzymes, variation between species, subspecies, and strains

Plasmodium inui
Cheong, W. H.; et al., 1965, Med. J. Malaya, v. 20 (1), 52
isolation of Plasmodium cynomolgi and Plasmodium inui from A[nopheles] b. balabacensis mosquitoes and exper. infections in rhesus monkeys: Perlis, Malaya

Plasmodium inui
Cheong, W. H.; et al., 1965, Med. J. Malaya, v. 20 (1), 76
Anopheles balabacensis balabacensis as vector of simian malaria in Malaya

Plasmodium inui-like
Das, S. K.; Strauss, J. M.; and Chee, C. T., 1966, Med. J. Malaya, v. 20 (4), 323
Plasmodium inui-like infection presumably producing fatal illness in Indian leaf monkey (Presbytis johnii): National Zoo, Malaysia (natural habitat in Southern India)

Plasmodium inui
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Plasmodium inui
Wyler, D. J.; Miller, L. H.; and Schmidt, L. H., 1977, J. Infect. Dis., v. 135 (1), 86-93
Plasmodium inui, intact and splenectomized rhesus monkeys (exper.), role of spleen in host defense and chronicity in malaria infection, results suggest that the spleen plays a protective role during the acute infection and a suppressive role during the chronic phase

Plasmodium juxtanucleare
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Francolinus sephaena: Ethiopia

Plasmodium juxtanucleare
Bisseru, B.; and Lim, K. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 412 [Demonstration]
Corvus splendens protegatus (blood): Klang, Selangor, Malaysia

Plasmodium juxtanucleare
Fernando, M. A.; and Dissanaike, A. S., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 144 [Demonstration]
Gallus gallus spadiceus: Tasek Berah, State of Pahang
Aedes togoi (exper.)
Culex sitiens "

Plasmodium juxtanucleare, Versiani & Gomes, 1941, illus.
Fernando, M. A.; and Dissanaike, A. S., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (1), 25-32
Plasmodium gallinaceum and P. juxtanucleare, developmental and morphologic comparisons, pleomorphism of gametocytes and erythrocytic schizonts of P. gallinaceum
Gallus gallus spadiceus (blood): Peninsular Malaysia

Plasmodium juxtanucleare
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Bambusicola thoracica sonorivox (blood): Taiwan

Plasmodium juxtanucleare
Shanta, C. S.; and Wan, S. P., 1976, Malaysian Vet. J., v. 6 (2), 45-49
Plasmodium juxtanucleare, chickens (exper.), pyrimethamine at various concentrations, not effective in eliminating parasite from peripheral circulation, only useful for reducing parasitaemia in younger birds thereby lowering morbidity and mortality rates

Plasmodium juxtanucleare
Shanta, C. S.; Wan, S. P.; and Kwong, K. H., 1977, Kajian Vet., v. 9 (1), 14-18
Plasmodium juxtanucleare, chickens (exper.), sulfadoxine + pyrimethamine, good results

Plasmodium knowlesi
Angus, M. G. N.; Fletcher, K. A.; and Maegraith, B. G., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 419-427
Plasmodium knowlesi-infected Macaca mulatta, changes in liver lipids

Plasmodium knowlesi
Angus, M. G. N.; Fletcher, K. A.; and Maegraith, B. G., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 429-439
Plasmodium knowlesi-infected Macaca mulatta, changes in erythrocyte lipids

Plasmodium knowlesi
Areekul, S., 1972, Southeast Asian J. Trop. Med. and Pub. Health, v. 3 (4), 617-618
Plasmodium knowlesi in Macaca mulatta (exper.), infected red cells travel at same rate as normal red cells with no evidence of being trapped in capillaries of brains of infected monkeys when compared with normal controls

P[lasmodium] knowlesi
Areekul, S.; et al., 1972, Southeast Asian J. Trop. Med. and Pub. Health, v. 3 (4), 505-510
P[lasmodium] falciparum in humans, P. knowlesi in Macaca mulatta, serum haptoglobin levels during course of infection and after treatment, fall and rise of haptoglobin levels were in parallel with changes of hemoglobin concentrations, alterations probably due to hemolysis and increased phagocytic activity

Plasmodium knowlesi
Areekul, S.; et al., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (4), 474-480
Plasmodium knowlesi in Macaca mulatta, alteration of liver blood flow and phagocytic activity of reticuloendothelial system in infected monkeys with return to normal value after recovery

P[lasmodium] knowlesi
Areekul, S.; et al., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (3), 452 [Demonstration]
P[lasmodium] spp., erythrocyte adenosine triphosphate content compared in controls and infected man, monkeys and mice

Plasmodium knowlesi
Areekul, S.; Chantachum, Y.; and Thaijongrak, K., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (4), 598-600
Plasmodium knowlesi infected Macaca mulatta, measurement of increased erythrophagocytic activity during infection

Plasmodium knowlesi
Bannister, L. H.; et al., 1975, J. Protozool., v. 22 (3), 48A [Abstract]
Plasmodium knowlesi merozoites, invasion of erythrocytes, electron microscopic study

Plasmodium knowlesi
Bannister, L. H.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 5 [Demonstration]
Plasmodium knowlesi merozoites, fine structure and invasive behavior

Plasmodium knowlesi
Bannister, L. H.; Butcher, G. A.; and Mitchell, G. H., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 13 [Demonstration]
Plasmodium knowlesi, extracellular merozoites, description of cell coat and dense bodies

Plasmodium knowlesi
Bannister, L. H.; Butcher, G. A.; and Mitchell, G. H., 1977, J. Protozool., v. 24 (2), Suppl., 6A [Abstract]
Plasmodium knowlesi, P. yoelii, extracellular erythrocytic merozoites, nature of cell surface, adhesion with other cells

Plasmodium knowlesi, illus.
Brown, K. N., 1977, Advances Exper. Med. and Biol., v. 93, 5-25
antigenic variation in malaria, review: Plasmodium knowlesi, schizont-infected cell agglutination test, protective variant-specific antibodies, induction of antigenic variation, antigenic variation and the cell cycle, antigenic variation and red cell penetration, variant-specific antibody synthesis and protective immunity; Plasmodium berghei, lymphocyte subpopulations and immunity transcending antigenic variation; gametocytes, antigenic variation, and protection; antigenic variation and immunization against malaria

Plasmodium knowlesi
Brown, K. N.; and Hills, L. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 139-142
Plasmodium knowlesi, variant-specific antibodies detected in exper. Macaca mulatta, one inducing antigenic variation and one with parasiticidal response

Plasmodium knowlesi
Butcher, G. A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 554-555 [Letter]
Plasmodium knowlesi, unsuccessful attempt to demonstrate stimulus that induces synthesis of new antigens by parasite in response to a development of immunity by host

Plasmodium knowlesi
Butcher, G. A.; Bannister, L. H.; and Mitchell, G. H., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 11 [Demonstration]
Plasmodium knowlesi, immune damage to intracellular parasites, morphologic features

Plasmodium knowlesi
Butcher, G. A.; Mitchell, G. H.; and Cohen, S., 1973, Nature (5410), v. 244, 40-42
Plasmodium knowlesi, ability of serum and of red cells from susceptible and resistant hosts to support growth and multiplication, properties of serum could not be correlated with susceptibility or resistance of donor species but properties of host erythrocytes did play a role, implications for mechanism of host specificity

Plasmodium knowlesi
Cabrera, E. J.; et al., 1976, Ztschr. Parasitenk., v. 50 (1), 31-42
Plasmodium knowlesi, monkeys immunized by antigen from infected erythrocytes, delayed dermal hypersensitivity response, protection against challenge infection; preliminary biochemical analysis of antigen

Plasmodium knowlesi
Cabrera, E. J.; Barr, M. L.; and Silverman, P. H., 1977, Infect. and Immun., v. 15 (2), 461-465
Plasmodium knowlesi-vaccinated Macaca mulatta, protection against challenge with heterologous strain present even 4 years after immunization schedule had been completed

Plasmodium knowlesi
Carter, R.; and Voller, A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 14-15 [Demonstration]
Plasmodium spp., survey of electrophoretic forms of 4 enzymes, variation between species, subspecies, and strains

Plasmodium knowlesi
Chen, D. H.; Nussenzweig, R. S.; and Collins, W. E., 1976, J. Parasitol., v. 62 (4), 636-637
rat antisera against sporozoites of 6 primate Plasmodium spp. reacted in circum-sporozoite precipitation tests only with sporozoites of homologous species, geographically different strains of same species cross-reacted intensely however

P[lasmodium] knowlesi
Cohen, S., 1976, Ztschr. Immunitaetsforsch., v. 152 (2), 80-81 [Abstract]
P[lasmodium] knowlesi, successful vaccination of rhesus monkeys

Plasmodium knowlesi
Cohen, S.; Butcher, G. A.; and Mitchell, G.H., 1977, Advances Exper. Med. and Biol., v. 93, 89-112
immunization against erythrocytic forms of malaria parasites, review: malaria life cycle; innate and acquired resistance to malaria (specific malarial antibody, protective malarial antibody, synergistic action of malarial antibody and cells, role of specific cell-mediated immunity in malaria, isolation and properties of merozoites); vaccination against erythrocytic forms of malaria (vaccination and challenge using undefined and defined variants of Plasmodium knowlesi, adjuvant requirement for successful vaccination)

Plasmodium knowlesi
Collins, W. E.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (3), 373-376
Plasmodium knowlesi in Macaca mulatta (exper.), attempted immunization using heat-stable serum-soluble antigens, protection incomplete but with fewer deaths and reduced maximum parasitemia than in nonimmunized or Freund's adjuvant-immunized monkeys

Plasmodium knowlesi
Conklin, K. A.; et al., 1973, J. Protozool., v. 20 (5), 683-688
Plasmodium knowlesi, intraerythrocytic stages, incorporation of nucleic acid precursors, periodicity of DNA synthesis

Plasmodium knowlesi, illus.
Cowper, S. G., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 415-425
continuous flow apparatus for in vitro maintenance: Schistosoma mansoni, S. haematobium, survival time, carbohydrate metabolism; Plasmodium knowlesi, morphology and carbohydrate metabolism; preliminary attempts to cultivate Trypanosoma vivax and Babesia canis

Plasmodium knowlesi
Cox, R. A.; Godwin, E.; and Hastings, J. R. B., 1976, Biochem. J., v. 155 (3), 465-475
Plasmodium knowlesi, ribosomal ribonucleic acid, spectroscopic evidence for uneven distribution of adenine and uracil residues, possible evolutionary significance

Plasmodium knowlesi
Dave, D.; Gutteridge, W. E.; and Richards, W. H. G., 1976, Parasitology, v. 73 (2), xvii [Abstract]
Plasmodium spp., conversion of dihydroorotate to orotate, activities were irreversible, required O_2, and were inhibited by CN^- and Antimycin A, no involvement of reduced pteridines or pyridine nucleotides detected, this area of metabolism may be important site for selective drug action

Plasmodium knowlesi
Ferone, R., 1973, J. Protozool., v. 20 (3), 459-464
Plasmodium spp., enzymic synthesis of dihydropteroate and dihydrofolate

Plasmodium knowlesi
Fletcher, K. A.; Canning, M. V.; and Theakston, R. D. G., 1977, Ann. Trop. Med. and Parasitol., v. 71 (2), 125-130
Plasmodium berghei-infected mice, rats and hamsters (all exper.) and P. knowlesi-infected monkeys (exper.), absence of glucose-6-phosphate activity of "parasite origin" and presence of 6-phosphogluconate dehydrogenases of "parasite origin" in erythrocytes of infected animals, possible relationships in erythrocyte metabolism and parasite growth

Plasmodium knowlesi
Hommel, M.; McColm, A. A.; and Trigg, P. I., 1977, J. Protozool., v. 24 (4), 40A-41A [Abstract]
Plasmodium knowlesi, reinvasion in vitro into erythrocytes pretreated with membrane-active drugs

Plasmodium knowlesi
Houba, V.; Faulk, W. P.; and Matola, Y. G., 1974, Clin. and Exper. Immunol., v. 18 (1), 89-93
heterophilic antibodies to sheep erythrocytes identified in sera from both malarious and non-malarious subjects in Tanzania, in patients with sickle-cell anemia but not exposed to malaria (U.S.A.), and in rhesus monkeys infected with Plasmodium knowlesi and P. cynomolgi bastianellii, observations do not support concept that heterophilic antibodies are integrally associated with host resistance to malaria

Plasmodium knowlesi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium knowlesi
Levy, M. R.; Siddiqui, W. A.; and Chou, S. C., 1974, Nature (5442), v. 247, 546-549
Plasmodium falciparum, P. knowlesi, acid protease activity in parasites and in ghosts of their respective host red cells, possible value of protease inhibitors in inhibiting growth of malarial parasites

Plasmodium knowlesi, illus.
Lunde, M. N.; and Powers, K. G., 1976, Ann. Trop. Med. and Parasitol., v. 70 (3), 283-291
Plasmodium knowlesi, comparison of methods of preparing malarial hemagglutination antigen

Plasmodium knowlesi
McClean, S.; et al., 1976, Analyt. Chim. Acta, v. 82 (1), 175-185
phospholipid composition of normal and infected rhesus monkey erythrocyte membranes and of the parasite itself

Plasmodium knowlesi
McColm, A. A.; Shakespeare, P. G.; and Trigg, P. I., 1976, Parasitology, v. 73 (2), xvii [Abstract]
Plasmodium knowlesi, protein synthesis in vitro

Plasmodium knowlesi
McColm, A. A.; Shakespeare, P. G.; and Trigg, P. I., 1977, J. Protozool., v. 24 (4), 40A [Abstract]
Plasmodium knowlesi, proteins released by schizonts cultivated in vitro

P[lasmodium] knowlesi
Maskrey, P.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 15-16 [Demonstration]
P[lasmodium] knowlesi-infected rhesus monkeys, cytotoxic substances in sera

Plasmodium knowlesi, illus.
Mason, S. J.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (2), 195-197
Plasmodium knowlesi, surface coat on merozoites, transmission electron microscopy supports theory of origination of coat from parasite rather than from adsorbed serum protein of culture medium

Plasmodium knowlesi
Mason, S. J.; et al., 1977, Brit. J. Haematol., v. 36 (3), 327-335
Plasmodium knowlesi, evaluation of role of Duffy blood group negative erythrocytes in host resistance to invasion by P. knowlesi merozoites

Plasmodium knowlesi
Migasena, P.; and Maegraith, B. G., 1970, Southeast Asian J. Trop. Med. and Pub. Health, v. 1 (4), 559 [Demonstration]
Plasmodium knowlesi in Macaca mulatta, barriers to movement of albumin from plasma to cerebrospinal fluid and from spinal fluid to plasma resulting from infection

Plasmodium knowlesi
Miller, L. H.; et al., 1977, J. Exper. Med., v. 146 (1), 277-281
Plasmodium falciparum, P. knowlesi, susceptibility of human erythrocytes lacking various blood group antigens to invasion, differential effect of enzyme treatment of human erythrocytes on invasion, evidence for difference in erythrocyte surface receptors for these two parasites

Plasmodium knowlesi, illus.
Miller, L. H.; Aikawa, M.; and Dvorak, J. A., 1975, J. Immunol., v. 114 (4), 1237-1242
Plasmodium knowlesi, immune serum agglutinated merozoites in culture, agglutinated merozoites attached to erythrocytes but were unable to invade, merozoite agglutination was caused by binding of surface coats on adjacent parasites

Plasmodium knowlesi
Miller, L. H.; McAuliffe, F. M.; and Mason, S. J., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, Pt. 2), 204-208
erythrocyte receptors for malaria merozoites: Duffy blood group system and invasion by Plasmodium knowlesi and P. vivax, erythrocyte requirements for invasion by P. falciparum, workshop report

Plasmodium knowlesi
Miller, L. H.; Powers, K. G.; and Shiroishi, T., 1977, Exper. Parasitol., v. 41 (1), 105-111
Plasmodium knowlesi in rhesus monkeys, no correlation between functional immunity and results of 2 in vitro tests (schizont-infected cell agglutination test; suppression of merozoite invasion by immune serum)

Plasmodium knowlesi
Mitchell, G. H., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 281-282
Plasmodium knowlesi malaria in rhesus monkeys (exper.), techniques for isolation of merozoites for use in experimental vaccines, review of current status, symposium presentation

Plasmodium knowlesi
Mitchell, G. H.; et al., 1977, Clin. and Exp. Immunol., v. 28 (2), 276-279
Plasmodium knowlesi, rhesus monkeys, effectiveness of freeze-dried merozoite vaccine in protecting against homologous or heterologous variant blood challenge

Plasmodium knowlesi
Phillips, R. S.; Trigg, P. I.; and Gutteridge, W. E., 1973, Isotopes and Radiation Parasitol. III, 185-186
Plasmodium knowlesi-infected red cells exposed to gamma irradiation and incubated in in vitro culture, growth, rate of incorporation of radiotracers into parasite DNA, RNA, and protein, brief note

Plasmodium knowlesi, illus.
Powers, K. G.; Aikawa, M.; and Nugent, K. M., 1976, Exper. Parasitol., v. 40 (1), 13-24
Plasmodium knowlesi in rhesus monkeys, clinical and morphologic effects of clindamycin and its N-demethyl-4'-pentyl analog

Plasmodium knowlesi
Richards, W. H. G.; et al., 1977, Parasitology, v. 74 (2), 191-198
Plasmodium knowlesi, merozoite vaccination of Macaca mulatta, immunity to sporozoite (mosquito-transmitted) challenge even if challenge strain was different from that used for vaccination, implications for development of human malaria vaccine

Plasmodium knowlesi
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Plasmodium knowlesi
Richards, W. H. G.; and Williams, S. G., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 179-190
Plasmodium berghei, in vitro evaluation of drug activities using whole blood vs. leucocyte-free blood-dilution cultures and ^{3}H-leucine uptake, comparison with P. knowlesi

Plasmodium knowlesi
Richards, W. H. G.; and Williams, S. G., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 249-250
technique for removal of leucocytes from malaria infected blood

Plasmodium knowlesi
Schenkel, R. H.; et al., 1975, J. Parasitol., v. 61 (3), 549-550
Plasmodium knowlesi, rhesus monkeys, immunization with lyophilized antigen plus Adjuvant 65 and BCG afforded same protection as antigen plus Freund's Complete Adjuvant, results suggest important role for cell-mediated immunity in vaccine-induced protection against malaria

Plasmodium knowlesi
Schmidt, L. H.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 612-622
Plasmodium knowlesi, differences in virulence for Macaca irus strains depending on their Philippine or Malayan origins, possible application to production of merozoite vaccines and evaluation of efficacy

Plasmodium knowlesi
Shakespeare, P. G.; and Trigg, P. I., 1973, J. Protozool., v. 20 (4), 527-528
Plasmodium knowlesi, glucose catabolism, changes during life cycle

Plasmodium knowlesi
Shakespeare, P. G.; and Trigg, P. I., 1973, Nature (5391), v. 241, 538-540
Plasmodium knowlesi, glucose catabolism in normal rhesus monkey erythrocytes, in parasitized erythrocytes, and in parasites "freed" from their host erythrocytes

Plasmodium knowlesi
Sherman, I. W., 1974, J. Protozool., v. 21 (3), 452 [Abstract]
Plasmodium knowlesi cell-free system, requirements for optimum protein synthesis, effect of various drugs

Plasmodium knowlesi
Sherman, I. W., 1976, Comp. Biochem. and Physiol., v. 53 (4B), 447-450
Plasmodium knowlesi ribosomes, development of highly active cell-free protein synthesizing system, use in assaying antimalarials

Plasmodium knowlesi, illus.
Sherman, I. W.; et al., 1975, J. Protozool., v. 22 (4), 568-572
Plasmodium knowlesi, ribosomes, isolation technique, characterization

Plasmodium knowlesi
Siddiqui, W. A.; Schnell, J. V.; and Richmond-Crum, S. M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 387-391
susceptibility of Aotus trivirgatus (exper.) and maintenance of infection by serial passages

Plasmodium knowlesi
Smith, C. C.; McCormick, G. J.; and Canfield, C. J., 1976, Exper. Parasitol., v. 40 (3), 432-437
Plasmodium knowlesi, in vitro biosynthesis of methionine

Plasmodium knowlesi
Smith, C. C.; McCormick, G. J.; and Canfield, C. J., 1977, Exper. Parasitol., v. 43 (1), 248-254
Plasmodium knowlesi, in vitro biosynthesis of methionine and thymidylic acid, related folate metabolism

Plasmodium knowlesi, illus.
Speer, C. A.; Silverman, P. H.; and Barr, M.L., 1976, J. Protozool., v. 23 (3), 437-442
Plasmodium knowlesi, antigen used in vaccination of rhesus monkeys, ultrastructural examination of material from various steps obtained in French pressure cell technic of preparing antigen from infected red cells

Plasmodium knowlesi
Speer, C. A.; Silverman, P. H.; and Schiewe, S. G., 1976, Ztschr. Parasitenk., v. 50 (3), 237-244
Plasmodium berghei, P. vinckei vinckei, P. coatneyi, P. knowlesi, attempts to cultivate erythrocytic stages in various cell lines, success only with P. berghei in mouse Leydig cell testicular tumor cultures

Plasmodium knowlesi
Speer, C. A.; Silverman, P. H.; and Schiewe, S. G., 1976, J. Parasitol., v. 62 (5), 657-663
Plasmodium berghei, erythrocytic stages, cultivation in primary cultures of rodent bone marrow cells, unsuccessful with P. vinckei vinckei and P. knowlesi

Plasmodium knowlesi
Sucharit, P.; and Reid, H. A., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (1), 15-18
Plasmodium knowlesi-infected Macaca mulatta, blood value comparisons with normal controls, multiple coagulation defects, increased fibrinogen/fibrin degradation products and evidence of intravascular coagulation in monkeys dying from infection

Plasmodium knowlesi
Sucharit, P.; and Reid, H. A., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (2), 168-171
Plasmodium knowlesi, fibrinogen catabolism compared in normal and infected monkeys

Plasmodium knowlesi
Theakston, R. D. G.; and Fletcher, K. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 16-17 [Demonstration]
Plasmodium spp., 6-phosphogluconate dehydrogenase in infected erythrocytes, activity present in parasite cytoplasm as well as in host cell

Plasmodium knowlesi
Trigg, P. I.; and Shakespeare, P. G., 1975, J. Protozool., v. 22 (3), 57A [Abstract]
Plasmodium knowlesi, factors affecting invasion of erythrocytes in vitro, results demonstrate that maintenance of integrity of surface of erythrocyte in culture is necessary prerequisite for efficient culture system

Plasmodium knowlesi
Trigg, P. I.; and Shakespeare, P. G., 1976, Parasitology, v. 73 (2), 149-160
changes in uninfected rhesus monkey erythrocytes incubated in vitro (osmotic fragility, acetylthiocholinesterase activity, glucose catabolism, ATP content) in relation to susceptibility of these cells to invasion by Plasmodium knowlesi, results suggest that maintenance of erythrocyte surface integrity is necessary prerequisite for efficient culture system for malaria parasite

Plasmodium knowlesi
Wyler, D. J.; and Gallin, J. I., 1977, J. Immunol., v. 118 (2), 478-484
Plasmodium spp. in mice and monkeys, spleen-derived mononuclear cell chemotactic factor as a possible mechanism for splenic macrophage accumulation

Plasmodium lemuris Huff & Hoogstraal, 1963
Garnham, P. C. C.; and Uilenberg, G., 1975, Ann. Parasitol., v. 50 (4), 409-418
as syn. of P. foleyi Buck et al., 1952

Plasmodium lophurae
Beach, D. H.; Sherman, I. W.; and Holz, G. G., jr., 1977, J. Parasitol., v. 63 (1), 62-75
Plasmodium lophurae, lipids of parasite and of erythrocytes and plasma of normal and infected Pekin ducklings

Plasmodium lophurae
Brohn, F. H., 1974, J. Protozool., v. 21 (3), 434 [Abstract]
Plasmodium lophurae, enzymes required for coenzyme A biosynthesis detected in host duck erythrocytes but parasite extracts show no activity, coenzyme A required by parasite is synthesized entirely in host erythrocyte

Plasmodium lophurae
Dimopoullos, G. T.; and Finerty, J. F., 1976, Am. J. Vet. Research, v. 37 (6), 693-695
Anaplasma marginale, Plasmodium lophurae, P. berghei, cross reactivity in passive haemagglutination tests

Plasmodium lophurae
Dykstra, W. G.; and Meszoely, C. A., 1975, J. Protozool., v. 22 (3), 17A [Abstract]
Plasmodium lophurae intraerythrocytic stages, quinine-induced morphological changes apparently specific for feeding stages

Plasmodium lophurae
Ferone, R., 1973, J. Protozool., v. 20 (3), 459-464
Plasmodium spp., enzymic synthesis of dihydropteroate and dihydrofolate

Plasmodium lophurae
Hart, L. T.; Dimopoullos, G. T.; and Mandhare, K. S., 1973, Avian Dis., v. 17 (4), 752-757
Plasmodium lophurae, pyrimethamine, effect on in vitro incorporation of sodium acetate by lipids of blood from normal and infected ducks, inhibition of lipolysis in both normal and infected blood

'lasmodium lophurae
Holz, G. G., jr.; Beach, D. H.; and Sherman, I. W., 1977, J. Protozool., v. 24 (4), 566-574
octadecenoic fatty acid content of lipids of erythrocytes and plasmas of normal and Plasmodium lophurae-infected Pekin ducklings and of P. lophurae itself, hemolytic properties of octadecenoic fatty acids

lasmodium lophurae
Kilejian, A., 1974, J. Protozool., v. 21 (3), 434 [Abstract]
Plasmodium lophurae, mitochondrial DNA

Plasmodium lophurae
Kilejian, A., 1975, J. Protozool., v. 22 (3), 40A [Abstract]
Plasmodium coatneyi, P. lophurae, DNA properties

Plasmodium lophurae, illus.
Kilejian, A., 1976, J. Protozool., v. 23 (2), 272-277
Plasmodium lophurae, histidine-rich protein, possible role in merozoite penetration

Plasmodium lophurae, illus.
Langreth, S. G., 1976, J. Protozool., v. 23 (2), 215-223
Babesia microti vs. Plasmodium lophurae, feeding mechanisms of parasites freed of host erythrocytes, ferritin uptake of in vitro culture system

Plasmodium lophurae, illus.
Langreth, S. G.; and Trager, W., 1973, J. Protozool., v. 20 (5), 606-613
Plasmodium lophurae, extracellular development in vitro, light and electron microscopy, incorporation of methionine or proline, observation of abnormalities which may indicate limits of extracellular cultivation in vitro (loss of host-derived outer parasite membrane and apparent reduction in feeding activity via food vacuoles)

Plasmodium lophurae
Langreth, S. G.; and Trager, W., 1975, J. Protozool., v. 22 (3), 23A [Abstract]
Babesia microti, Plasmodium lophurae, comparison of feeding mechanisms

Plasmodium lophurae
Mandhare, K. S.; Dimopoullos, G. T.; and Hart, L. T., 1973, Avian Dis., v. 17 (3), 617-622
Plasmodium lophurae-infected ducks, in vitro incorporation of sodium acetate into fatty acids of lipid classes of blood plasma and erythrocytes

Plasmodium lophurae, illus.
Meszoely, C. A. M.; et al., 1975, Proc. 33. Ann. Meet. Electron Microsc. Soc. America, 654-655
Plasmodium lophurae, pellicular complex of exoerythrocytic merozoites from turkey brain monolayer cell culture, electron microscopy of freeze-fractures

Plasmodium lophurae
Platzer, E. G., 1974, J. Protozool., v. 21 (2), 400-405
Plasmodium lophurae, dihydrofolate reductase in parasite and in duckling erythrocytes, temporal changes in activity, isolation, properties, sensitivity to pyrimethamine inhibition

Plasmodium lophurae
Platzer. E. G.. 1977. Life Sc., v. 20 (8), 1417-1423
Plasmodium lophurae, subcellular distribution of serine hydroxymethyltransferase, cytochrome oxidase, glutamate dehydrogenase, and malate dehydrogenase

Plasmodium lophurae
Platzer, E. G.; and Campuzano, H. C., 1976, J. Protozool., v. 23 (2), 282-286
Plasmodium lophurae, serine hydroxymethyltransferase, partial purification and characterization

Plasmodium lophurae
Sherman, I. W.; and Jones, L. A., 1976, J. Protozool., v. 23 (2), 277-281
Plasmodium lophurae, reliable procedure for isolation of active ribosomes, optimal requirements for in vitro synthesis of proteins, effects of variety of drugs on duck reticulocyte and plasmodial ribosomes in cell-free protein-synthesizing systems (restricted selectivity not obtained)

Plasmodium lophurae
Sherman, I. W.; and Jones, L. A., 1977, J. Protozool., v. 24 (2), 331-334
Plasmodium lophurae, ribosomes, isolation and characterization, results suggest that host ribosomes do not contribute to ribosome biogenesis in this malaria, but rather parasite rRNA is the transcription product of plasmodial cistrons

Plasmodium lophurae
Sherman, I. W.; and Tanigoshi, L., 1974, J. Protozool., v. 21 (4), 603-607
Plasmodium lophurae-infected erythrocytes, enhanced glucose transport

Plasmodium lophurae
Warner, V. D.; et al., 1976, J. Pharm. Sc., v. 66 (3), 418-419
Plasmodium lophurae, turkeys, synthesis and evaluation of isopropyl [(4-chlorophenyl) amino]iminomethylcarbamimidate, compared with chloroguanide

Plasmodium malariae
Anderson, R. I.; and Buck, A. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 447-456
complement levels in residents of rural village in relation to wide variety of clinical, laboratory, and epidemiological factors including parasitic diseases: Ouli Bangala, Republic of Chad

Plasmodium malariae
Ayala, S. C.; and Sarmiento, G., 1976, Actualidades Biol., v. 5 (17), 55-61
Plasmodium, man and animals, review: Colombia

Plasmodium malariae
Bell, D. R.; and Howells, R. E., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 1-14
malaria, blood film and spleen examination results in relation to age, chloroquine and cycloguanil sensitivity of Plasmodium falciparum; Schistosoma haematobium, prevalence and intensity in relation to age and sex: Malumfashi District, North Central State of Nigeria

Plasmodium malariae
Bidwell, D.; and Voller, A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 6 [Demonstration]
Plasmodium spp., counter immunoelectrophoresis studies on development of precipitating antibodies

Plasmodium malariae
Bolton, J. M., 1972, Med. J. Malaysia, v. 27 (1), 10-19
review of malarial control program among Orang Asli aborigines, prophylactic distribution of chloroquine and pyrimethamine and frequent spraying of homes with DDT implimented, discussion of problems involved: West Malaysia jungle areas

Plasmodium malariae
Bourgeade, A.; and Bertrand, E., 1972, Medecine Afrique Noire, v. 19 (4), 377-380
quinine as drug of choice in human malarias, indications for use

Plasmodium malariae
Brown, P.; et al., 1976, Am. J. Trop. Med. and Hyg., v. 25 (6), 775-783
Plasmodium, 4 human spp., remote island populations, fluorescent antibody prevalence patterns, comparison with published parasite survey data based on blood smears and medical examinations, evaluation of epidemiological usefulness: New Hebrides; Solomons; Western Carolines (malaria entirely absent); New Guinea

Plasmodium malariae
Bruce-Chwatt, L. J.; and de Zulueta, J., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (3), 232-240
human malaria, details of eradication program to present stage of certification by WHO as area where malaria has been eradicated, emphasis on need for continued vigilance because of increased travel and imported cases: Portugal

Plasmodium malariae
Butler, T.; Warren, K. S.; and Mahmoud, A. A., 1976, J. Infect. Dis., v. 133 (6), 721-726
human malarias, algorithms in the diagnosis and management

Plasmodium malariae
Charters, A. D.; and Davis, R. E., 1974, Med. J. Australia, v. 1 (14), 527-530
statistical review of human malaria infections imported into Western Australia, increasing incidence, drug resistance to commonly used antimalarials in 4 persons

Plasmodium malariae
Collins, W. E.; Contacos, P. G.; and Richardson, B. B., 1975, J. Parasitol., v. 61 (5), 941-943
Plasmodium malariae, comparative infectivity studies with 5 species of anophelines feeding on infected Aotus trivirgatus; results of passages in A. trivirgatus
Anopheles freeborni
A. balabacensis balabacensis
A. atroparvus
A. stephensi
A. maculatus
(all exper.)

Plasmodium malariae
Cook, G. C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 143-145
Plasmodium falciparum, P. malariae in humans, possible correlations between malarial infection, host malnutrition and immunoglobulin G concentrations

Plasmodium malariae
Crane, G. G.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 308-314
malarial antibodies compared in New Guineans with tropical splenomegaly syndrome and normal controls; results of study cannot implicate any one species of Plasmodium in production of this pathological condition

Plasmodium malariae
Degremont, A. ; and Weiss, N., 1975, Praxis, Bern, v. 64 (18), 553-555
human malarias, evaluation of immunofluorescence as adjunct to diagnostic serology

Plasmodium malariae
Dondero, T. J., jr.; Parsons, R. E.; and Ponnampalam, J. T., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 149-154
Plasmodium falciparum, appreciable chloroquine and chloroquine combined with pyrimethamine drug resistance discovered in survey of peninsular area; no evidence of resistance with P. vivax and P. malariae: Malaysia

Plasmodium malariae
Edrissian, G. H., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (6), 491-493
Plasmodium malariae and P. vivax in humans, importance of blood transfusions as means of transmission and need for stringent control measures: Iran

Plasmodium malariae
Elslager, E. F., 1974, Progr. Drug Research, v. 18, 99-172
human malaria and filariasis, new perspectives on chemotherapy, extensive review

P[lasmodium] malariae
zum Felde, I.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (4), 477-484
P[lasmodium] falciparum, P. vivax, P. ovale, P. malariae, human, indirect immunofluorescence test with same 4 Plasmodium spp. plus P. fieldi as antigen, species-specific and cross-reactions, effect of therapy, implications for immunodiagnosis

Plasmodium malariae
Gibson, J. J., jr.; Brodsky, R. E.; and Schultz, M. G., 1974, J. Infect. Dis., v. 130 (5), 553-555
increased incidence of malaria in the United States probably reflects increased foreign travel and improved reporting of infections

Plasmodium malariae
Gilles, H. M., 1975, Medecine et Malad. Infect., v. 5 (12), special no., 602-608
relationship between Plasmodium malariae and nephrotic syndrome in children, epidemiology, clinical aspects

Plasmodium malariae
Gueye, I.; and Michel, R., 1973, Medecine Afrique Noire, v. 20 (10), 753-766
guidelines for mass prophylactic therapy against human forms of malaria

P[lasmodium] malariae
Harinasuta, T.; Gilles, H. M.; and Sandosham, A. A. [Editors], 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (4), 641-678
human malaria in southeast Asia, scientific group meeting with discussion on: epidemiology, clinical features, pathophysiology, genetic factors, immunology, diagnosis, chemotherapy, control measures

Plasmodium malariae
Hendrickse, R. G., 1975, Paediat. Indonesiana, v. 15 (3-4), 83-92
Plasmodium malariae in children as cause of immune complex nephritis probably involving an auto-immune process, treatment efforts still unsatisfactory

Plasmodium malariae
Hendrickse, R. G., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 268-273
aspects of tropical pediatrics, importance of malaria, review

Plasmodium malariae
Houba, V.; et al., 1976, Clin. Immunol. and Immunopathol., v. 6 (1), 1-12
Plasmodium brasilianum, P. malariae, P. falciparum, humans, Aotus monkeys, increased binding of malarial antibodies with sera, faster disappearance of this antibody from circulation, and its increased deposition in renal glomeruli, serum complement levels during progress of infection, important role of P. malariae in pathogenesis of chronic progressive nephropathies occurring in malarial areas

Plasmodium malariae
Houba, V.; Faulk, W. P.; and Matola, Y. G., 1974, Clin. and Exper. Immunol., v. 18 (1), 89-93
heterophilic antibodies to sheep erythrocytes identified in sera from both malarious and non-malarious subjects in Tanzania, in patients with sickle-cell anemia but not exposed to malaria (U.S.A.), and in rhesus monkeys infected with Plasmodium knowlesi and P. cynomolgi bastianellii, observations do not support concept that heterophilic antibodies are integrally associated with host resistance to malaria

Plasmodium malariae
Hutt, M. S. R.; and Voller, A., 1976, J. Trop. Med. and Hyg., v. 79 (5), 106-110
histopathology of hepatic changes in exper. Aotus monkeys and possible role of liver in immunologic response to malaria

Plasmodium malariae
van der Kaay, H. J.; and Darwish, R. O., 1975, Riv. Parassitol., Roma, v. 36 (1), 1-6
Plasmodium vivax, P. falciparum, P. malariae, spleen and blood examinations, vector survey, feasibility study for possible control measures: Bokondini, Irian Jaya, Indonesia

Plasmodium malariae
Kean, B. H.; and Reilly, P. C., jr., 1976, Am. J. Med., v. 61 (2), 159-164
recommendations for diagnosis, prophylaxis and treatment of human malaria with epidemiologic review of recently treated infections in persons who had travelled or lived in endemic areas: New York City

Plasmodium malariae
Kibukamusoke, J. W., 1970, Mater. Med. Pol. (4), v. 2 (3-4), 44-48
Plasmodium malariae, P. falciparum, causal relationship between human malaria and nephrotic syndrome in tropical areas, pathology, therapy

Plasmodium malariae
Laing, A. B. G., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 133-138
Plasmodium falciparum, hospital trials with Gambian children testing BRL 50216 alone or in combinations with sulphadimethoxine or sulphafurazole (acting as potentiators); some response also with P. malariae; sporontocidal effect only when BRL 50216 given in presence of potentiators

Plasmodium malariae
Laing, A. B. G.; Gooi, H. C.; and Gelber, R. H., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 165-166 [Letter]
Plasmodium malariae in man being treated for leprosy with dapsone, apparent quartan malarial dapsone-resistant strain

Plasmodium malariae, illus.
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium malariae
Leeflang, P.; et al., 1976, Internat. J. Parasitol., v. 6 (2), 159-161
survey of 173 males using indirect fluorescent antibody technique with Babesia bovis, B. bigemina, and B. ratti as antigens, babesial antibodies detected in 54%, no correlation found with mode of living, contact with livestock, incidence of malaria parasitaemias or malarial antibodies, no Babesia organisms found in blood smears or by animal subinoculation, source of antibodies unknown but subclinical infection not likely: Nigeria

Plasmodium malariae
Lopez, C. E.; and Schultz, M. G., 1977, J. Infect. Dis., v. 135 (5), 875-876 [Letter]
incidence of human transfusion malaria and standards for blood donor selection

Plasmodium malariae
Mannweiler, E.; et al., 1976, Munchen. Med. Wchnschr., v. 118 (36), 1139-1144
Plasmodium spp., homologous and heterologous malaria pathogens used as antigens to determine malaria antibodies by direct immunofluorescence, pre- and post-treatment with chloroquine compared

Plasmodium malariae
Mathews, H. M.; and Dilworth, D. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (2), 351-352
Plasmodium brasilianum, use as antigen in indirect hemagglutination test for diagnosing human P. falciparum, P. vivax, P. ovale, P. malariae

Plasmodium malariae
Mathews, H. M.; Lobel, H. O.; and Breman, J. G., 1976, Am. J. Trop. Med. and Hyg., v. 25 (2), 217-220
Plasmodium falciparum, P. malariae, infants and young children, prevalence of malaria antibody evaluated using indirect hemagglutination test with P. falciparum antigen and filter paper blood specimens, slight decline in 6- to 8-month-old children with no demonstrable parasitemia but those older than 10 months had similar antibody levels regardless of presence or absence of demonstrated parasites in blood smears: Ivory Coast

Plasmodium malariae
Maulitz, R. M.; Marr, J. S.; and Shookhoff, H. B., 1976, Am. J. Trop. Med. and Hyg., v. 25 (1), 1-4
Plasmodium malariae, human, 2 cases of transfusion-induced malaria, donors were Greek immigrants with latent infection who responded to special blood drives as result of 1974 Turkish-Greek war on Cyprus, need for alertness when natural or man-made disasters result in demographically unusual and large drives to collect blood: New York City

Plasmodium malariae
Miller, L. H., 1974, Transmis. Disease and Blood Transfus., 241-266
human malarias, control measures established to prevent transmission via infected blood donors; review of life cycles and clinical aspects of malaria infections

Plasmodium malariae
Miller, L. H., 1976, J. Infect. Dis., v. 133 (6), 727 [Letter]
Plasmodium malariae, human transfusion malaria, rules for donor selection should be reevaluated every few years after evaluation of epidemiologic and public health surveillance data, including that pertaining to immigrant blood donors: United States

Plasmodium malariae
Miller, L. H., 1977, J. Infect. Dis., v. 135 (5), 855-864
human malarias, current prospects and problems in developing a vaccine for malaria immunization

Plasmodium malariae
Moore, D. L.; Heyworth, B.; and Brown, J., 1974, Clin. and Exper. Immunol., v. 17 (4), 647-656
phytohaemagglutinin-induced lymphocyte transformation in leucocyte cultures from malarious, malnourished, and control Gambian children, some depression of response to low doses of PHA in children with malaria

Plasmodium malariae
Morel-Maroger, L.; et al., 1975, Medecine et Malad. Infect., v. 5 (12), special no., 609-616
presence or absence of anti-Plasmodium falciparum and/or anti-P. malariae antibodies in patients with different types of glomerulo-nephritis: Senegal; Cameroun

Plasmodium malariae
Nakabayashi, T.; et al., 1973, Nettai Igaku (Trop. Med.), v. 15 (3), 154-168
Plasmodium spp. in humans, epidemiologic survey to determine prevalence rate in rural areas of the Philippines

P[lasmodium] malariae
Nakabayashi, T.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (11), 247-251
human malaria, statistical survey of cases imported into Japan from 1972-1974

P[lasmodium] malariae
Neri, I.; and Cavallini, C., 1972, Parassitologia, v. 14 (2-3), 325-327
P[lasmodium] malariae, transmitted by blood transfusion; advice to choose only blood donors born after malaria eradication, 1952: Sardinia

Plasmodium malariae
Ohlrogge, R.; and Schlipkoeter, H. W., 1973, Med. Microbiol. and Immunol., v. 158 (4), 267-274
deviations in erythrogram before and after treatment of human malaria infections with resochin, no such changes in hookworm and other worm infections: Liberia; Togo

Plasmodium malariae
Otieno, L. H.; et al., 1971, Trop. and Geogr. Med., v. 23 (4), 369-375
human Plasmodium spp., dynamics of epidemiologic survey in highly endemic malarious area, data show decreasing incidence in recent years: north-eastern Tanzania

Plasmodium malariae
Papua N. Guinea Med. J., v. 17 (1), 1974, 1-115
multiple articles focusing on human malaria in Papua New Guinea (epidemiology, public health, control programs, vectors, diagnosis, toxicity of antimalarials, statistics of surveys)

Plasmodium malariae
Payne, D.; et al., 1976, Bull. World Health Organ., v. 54 (4), 369-377
impact of control measures on malaria transmission and general mortality, extensive survey, results show relationship between efficacy of antivectorial measures (fenithrothion spraying) and decrease in general mortality and infant mortality in Kenya

Plasmodium malariae
Pull, J. H.; and Gramiccia, G., 1976, WHO Chron., v. 30 (7), 286-289
review of 6 years of research activity on malaria control carried out in North Nigeria and proposals for the future

Plasmodium malariae
Radford, A. J.; Van Leeuwen, H.; and Christian, S. H., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 11-23
changes in distribution and determinants of malaria that occurred with the development process in the highlands of Papua New Guinea between 1930 and 1970

Plasmodium malariae
Ringelhann, B.; et al., 1976, Am. J. Hum. Genet., v. 28 (3), 270-279
possible relationships between hemoglobin types and human malarial infection rate, parasite species, parasite density, host age and sex; correlations with transplacental and passive immunity

Plasmodium malariae
Robinson, M. J.; and Eykyn, S., 1977, Brit. Med. J. (6103), v. 2, 1667 [Letter]
childhood malaria, increasing incidence with increased travel to and from endemic areas, diagnostic problem in children: United Kingdom

Plasmodium malariae
Roy, R. G.; Chakrapani, K. P.; and Zacharias, J., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 447 [Letter]
statistics indicate eradication in Kerala and Tamilnadu States, India

Plasmodium malariae
Shaw, P. K.; Brodsky, R. E.; and Schultz, M. G., 1976, J. Infect. Dis., v. 133 (1), 95-101
malaria surveillance, statistics for 1974, increasing incidence with increasing international travel: United States

Plasmodium malariae
Soothill, J. F.; Smith, M. D.; and Morgan, A. G., 1975, Symposia Brit. Soc. Parasitol., v. 13, 59-68
association of parasites with nephrotic syndrome, genetically and environmentally determined host variation may be the immunodeficiency underlying proneness to chronic soluble complex disease, extensive review with emphasis on Schistosoma spp., Plasmodium malariae, and some preliminary experiments with Trypanosoma brucei in mice

Plasmodium malariae
Sulzer, A. J.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 55-58
malaria, human, diagnosis, indirect fluorescent antibody test using multi-species thick smear antigen containing equal proportions of Plasmodium vivax, P. falciparum, and P. brasilianum (latter serologically equivalent in IFA test to P. malariae), circumvents need for multiple testing with several antigen species

Plasmodium malariae
Sulzer, A. J.; and Latorre, C. R., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 553 [Letter]
primate malarias, method for in vitro production of schizont antigens for use in indirect immunofluorescence tests for malarial antibodies

Plasmodium malariae
Tapper, M. L.; and Armstrong, D., 1976, Arch. Int. Med., Chicago, v. 136 (7), 807-810
blood transfusion induced malarial infections in 2 patients with neoplastic disease, case reports, implications for alterations of immunologic status

Plasmodium malariae
Thomas, V.; and Dissanaike, A. S., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 602-606
Plasmodium spp., detection of malaria endemicity among Orang Asli aborigines using Plasmodium falciparum and P. brasilianum antigens and indirect fluorescent antibody test (IFA), age dependent increase in number of positive results, IFA valuable as adjunct to blood slide examination especially when parasites are at very low levels: Malaysia

Plasmodium malariae
Truong-Minh-Ky, D.; et al., 1976, Medecine Afrique Noire, v. 23 (4), 215-223
Plasmodium malariae, Plasmodium falciparum, survey of village infants for presence of malarial antibodies: Abidjan

Plasmodium malariae
Tsukamoto, M.; et al., 1975, Nettai Igaku (Trop. Med.), v. 17 (1), 13-26
Plasmodium spp. in humans, field trials of biochemical analysis of blood and urine, value comparisons before and after antimalarial therapy: Philippines

Plasmodium malariae
Voller, A., 1975, Symposia Brit. Soc. Parasitol., v. 13, 69-84
immunopathology of malaria, extensive review: immunosuppression and auto-immunity; tropical splenomegaly syndrome; nephrotic syndrome

Plasmodium malariae
Welch, S. G.; McGregor, I. A.; and Williams, K., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 295-296
survey of 2 rural Gambian villages showed total absence of Duffy blood group antigens and absence of Plasmodium vivax infection although P. falciparum, P. malariae and P. ovale were present; findings consistant with theory that absence of Duffy phenotypes constitutes basis of innate resistance to P. vivax

Plasmodium matutinum
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
Erithacus rubecula (blood): Great Britain

Plasmodium merulae
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Turdus merula (blood): Berkshire, England

Plasmodium mexicanum, illus.
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Sceloporus occidentalis
S. graciosus (exper)
S. jarrovi (exper.)
S. magister (exper.)
Gerrhonotus multicarinatus (nat. and exper.)
(blood of all): all from central California

Plasmodium mexicanum
Moore, J.; and Sinden, R. E., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 13-14 [Demonstration]
Plasmodium mexicanum, fine structure

Plasmodium minasense
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis capito
A. limifrons
A. frenatus
all from Panama

Plasmodium morulum
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Mabuya mabouya: Panama

Plasmodium nucleophilum
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Sylvietta brachyura
Terpsiphone viridis
Ploceus galbula
P. intermedius
P. cucullatus
all from Ethiopia

Plasmodium nucleophilum
Manwell, R. D., 1974, J. Protozool., v. 21 (3), 423 [Abstract]
starlings (blood)

Plasmodium nucleophilum
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Sturnus contra
S. pagadorum
Cosmopsarus regius
Lamprotornis chalybaeus
Spreo superbus
Euplectes hordeacea
Sporopipes squamifrons
Sturnus javanicus
(blood of all): all imported to U. S., origin unknown

Plasmodium nucleophilum toucani [subsp. n.], illus.
Manwell, R. D.; and Sessler, G. J., 1971, J. Protozool., v. 18 (4), 570-574 [For complete author reference, see Suppl. 20, Part 1]
[lapsus p. 570 as P. nucleophilum tucani]
Ramphastos swainsonii (blood)
Serinus canarius (exper.)
Pteroglossus torquatus (exper.)

Plasmodium nucleophilum toucani
Manwell, R. D., 1974, J. Protozool., v. 21 (3), 423 [Abstract]
Ramphastos s. swainsoni (blood)

Plasmodium nucleophilum toucani Manwell & Sessler 1971
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Syn.: Plasmodium huffi Muniz, Soares & Battista pro parte
Andegina laminorostris
Ramphastos s. swainsonii
(blood of all): all imported to U. S., origin unknown

Plasmodium nucleophilum tucani [lapsus p. 570 for P. nucleophilum toucani [subsp. n.]]
Manwell, R. D.; and Sessler, G. J., 1971, J. Protozool., v. 18 (4), 570-574 [For complete author reference, see Suppl. 20, Part 1]

Plasmodium octamerium
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Estrilda paludicola
E. astrild
Euplectes macrourus
all from Ethiopia

Plasmodium octamerium
Manwell, R. D., 1974, J. Protozool., v. 21 (3), 423 [Abstract]
Vidua macroura (blood): Africa

Plasmodium octamerium Manwell, 1968
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Lamprotornis chalybaeus
Euplectes orix orix
Steganura/Vidua macroura
(blood of all): all imported to U. S., origin unknown

Plasmodium ovale
Bell, D. R.; and Howells, R. E., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 1-14
malaria, blood film and spleen examination results in relation to age, chloroquine and cycloguanil sensitivity of Plasmodium falciparum; Schistosoma haematobium, prevalence and intensity in relation to age and sex: Malumfashi District, North Central State of Nigeria

Plasmodium ovale
Brown, P.; et al., 1976, Am. J. Trop. Med. and Hyg., v. 25 (6), 775-783
Plasmodium, 4 human spp., remote island populations, fluorescent antibody prevalence patterns, comparison with published parasite survey data based on blood smears and medical examinations, evaluation of epidemiological usefulness: New Hebrides; Solomons; Western Carolines (malaria entirely absent); New Guinea

Plasmodium ovale
Butler, T.; Warren, K. S.; and Mahmoud, A. A., 1976, J. Infect. Dis., v. 133 (6), 721-726
human malarias, algorithms in the diagnosis and management

Plasmodium ovale
Conradie, M.; and Jacobs, P., 1976, Brit. Med. J. (6028), v. 2, 181
suggested technique for identification of scanty numbers of malarial parasites in human blood

P[lasmodium] ovale
zum Felde, I.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (4), 477-484
P[lasmodium] falciparum, P. vivax, P. ovale, P. malariae, human, indirect immunoflourescence test with same 4 Plasmodium spp. plus P. fieldi as antigen, species-specific and cross-reactions, effect of therapy, implications for immunodiagnosis

Plasmodium ovale
Gibson, J. J., jr.; Brodsky, R. E.; and Schultz, M. G., 1974, J. Infect. Dis., v. 130 (5), 553-555
increased incidence of malaria in the United States probably reflects increased foreign travel and improved reporting of infections

Plasmodium ovale
Gratecos, N., 1972, Marseille Med., v. 109 (5), 361-369
Plasmodium spp., Leishmania donovani, human splenomegaly, differential diagnosis, clinical review

Plasmodium ovale
Gueye, I.; and Michel, R., 1973, Medecine Afrique Noire, v. 20 (10), 753-766
guidelines for mass prophylactic therapy against human forms of malaria

Plasmodium ovale
Gundelfinger, B. F.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 393-396
statistics of survey for prevalence of human Plasmodium spp. and response of Plasmodium falciparum and P. vivax to standard dose of chloroquine; first record of P. ovale reported in Timor, Indonesia

Plasmodium ovale
van der Kaay, H. J.; and Zuidema, P. J., 1976, Nederl. Tijdschr. Geneesk., v. 120 (20), 884-886
human Plasmodium spp., recommendations for malaria prophylaxis during travel to endemic areas

Plasmodium ovale
Kean, B. H.; and Reilly, P. C., jr., 1976, Am. J. Med., v. 61 (2), 159-164
recommendations for diagnosis, prophylaxis and treatment of human malaria with epidemiologic review of recently treated infections in persons who had travelled or lived in endemic areas: New York City

Plasmodium ovale
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium ovale
Mannweiler, E.; et al., 1976, Munchen. Med. Wchnschr., v. 118 (36), 1139-1144
Plasmodium spp., homologous and heterologous malaria pathogens used as antigens to determine malaria antibodies by direct immunofluorescence, pre- and post-treatment with chloroquine compared

Plasmodium ovale
Mathews, H. M.; and Dilworth, D. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (2), 351-352
Plasmodium brasilianum, use as antigen in indirect hemagglutination test for diagnosing human P. falciparum, P. vivax, P. ovale, P. malariae

Plasmodium ovale
Miller, L. H., 1974, Transmis. Disease and Blood Transfus., 241-266
human malarias, control measures established to prevent transmission via infected blood donors; review of life cycles and clinical aspects of malaria infections

Plasmodium ovale
Miller, L. H., 1977, J. Infect. Dis., v. 135 (5), 855-864
human malarias, current prospects and problems in developing a vaccine for malaria immunization

P[lasmodium] ovale
Nakabayashi, T.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (11), 247-251
human malaria, statistical survey of cases imported into Japan from 1972-1974

Plasmodium ovale
Otieno, L. H.; et al., 1971, Trop. and Geogr. Med., v. 23 (4), 369-375
human Plasmodium spp., dynamics of epidemiologic survey in highly endemic malarious area, data show decreasing incidence in recent years: north-eastern Tanzania

Plasmodium ovale, illus.
Sawyer, D. M.; and Wadsworth, L. D., 1977, Canad. Med. Ass. J., v. 117 (8), 923
Plasmodium ovale in woman who had received recent blood transfusion, donor of blood had travelled earlier to Africa in the company of another person who had had 3 clinical attacks of malaria, clinical case report, emphasis on diagnostic awareness: Canada

Plasmodium ovale
Shaw, P. K.; Brodsky, R. E.; and Schultz, M. G., 1976, J. Infect. Dis., v. 133 (1), 95-101
malaria surveillance, statistics for 1974, increasing incidence with increasing international travel: United States

P[lasmodium] ovale
Topley, E.; Knight, R.; and Woodruff, A. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 51-54
malaria patients, direct antiglobulin test and immunoconglutinin titres, possible significance of results in understanding mechanism of anemia

Plasmodium ovale
Welch, S. G.; McGregor, I. A.; and Williams, K., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 295-296
survey of 2 rural Gambian villages showed total absence of Duffy blood group antigens and absence of Plasmodium vivax infection although P. falciparum, P. malariae and P. ovale were present; findings consistant with theory that absence of Duffy phenotypes constitutes basis of innate resistance to P. vivax

Plasmodium paranucleophilum
Manwell, R. D., 1974, J. Protozool., v. 21 (3), 423 [Abstract]
Tachyphonus sp. (blood)

Plasmodium paranucleophilum Manwell & Sessler, 1971
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Tachyphonus sp. (blood): imported to U.S., origin unknown

Plasmodium (Giovannolaia) pedioecetii Shillinger 1942, emend. Stabler et al., 1973, illus.
Stabler, R. M.; and Kitzmiller, N. J., 1976, J. Parasitol., v. 62 (4), 539-544
Plasmodium pedioecetii, redescription, periodicity, no synchronicity of parasite development
Alectoris chukar (exper.)
Colinus virginianus (nat. and exper.)
Coturnix coturnix (exper.)
Serinus canaria (exper.)
Dendragapus obscurus
Pedioecetes phasianellus
Oreortyx pictus
Lophortyx gambelii
Perdix perdix
(blood of all): all from Colorado

Plasmodium pinottii Muniz and Soares 1953
Manwell, R. D.; and Allen, C. S., 1976, J. Parasitol., v. 62 (1), 110
Plasmodium pinottii, internal hemorrhages as possible important factor in death of infected Columba livia and Streptopelia risoria despite reduced blood clotting time in pigeons

Plasmodium pitheci
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium polare Manwell
Meyer, C. L.; and Bennett, G. F., 1976, Canad. J. Zool., v. 54 (2), 133-141
Plasmodium circumflexum, P. polare, sporogony
Mansonia perturbans (exper.)
Culiseta morsitans (exper.)
Anas platyrhynchos (exper.)
Anser cygnoides (exper.)
Anas discors (blood): New Brunswick

Plasmodium polare
Stabler, R. M.; and Kitzmiller, N. J., 1976, J. Parasitol., v. 62 (4), 539-544
Petrochelidon pyrrhonata (blood): Colorado

Plasmodium praecox
Djudovic, P.; et al., 1976, Pharmazie, v. 31 (12), 845-849
quinoline-3-, 6-methoxyquinoline-3-, and quinoxaline-2-derivatives, synthesis, activity against drug-sensitive and resistant strains of Plasmodium spp. in white mice and canaries, comparison with known antimalarials

Plasmodium reichenowi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium relictum
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Motacilla flava
Anthus similis
Pycnonotus barbatus
Nilaus afer
Dryoscopus gambensis
Laniarius aethiopicus
Malaconotus sulfureopectus
Turdus pelios
Turdoides leucopygius
Locustella luscinioides
Sylvia communis
Cisticola brachyptera
Prinia subflava
Serinus nigriceps
Ploceus galbula
P. cucullatus
Quelea erythrops
Q. quelea
Passer griseus
P. eminibey
Lamprotornis chalybaeus
all from Ethiopia

Plasmodium relictum
Ball, G. H.; and Chao, J., 1976, Exper. Parasitol., v. 39 (1), 115-118
Plasmodium relictum, utilization of amino acids by developing oocysts in presence of metabolizing insect cells

Plasmodium relictum
Bennett, G. F.; and Borrero H., J. I., 1976, J. Wildlife Dis., v. 12 (3), 454-458
Schiffornis turdinus (blood): Colombia

Plasmodium relictum
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Lanius collaris: Kenya
Cyanomitra olivacea: Tanzania
Guttera pucherani: Tanzania
Lamprotornis chalybaeus: Kenya

Plasmodium (Haemamoeba) relictum (Grassi & Feletti, 1891)
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Picoides m. mahrattensis: Godavari River Basin, Maharashtra

Plasmodium relictum Grassi et Feletti, 1891
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Columba livia]
[Merops apiaster]
[Hirundo rustica]
[Motacilla alba]
[Lanius minor]
[Lanius cristatus]
[Emberiza melanocephala]
[Emberiza hortulana]
[Chloris chloris]
[Passer domesticus]
all from north-east Azerbaidzhan

Plasmodium relictum-cathemerium
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Anas c. crecca
Lophura swinhoei
Capella gallinago
Streptopelia chinensis formosae
Ketupa flavipes
Apus affinis kuntzi
Hirundo daurica formosae
Pomatorhinus ruficollis musicus
Turdus c. chrysolaus
T. pallidus
Hypothymis azurea obholseri
Muscicapa rufilata
Anthus hodgsoni yunnanensis
Motacilla alba leucopsis
Lanius cristatus lucionensis
L. schach formosae
(blood of all): all from Taiwan

Plasmodium relictum (Grassi & Feletti)
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Liothrix luteus (blood): imported to U.S., origin unknown

Plasmodium relictum
Manwell, R. D.; and Sessler, G. J., 1973, J. Protozool., v. 20 (3), 363-364
Liothrix luteus (blood): imported from Southeast Asia

Plasmodium (Haemamoeba) relictum
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Luscinia luscinia (blood): Kenya
Ploceus cucullatus (blood): Kariobangi, Nairobi

Plasmodium rhadinurum
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Iguana iguana: Panama

Plasmodium roussetti
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium rouxi
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Pycnonotus barbatus
Dryoscopus gambensis
Tchagra senegala
Cossypha semirufa
Turdus pelios
Cisticola galactotes
Prinia subflava
Batis orientalis
Serinus mozambicus
Ploceus baglafecht
P. velatus
Euplectes ardens
E. macrourus
E. franciscanus
all from Ethiopia

Plasmodium rouxi
Bennett, G. F.; and Blancou, J., 1974, J. Wildlife Dis., v. 10 (3), 239-240
Margaroperdix madagascarensis (blood): Madagascar

Plasmodium rouxi
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Cyanomitra olivacea: Tanzania
Hyphanturgis ocularis: Tanzania
Pycnonotus barbatus: Kenya
P. xanthopygus: Tanzania
Quelea erythrops: "

Plasmodium rouxi
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Apus affinis kuntzi
Hypsipetes madagascariensis nigerrimus
Spizixos semitorques cinereicapillus
Turdus c. chrysolaus
(blood of all): all from Taiwan

Plasmodium rouxi Sergent, Sergent & Catani
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Ramphastos s. sulfuratus (blood): imported to U.S., origin unknown

Plasmodium sandoshami, illus.
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium scelopori sp. nov., illus.
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears, for detection: Middle America
Sceloporus teapensis: Blancaneaux Lodge, Belize
S. variabilis: Guanacaste Province, Costa Rica; Honduras

Plasmodium shortti (Jayawardene, 1963)
Nelson, P., 1971, Tr. Roy. Soc. Trop. Med. and Hyg., v. 65 (5), 695-696 [Letter]
Anopheles elegans established as vector of simian malaria in Ceylon
rhesus monkey (exper.)

Plasmodium simiovale, illus.
Aikawa, M.; Hsieh, C.-L.; and Miller, L. H., 1977, J. Parasitol., v. 63 (1), 152-154
Plasmodium simiovale- and P. fieldi-infected Macaca mulatta, ultrastructural alterations in cell membrane of erythrocytes, caveola-vesicle complexes likely correspond to Schüffner's dots, electron dense excrescences at protruded tip of infected erythrocytes

Plasmodium simiovale
Chen, D. H.; Nussenzweig, R. S.; and Collins, W. E., 1976, J. Parasitol., v. 62 (4), 636-637
rat antisera against sporozoites of 6 primate Plasmodium spp. reacted in circum-sporozoite precipitation tests only with sporozoites of homologous species, geographically different strains of same species cross-reacted intensely however

Plasmodium simium
Deane, L. de M., 1976, Scient. Publication (317). Pan Am. Health Organ., 144-163
Plasmodium brasilianum, P. simium, monkeys, epidemiology: Brazil

Plasmodium simium
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium simium
Rossan, R. N.; Baerg, D. C.; and Young, M. D., 1975, J. Parasitol., v. 61 (4), 768-769
Aotus
Saimiri sciureus
Saguinus geoffroyi
Alouatta villosa
Cebus capucinus
Ateles fusciceps
Ateles geoffroyi
(all exper.)

Plasmodium simium, illus.
Seed, T. M.; et al., 1976, Exper. Parasitol., v. 39 (2), 262-276
Plasmodium simium, ultrastructure of erythrocytic trophozoites, schizonts, merozoites, and gametocytes; pathological changes in host cell structure

Plasmodium sylvaticum
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium tenue
Manwell, R. D., 1974, J. Protozool., v. 21 (3), 423 [Abstract]
Liothrix luteus (blood)

Plasmodium tenue
Manwell, R. D.; Allen, C. S.; and Kuntz, R. E., 1976, J. Protozool., v. 23 (4), 571-576
Garrulax canorus taewanus (blood): Taiwan

Plasmodium tenue Laveran & Marullaz
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Liothrix luteus (blood): imported to U.S., origin unknown

Plasmodium (Novyella) tenue
Manwell, R. D.; and Sessler, G. J., 1973, J. Protozool., v. 20 (3), 363-364
Liothrix luteus (blood): imported from Southeast Asia

Plasmodium traguli, illus. (exper.)
Hoo, C. C.; and Sandosham, A. A., 1968, Med. J. Malaya, v. 22 (4), 299-301
morphology of early forms of Hepatocystis fieldi and exper. Plasmodium traguli in Tragulus javanicus

Plasmodium traguli
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium tropiduri
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis humilis: Costa Rica
A. limifrons: Costa Rica; Panama
A. lemurinus: Costa Rica
A. lionotus: Costa Rica; Panama
A. biporcatus: Costa Rica; Panama
Anolis sp. indet.: Costa Rica
A. poecilopus: Panama
A. frenatus: Panama
A. pentaprion: Panama
Corytophanes cristatus: Panama

Plasmodium vaughani
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Turtur chalcospilos
Merops albicollis
Tchagra minuta
T. senegala
Laniarius aethiopicus
Turdus pelios
Acrocephalus gracilirostris
Cisticola natalensis
Camaroptera brevicaudata
Terpsiphone viridis
Nectarinia cuprea
Estrilda astrild
Lagonosticta senegala
Amblyospiza albifrons
Ploceus cucullatus
P. superciliosus
P. ocularis
Euplectes macrourus
Passer griseus
P. eminibey
Petronia dentata
all from Ethiopia

Plasmodium vaughani
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Buccanodon leucotis: Tanzania
Lanius collurio: Tanzania
Bradornis pumilus: Tanzania
Musophago violacea: Zaire
Anthropetes collaris: Tanzania
Cyanomitra olivacea: Tanzania
Coliuspasser hartlaubi: Zaire
Othypantes stuhlmanni: Tanzania

Plasmodium vaughani
Crewe, S. M., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 477-478
Aedomyia as possible vector of avian malaria
Streptopelia senegalensis: Nigeria

Plasmodium vaughani Novy & MacNeal
Manwell, R. D.; and Rossi, G. S., 1975, J. Protozool., v. 22 (1), 124-127
Turdus fulviventris (blood): imported to U.S., origin unknown

Plasmodium (Novyella) vaughani Novy and MacNeal, 1904
Nandi, N. C., 1976, Indian J. Animal Health, v. 15 (2), 137-138
Acridotheres t. tristis: Godavari River Basin, Maharashtra

Plasmodium (Novyella) vaughani Novy & MacNeal, 1904
Nandi, N. C.; and Mandal, A. K., 1977, Indian J. Animal Sc., v. 47 (9), 558-561
Ploceus philippinus: Orissa, India

Plasmodium vaughani Novy et MacNeal, 1904
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Parus major]
[Passer domesticus]
[Sturnus vulgaris]
[Garrulus glandarius]
all from north-east Azerbaidzhan

Plasmodium (Novyella) vaughani merulae n. subsp., illus.
Corradetti, A.; and Scanga, M., 1972, Parassitologia, v. 14 (1), 85-93
description of pre-erythrocytic cycle
Turdus merula (nat. and exper.) (sangue):
Italia (Lombardia, Emilia, Toscana, Lazio, Marche)
Culex pipiens (exper.)

Plasmodium vinckei
Buengener, W., 1974, Tropenmed. u. Parasitol., v. 25 (3), 309-312
Plasmodium berghei, P. vinckei, P. chabaudi, mice, rats, allopurinol stimulated multiplication of plasmodia and infections caused death more rapidly or in more animals, this effect could result from hypoxanthine concentration being limiting factor for parasite multiplication, in these parasite-host combinations host-purine-dependent parasites would have a selective advantage over non-dependent ones

Plasmodium vinckei
Buengener, W., 1974, Tropenmed. u. Parasitol., v. 25 (4), 464-468
Plasmodium vinckei, rats, allopurinol treatment, influence on length of asexual cycle and multiplication rate of parasites

Plasmodium vinckei
Buengener, W., 1976, Tropenmed. u. Parasitol., v. 27 (1), 79-81
influence of xanthine oxidase on duration of asexual cycle of Plasmodium vinckei in the mouse

Plasmodium vinckei like
Carter, R.; and Walliker, D., 1974, J. Protozool., v. 21 (3), 449 [Abstract]
Thamnomys rutilans (blood): Central African Republic

Plasmodium vinckei
Chance, M. L.; and Warhurst, D. C., 1973, J. Protozool., v. 20 (4), 524
Plasmodium berghei berghei, P. berghei yoelii, P. chabaudi, P. vinckei, DNA relationships in the subgenus Vinckeia

Plasmodium vinckei
Clark, I. A.; Allison, A. C.; and Cox, F. E., 1976, Nature, London (5541), v. 259, 309-311
Babesia, Plasmodium, protection of mice by previous infection with BCG, possible mechanism

Plasmodium vinckei, illus.
Clark, I. A.; Cox, F. E. G.; and Allison, A. C., 1977, Parasitology, v. 74 (1), 9-18
mice pre-treated with killed Corynebacterium parvum, completely resistant to infection with Babesia microti or B. rodhaini, protected from death caused by Plasmodium vinckei or P. chabaudi infection, no antibody detected, suggested that non-specific soluble mediator may play important role in protection observed

Plasmodium vinckei
Cottrell, B.J.; Playfair, J.H.L.; and de Sousa, B., 1977, Exper. Parasitol., v. 43 (1), 45-53
Plasmodium yoelii, Plasmodium vinckei, mice, effects of nonspecific immunostimulation with a variety of agents

Plasmodium vinckei
Fitch, C. D.; Chevli, R.; and Gonzalez, Y., 1975, J. Pharmacol. and Exper. Therap., v. 195 (3), 389-396
Plasmodium berghei and P. vinckei chloroquine-resistant and -susceptible strains, P. berghei yoelii spontaneously resistant to chloroquine, variations of substrate-stimulated chloroquine accumulation

Plasmodium vinckei
Heischkeil, R., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (4), 505-516
Plasmodium vinckei, chemotherapeutic activity of selected antagonists of folic acid metabolism with special reference to structure-activity relationships, comparison with some standard antimalarials; some compounds also exert noxious effect on host

Plasmodium vinckei
Heischkeil, R., 1974, Tropenmed. u. Parasitol., v. 25 (1), 105-115
Plasmodium vinckei, mice, purine and pyrimidine derivatives, antimetabolic activity against structurally analogous vitamins of the B complex and precursors of nucleic acids, plasmodiostatic activity compared to some standard antimalarials

Plasmodium vinckei
Jung, A.; et al., 1975, Tropenmed. u. Parasitol., v. 26 (1), 27-34
Plasmodium vinckei, DNA, RNA, and lipid synthesis, specific activity of glucose-6-phosphate dehydrogenase and glucose-6-phosphatase in parasite developmental stages

Plasmodium vinckei et ssp.
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium vinckei
Nickel, P.; et al., 1977, Arch. Pharm., v. 310 (2), 81-89
Plasmodium vinckei, mice, 2-, 3- and 4-chloro derivatives of 6-(4-diethylamino-1-methylbutylamino)-5,8-dimethoxyquinoline, only slight activity

Plasmodium vinckei
Nickel, P.; Zimmermann, R.; and Fink, E., 1977, Arch. Pharm., v. 310 (7), 529-536
Plasmodium vinckei in mice (exper.), synthesis and characterization of 2- and 4-hydroxymethyl derivatives of 6-(4-diethylamino-1-methylbutylamino)-5,8-dimethoxy-quinoline, hydroxylation reduces toxicity without influencing antimalarial activity

Plasmodium vinckei
Picard-Maureau, A.; et al., 1975, Tropenmed. u. Parasitol., v. 26 (4), 405-416
Plasmodium vinckei-infected mouse erythrocytes, protein content, glutathione concentration, enzyme activity, correlation with stage of parasite development, significance of results for metabolism of malaria parasites and for possible adaptation to mosquito

Plasmodium vinckei
Playfair, J. H. L.; de Souza, J. B.; and Cottrell, B. J., 1977, Immunology, v. 33 (4), 507-515
Plasmodium yoelii, P. vinckei, P. berghei, mice, regime of killed homologous vaccine plus Bordetella pertussis adjuvant, differences between species in effectiveness of protection, some cross-protection but largely species-specific, passive transfer of immunity to P. yoelii by serum or spleen cells

Plasmodium vinckei
Playfair, J. H. L.; de Souza, J. B.; and Cottrell, B. J., 1977, Immunology, v. 32 (5), 681-687
Plasmodium spp., Babesia microti, mouse helper T-cell response to parasites measured by using them as carriers for a standard hapten, results show extensive cross-reaction between the 4 parasites as carriers apparently unrelated to known serological cross-reactions but are against the idea that helper T-cells are exclusively responsible for resistance

Plasmodium vinckei
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Plasmodium vinckei
Seitz, H. M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 88-90
Plasmodium vinckei, P. berghei, counter-current immunoelectrophoresis used to detect malarial antigens and antibodies in rat and mouse sera, more sensitive than double diffusion methods

Plasmodium vinckei
Zuckerman, A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 793-829
rodent malaria, immunology, review

Plasmodium vinckei brucechwatti subsp. n., illus.
Killick-Kendrick, R., 1975, Ann. Parasitol., v. 50 (3), 251-264
morphology, development, sporogony obtained in Anopheles spp. but sporozoites in salivary glands never infective
Syn.: P. vinckei chwatti Garnham, 1973
Thamnomys rutilans (blood): edge of Ilaro Forest Reserve near the village of Ilobi, Western State, Republic of Nigeria, Western Nigeria
white mice (blood) (exper.)
Anopheles stephensi (exper.)
A. quadrimaculatus (exper.)
A. labranchiae atroparvus (exper.)

Plasmodium vinckei chabaudi
Coombs, G. H.; and Gutteridge, W. E., 1975, J. Protozool., v. 22 (4), 555-560
Plasmodium vinckei chabaudi rat-adapted strain, in vitro system (based on rocker dilution technic) that supports intraerythrocytic growth from ring to schizont stages, some reinvasion obtained but associated with decrease in parasite numbers, lactate production, glucose utilization, ^{3}H-leucine and ^{3}H-adenosine incorporation

Plasmodium vinckei chabaudi Landau, 1965
Killick-Kendrick, R., 1971, Symposia Brit. Soc. Parasitol., v. 9, 39-64
murine malaria parasites: collection of strains from rodents; collection of strains from mosquitoes; establishment of strains in the laboratory; maintenance by cyclical transmission, extensive review

Plasmodium v[inckei] chabaudi
McDonald, V.; and Phillips, R. S., 1975, J. Protozool., v. 22 (3), 54A [Abstract]
Plasmodium v[inckei] chabaudi, mice, adoptive transfer of whole spleen cell population vs. populations enriched with either T- or B-lymphocytes

Plasmodium vinckei chabaudi
Shungu, D. M.; and Arnold, J. D., 1971, Tr. Roy. Soc. Trop. Med. and Hyg., v. 65 (5), 684-685 [Letter]
Plasmodium vinckei chabaudi adapted to rats (exper.), synchronization of growth and reproduction by photoperiodic rhythm

Plasmodium vinckei chabaudi, illus.
Sinden, R. E.; and Garnham, P. C. C., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (5), 631-637
Plasmodium spp., sporozoite ultrastructure within oocyst and salivary glands, incidence of micropores and possible relationship to infectivity, sporozoite morphogenesis, abnormalities in development

Plasmodium vinckei chabaudi
Walliker, D.; and Carter, R., 1973, J. Protozool., v. 20 (4), 529
Plasmodium vinckei chabaudi, demonstration of genetic recombination between two lines

Plasmodium vinckei chwatti Garnham, 1973 (nomen nudum)
Killick-Kendrick, R., 1975, Ann. Parasitol., v. 50 (3), 251-264
as syn. of P. vinckei brucechwatti subsp. nov.

Plasmodium vinckei lentum Landau, Michel, Adam and Boulard, 1970, illus.
Carter, R.; and Walliker, D., [1977], Ann. Parasitol., v. 51 (6), 1976, 637-646
description of the blood-forms, sporogony and enzyme forms
laboratory mice (exper.)
Grammomys surdaster (exper.)
Anopheles stephensi (exper.)
Thamnomys rutilans: near the village of N'ganga Lingolo, 15 km west of Brazzaville, Congo

Plasmodium vinckei lentum Landau, Michel and Boulard, 1970
Killick-Kendrick, R., 1971, Symposia Brit. Soc. Parasitol., v. 9, 39-64
murine malaria parasites: collection of strains from rodents; collection of strains from mosquitoes; establishment of strains in the laboratory; maintenance by cyclical transmission, extensive review

Plasmodium (Vinckeia) vinckei petteri subsp. nov., illus.
Carter, R.; and Walliker, D., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 187-196
description of blood stages, sporogony, and enzyme forms; "From the results reported here it is now clear from the account in her paper (1965) that Landau was describing blood forms of both P. chabaudi and P. v. petteri."
Thamnomys rutilans (mature erythrocytes): near Bangui, Central African Republic

Plasmodium vinckei vinckei
Hargreaves, B. J.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 289-299
Plasmodium berghei yoelii, mice, protective immunity induced by mild parasite strains against virulent line, effect of increased inocula and time course of infection, further studies testing cross-protection against P. b. berghei and P. v. vinckei

Plasmodium vinckei vinckei Rodhain, 1952
Killick-Kendrick, R., 1971, Symposia Brit. Soc. Parasitol., v. 9, 39-64
murine malaria parasites: collection of strains from rodents; collection of strains from mosquitoes; establishment of strains in the laboratory; maintenance by cyclical transmission, extensive review

Plasmodium vinckei vinckei
Speer, C. A.; Silverman, P. H.; and Schiewe, S. G., 1976, J. Parasitol., v. 62 (5), 657-663
Plasmodium berghei, erythrocytic stages, cultivation in primary cultures of rodent bone marrow cells, unsuccessful with P. vinckei vinckei and P. knowlesi

Plasmodium vinckei vinckei
Speer, C. A.; Silverman, P. H.; and Schiewe, S. G., 1976, Ztschr. Parasitenk., v. 50 (3), 237-244
Plasmodium berghei, P. vinckei vinckei, P. coatneyi, P. knowlesi, attempts to cultivate erythrocytic stages in various cell lines, success only with P. berghei in mouse Leydig cell testicular tumor cultures

Plasmodium vivax
Ambroise-Thomas, P.; et al., 1976, Bull. World Health Organ., v. 54 (4), 355-367
human malaria, extensive sero-epidemiologic survey (6 surveys at 6-month intervals using peripheral blood examination and fluorescent antibody technique) to evaluate past and present status of malarial infection in Tunisia

Plasmodium vivax
Asch, A. J., 1976, Canad. Med. Ass. J., v. 115 (5), 405-406
increasing incidence of malarial infections due to increased travel to and immigration from endemic areas, emphasis on diagnostic awareness; transplacental infection reported in 3-week-old twins: Toronto

Plasmodium vivax
Avery, J. G., 1975, Papua N. Guinea Med. J., v. 18 (2), 124-125 [Letter]
progress report of the human malaria eradication program in the British Solomon Islands

Plasmodium vivax
Ayala, S. C.; and Sarmiento, G., 1976, Actualidades Biol., v. 5 (17), 55-61
Plasmodium, man and animals, review: Colombia

Plasmodium vivax
Baerg, D. C.; and Rossan, R. N., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 167-168
Plasmodium vivax, development of exoerythrocytic stage in Saguinus geoffroyi (exper.)

Plasmodium vivax
Biagi Finisola, F.; and Salmon Rodriguez, L., 1975, Bol. Med. Hosp. Inf., v. 32 (4), 691-698
Plasmodium vivax infection in infant, probable placental transmission: Mexico

Plasmodium vivax
Black, R. H., 1973, Med. J. Australia, v. 1 (26), 1265-1270
statistical survey of malaria prevalence in Australian army in South Vietnam; successful use of proguanil in conjunction with dapsone for chloroquine-resistant Plasmodium falciparum strain

Plasmodium vivax
Bolton, J. M., 1972, Med. J. Malaysia, v. 27 (1), 10-19
review of malarial control program among Orang Asli aborigines, prophylactic distribution of chloroquine and pyrimethamine and frequent spraying of homes with DDT implimented, discussion of problems involved: West Malaysia jungle areas

Plasmodium vivax
Bourgeade, A.; and Bertrand, E., 1972, Medecine Afrique Noire, v. 19 (4), 377-380
quinine as drug of choice in human malarias, indications for use

Plasmodium vivax
Bradbury, A. J., 1977, Brit. Med. J. (6087), v. 2, 613
Plasmodium vivax, congenital malaria in one non-identical twin, probably resulting from placental damage at time of delivery: London, mother had lived previously in India

Plasmodium vivax
Brown, P.; et al., 1976, Am. J. Trop. Med. and Hyg., v. 25 (6), 775-783
Plasmodium, 4 human spp., remote island populations, fluorescent antibody prevalence patterns, comparison with published parasite survey data based on blood smears and medical examinations, evaluation of epidemiological usefulness: New Hebrides; Solomons; Western Carolines (malaria entirely absent); New Guinea

Plasmodium vivax
Bruce-Chwatt, L. J.; and de Zulueta, J., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (3), 232-240
human malaria, details of eradication program to present stage of certification by WHO as area where malaria has been eradicated, emphasis on need for continued vigilance because of increased travel and imported cases: Portugal

Plasmodium vivax
Butler, T.; Warren, K. S.; and Mahmoud, A. A., 1976, J. Infect. Dis., v. 133 (6), 721-726
human malarias, algorithms in the diagnosis and management

Plasmodium vivax
Cabrera, B. D.; and Ramos, O. L., 1972, Southeast Asian J. Trop. Med. and Pub. Health, v. 3 (4), 501-504
Plasmodium vivax, P. falciparum, humans, trials with low dose sulfamonomethoxine, good results as causal prophylactic drug: Palawan, Philippines

Plasmodium vivax
Charoenlarp, P.; and Harinasuta, T., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (1), 135-137
Plasmodium vivax, humans, malarial relapse in 2 persons after conventional course of primaquine and chloroquine, cures obtained with increased dosages of both drugs: Thailand

Plasmodium vivax
Charters, A. D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 291-292 [Letter]
Plasmodium falciparum, P. vivax, technique for collecting multiple thick films on same slide when carrying out field surveys; comparison of high altitude endemicity records of areas in Ethiopia and Kenya

Plasmodium vivax
Charters, A. D.; and Davis, R. E., 1974, Med. J. Australia, v. 1 (14), 527-530
statistical review of human malaria infections imported into Western Australia, increasing incidence, drug resistance to commonly used antimalarials in 4 persons

Plasmodium vivax
Chen, D. H.; Nussenzweig, R. S.; and Collins, W. E., 1976, J. Parasitol., v. 62 (4), 636-637
rat antisera against sporozoites of 6 primate Plasmodium spp. reacted in circum-sporozoite precipitation tests only with sporozoites of homologous species, geographically different strains of same species cross-reacted intensely however

Plasmodium vivax
Clyde, D. F.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 397-401
immunization of man against sporozoite-induced Plasmodium falciparum and P. vivax by inoculation with X-irradiated attenuated sporozoites administered by mosquito bite, species specificity of antigen and antibody, no increase in levels of immunoglobulins G and M

Plasmodium vivax
Clyde, D. F.; et al., 1975, Antimicrob. Agents and Chemotherapy, v. 9 (3), 384-386
sporozoite-induced human malaria (exper.), suppressive activity of mefloquine, more effective against Plasmodium falciparum than against P. vivax

Plasmodium vivax
Clyde, D. F.; and McCarthy, V. C., 1977, Am. J. Trop. Med. and Hyg., v. 26 (3), 562-563
Plasmodium vivax in humans (exper.), radical cure without relapse obtained by treating with primaquine 60 mg over 7 rather than 14 days after previously administering chloroquine

Plasmodium vivax
Clyde, D. F.; McCarthy, V. C.; and Miller, R. M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 167-168 [Letter]
Plasmodium vivax, inactivity of RC-12 as causal prophylactic and relapse inhibitor when administered alone or with chloroquine in experimental human trials

Plasmodium vivax
Collins, W. E.; et al., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 372-375
Plasmodium vivax different strains, differential development in Anopheles maculatus and A. freeborni, differences in ability of A. freeborni to convert gut infections to salivary gland infections could have marked effect on transmission potential with this parasite-vector system, implications for introduction of malaria from Vietnam or Central America to United States

Plasmodium vivax
Collins, W. E.; et al., 1976, J. Parasitol., v. 62 (2), 190-194
comparative infectivity of 2 strains of Plasmodium vivax to 4 strains of Anopheles albimanus as well as to A. freeborni and A. maculatus (all exper.)

Plasmodium vivax
Collins, W. E.; et al., 1977, J. Parasitol., v. 63 (1), 52-56
Plasmodium falciparum, Santa Lucia strain isolated from El Salvador, sensitive to commonly used antimalarial drugs, established in Aotus trivirgatus griseimembra (exper.), successful transmission from monkey to monkey both by transfusion of infected blood and through bites of infected Anopheles freeborni, A. maculatus, and A. albimanus (all exper.), description of course of infection, higher parasitemia and mortality in animals previously exposed to Plasmodium vivax, parasitemia in animals exposed to P. vivax and P. cynomolgi lower than those exposed to P. vivax only

Plasmodium vivax
Collins, W. E.; Cedillos, R. A.; and Warren, M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1105-1107
Plasmodium vivax, passage of malaria antibodies from mothers to infants, survey in endemic area using the fluorescent antibody test: El Salvador

Plasmodium vivax
Collins, W. E.; Lunde, M. N.; and Skinner, J. C., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 412-416
measurement of developing antibodies to Plasmodium vivax comparing results of indirect hemagglutination and indirect fluorescent antibody tests, effects of short-term infections on development and persistence of antibody response and effect of relapse on response

Plasmodium vivax
Crane, G. G.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 308-314
malarial antibodies compared in New Guineans with tropical splenomegaly syndrome and normal controls; results of study cannot implicate any one species of Plasmodium in production of this pathological condition

Plasmodium vivax
Degremont, A. ; and Weiss, N., 1975, Praxis, Bern, v. 64 (18), 553-555
human malarias, evaluation of immunofluorescence as adjunct to diagnostic serology

Plasmodium vivax
Devlin, H. R.; and Bannatyne, R. M., 1977, Canad. Med. Ass. J., v. 116 (1), 20 [Letter]
Plasmodium vivax malaria in 3-week-old twins, mother treated for malaria prior to and during pregnancy but means of transmission to infants uncertain, successfully treated with chloroquine: Toronto, Canada (mother immigrated from India)

Plasmodium vivax, illus.
Doby, J. M.; and Barker, R., 1976, Compt. Rend. Soc. Biol., Paris, v. 170 (3), 661-665
Plasmodium vivax, human liver cell cultures inoculated with North Korean strain, description of some rare intracytoplasmic forms, probably connected with pre-erythrocytic cycle

Plasmodium vivax
Dondero, T. J., jr.; et al., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 144-145 [Demonstration]
invasion of child host's red blood cells by both sexual and asexual forms: Pahang, Peninsular Malaysia

Plasmodium vivax
Dondero, T. J., jr.; et al., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (4), 574-578
Plasmodium falciparum, P. vivax, human malaria prevalence survey revealed no evidence of chloroquine resistance in surveyed areas of North Sumatra, Indonesia

P[lasmodium] vivax
Dondero, T. J., jr.; Parsons, R. E.; and O'Holohan, D. R., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (4), 488-494
P[lasmodium] falciparum, increased frequency of chloroquine resistance during 2-year survey of employees of rubber estate; P. vivax did not exhibit similar increase: Paroi Estate, near Kuala Lumpur, Malaysia

Plasmodium vivax
Dondero, T. J., jr.; Parsons, R. E.; and Ponnampalam, J. T., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 149-154
Plasmodium falciparum, appreciable chloroquine and chloroquine combined with pyrimethamine drug resistance discovered in survey of peninsular area; no evidence of resistance with P. vivax and P. malariae: Malaysia

[Plasmodium] vivax
Ebisawa, I.; and Muto, T., 1972, Jap. J. Exper. Med., v. 42 (4), 415-417
survey of healthy Laotians employed in endemic vivax malarial area for possible primaquine sensitivity (genetic metabolic deficiency) using methemoglobin reduction test; Japanese similarly tested

Plasmodium vivax
Edrissian, G. H., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (6), 491-493
Plasmodium malariae and P. vivax in humans, importance of blood transfusions as means of transmission and need for stringent control measures: Iran

Plasmodium vivax
Edrissian, G. H.; and Afshar, A., 1973, Pahlavi Med. J., v. 4 (4), 533-544
human malarias, detection of asymptomatic and scanty parasitemia using combined indirect fluorescent antibody technique and parasite concentration from thick and thin blood films, use in mass surveys: Iran

Plasmodium vivax
Elslager, E. F., 1974, Progr. Drug Research, v. 18, 99-172
human malaria and filariasis, new perspectives on chemotherapy, extensive review

Plasmodium vivax
Eshghi, N.; et al., 1976, Trop. and Geogr. Med., v. 28 (1), 41-44
Plasmodium falciparum, P. vivax, role of Anopheles fluviatilis as vector: southern regions of Iran

Plasmodium vivax
Fajardo, L. F., 1973, Nature (5405), v. 243, 298-299
Plasmodium falciparum, P. vivax, human, ultrastructural changes in blood platelets; P. berghei, mice (exper.), presence of parasites in platelets

P[lasmodium] vivax
zum Felde, I.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (4), 477-484
P[lasmodium] falciparum, P. vivax, P. ovale, P. malariae, human, indirect immunoflourescence test with same 4 Plasmodium spp. plus P. fieldi as antigen, species-specific and cross-reactions, effect of therapy, implications for immunodiagnosis

Plasmodium vivax
Fleming, K., 1974, Med. J. Australia, v. 2 (23), 834-837
review of Plasmodium falciparum and Plasmodium vivax case reports and clinical findings in Australian soldiers fighting in South Vietnam, problems encountered

Plasmodium vivax
Gelpi, A. P.; and King, M. C., 1976, Science, v. 191 (4233), 1284
Plasmodium falciparum, P. vivax, Duffy blood group antigens, sickle cell trait carriers and resistance to malaria: Saudi Arabia

Plasmodium vivax
Ghose, A. C.; and Chowdhury, A. B., 1977, Indian J. Med. Research, v. 66 (4), 566-569
human malaria and kala-azar, immunoglobulin levels of persons with known infections compared with normal controls

Plasmodium vivax
Gibson, J. J., jr.; Brodsky, R. E.; and Schultz, M. G., 1974, J. Infect. Dis., v. 130 (5), 553-555
increased incidence of malaria in the United States probably reflects increased foreign travel and improved reporting of infections

Plasmodium vivax
Glew, R. H.; et al., 1974, J. Infect. Dis., v. 129 (4), 385-390
Plasmodium falciparum in young girl, strain with resistance to chloroquine, quinacrine, pyrimethamine and chlorguanide, radical cure obtained with pyrimethamine and sulfadiazine in combination; delayed primary attack with Plasmodium vivax: Maryland (had traveled and engaged in field work in remote area of eastern Colombia)

Plasmodium vivax
Glew, R. H.; and Levin, J., 1975, Proc. Soc. Exper. Biol. and Med., v. 148 (2), 508-510
Plasmodium falciparum, P. vivax in humans (exper.), P. coatneyi in Macaca mulatta (exper.), unable to demonstrate the presence of endotoxins in blood of either humans or monkeys infected with malaria in studies using the Limulus amebocyte lysate test

Plasmodium vivax
Gratecos, N., 1972, Marseille Med., v. 109 (5), 361-369
Plasmodium spp., Leishmania donovani, human splenomegaly, differential diagnosis, clinical review

Plasmodium vivax
Grindel, J. M.; Tilton, P. F.; and Shaffer, R. D., 1977, J. Pharm. Sc., v. 66 (6), 834-837
human malarias, quantitation of mefloquine in blood, plasma, and urine using high-pressure liquid chromatography, suitable assay for study of pharmacokinetics in humans

Plasmodium vivax
Gueye, I.; and Michel, R., 1973, Medecine Afrique Noire, v. 20 (10), 753-766
guidelines for mass prophylactic therapy against human forms of malaria

Plasmodium vivax
Gundelfinger, B. F.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 393-396
statistics of survey for prevalence of human Plasmodium spp. and response of Plasmodium falciparum and P. vivax to standard dose of chloroquine; first record of P. ovale reported in Timor, Indonesia

P[lasmodium] vivax
Harinasuta, T.; Gilles, H. M.; and Sandosham, A. A. [Editors], 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (4), 641-678
human malaria in southeast Asia, scientific group meeting with discussion on: epidemiology, clinical features, pathophysiology, genetic factors, immunology, diagnosis, chemotherapy, control measures

P[lasmodium] vivax
Jaroonvesama, N.; et al., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (3), 419-424
human malaria, degree of changes in coagulation and serum fibrin degradation products varied with severity of infection in falciparum malaria and not at all in vivax malaria, therefore possibly result rather than cause of pathogenesis

Plasmodium vivax
Jeffery, G. M.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 402-411
application of indirect fluorescent antibody test findings to assessment of endemicity of Plasmodium falciparum and P. vivax in malarial surveillance areas, climate, geography and area development as additional varying factors: Mato Grosso State, Brazil

Plasmodium vivax
Johnson, A., jr., 1975, Mil. Med., v. 140 (12), 853-856
Plasmodium vivax, P. falciparum, military personnel, health education, preventive and control measures: U.S. military base, Vietnam

Plasmodium vivax
van der Kaay, H. J.; and Darwish, R. O., 1975, Riv. Parassitol., Roma, v. 36 (1), 1-6
Plasmodium vivax, P. falciparum, P. malariae, spleen and blood examinations, vector survey, feasibility study for possible control measures: Bokondini, Irian Jaya, Indonesia

Plasmodium vivax
van der Kaay, H. J.; and Zuidema, P. J., 1976, Nederl. Tijdschr. Geneesk., v. 120 (20), 884-886
human Plasmodium spp., recommendations for malaria prophylaxis during travel to endemic areas

Plasmodium vivax
Kaplan, M. H.; and Bernstein, L. S., 1974, Mil. Med., v. 139 (6), 444-448
Plasmodium vivax, U.S. soldiers in Vietnam, improved care with primaquine combined with chloroquine therapy; survey of servicemen shows frequent mixed hookworm infections, increased serum IgM levels and decreased cholesterol levels

Plasmodium vivax
Kean, B. H.; and Reilly, P. C., jr., 1976, Am. J. Med., v. 61 (2), 159-164
recommendations for diagnosis, prophylaxis and treatment of human malaria with epidemiologic review of recently treated infections in persons who had travelled or lived in endemic areas: New York City

Plasmodium vivax
Lainson, R.; et al., 1976, Acta Amazonica, v. 6 (4), 55-60
humans: Municipio of Aripuana, Mato Grosso State, Brazil

Plasmodium vivax
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium vivax
Lewis, A.-N.; and Ponnampalam, J. T., 1975, Ann. Trop. Med. and Parasitol., v. 69 (1), 1-12
Plasmodium falciparum, P. vivax, suppression of malaria with monthly administration of combined sulphadoxine and pyrimethamine, compared with control groups on placebo and on weekly chloroquine, no serious side effects observed

Plasmodium vivax
McCarthy, V. C.; and Clyde, D. F., 1977, Exper. Parasitol., v. 41 (1), 167-171
Plasmodium vivax, human, correlation of circumsporozoite precipitation reaction with sporozoite-induced protective immunity

P[lasmodium] vivax
Maffi, M.; et al., 1975, Riv. Parassitol., Roma, v. 36 (4), 233-254
malaria, prevalence survey of highland natives using enlargement of spleen as sign of infection, considerations for control: around Bokondini (Jayawijaya), Irian Jaya, Indonesian New Guinea

Plasmodium vivax
Mahajan, R. C.; and Dutta, D. V., 1976, Indian J. Med. Research, v. 64 (5), 652-655
detection of malarial antibodies in humans with non-cirrhotic portal fibrosis, discussion of malaria as possible etiologic factor in hepatic disease

Plasmodium vivax
Mannweiler, E.; et al., 1976, Munchen. Med. Wchnschr., v. 118 (36), 1139-1144
Plasmodium spp., homologous and heterologous malaria pathogens used as antigens to determine malaria antibodies by direct immunofluorescence, pre- and post-treatment with chloroquine compared

Plasmodium vivax
Mannweiler, E.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (1), 23-25
Plasmodium falciparum, P. vivax, strain related differences in immunodiagnosis, antigen variations as cause of antibody titer differences

Plasmodium vivax
Mason, J., 1975, Am. J. Trop. Med. and Hyg., v. 24 (4), 581-585
Plasmodium vivax, seasonal pattern and patterns of recurrences in high-incidence coastal area, relapse pattern observed may provide for parasite survival through prolonged dry season of low transmission potential: El Salvador

Plasmodium vivax
Mason, J.; and Hobbs, J., 1977, Bull. Pan Am. Health Organ., v. 11 (1), 17-30
malaria field studies, 1971-1973, incidence, seasonal occurrence, densities in regard to host age and sex: coastal area of El Salvador

Plasmodium vivax
Mathews, H. M.; and Dilworth, D. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (2), 351-352
Plasmodium brasilianum, use as antigen in indirect hemagglutination test for diagnosing human P. falciparum, P. vivax, P. ovale, P. malariae

Plasmodium vivax
Mathews, H. M.; Fried, J. A.; and Kagan, I. G., 1975, Am. J. Trop. Med. and Hyg., v. 24 (3), 417-422
evaluation by indirect hemagglutination test of soluble antigens prepared from Plasmodium falciparum and P. vivax, measurement of detection rate in heterologous and homologous Plasmodium spp.

Plasmodium vivax
Miller, L. H., 1974, Transmis. Disease and Blood Transfus., 241-266
human malarias, control measures established to prevent transmission via infected blood donors; review of life cycles and clinical aspects of malaria infections

Plasmodium vivax
Miller, L. H., 1977, J. Infect. Dis., v. 135 (5), 855-864
human malarias, current prospects and problems in developing a vaccine for malaria immunization

Plasmodium vivax
Miller, L. H.; et al., 1976, N. England J. Med., v. 295 (6), 302-304
Plasmodium vivax, resistance factor in African and American blacks, Duffy determinants on erythrocyte surface required for invasion of erythrocyte by vivax merozoites (Duffy-blood-group-negative human erythrocytes resistant to invasion)

Plasmodium vivax
Miller, L. H.; McAuliffe, F. M.; and Mason, S. J., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, Pt. 2), 204-208
erythrocyte receptors for malaria merozoites: Duffy blood group system and invasion by Plasmodium knowlesi and P. vivax, erythrocyte requirements for invasion by P. falciparum, workshop report

Plasmodium vivax
Nakabayashi, T.; et al., 1973, Nettai Igaku (Trop. Med.), v. 15 (3), 154-168
Plasmodium spp. in humans, epidemiologic survey to determine prevalence rate in rural areas of the Philippines

P[lasmodium] vivax
Nakabayashi, T.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (11), 247-251
human malaria, statistical survey of cases imported into Japan from 1972-1974

Plasmodium vivax
O'Holohan, D. R., 1975, Med. J. Malaysia, v. 29 (3), 157-162
statistical survey of individual physician's practice for current trends in human malarial infections, review of presenting clinical symptoms: Seremban, West Malaysia

Plasmodium vivax
O'Holohan, D. R., 1976, J. Trop. Med. and Hyg., v. 79 (9), 191-196
Plasmodium vivax, P. falciparum, human, analysis of clinical and laboratory findings of 1000 cases, need for diagnostic awareness and repeated blood smears in endemic areas: Malaysia

Plasmodium vivax
Otieno, L. H.; et al., 1971, Trop. and Geogr. Med., v. 23 (4), 369-375
human Plasmodium spp., dynamics of epidemiologic survey in highly endemic malarious area, data show decreasing incidence in recent years: north-eastern Tanzania

Plasmodium vivax
Ough, R. N., 1976, Canad. Med. Ass. J., v. 115 (12), 1196 [Letter]
Plasmodium vivax, increasing incidence of malaria imported into non-endemic areas, case reports, need for differential diagnostic awareness: British Columbia, Canada

Plasmodium vivax
Palmer, A., 1976, Brit. J. Clin. Pract., v. 30 (1), 19-20
Plasmodium vivax, increased incidence of malaria with increased travel, implications if associated with pregnancy, need for diagnostic awareness: United Kingdom

Plasmodium vivax
Papua N. Guinea Med. J., v. 17 (1), 1974, 1-115
multiple articles focusing on human malaria in Papua New Guinea (epidemiology, public health, control programs, vectors, diagnosis, toxicity of antimalarials, statistics of surveys)

Plasmodium vivax
Partono, F.; et al., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (4), 484-486
epidemiologic survey in South Sulawesi for human malarias and filariasis conducted 8 and 22 months after arrival of transmigrants from Central Java showed decreased incidence of malaria and increased filariasis, discussion of associated problems and effects on populace: Indonesia

Plasmodium vivax
Pearlman, E. J.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1108-1115
Plasmodium falciparum, P. vivax, chloroquine resistant strains, field trials comparing efficacy of pyrimethamine combined with either sulfadoxine or diformyldapsone: Thailand

Plasmodium vivax
Pearlman, E. J.; Thiemanun, W.; and Castaneda, B. F., 1975, Am. J. Trop. Med. and Hyg., v. 24 (6, pt. 1), 901-909
Plasmodium falciparum, P. vivax, chemosuppressive field trials testing efficacy of diformyldapsone alone and in combination with pyrimethamine, comparisons with dapsone and pyrimethamine: Thailand

Plasmodium vivax
Pettyjohn, F. S., 1975, Mil. Med., v. 140 (8), 535-537
Plasmodium vivax, P. falciparum, human, centrifuged vacutainer of venous blood procedure of choice in quantitative comparisons of methods of thin blood smear preparations for diagnosis

Plasmodium vivax
Ponnampalam, J. T.; Seow, C. L.; and Roy, O. S., 1976, J. Trop. Med. and Hyg., v. 79 (10), 220-225
human malaria prophylaxis, controlled study comparing efficacy of dapsone combined with pyrimethamine (Maloprim) to that of chloroquine for use in long-term chemosuppression of infection: Malaysia

Plasmodium vivax
Porter, J. A., jr., 1971, Am. J. Trop. Med. and Hyg., v. 20 (4), 535-538
Aotus trivirgatus established as successful laboratory host for the study of human Plasmodium vivax malaria

Plasmodium vivax
Radford, A. J.; Van Leeuwen, H.; and Christian, S. H., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 11-23
changes in distribution and determinants of malaria that occurred with the development process in the highlands of Papua New Guinea between 1930 and 1970

[Plasmodium] vivax
Ramachandran, S.; and Perera, M. V. F., 1976, J. Trop. Med. and Hyg., v. 79 (9), 207-210
Plasmodium falciparum, P. vivax, humans, jaundice and hepatomegaly associated with attacks of primary uncomplicated malaria

Plasmodium vivax
Ramos, O. L.; and Cabrera, B. D., 1972, Southeast Asian J. Trop. Med. and Pub. Health, v. 3 (4), 562-568
Plasmodium falciparum, P. vivax, humans, evaluation of blood schizonticidal and radical curative activities of sulfamonomethoxine alone or combined with pyrimethamine in strains resistant and sensitive to chloroquine

Plasmodium vivax
Robinson, M. J.; and Eykyn, S., 1977, Brit. Med. J. (6103), v. 2, 1667 [Letter]
childhood malaria, increasing incidence with increased travel to and from endemic areas, diagnostic problem in children: United Kingdom

Plasmodium vivax
Rossan, R. N.; and Baerg, D. C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 471-472
Plasmodium vivax in Saimiri sciureus (exper.), morphology and development of exoerythrocytic stages of infection

P[lasmodium] vivax
Roy, R. G.; et al., 1976, Indian J. Med. Research, v. 64 (10), 1451-1455
Plasmodium vivax, P. falciparum, humans, nirmalaria (indigenous compound of 4 herbal ingredients) unsuccessful in trials as inhalation therapy: India

Plasmodium vivax
Rutledge, L. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 451 [Letter]
comments on previous authors' interpretations of incubation and prepatent periods

Plasmodium vivax
Rutter, N., 1977, Brit. Med. J. (6098), v. 2, 1335-1336
malaria infections in children born and living in non-malarious areas who have contracted infections upon visiting malarious areas with their immigrant parents who possibly have some natural immunity to malaria although their children do not, importance of diagnostic awareness: Nottingham, England

Plasmodium vivax
Rytel, M. W.; Rose, H. D.; and Stewart, R. D., 1973, Proc. Soc. Exper. Biol. and Med., v. 144 (1), 122-123
Trypanosoma cruzi, Plasmodium falciparum, P. vivax, humans, lack of circulating humoral interferon in serum of persons with acute illness

Plasmodium vivax
Schmidt, L. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 446-474
Plasmodium falciparum and Plasmodium vivax in Aotus trivirgatus as model systems for chemotherapeutic studies, handling of animals, course of infections and response to standard antimalarial drugs compared with that in man, use of model in assessing activities of new agents and drug resistance

Plasmodium vivax
Schmidt, L. H.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (5, part 1), 837-849
Plasmodium cynomolgi in Macaca mulatta, P. vivax and P. falciparum in Aotus trivirgatus griseimembra (both exper.), therapeutic potentiation with combinations of pyrimethamine and sulfonamides, good response in pyrimethamine susceptible and resistant strains except for Malayan Camp and Vietnam Smith strains of P. falciparum

Plasmodium vivax, illus.
Schultz, M. G., 1977, Postgrad. Med., v. 62 (2), 121-125

Plasmodium vivax
Shanmugam, C. A. K.; et al., 1976, Indian J. Med. Research, v. 64 (9), 1314-1317
Plasmodium falciparum, P. vivax, humans treated with chloroquine and/or primaquine, follow-up for chloroquine sensitivity status: Tamil Nadu State, India

Plasmodium vivax
Shaw, P. K.; Brodsky, R. E.; and Schultz, M. G., 1976, J. Infect. Dis., v. 133 (1), 95-101
malaria surveillance, statistics for 1974, increasing incidence with increasing international travel: United States

Plasmodium vivax
Shute, P. G.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 474-481
Plasmodium vivax, delayed incubation in North Korean strain characterized by predominantly prolonged prepatent period, study shows that number of sporozoites influences length of prepatent period

Plasmodium vivax
Singal, M.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 1-9
Plasmodium vivax, outbreak of introduced (Punjabi immigrants to agricultural area) malaria with likelihood of secondary transmission, coincidentally high numbers of Anopheles freeborni (effective P. vivax vectors) in transmission area: California

Plasmodium vivax
Sulzer, A. J.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 55-58
malaria, human, diagnosis, indirect fluorescent antibody test using multi-species thick smear antigen containing equal proportions of Plasmodium vivax, P. falciparum, and P. brasilianum (latter serologically equivalent in IFA test to P. malariae), circumvents need for multiple testing with several antigen species

Plasmodium vivax
Sulzer, A. J.; and Latorre, C. R., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 553 [Letter]
primate malarias, method for in vitro production of schizont antigens for use in indirect immunofluorescence tests for malarial antibodies

Plasmodium vivax
Szarfman, A.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 19-24
evaluation of specificity of EVI factor observed in Trypanosoma cruzi cases using indirect fluorescent antibody test on sera from patients with other parasitic diseases, immunofluorescence of anti-skeletal muscle antibody from leishmaniasis

Plasmodium vivax
Talathi, S. B.; Joshi, N. C.; and Sood, S. R., 1973, Pediat. Clin. India, v. 8 (4), 213-214
Plasmodium vivax malaria in neonate following exchange transfusion for Rh incompatibility, successful treatment with amodiaquin, clinical case report, suggested control measures: Bombay, India

Plasmodium vivax
Tapper, M. L.; and Armstrong, D., 1976, Arch. Int. Med., Chicago, v. 136 (7), 807-810
blood transfusion induced malarial infections in 2 patients with neoplastic disease, case reports, implications for alterations of immunologic status

P[lasmodium] vivax
Thaver, S.; and Draper, C. C., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 8 [Demonstration]
P[lasmodium] falciparum, P. vivax and T[rypanosoma] cruzi antibodies tested for reliability of epidemiologic field survey collection methods, use of chromatography paper for collecting capillary blood recommended

Plasmodium vivax
Thompson, D.; et al., 1977, Pediatrics, Am. Acad. Pediat., v. 60 (2), 209-212
Plasmodium vivax, splenomegaly and anemia in an infant diagnosed as congenital malaria, case report: Los Angeles, California (mother had left Michoacan, Mexico shortly before delivery)

Plasmodium vivax
Thompson, R. G.; et al., 1977, Brit. Med. J. (6074), v. 1, 1468 [Letter]
human malaria, increasing number of infections seen in travelers returning from endemic areas, suggested prophylactic measures: United Kingdom

P[lasmodium] vivax
Topley, E.; Knight, R.; and Woodruff, A. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 51-54
malaria patients, direct antiglobulin test and immunoconglutinin titres, possible significance of results in understanding mechanism of anemia

Plasmodium vivax
Tsukamoto, M.; et al., 1975, Nettai Igaku (Trop. Med.), v. 17 (1), 13-26
Plasmodium spp. in humans, field trials of biochemical analysis of blood and urine, value comparisons before and after antimalarial therapy: Philippines

Plasmodium vivax
Ungureanu, E.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 482-483
Plasmodium vivax, tropical and temperate strain differences probably resulting from production of 2 types of sporozoites, type I in tropical strain eliciting short prepatent period and type II in temperate strain eliciting long prepatent period, relapse of P. vivax possibly delayed parasitemia from type II sporozoites

Plasmodium vivax
Warren, M.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 607-611
differences in susceptibility to Plasmodium vivax and P. falciparum of inbred morphologic phenotypes of Anopheles albimanus vector mosquitoes, significance in epidemiologic surveys and development of vector control measures: El Salvador

Plasmodium vivax
Welch, S. G.; McGregor, I. A.; and Williams, K., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 295-296
survey of 2 rural Gambian villages showed total absence of Duffy blood group antigens and absence of Plasmodium vivax infection although P. falciparum, P. malariae and P. ovale were present; findings consistant with theory that absence of Duffy phenotypes constitutes basis of innate resistance to P. vivax

Plasmodium vivax
Young, M. D.; Baerg, D. C.; and Rossan, R. N., 1976, Lab. Animal Sc., v. 26 (6, pt. 2), 1131-1137
Plasmodium falciparum, P. vivax, induced malarias in Aotus monkeys, transmission to other New World monkeys and man, use as biological model, literature review

Plasmodium vivax hibernans
Carter, R.; and Voller, A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 14-15 [Demonstration]
Plasmodium spp., survey of electrophoretic forms of 4 enzymes, variation between species, subspecies, and strains

Plasmodium vivax hibernans, illus.
Sinden, R. E.; and Garnham, P. C. C., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (5), 631-637
Plasmodium spp., sporozoite ultrastructure within oocyst and salivary glands, incidence of micropores and possible relationship to infectivity, sporozoite morphogenesis, abnormalities in development

Plasmodium voltaicum
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium yoelii
Bannister, L. H.; Butcher, G. A.; and Mitchell, G. H., 1977, J. Protozool., v. 24 (2), Suppl., 6A [Abstract]
Plasmodium knowlesi, P. yoelii, extracellular erythrocytic merozoites, nature of cell surface, adhesion with other cells

Plasmodium yoelii
Belehu, A.; Poulter, L. W.; and Turk, J. L., 1976, Infect. and Immun., v. 14 (2), 457-462
hamsters, effect of Plasmodium berghei or P. yoelii malaria on concurrent Leishmania enriettii infection depended on relative timing of two infections

Plasmodium yoelii
Brown, A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 284 [Demonstration]
Plasmodium yoelii, technique for isolation of microgametes of malaria parasites, potential use in genetic studies

Plasmodium yoelii
Brown, I. N.; Watson, S. R.; and Sljivic, V. S., 1977, Infect. and Immun., v. 16 (2), 456-460
antibody response in vitro of spleen cells from Plasmodium yoelii-infected mice, response to sheep erythrocytes enhanced at early stage of infection and depressed at later intervals, cell fractionation experiments indicated a defect of macrophage function, response to dinitrophenylated Ficoll remained normal

Plasmodium yoelii
Cottrell, B.J.; Playfair, J.H.L.; and de Sousa, B., 1977, Exper. Parasitol., v. 43 (1), 45-53
Plasmodium yoelii, Plasmodium vinckei, mice, effects of nonspecific immunostimulation with a variety of agents

Plasmodium yoelii, illus.
Jackson, P. R.; et al., 1977, J. Parasitol., v. 63 (4), 593-598
Plasmodium yoelii, Trypanosoma gambiense, and Trypanosoma equiperdum, cytofluorograf detection of parasites by laser excited fluorescence of stained rodent blood

Plasmodium yoelii, illus.
Jayawardena, A. N.; et al., 1977, Immunology, v. 32 (6), 849-859
Plasmodium berghei yoelii (P. yoelii), CBA mice, immunological response, general characteristics and effects of T-cell deprivation and reconstitution with thymus grafts

Plasmodium yoelii
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Plasmodium yoelii, illus.
McAlister, R. O.; and Gordon, D. M., 1976, J. Parasitol., v. 62 (4), 639-641
rapid method for recovery of staining properties and morphology of Plasmodium yoelii-infected mouse erythrocytes following washing and culture in vitro

Plasmodium yoelii
McAlister, R. O.; and Gordon, D. M., 1976, J. Parasitol., v. 62 (5), 664-669
Plasmodium berghei- or P. yoelii-parasitized mouse erythrocytes, differential fractionation by ultracentrifugation using preformed isodensity gradients of discontinuous type (Stractan II as gradient material), produces significant fractionation of various erythrocytic stages, yields cell suspensions both morphologically and physiologically intact

Plasmodium yoelii
Playfair, J. H. L.; de Souza, J. B.; and Cottrell, B. J., 1977, Immunology, v. 32 (5), 681-687
Plasmodium spp., Babesia microti, mouse helper T-cell response to parasites measured by using them as carriers for a standard hapten, results show extensive cross-reaction between the 4 parasites as carriers apparently unrelated to known serological cross-reactions but are against the idea that helper T-cells are exclusively responsible for resistance

Plasmodium yoelii
Playfair, J. H. L.; de Souza, J. B.; and Cottrell, B. J., 1977, Immunology, v. 33 (4), 507-515
Plasmodium yoelii, P. vinckei, P. berghei, mice, regime of killed homologous vaccine plus Bordetella pertussis adjuvant, differences between species in effectiveness of protection, some cross-protection but largely species-specific, passive transfer of immunity to P. yoelii by serum or spleen cells

Plasmodium yoelii
Tarzaali, A.; Viens, P.; and Quevillon, M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (3), 520-524
Plasmodium yoelii-infected mice, depressed capacity to build up immune response to diphtheria vaccine and tetanus toxin, immunodepression overcome by Freund's adjuvant or pertussis bacilli, results suggest that diphtheria and tetanus vaccine should be given in association with pertussis vaccine in malaria endemic areas

Plasmodium yoelii
Wedderbun, N.; and Dracott, B. N., 1977, Clin. and Exper. Immunol., v. 28 (1), 130-137
mice with acute and chronic malaria, suppression of immune response to type III pneumococcal polysaccharide

Plasmodium yoelii (P. berghei yoelii)
Wells, R. A.; et al., 1977, Exper. Parasitol., v. 41 (2), 472-479
Plasmodium yoelii, inbred model for protective immunization against malaria in BALB/c mice, irradiated blood forms of two lines of 17X strain used as immunogen, "most complete protection of mice against a lethal challenge with malaria parasite in the absence of previous active infection that has, to our knowledge, been thus far reported."

Plasmodium yoelii nigeriensis
Momen, H., 1976, Parasitology, v. 73 (2), xvi [Abstract]
Plasmodium spp., Babesia spp., Anthemosoma garnhami, carbohydrate metabolism: enzyme activity, glucose catabolism

Plasmodium yoelii nigeriensis
Peters, W.; et al., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 407-418
Plasmodium berghei, chloroquine sensitive and resistant strains, or mixed infections with P. yoelii nigeriensis, efficacy and mode of action of mefloquine in mice (exper.), evidence of potent blood schizontocide with effects enhanced by addition of pyrimethamine, sulfaphenazole or primaquine

Plasmodium yoelii nigeriensis
Peters, W.; Davies, E. E.; and Robinson, B. L., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 311-328
Plasmodium yoelii nigeriensis-mouse model for screening drugs for causal prophylactic activity

Plasmodium yoelii nigeriensis
Sinden, R. E., 1975, J. Protozool., v. 22 (3), 56A [Abstract]
Plasmodium yoelii nigeriensis, microgametogenesis, oocyst maturation, sporozoite encystment, scanning electron microscopy

Plasmodium yoelii nigeriensis, illus.
Sinden, R. E.; Canning, E. U.; and Spain, B., 1976, Proc. Roy. Soc. London, B., Biol. Sc. (1110), v. 193, 55-76
Plasmodium yoelii nigeriensis, transmission electron microscopy, ultrastructure of micro- and macrogametocytes, nuclear and cytoplasmic changes during gametogenesis, fertilization

Plasmodium yoelii nigeriensis
Sinden, R. E.; and Croll, N. A., 1974, J. Protozool., v. 21 (3), 452-453 [Abstract]
Plasmodium yoelii nigeriensis, kinetics of movement of microgametes, recognition of macrogamete and fertilization

Plasmodium yoelii nigeriensis
Suzuki, M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 20 [Demonstration]
Plasmodium berghei berghei, P. yoelii nigeriensis, lung pathology in mice (exper.)

Plasmodium yoelii yoelii
Carter, R.; and Walliker, D., 1975, Ann. Trop. Med. and Parasitol., v. 69 (2), 187-196
Thamnomys rutilans (reticulocytes): near Bangui, Central African Republic

Plasmodium yoelii yoelii
McBride, J. S.; Micklem, H. S.; and Ure, J. M., 1977, Immunology, v. 32 (5), 635-644
Plasmodium yoelii yoelii, P. berghei, mice, depression of antibody response to type III pneumococcal polysaccharide

Plasmodium yoelii yoelii
Walliker, D.; et al., 1976, Parasitology, v. 72 (2), 183-194
Plasmodium yoelii yoelii, genetic basis of sudden appearance of virulence

Plasmodium yoelii yoelii
Walliker, D.; and Sanderson, A., 1977, J. Protozool., v. 24 (4), 39A [Abstract]
Plasmodium yoelii yoelii, genetics of virulence

Plasmodium youngi
Landau, I.; Miltgen, F.; and Chabaud, A. G., 1976, Ann. Parasitol., v. 51 (2), 175-187
Haemosporidia of mammals, classification into 3 groups based on morphology of gametocytes, correlation with morphology of tissue schizonts, host range, hypothesis on evolution

Pleistophora Gurley, 1893
Baxter, K. N.; Rigdon, R. H.; and Hanna, C., 1970, J. Invert. Path., v. 16 (2), 289-291
[For complete author reference see Supplement 19, Part 1]
Syn.: Plistophora Labbe, 1889 [i.e. 1899]

Pleistophora
Nagel, M. L.; and Hoffman, G. L., 1977, J. Parasitol., v. 63 (1), 160-162
characteristics of various Pleistophora spp. occurring in fish gonads

Pleistophora sp.
Iversen, E. S.; and Kelly, J. F., 1976, J. Invert. Path., v. 27 (3), 407-408
Penaeus duorarum: Biscayne Bay

Pleistophora sp.
Maddox, J. V., 1973, Misc. Publication Entom. Soc. Am., v. 9 (2), 99-104
persistence of Microsporida, resistance of spores to environmental conditions, transmission and dispersal, host range and infectivity, effect of temperature on development
Crambus trisectus

Pleistophora sp., illus.
Overstreet, R. M.; and Howse, H. D., 1977, Ann. N. York Acad. Sc., v. 298, 427-462
helminths and protozoans of estuarine fishes, incidence and intensity; possible relationships with water pollutants; tumorous growths
Leiostomus xanthurus: estuaries of Mississippi

Pleistophora, probably n. sp., illus.
Sanders, R. D.; and Poinar, G. O., jr., 1976, J. Invert. Path., v. 28 (1), 109-119
Pleistophora probably n. sp. in Aedes sierrensis, development and fine structure, potential as biological control agent
Aedes sierrensis (posterior midgut epithelium) (nat. and exper.): Strawberry Canyon, University of California, Berkeley, campus

Pleistophora sp.
Vavra, J., 1975, J. Protozool., v. 22 (3), 69A [Abstract]
Pleistophora sp., presence of scindosome--a new microsporidian organelle

Pleistophora sp., illus.
Wilson, G. G., 1975, Canad. J. Zool., v. 53 (12), 1799-1802
description
Choristoneura fumiferana (gut): Ontario, Canada [and/or] northern Minnesota, U.S.A.

Pleistophora crangoni n. sp., illus.
Breed, G. M.; and Olson, R. E., 1977, J. Invert. Path., v. 30 (3), 387-405
Pleistophora crangoni n. sp. in Crangon spp., prevalence and intensity in relation to season and host size, parasitic castration of females, shift in host sex ratio toward females, unsuccessful laboratory transmission experiments, spread of infection through shrimp, infected shrimp succumbed to low oxygen stress before uninfected shrimp, histopathology
Crangon franciscorum: vicinity of Yaquina Bay, Oregon
C. nigricauda: vicinity of Yaquina Bay, Oregon
C. nigromaculata: vicinity of Yaquina Bay, Oregon
C. stylirostris: vicinity of Yaquina Bay, Oregon (Seal Rock)
(skeletal muscle of all)

Pleistophora debaisieuxi Jirovec
Boemare, N.; and Maurand, J., 1976, Bull. Soc. Zool. France, v. 101 (3), 377-385
Thelohania fibrata, Pleistophora debaisieuxi, respiratory metabolism of healthy and infected Tetisimulium bezzii larvae compared

Pleistophora debaisieuxi Jirovec, 1940, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Pleistophora debaisieuxi
Vavra, J., 1977, J. Protozool., v. 24 (2), Suppl., 17A [Abstract]
microsporidian mitotic apparatus

Pleistophora debaisieuxi
Weiser, J., 1975, J. Protozool., v. 22 (3), 70A [Abstract]
microsporidians in simuliids (including Pleistophora debaisieuxi in Odagmia ornata; and Glugea), provocation of neoplastic xenoma

Pleistophora debaisieuxi, illus.
Weiser, J., 1976, Ztschr. Parasitenk., v. 48 (3-4), 263-270
Pleistophora debaisieuxi, xenomas (xenoparasitic complexes or parasite-host interactions) in blackflies, Odagmia ornata (fat body), syncytial and neoplastic types, morphology: Kokorin near Prague

Pleistophora debaisieuxi
Weiser, J.; and Zika, Z., 1974, J. Protozool., v. 21 (3), 477 [Abstract]
Pleistophora debaisieuxi, stages in sporogony

Pleistophora hyperparasitica R. Codreanu, 1967
Codreanu, R.; and Codreanu-Balcescu, D., 1975, J. Protozool., v. 22 (3), 77A-78A [Abstract]
hyperparasitic in Enterocystis rhithrogenae which in turn is parasitizing Rhithrogena semicolorata: environs de la Station Zoologique de Sinaia, Carpates Meridionales, Roumanie

Pleistophora hyperparasitica R. Codreanu, 1967
Codreanu-Balcescu, D.; and Codreanu, R., 1976, J. Protozool., v. 23 (4), 8A [Abstract]
Pleistophora hyperparasitica (hyperparasite in Enterocystis rhithrogenae), ultrastructure: Sinaia, Roumanie

Pleistophora oncoperae sp. n., illus.
Milner, R. J.; and Beaton, C. D., 1977, J. Invert. Path., v. 29 (2), 133-140
pathology
Oncopera alboguttata
O. rufobrunnea
(muscle, fat body, connective tissue of all): all from Ebor, New South Wales, Australia

Pleistophora oncoperae (Milner & Beaton, in press)
Milner, R. J., 1977, J. Austral. Entom., v. 16 (1), 21-26
Pleistophora oncoperae in Oncopera alboguttata and O. rufobrunnea, incidence, mixed infection with virus in O. alboguttata responsible for dramatic decrease in population density: Ebor Dorrigo region of N.S.W.

Pleistophora ovariae
Nagel, M. L.; and Hoffman, G. L., 1977, J. Parasitol., v. 63 (1), 160-162
brief description
Pimephales promelas (ovary): Arkansas

Pleistophora ovariae
Nagel, M. L.; and Summerfelt, R. C., 1977, Progr. Fish Cult., v. 39 (1), 18-23
Pleistophora ovariae in Notemigonus crysoleucas (ovaries), nitrofurazone, efficacy at various drug levels, high levels may inhibit host gonad development: culture ponds adjacent to Lake Carl Blackwell, Oklahoma

Pleistophora schubergi, illus.
Weiser, J., 1976, J. Invert. Path., v. 28 (1), 147-149
microsporidian spores, staining nuclei, modified technique
Mamestra brassicae (exper.)

Pleistophora schubergi
Wilson, G. G., 1977, Canad. Entom., v. 109 (7), 1021-1022
Nosema disstriae, Pleistophora schubergi, possible biological control agents of Malacosoma disstria (exper.), host survival

Pleistophora simulii (Lutz and Splendore)
Ebsary, B. A.; and Bennett, G. F., 1975, Canad. J. Zool., v. 53 (8), 1058-1062
Simulium tuberosum
Simulium venustum
Simulium vittatum
all from insular Newfoundland

Pleistophora simulii (Lutz and Splendore, 1904) Jirovec, 1943, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Pleurocystis eiseniellae n. sp., illus.
Ormieres, R., 1977, Ann. Parasitol., v. 52 (2), 109-116
life cycle
Eiseniella tetraedra (coelome): environs de Sete, France

Pleurozyga buetschlii Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria buetschlii (Mingazzini, 1891) Ormieres, 1965 emend.

Pleurozyga clavellinae (von Kolliker, 1848) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria clavellinae (von Kolliker, 1848)

Pleurozyga distapliae Mingazzini, 1891
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria distapliae (Mingazzini, 1891) Ormieres, 1965

Pleurozyga phallusiae (von Kolliker, 1848) Labbe, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
nomen inquirenda

Pleurozyga planariae Mingazzini, 1893
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Monocystella planariae (Mingazzini, 1893) comb. nov.

Plistophora Labbe, 1889 [i.e. 1899]
Baxter, K. N.; Rigdon, R. H.; and Hanna, C., 1970, J. Invert. Path., v. 16 (2), 289-291
[For complete author reference see Supplement 19, Part 1]
as syn. of Pleistophora Gurley, 1893

Plistophora sp.
Rawstron, R. R., 1971, Calif. Fish and Game, v. 57 (4), 253-256
Salmo gairdneri (gills): Darrah Springs Hatchery, Shasta County, California

Plistophora anguillarum, illus.
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Plistophora anguillarum spores, electron microscopy
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Whisnant, J. K.; and Buckley, R. H., 1976, National Cancer Inst. Monograph (43), 211-217
Pneumocystis carinii pneumonia, successful pyrimethamine-sulfadiazine therapy in infants with X-linked immunodeficiency with hyper-IgM

Pneumocystis carinii, illus.
Williams, D. M.; Krick, J. A.; and Remington, J. S., 1976, Am. Rev. Resp. Dis., v. 114 (3), 593-627
parasitic pulmonary infections in the compromised host, immunopathologic aspects, clinical manifestations, diagnosis and treatment, review

Pneumocystis carinii
Wilson, D. E.; and Hirst, S. M., 1977, Wildlife Monogr. (54), Suppl., 3-111
Hippotragus niger: Percy Fyfe Nature Reserve, South Africa

Pneumocystis carinii
Wing, E. J.; and Remington, J. S., 1977, West. J. Med., San Francisco, v. 126 (1), 14-31
role of cell-mediated immunity (and enhancement by levamisole) in host defense against human parasitic diseases, extensive review

Pneumocystis carinii
Wolff, L. J.; et al., 1977, Pediatrics, Am. Acad. Pediat., v. 60 (1), 41-45
Pneumocystis carinii as cause of interstitial pneumonia in immunocompromised children, diagnosis by open lung biopsy, statistics of clinical cases

Pneumocystis carinii
Young, L. S.; and Lau, W. K., 1977, N. England J. Med., v. 296 (1), 48 [Letter]
Pneumocystis carinii pneumonia in immunologically compromised patients, recommendations and suggestions for co-trimoxazole therapy

Pneumocystis carinii, illus.
Young, R. C.; Bennett, J. E.; and Chu, E. W., 1976, Lancet, London (7994), v. 2, 1082-1083
Pneumocystis carinii, human pneumonia, differential diagnosis from similar mimicking diseases

Pneumocystis carinii
Young, R. C.; and DeVita, V. T., jr., 1976, National Cancer Inst. Monograph (43), 193-200
Pneumocystis carinii pneumonia in humans, randomized clinical trial comparing relative merits of pentamidine isethionate and pyrimethamine + sulfadiazine in treatment, comparative toxicity

Pneumocystis jiroveci n. sp., illus.
Frenkel, J. K., 1976, National Cancer Inst. Monograph (43), 13-30
"there is justification to regard Pneumocystis from rats and man as different species. . . that from humans is designated P. jiroveci n. sp. . . . no significant morphologic differences between the two species are recognized; the separation rests on apparent species specificity and serologic differences.", morphology, physiology and immunology in relation to pathology

Polychromophilus sp.
Adam, J. P.; and Landau, I., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 5-6 [Demonstration]
developmental stages in nycteribiid fly infesting insectivorous bats
Miniopterus minor minor
Rhinolophus clivosus sylvestris
R. landeri
R. adami
Hipposideros caffer
Penicillidia fulvida
all from caves in Congo-Brazzaville

Polychromophilus [sp.]
Ewers, W. H., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 88-89 [Demonstration]
Hipposideros galerita
Miniopterus tristis
M. medius
Tadarida sp.
Emballonura sp.
all from New Guinea

Polychromophilus sp.
Landau, I.; et al., 1976, Ann. Parasitol., v. 51 (3), 303-307
Hipposideros bicolor (sang): Limestone Cave (Ulu Gombak), grotte des environs de Kuala Lumpur, Malaisie

Polychromophilus melanipherus
Garnham, P. C. C., 1973, Tr. Roy Soc. Trop. Med. and Hyg., v. 67 (1), 2-3 [Demonstration]
Miniopterus australis: Gomanton Cave, Sabah

Polychromophilus murinus
Garnham, P. C. C., 1973, Tr. Roy Soc. Trop. Med. and Hyg., v. 67 (1), 2-3 [Demonstration]
Myotis horsfieldi: Ampang Water Reservoir, Selangor

Polycycla
Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Urceolariidae

Polymastigina [sp.], illus.
  Jabamoni, R.; et al., 1977, Obst. and Gynec., v. 50 (2), 224-227
    Polymastigina [sp.], transient colonization of human endocervix by multiflagellated protozoan discovered incidentally during evaluation for infertility, case report: Maywood, Illinois

Polyrhabdina Mingazzini, 1891
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Lecudinidae, revision and checklist of species
    diagnosis

Polyrhabdina annulata (Greeff, 1885) Labbe, 1899
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium annulata (Greeff, 1885) comb. nov.

Polyrabdina annulata (Greeff, 1885) Mingazzini, 1893
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium annulata (Greeff, 1885) comb. nov.

Polyrhabdina bifurcata Mackinnon & Ray, 1931 emend. Reichenow, 1932
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Polyrhabdina spionis var. bifurcata Mackinnon & Ray, 1931

Polyrhabdina bogolepovae Levine, 1971
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lecudina bogolepovae (Levine, 1971) comb. nov.

Polyrhabdina brasili Caullery & Mesnil, 1914
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Polyrhabdina cirratuli Bogolepova, 1953
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lecudina bogolepovae (Levine, 1971) comb. nov.

Polyrhabdina cirratuli Mingazzini, 1893 of Labbe (1899) pro parte
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium cirratuli (Lankester, 1866) Caullery & Mesnil, 1899

Polyrhabdina cirratuli (Lankester) of Labbe (1899) pro parte
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium fallax MacGregor & Thomasson, 1965

Polyrhabdina eunicae (Lankester, 1866) Labbe, 1899
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lecudina eunicae (Lankester, 1866) comb. nov.

Polyrhabdina minuta Ganapati, 1946
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Polyrhabdina nereicola Bogolepova, 1953
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lecudina nereicola (Bogolepova, 1953) comb. nov.

Polyrhabdina polydorae (Leger, 1893) Caullery & Mesnil, 1914
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lecudina polydorae (Leger, 1893) Kamm, 1922

Polyrhabdina pygospionis Caullery & Mesnil, 1914
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    nomen nudum

Polyrhabdina sabellae (Lankester, 1863) Labbe, 1899
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium sabellae (Lankester, 1863) Ray, 1930

Polyrhabdina serpulae (Lankester, 1863) Mingazzini, 1893 of Labbe (1899)
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium serpulae (Lankester, 1863) Caullery & Mesnil, 1899

Polyrabdina serpulae (Lankester, 1863) Mingazzini, 1893
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium serpulae (Lankester, 1863) Caullery & Mesnil, 1899

Polyrhabdina spionis (von Kolliker, 1848) Mingazzini, 1891
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    synonymy

Polyrabdina spionis Mingazzini, 1871 pro parte
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Selenidium spionis (von Kolliker, 1845) Ray, 1930

Polyrhabdina spionis var. bifurcata Mackinnon & Ray, 1931
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Polyrhabdina bifurcata Mackinnon & Ray, 1931 emend. Reichenow, 1932

Polyrhabdina vella Johnson, 1968
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    nomen nudum, possibly lapsus calami

Pontesia Hasselmann, 1926
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Lecudinidae, revision and checklist of species
    diagnosis

Pontesia kelschi Hasselmann, 1926
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Porospora pisae Leger & Duboscq, 1911
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Paraophioidina pisae (Leger & Duboscq, 1911) comb. nov.

Prosicuophora macropharyngeus (Bezzenberger, 1904) de Puytorac & Oktem, 1967
Albaret, J.-L., 1973, J. Protozool., v. 20 (1), 51-57
as syn. of Sicuophora macropharyngea (Bezzenberger, 1904) comb. nov.

Proteromonas
Brugerolle, G.; and Joyon, L., 1975, J. Protozool., v. 22 (3), 59A [Abstract]
Proteromonas, Karotomorpha, ultrastructural cytology

Proteromonas lacertae-viridis (Grassi 1879)
Kulda, J., 1973, J. Protozool., v. 20 (4), 536-537
Proteromonas lacertae-viridis, axenic cultivation

Proteromonas lacertaeviridis
Kurashvili, B. E., 1975, Izvest. Akad. Nauk Gruzinsk. SSR, s. Biol., v. 1 (4), 317-320
antagonistic and synergetic interrelationships between intestinal parasites

Protistan, unidentified, likely belonging to subclass Coccidiida, illus.
Wolf, P. H., 1977, J. Invert. Path., v. 29 (2), 244-246
Crassostrea echinata (ova): Northern Territory, Australia

Protococcidiorida Kheisin, 1956
Levine, N. D., 1973, J. Protozool., v. 20 (5), 548-549
Apicomplexa, Sporozoasida; Coccidiasina includes: Eleutheroschizonidae; Grelliidae fam. nov.

Protoopalina
Sandon, H., 1976, Tr. Am. Micr. Soc., v. 95 (3), 357-366
opalinids with special reference to Protoopalina, problems in specific identification, speculations about evolution

Protoopalina pseudonutti n. sp., illus.
Sandon, H., 1976, Tr. Am. Micr. Soc., v. 95 (3), 357-366
opalinids with special reference to Protoopalina, problems in specific identification, speculations about evolution
Bufo regularis: Cairo

Protoopalina stevensoni, illus.
Sandon, H., 1976, Tr. Am. Micr. Soc., v. 95 (3), 357-366
opalinids with special reference to Protoopalina, problems in specific identification, speculations about evolution
Bufo regularis: Cairo; Khartoum

Protoradiophrya armata Rossolimo et Pertzewa, 1929
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya secans (Stein 1859) Stein, 1861

Protoradiophrya fissispiculata Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya fissispiculata (Cheissin, 1930)

Protoradiophrya minuta, Miyashita, 1933
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya minuta (Miyashita), 1933

Protoradiophrya werestchagini [sic], Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Hoplitophrya werestschagini (Cheissin, 1930) Heidenreich, 1935

Protoradiophryopsis Georgevitch, 1964
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Hoplitophryinae

Protoradiophryopsis ochridensis Georgevitch, 1950, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Criodrilus ochridensis

Protozoa
Brooks, M. A., 1976, Invert. Tissue Cult. Research Applic. (Maramorosch), 181-199
parasite transmission, applications of insect tissue culture, review and prospects

Protozoa
Chiodini, P. L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 437 [Demonstration]
description of apparatus for microscopic observation of protozoan cultures

Protozoa
Cordero del Campillo, M.; and Rojo Vazquez, F. A., 1977, An. Fac. Vet. Leon, Oviedo, v. 21 (21), 1975, 33-40
Indice-Catalogo de Zooparasitos Ibericos, I. Protozoos, II. Trematodos, additions and corrections

Protozoa
Diamond, L. S.; and Mattern, C. F. T., 1976, Advances Virus Research, v. 20, 87-112
protozoal viruses, review

Protozoa
Dunlap, J. S., 1970, Dis. Swine (Dunne), 3. ed., 745-762
swine, diagnosis, pathology, control, textbook

Protozoa
Dunlap, J. S., 1975, Dis. Swine (Dunne). 4. ed., 816-832
Protozoa of swine, review

Protozoa, illus.
Ferguson, H. W.; and Adair, B. M., 1977, Vet. Rec., v. 100 (8), 158-159
Protozoa in Salmo gairdneri, possible agent of proliferative kidney disease: northern Irish farm

Protozoa
Hanson, E. D., 1976, J. Protozool., v. 23 (1), 4-12
major evolutionary trends in animal protists

Protozoa
Herkner, H., 1976, Prakt. Tierarzt , v. 57, Sondernummer, 28-30
protozoan parasites of aquarium fishes, control, brief review

Protozoa
Hutner, S. H., 1977, J. Protozool., v. 24 (3), 475-478
essay and reviews of recent symposia on protozoan chemotherapy

Protozoa
Leeflang, P., 1977, Trop. Animal Health and Prod., v. 9 (3), 147-152
protozoa, tick-borne diseases, domestic animals, 1923-1966, historical review: northern Nigeria

Protozoa
Long, P. L., 1974, Folia Vet. Latina, v. 4 (2), 289-300
pathogenic protozoa, cultivation in chicken embryos, review

Protozoa
Marsot, P.; and Couillard, P., 1973, J. Protozool., v. 20 (1), 105-106
use of protamine-coated slides for immobilizing Protozoa for observation

Protozoa
Noble, E. R., 1974, J. Protozool., v. 21 (1), 1-4
3 ecologic approaches to study of protozoan parasitism: studies of energy flow between parasite and host; systems analysis; studies of nutrition through analysis of parasite's environment (Past President's address, Soc. Protozool.)

Protozoa
Ollenschlaeger, B., 1975, Fisch u. Umwelt (1), 35-44
blood parasites of economically important fishes, species, importance, recommendations for therapy, review

Protozoa
Polianskii, I. I.; and Raikov, I. B., 1976, Tr. Am. Micr. Soc., v. 95 (3), 314-326
polymerization and oligomerization phenomena in protozoan evolution

Protozoa
Polianskii, Iu. I.; and Raikov, I. B., 1977, Zhurnal Obshch. Biol., v. 38 (3), 325-335
Protozoa, evolution, polymerization and oligomerization processes in nuclei and organelles, particularly ciliae, theoretical review

Protozoa
Spoon, D. M.; Feise, C. O. II; and Youn, R. S., 1977, J. Protozool., v. 24 (3), 471-474
poly(ethylene oxide), new slowing agent for Protozoa

Protozoa
Vickerman, K., 1973, J. Protozool., v. 20 (4), 529
Protozoa, life cyclical changes in physiology

Protozoa [sp.], resembling Sarcocystis spp.
Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
blood parasites of wild Bovidae, incidence, epizootiological importance
Aepyceros melampus
Damaliscus lunatus
(blood of all): all from Ngamiland, Botswana

Protozoa [sp.], illus.
Daniels, S. B.; Herman, R. L.; and Burke, C. N., 1976, J. Protozool., v. 23 (3), 402-410
unidentified protozoon in Salmo gairdneri (epithelium) exposed for as little as 1 hour to water known to contain infective stages of Myxosoma cerebralis, fine structure and possible life cycle stages, relationship to M. cerebralis remains unresolved

Protozoa [spp.]
Heydorn, A. O.; and Mehlhorn, H., 1977, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 237 (1), 124-137
Protozoa [spp.], two types of giant (globidial) schizonts, light and electron microscopy
sheep (jejunum)

Protozoa [sp.], illus.
O'Brien, D. J.; McArdle, J.; and Doyle, J., 1977, Irish Vet. J., v. 31 (3), 46-48
protozoan-like organism in rainbow trout (kidney), pathology: Co. Wicklow

Protozoa [sp.], unidentified
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Leptobrachium hasselti: Malayan Peninsula

Protozoa [sp.], illus.
Woodard, J. C.; et al., 1977, Vet. Path., v. 14 (4), 338-350 [See also Besnoitia-like organism, Simpson, C. F.; Woodard, J. C.; and Forrester, D. J., 1977]
protozoan, disease syndrome, histopathology, transmission studies unsuccessful
Calidris canutus (aorta, wall of small intestine): St. Petersburg, Florida

Protrichomonadinae Honigberg & Kulova, 1969
Camp, R. R.; Mattern, C. F.; and Honigberg, B. M., 1974, J. Protozool., v. 21 (1), 69-82
Trichomonadida Kirby, emend.; Monocercomonadidae Kirby, emend.

Protrichomonas legeri
Brugerolle, G., 1976, J. Protozool., v. 23 (4), 5A [Abstract]
ultrastructure
Box boops (oesophage)

Pseudolankesteria Ormieres, 1965
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Monocystella Valkanov, 1935

Pseudolankesteria cyclopori (Poisson, 1921) Ormieres, 1965
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Monocystella cyclopori (Poisson, 1921) comb. nov.

Pseudolankesteria planariae (Mingazzini, 1893) Ormieres, 1965
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Monocystella planariae (Mingazzini, 1893) comb. nov.

Pseudolimax butschlii
  Timon-David, P.; and Andrac, A., 1975, Microbia, v. 1 (4), Suppl., 1-8
    amebas in fecal matter, techniques of fecal examination, characters for identification, review

Pseudomonocystis sp.
  Marchal, M., 1976, Compt. Rend. Soc. Biol., Paris, v. 170 (2), 295-299
    Pseudomonocystis sp.-infected Melolontha melolontha, increased susceptibility to exper. infection with low or medium doses of spores of Beauveria brongniartii

Psorospermium sp. Virchow, 1860 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora hominis (Railliet & Lucet, 1891) comb. nov.

Psorospermium sp. Virchow, 1860 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora suihominis nom. nov. [i.e., n. sp.]

Pterospora Labbe & Racovitza, 1897
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Urosporidae
    checklist of species

Pterospora clymenellae (Porter, 1897) Dogiel, 1909
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Pterospora (?) demani Micoletzky, 1921
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    nomen inquirendum

Pterospora petaloprocti n. sp., illus.
  Ormieres, R., 1977, Ann. Parasitol., v. 52 (2), 109-116
    life cycle
    Petaloproctus terricola (coelome): environs de Sete, France

Pythonella
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    key

Pyxinia frenzeli Laveran et Mesnil, illus.
  Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
    description
    Attagenus pellio (intestine): Poland

Pyxinia frenzeli
  Schwalbe, C. P.; and Baker, J. E., 1976, J. Invert. Path., v. 28 (1), 11-15
    Pyxinia frenzeli-infected vs. uninfected Attagenus megatoma, no significant differences in starvation-mediated alterations of nutrient reserves, data suggest P. frenzeli is commensal or mutalist rather than pathogen

Pyxinia rubecula, illus.
  Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
    ultrastructure
    Dermestes lardarius

Radiophrya Rossolimo, 1926
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Radiophryinae nov. comb.

Radiophrya baikalensis Cheissin, 1928
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Anthonyella hoplites (Rossolimo, 1926) Delphy, 1936

Radiophrya biacuta de Puytorac, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Nais obtusa

Radiophrya brasili (Leger et Duboscq, 1904) Heidenreich, 1935 p. p.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Durchoniella brasili (Leger et Duboscq, part. 1894) de Puytorac, 1954

Radiophrya communissima Katashima, 1951
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mimophrya communissima (Katashima 1951) de Puytorac, 1970

Radiophrya deronis Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Dero limosa

Radiophrya enchytraei (Meier, 1954) Lom, 1959
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Enchytraeus buchholzi
E. albidus

Radiophrya gracilis Rossolimo, 1926
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
"peut-etre" as syn. of Radiophrya prolifera (Clap. et Lachm., 1858), Rossolimo, 1926

Radiophrya hoplites Rossolimo, 1926
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Anthonyella hoplites (Rossolimo, 1926) Delphy, 1936

Radiophrya intermedia de Puytorac, 1955
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Helella intermedia (de Puytorac, 1955) Kaczanowski, 1961

Radiophrya limnodrili Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Limnodrilus arenarius

Radiophrya lumbriculi, Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Anthonyella lumbriculi (Cheissin, 1930) Delphy, 1936

Radiophrya naidos (Schmidt, 1846) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Nais elinguis
N. variabilis

Radiophrya ochridana de Puytorac, 1959, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubifex oligosetosus

Radiophrya olivieri de Puytorac, 1954, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Pachydrilus

Radiophrya ovata Katashima, 1951
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Mimophrya ovata (Katashima, 1951) de Puytorac, 1970

Radiophrya pachydrili (Claparede, 1861) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Pachydrilus verrucosus
P. pagenstecheri

Radiophrya peloscolecis Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Peloscolex ferox

Radiophrya prolifera (Clap. et Lachm., 1858), Rossolimo, 1926, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Enchytreus albidus

Radiophrya prolifera Rossolimo, 1926
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya styloscolecis (Heidenreich, 1935)

Radiophrya pungens (Stein, 1861) Heidenreich, 1935
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Tubifex tubifex

Radiophrya rara, Katashima, 1951 [sic]
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Dicontophrya rara (Katashima, 1952 [sic]) de Puytorac, 1970

Radiophrya rossolimoi, Cheissin, 1930
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Juxtaradiophrya rossolimoi (Cheissin, 1930)

Radiophrya sagittata Meier, 1954
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubifex tubifex
T. barbatus

Radiophrya tubificis (Katashima, 1952)
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubifex hattai

Radiophrya tubificis Rossolimo, 1929
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Radiophrya pungens (Stein, 1861) Heidenreich, 1935

Radiophryidae n. fam.
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryoidea nv. super-fam.
includes: Radiophyrinae nov. comb.; Anthonyellinae n. s. fam.; Metaradiophryinae n. s. fam.; Acanthodiophryinae n. s. fam.; Durchoniellinae n. s. fam.; Eudrilophryinae; Metaracoelophryinae n. sous-fm.

Radiophryinae (Rossolimo, 1929) nov. comb.
de Puytorac, Pierre, 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam.
includes: Radiophrya; Mrazekiella; Hellella; Desmophrya; Mimophrya

Radiophryoides Lom, 1956
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Hoplitophryidae, Juxtaradiophryinae n. s. fam.

Radiophryoides aelosomatis (Anderson, 1888) Lom, 1956
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Aelosoma chlorstictum
A. hemprichi

Radiophryoides komarecki Lom, 1956, illus.
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Aelosoma hemprichi

Radiophryopsis Georgevitch, 1941
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Radiophryidae n. fam., Anthonyellinae n.s. fam.

Radiophryopsis acanthostephanos Georgevitch, 1941
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Tubificidae

Ramicephalus sp., illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Holochlora sp.: Khao Yai, Thailande

Ramicephalus albertianus (Theod., Orm. et Jolivet)
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Acrida sp. (caecums digestifs): Cote d'Ivoire

Ramicephalus amphoriformis (Bush)
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Patanga succincta: Lam Narai, Thailande
Trilophidia annulata: Khao Yai, Thailande

Ramicephalus bodenheimeri (Theodorides, Ormieres et Jolivet)
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Homerocoryphus nitidulus ssp. vicinus: Gabon

Ramicephalus ozakii Obata
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Chlaeniellus inops: Coree du sud

Ramicephalus wellmeri (Theodorides), illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Dolichus halensis: Coree du sud

Rasajeyna n. gen.
Beesley, J. E., 1977, Parasitology, v. 74 (3), 273-283
Adeleina, Adeleidae
mt: Rasajeyna nannyla n. gen., n. sp.

Rasajeyna nannyla n. gen., n. sp. (mt), illus.
Beesley, J. E., 1977, Parasitology, v. 74 (3), 273-283
life cycle
Tipula paludosa (epithelial cells of caecum and mid-gut)

Rhizinia oblongata Hammerschmidt, 1838
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stylocephalus oblongatus (Hammerschmidt)

Rhopalonia lithobii sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Lithobius calcaratus (intestine): Siemianice, Poland

Sarcocystidae Poche 1913
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
includes: Sarcocystinae; Toxoplasmatinae
diagnosis

Sarcocystinae Poche 1913
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Sarcocystidae
includes: Sarcocystis; Frenkelia
diagnosis

Sarcocystis
Beier, T. V.; Siim, I. K.; and Hutchison, W. M., 1977, Tsitologiia, v. 19 (12), 1362-1368
Eimeriidae, Isosporinae

Sarcocystis
D'Haese, J.; Mehlhorn, H.; and Peters, W., 1976, Ztschr. Parasitenk., v. 50 (2), 198-199
coccidia, negative staining to show characteristic structures

Sarcocystis
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
"Endorimospora [Tadros and Laarman] cannot be accepted in place of Sarcocystis"

Sarcocystis
Dubey, J. P.; and Streitel, R. H., 1976, J. Parasitol., v. 62 (5), 828-830
Sarcocystis: dogs but not cats shed sporocysts after being fed ovine tissue from Iowa, neither dogs nor cats did with ovine tissue from Ohio although cysts were demonstrated in it; neither dogs nor cats shed after porcine tissue; both dogs and cats shed after bovine tissue

Sarcocystis
Duszynski, D. W.; and Speer, C. A., 1976, Ztschr. Parasitenk., v. 48 (3-4), 191-197
Isospora arctopitheci, I. bigemina, sporozoite excystation, lack of Stieda body, comparison with other Isospora, Eimeria and Sarcocystis species

Sarcocystis
Fayer, R., 1975, Proc. Helminth. Soc. Washington, v. 42 (2), 138-140
Sarcocystis-infected beef obtained at slaughter or from retail food stores, effect of refrigeration, cooking, and freezing on infectivity to dogs, "the potential for transmission of Sarcocystis to humans and their pets by fresh beef clearly exists."

Sarcocystis
Fayer, R.; and Lunde, M. N., 1977, J. Parasitol., v. 63 (3), 438-442
Sarcocystis from dogs, calves (exper.), changes in serum and plasma proteins and in IgG and IgM antibodies

Sarcocystis, illus.
Frelier, P.; et al., 1977, Science (4284), v. 195, 1341-1342
Sarcocystis, 8 yearling dairy heifers, clinical and hematologic findings, histologic demonstration of schizonts, serologic evidence confirmed diagnosis, resident farm dog incriminated as source of infection, possible economic impact of acute sarcocystosis causing poor growth rate or death: Seneca County, central New York State

Sarcocystis Lankester 1882
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Sarcocystidae, Sarcocystinae
diagnosis, life cycle pattern

Sarcocystis
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
synonymy, definition

Sarcocystis Lankester 1882
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Eimeriidae
diagnosis; nomenclature reviewed
Syn.: Hoareosporidium Pande, Bhatia, and Chauhan 1972
"synonymize Endorimospora Tadros and Laarman, 1976 with Sarcocystis in part and Frenkelia in part."

Sarcocystis
Levine, N. D.; and Cechner, P. E., 1973, J. Protozool., v. 20 (4), 510-511
Eimeria spp. and Isospora canis oocysts fed to pigs, no evidence that Sarcocystis infections resulted

Sarcocystis
Levine, N. D.; Cechner, P. E.; and Meyer, R. C., 1974, Indian Vet. J., v. 51 (1), 57-59
pigs fed oocysts of porcine Eimeria spp. and of Isospora canis, "Neither Sarcocystis nor any other protozoon was found, except for Eimeria spp. in the intestinal epithelium of pigs that had been inoculated with porcine Eimeria spp."

Sarcocystis, illus.
Lunde, M. N.; and Fayer, R., 1977, J. Parasitol., v. 63 (2), 222-225
Sarcocystis, soluble antigen prepared from zoites obtained by pepsin digestion techniques, indirect hemagglutination test and agar gel diffusion test in cattle, possible use in diagnosis, antigen did not cross-react with sera of Toxoplasma-positive humans and did not react with sera from Sarcocystis-infected dogs

Sarcocystis, resembling, illus.
Munday, B. L.; and Black, H., 1976, Ztschr. Parasitenk., v. 51 (1), 129-132
protozoans resembling Sarcocystis in brains of bovine foetuses and in placentas from aborted cows, pathological changes

Sarcocystis
Porchet-Hennere, E.; and Vivier, E., 1974, J. Protozool., v. 21 (3), 475 [Abstract]
Sarcocystis endozoite, negative staining, surface and apical structures

Sarcocystis
Supperer, R.; and Leibetseder, J., 1977, Wien. Tierarztl. Monatsschr., v. 64 (6-7), 188-189
Sarcocystis, Toxoplasma, raw meat and slaughtering offals should be cooked to prevent transmission of parasites to dog and cat

Sarcocystis
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
Sarcocystis and related coccidian parasites, general review with discussion of biological aspects of life cycles and new proposal for classification

Sarcocystis, illus.
Viles, J. M.; and Powell, E. C., 1976, Ztschr. Parasitenk., v. 49 (2), 127-132
Sarcocystis, murine, ultrastructure of cyst wall

Sarcocystis [spp.]
Arru, E.; and Cosseddu, A. M., 1974, Atti Soc. Ital. Sc. Vet., v. 28, 857-858
incidence in slaughtered sheep, swine, cattle and Equidae: all from Sardinia

Sarcocystis [sp.]
Ashford, R. W., 1975, J. Protozool., v. 22 (3), 48A [Abstract]
canary fed on feces from Accipiter nisus infected with sporocysts resembling Isospora buteonis was infected with young Sarcocystis cysts when examined 2 months later, postulated that raptor-passerine cycle may exist

Sarcocystis [sp.]
Brown, R. J.; Smith, A. W.; and Keyes, M. C., 1974, J. Wildlife Dis., v. 10 (1), 53
Callorhinus ursinus (masseter muscle): St. Paul Island, Pribilof Islands, Alaska

Sarcocystis spp.
Collins, G. H.; Charleston, W. A. G.; and Moriarty, K. M., 1976, N. Zealand Vet. J., v. 24 (6), 123-124 [Letter]
2 morphologically distinct types of sarcocysts in sheep skeletal muscles may represent separate species, successful transmission to cats

Sarcocystis sp.
Coombs, D. W.; and Springer, M. D., 1974, J. Wildlife Dis., v. 10 (4), 436-441
Sus scrofa domesticus x Sus scrofa cristatus (cardiac and skeletal muscle): Aransas National Wildlife Refuge, southern Texas

Sarcocystis spp., illus.
Cravero, G. C., 1976, Ann. Fac. Med. Vet. Torino, v. 23, 158-164
description
Ovis aries (brain)

Sarcocystis sp.
Davidson, W. R., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 211-217
epizootiologic and pathologic study of endoparasites of selected populations of gray squirrels
Sciurus carolinensis (muscle): Maryland; Georgia; North Carolina; Alabama; South Carolina; Virginia; Tennessee

Sarcocystis sp.
Dissanaike, A. S.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (3), 271 [Letter]
2 species of Sarcocystis demonstrated in Bubalus bubalis (esophageal muscle), sexual stages of S. fusiformis produced in cats (exper.) (small intestine) and of Sarcocystis sp. in dogs (exper.) (small intestine): Malaysia

Sarcocystis spp., illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis spp. Janitschke, Protz and Werner, 1976
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis sp., illus.
Durfee, P. T.; Leksana, S. B.; and Koesharjono, C., 1972, Southeast Asian J. Trop. Med. and Pub. Health, v. 3 (4), 621
swine (muscle tissues of diaphragm): Djakarta, Indonesia (slaughterhouse)

Sarcocystis sp., illus.
Erber, M.; and Boch, J., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (23), 449-450
Sarcocystis sp. found in wild boars (muscles), exper. infection of dogs, Vulpes vulpes, and Canis lupus lupus (sporocysts in feces), exper. infection carried through wild boars and piglets (muscles), and then back to dogs: Bavaria; Poland

Sarcocystis sp., illus.
Fayer, R., 1972, Science (4017), v. 175, 65-67
Sarcocystis sp., development of gametocytes, gametes, and cyst-like bodies in cell culture

Sarcocystis sp., illus.
Fayer, R.; and Thompson, D. E., 1975, J. Parasitol., v. 61 (3), 466-475
Sarcocystis sp., intracellular stages in cell culture, distribution of lipid, carbohydrate, protein, and nucleic acids, correlation with previously observed morphological features

Sarcocystis sp.
Ford, G. E., 1974, Austral. Vet. J., v. 50 (1), 38-39
Sarcocystis sp., prey-predator transmission in ovine sarcosporidiosis, specific pathogen free dogs fed fresh meat from lambs containing cysts

Sarcocystis [sp.]
Ghorbani, M.; Hafizi, A.; and Shegerfcar, T., 1973, Pahlavi Med. J., v. 4 (3), 389-397
pigs (diaphragm): Tehran, Iran

Sarcocystis sp.
Gregory, G. G.; and Munday, B. L., 1976, Austral. Vet. J., v. 52 (7), 317-320
feral cats: Tasmanian Midlands

Sarcocystis sp.
Grundmann, A. W.; and Lombardi, P. S., 1976, Proc. Helminth. Soc. Washington, v. 43 (1), 39-46
Ochotona princeps uinta: Uinta Mountains, Utah

Sarcocystis [sp.]
Hoppe, D. M., 1976, J. Wildlife Dis., v. 12 (1), 27-29
Anas platyrhynchos
A. clypeata
A. acuta
A. americana
A. discors
A. crecca
all from North Dakota

Sarcocystis [sp.]
Howells, R. E.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 438 [Demonstration]
Sarcocystis [sp.] from Dasypus novemcinctus (tongue, skeletal muscle), electron micrographs and histochemistry of cysts

Sarcocystis sp.
Hudkins-Vivion, G.; Kistner, T. P.; and Fayer, R., 1976, J. Wildlife Dis., v. 12 (1), 86-87
coccidia-free Canis familiaris and captive Canis latrans fed Sarcocystis-infected flesh from Bos taurus and Odocoileus h. hemionus, statistical difference in size of resulting sporocysts indicates Sarcocystis from Bos taurus and O. h. hemionus probably constitute distinct species with life cycle dependent on respective ruminant host and a canine host: Oregon

Sarcocystis [sp.], illus.
Kannan Kutty, M.; Das, A. M.; and Dissanaike, A. S., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (3), 400-401
incidental finding of Sarcocystis infection in Orang Asli native (muscle of oropharyngeal region) discovered during autopsy: Malaysia

Sarcocystis [sp.], illus.
Kannan Kutty, M.; and Dissanaike, A. S., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 503-504
human (biopsy specimen from larynx): West Malaysia

Sarcocystis sp., illus.
King, N. W., jr., 1976, Scient. Publication (317). Pan Am. Health Organ., 169-198

Sarcocystis sp. Golemansky and Ridzhakow (1975) of red fox Vulpes vulpes
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
probably as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Sarcocystis sp.
McConnell, E. E.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (6), 851-855
Papio ursinus (muscles): Kruger National Park, Republic of South Africa

Sarcocystis sp., illus.
McConnell, E. E.; et al., 1974, Onderstepoort J. Vet. Research, v. 41 (3), 97-168
pathological and parasitological survey of 100 free-ranging chacma baboons
Papio ursinus (skeletal muscle, tongue, heart): Kruger National Park, Transvaal

Sarcocystis [sp.]
Meads, E. B., 1976, Canad. Vet. J., v. 17 (10), 271 [Letter]
Sarcocystis [sp.], probable cause and probable identification of Protozoa in recent case and 1961 outbreak of Dalmeny disease, cattle: Ontario

Sarcocystis sp., illus.
Mehlhorn, H.; Heydorn, A. O.; and Janitschke, K., 1977, Ztschr. Parasitenk., v. 51 (2), 165-178
cysts, light and electron microscopy
Macaca mulatta (heart and skeletal muscles): research institutes, Schering AG, Berlin (captured in wild)

Sarcocystis sp., illus.
Mehlhorn, H.; Heydorn, A. O.; and Janitschke, K., 1977, Ztschr. Parasitenk., v. 51 (2), 165-178
cysts, light and electron microscopy
Papio cynocephalus (heart and skeletal muscles): research institutes, Schering AG, Berlin (captured in wild)

Sarcocystis sp., illus.
Mehlhorn, H.; Heydorn, A. O.; and Janitschke, K., 1977, Ztschr. Parasitenk., v. 51 (2), 165-178
cysts, light and electron microscopy
Saguinus (= Oedipomidas) oedipus (cardiac and skeletal muscles): Robert Koch Institute, Berlin (originated from Colombia)

Sarcocystis [sp.], illus.
Munday, B. L.; Humphrey, J. D.; and Kila, V., 1977, Avian Dis., v. 21 (4), 697-703
Sarcocystis [sp.], fowl, severe myositis, prevalence, pathology, successful transmission to a dog
domestic fowl (muscle): Papua New Guinea; Australia
dog (jejunum, feces) (exper.)

Sarcocystis [sp.]
O'Farrell, T. P., 1975, Am. Midland Naturalist, v. 93 (2), 377-387
Perognathus parvus
Peromyscus maniculatus
(muscles of all): all from Arid Lands Ecology Reserve, Benton County, Washington

Sarcocystis [sp.], illus.
Powell, E. C.; and McCarley, J. B., 1975, J. Parasitol., v. 61 (5), 928-931
murine sarcocysts fed to cats produced isosporan infection, resulting oocysts fed to mice produced sarcocysts, sarcocysts from exper.-infected mice fed to other cats again produced Isospora felis-like infection

Sarcocystis [sp.]
Prathap, K., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 615 [Letter]
Macaca irus (muscle): West Malaysia

Sarcocystis [sp.], strong resemblance to cyst of S. booliati, illus.
Prathap, K.; and Dissanaike, A. S., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 489
human, cyst observed in muscle biopsy specimen from student with ischaemic contracture: Penang, Malaysia

Sarcocystis sp.
Prestwood, A. K.; Pursglove, S. R.; and Hayes, F. A., 1976, J. Wildlife Dis., v. 12 (3), 380-385
survey of parasites of Odocoileus virginianus and Ovis aries on common range, deer unlikely reservoir host for sheep parasites
Ovis aries
Odocoileus virginianus
all from Hardy County, West Virginia

Sarcocystis sp., illus.
Rioseco B., H.; et al., 1976, Arch. Med. Vet., Valdivia, v. 8 (2), 122-123
Sarcocystis sp., Pudu pudu (skeletal and cardiac musculature), pathology: Chile

Sarcocystis [sp.] type A, illus.
Rzepczyk, C.; and Scholtyseck, E., 1976, Ztschr. Parasitenk., v. 50 (2), 137-150
Sarcocystis [sp.] type A; Sarcocystis [sp.] type B, light and electron microscopy, effect on hosts' cells, distinct species; no cellular response apparent in host; possible life histories
Rattus fuscipes (skeletal and subcutaneous muscle): South-East Queensland

Sarcocystis [sp.] type B, illus.
Rzepczyk, C.; and Scholtyseck, E., 1976, Ztschr. Parasitenk., v. 50 (2), 137-150
Sarcocystis [sp.] type A; Sarcocystis [sp.] type B, light and electron microscopy, effect on hosts' cells, distinct species; no cellular response apparent in host; possible life histories
Rattus fuscipes (skeletal and subcutaneous muscle): South-East Queensland

Sarcocystis sp., illus.
Scheuring, W.; and Madej, J. A., 1976, Med. Wet., v. 32 (7), 437-438
nutria (miesniowej): woj. Zielonogorskie

Sarcocystis, resembling S. fusiformis, illus.
Schmitz, J. A.; and Wolf, W. W., 1977, Vet. Path., v. 14 (5), 527-531
spontaneous fatal sarcocystosis in a calf (liver, kidney, lungs, heart), pathological changes: north central Oregon

Sarcocystis spp.
Seneviratna, P.; Edward, A. G.; and DeGiusti, D. L., 1975, Am. J. Vet. Research, v. 36 (3), 337-339
Sarcocystis spp., frequency of infection in feral and domesticated animals, high frequency of infection in sheep in summer
sheep
cattle
pigs
Didelphis virginiana
Procyon lotor
Quiscalus quiscula
all from Detroit metropolitan area, Michigan

Sarcocystis sp., illus.
Sibalic, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 148-152
humans with high Sabin-Feldman dye test titers thought to have toxoplasmosis found apparently also infected with Sarcocystis spp., specimens of their blood were injected into mice and Citellus citellus and Sarcocystis organisms recovered as cystozoites in peritoneal exudate or cysts in musculature; further verification positive using Sarcocystis fusiformis antigen for indirect immunofluorescence on human blood

Sarcocystis species
Tadros, W.; and Laarman, J. J., 1977, Trop. and Geogr. Med., v. 29 (3), 315 [Abstract]
epidemiology and epizootiology of Sarcocystis and related cyst-forming coccidia

Sarcocystis sp., illus.
Vetterling, J. M.; Pacheco, N. D.; and Fayer, R., 1973, J. Protozool., v. 20 (5), 613-621
Sarcocystis sp., fine structure of gametogony and oocyst formation

Sarcocystis aucheniae
Castro, J., 1974, Rev. Invest. Pecuarias, v. 3 (1), 91-92
Sarcocystis aucheniae, incidence in Lama glama (corazon, diafragma, esofago, pierna): Provincia de Puno

Sarcocystis bertrami
Arru, E.; and Cosseddu, A. M., 1976, Clin. Vet., Milano, v. 99 (7), 322-327
incidence in regularly slaughtered animals
donkeys
horses
all from Sardinia

Sarcocystis bertrami, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review
Syn.: Sarcocystis equicanis

Sarcocystis bertrami, Doflein, 1901
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis bertrami Doflein 1901, illus.
Dubey, J. P.; et al., 1977, J. Parasitol., v. 63 (3), 443-447
Syn.: Sarcocystis equicanis Rommel and Geisel 1975

Sarcocystis bertrami Doflein, 1901
Gadaev, A.; and Abidzhanov, A. A., 1976, Uzbek. Biol. Zhurnal (4), 57-59
brief description
[Equus caballus]: meat packing plants in Tashkent and Karshi, Uzbekistan

Sarcocystis bertrami Doflein 1901
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: 'Isospora rivolta' sporocysts of Gassner (1940) and Levine and Ivens (1965) pro parte; 'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) and Heydorn, Mehlhorn, and Gestrich (1975) pro parte; Sarcocystis equicanis Rommel and Geisel 1975; perhaps Hoareosporidium pellerdyi Pande, Bhatia, and Chauhan 1972
Equus caballus
Asinus asinus
Canis familiaris

Sarcocystis besnoiti Marotel 1912
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Besnoitia besnoiti (Marotel 1912) Henry 1913

Sarcocystis besnoiti Marotel, 1912
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora besnoiti (Marotel, 1912) comb. nov.

Sarcocystis bigemina (Stiles 1891) comb. n.
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Coccidium bigeminum Stiles 1891; Coccidium bigeminum var. canis Railliet and Lucet 1891; Isospora bigemina (Stiles 1891) Luhe 1906 pro parte; Lucetina bigemina (Stiles 1891) Henry and Leblois, pro parte
Canis familiaris

Sarcocystis blanchardi
Arru, E.; and Cosseddu, A. M., 1976, Clin. Vet., Milano, v. 99 (7), 322-327
incidence in regularly slaughtered animals
cattle: Sardinia

Sarcocystis blanchardi, Doflein, 1901, pro parte
Heydorn, A.-O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovicanis n. spec.

Sarcocystis blanchardi, Doflein, 1901, pro parte
Heydorn, A.-O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovifelis n. spec.

Sarcocystis blanchardi, Doflein, 1901, pro parte
Heydorn, A.-O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovihominis n. spec.

Sarcocystis blanchardi Doflein 1901
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
presumably as syn. of Sarcocystis fusiformis Railliet 1897

Sarcocystis booliati, illus.
Kan, S. P.; and Dissanaike, A. S., 1976, Internat. J. Parasitol., v. 6 (4), 321-326
Sarcocystis booliati, zoites and cyst wall, ultrastructure

Sarcocystis bovicanis n. spec.
Heydorn, A.-O.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
Syns.: Sarcocystis fusiformis, Railliet, 1897, pro parte; S. blanchardi, Doflein, 1901, pro parte; Miescheria cruzi, Hasselmann, 1926, pro parte; Isospora bigemina, Stiles, 1891; Luhe 1906, pro parte
Bos taurus
Canis lupus
C. latrans
Vulpes vulpes
Canis familiaris

Sarcocystis bovicanis
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
as syn. of Sarcocystis cruzi

Sarcocystis bovicanis
Heydorn, A. O., 1977, Arch. Lebensmittel.-Hyg., v. 28 (1), 27-31
Sarcocystis bovihominis, S. bovicanis, exper. human infection from raw beef, minor symptoms; S. suihominis, exper. human infection from raw pork, severe symptoms, parasite pathogenic or toxic

Sarcocystis bovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Sarcocystis bovicanis
Nedjari, T.; Jungmann, R.; and Hiepe, T., 1976, Monatsh. Vet.-Med., v. 31 (24), 946-947
Sarcocystis bovicanis, cattle (exper.), indirect fluorescent antibody test proved applicable for diagnosis

Sarcocystis bovicanis Heydorn, Gestrich, Mehlhorn & Rommel, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hirsuta (Moule, 1887) comb. nov.

Sarcocystis bovifelis n. spec.
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
Syns.: Sarcocystis fusiformis, Railliet, 1897, pro parte; S. blanchardi, Doflein, 1901, pro parte; Miescheria cruzi, Hasselmann, 1926, pro parte; Isospora bigemina, Stiles 1891, Luhe 1906, pro parte
". . . according to International Nomenclature Rules S. fusiformis and S. tenella constitute a pro parte situation and the old names must be retained for two selected species. In this case we would prefer to denote the cat-cattle and cat-sheep species with the old names adding the new names in brackets: S. fusiformis (bovifelis) and S. tenella (ovifelis)."
Bos taurus
Felis catus
F. silvestris

Sarcocystis bovifelis
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
as syn. of Sarcocystis hirsuta

Sarcocystis bovifelis Heydorn et al., 1975
Heydorn, A.-O.; Gestrich, R.; and Janitschke, K., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (6), 116-120
Sarcocystis bovifelis, material from infected calf failed to infect experimental rhesus monkey

Sarcocystis bovifelis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hirsuta Moule 1888

Sarcocystis bovihominis n. spec.
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
Syns.: Sarcocystis fusiformis, Railliet, 1897, pro parte; S. blanchardi, Doflein, 1901, pro parte; Miescheria cruzi, Hasselmann, 1926, pro parte; Isospora hominis, Railliet, Lucet, 1891, pro parte
Bos taurus
Homo sapiens
Macaca rhesus
Papio cynocephalus

Sarcocystis bovihominis
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
as syn. of Sarcocystis hominis

Sarcocystis bovihominis, illus.
Heydorn, A. O., 1977, Arch. Lebensmittel.-Hyg., v. 28 (1), 27-31
Sarcocystis bovihominis, S. bovicanis, exper. human infection from raw beef, minor symptoms; S. suihominis, exper. human infection from raw pork, severe symptoms, parasite pathogenic or toxic

Sarcocystis bovihominis
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
Sarcocystis bovihominis failure to infect pigs proves that it is a different species from S. suihominis n. sp.

Sarcocystis bovihominis Heydorn et al., 1975, illus.
Heydorn, A.-O.; Gestrich, R.; and Janitschke, K., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (6), 116-120
Sarcocystis bovihominis, sporocysts in feces of rhesus monkey, baboons, and humans, all experimentally infected with material from calf
Macaca rhesus
Papio cynocephalus
humans
(feces of all)

Sarcocystis bovihominis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hominis (Railliet and Lucet 1891) comb. n.

Sarcocystis bovihominis Heydorn, Gestrich, Mehlhorn & Rommel, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hominis (Railliet & Lucet, 1891) comb. nov.

Sarcocystis bubali Willey, Chalmers, and Marshall 1904
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis fusiformis Railliet 1897

Sarcocystis bubalis Dogiel 1916
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis fusiformis Railliet 1897

Sarcocystis cruzi, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review
Syn.: Sarcocystis bovicanis

Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis cruzi
Fayer, R., 1977, J. Parasitol., v. 63 (6), 1072-1075
Sarcocystis cruzi, parameters of sporocyst production by dogs fed experimentally and naturally infected beef: prepatent period, time of maximum production, numbers shed per dog and per gram of feces

Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Miescheria cruzi Hasselmann 1926 pro parte; Sarcocystis fusiformis Railliet 1897 of Babudieri (1932) and auctores, pro parte, (non) S. fusiformis (Railliet 1897) Bernard and Bauche 1912; probably Sarcocystis iturbei Vogelsang 1938; Sarcocystis marcovi Vershinin 1975 pro parte; Sarcocystis bovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975; 'free sporocysts of Isospora rivolta' of Gassner (1940) and Levine and Ivens (1965) pro parte; 'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) and Heydorn, Mehlhorn, and Gestrich (1975) pro parte; '?Cryptosporidium' of Bearup (1954) pro parte; probably Sarcocystis sp. Golemansky and Ridzhakow (1975) of red fox Vulpes vulpes; probably Cryptosporidium vulpis Wetzel 1938
Bos taurus
Canis familiaris
Canis latrans
Canis lupus
Vulpes vulpes
Procyon lotor
Hyaena crocuta

Sarcocystis cruzi, illus.
Pacheco, N. D.; and Fayer, R., 1977, J. Protozool., v. 24 (3), 382-388
Sarcocystis cruzi, fine structure of schizonts, method of schizogonic development

Sarcocystis cruzi Hasselmann, 1926 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora cruzi (Hasselmann, 1926) comb. nov.

Sarcocystis cuniculi Brumpt, 1913
Tadros, W.; and Laarman, J. J., 1977, Proc. K. Nederl. Akad. Wetensch., s. C, v. 80 (4), 351-352
Sarcocystis cuniculi in Felis catus after being fed sarcocyst-infected muscle from Oryctolagus cuniculus

Sarcocystis diminuta sp. nov., illus.
Howells, R. E.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (4), 463-474
morphology, histochemistry
Dasypus novemcinctus (skeletal and tongue muscle): Entre Rios, Minas Gerais, Brazil

Sarcocystis equicanis n. sp., illus.
Rommel, M.; and Geisel, O., 1975, Berl. u. Munchen. Tierarztl. Wchnschr., v. 88 (24), 468-471
Sarcocystis equicanis, incidence, stages in horse and dog, life cycle
horse (oesophageal muscles): Schlachthof Munchen
dog (feces) (exper.)

Sarcocystis equicanis
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
as syn. of Sarcocystis bertrami

Sarcocystis equicanis Rommel and Geisel 1975
Dubey, J. P.; et al., 1977, J. Parasitol., v. 63 (3), 443-447
as syn. of Sarcocystis bertrami Doflein 1901

Sarcocystis equicanis
Goebel, E., 1976, Ztschr. Parasitenk., v. 50 (2), 201
Sarcocystis equicanis, cyst stages, fine structure

Sarcocystis equicanis Rommel and Geisel 1975
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis bertrami Doflein 1901

Sarcocystis equicanis Rommel & Geisel, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora bertrami Doflein, 1901 comb. nov.

Sarcocystis fayeri, Dubey, Streitel, Stromberg and Toussant, 1977 [nom. nud.]
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis fayeri sp. n., illus.
Dubey, J. P.; et al., 1977, J. Parasitol., v. 63 (3), 443-447
horses (heart, diaphragm, esophagus): meat packing plant, Creston, Ohio
dogs (exper.) (feces, small intestine)

Sarcocystis fusiformis, illus.
Anpilogova, N. V., 1976, Dokl. Akad. Nauk Tadzhiksk. SSR, v. 19 (2), 65-66
description of sporocysts
Vulpes vulpes (feces): khr. Rangontau
Alopex lagopus (feces): fur farm preserve "Ramit"

Sarcocystis fusiformis
Aryeetey, M. E.; and Piekarski, G., 1976, Ztschr. Parasitenk., v. 50 (2), 109-124
Sarcocystis fusiformis-infected raw meat fed to humans, Isospora hominis sporocysts excreted in feces; no immunity to reinfection manifested as reduced number of sporocysts; antibodies detected by indirect immunofluorescence test, also present in survey of population (common in adults, present in children over 6 months of age); rats fed infected sausage showed negative reaction to Sarcocystis antigen

Sarcocystis fusiformis, illus.
Chauhan, P. P. S.; et al., 1977, Indian J. Exper. Biol., v. 15 (6), 492-494
Sarcocystis fusiformis cysts obtained from buffaloes, gametogonic development in dogs (exper.)

Sarcocystis fusiformis (Railliet, 1897)
Dissanaike, A. S.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (3), 271 [Letter]
2 species of Sarcocystis demonstrated in Bubalus bubalis (esophageal muscle), sexual stages of S. fusiformis produced in cats (exper.) (small intestine) and of Sarcocystis sp. in dogs (exper.) (small intestine): Malaysia

Sarcocystis fusiformis, illus.
Fayer, R.; and Johnson, A. J., 1974, Proc. Helminth. Soc. Washington, v. 41 (1), 105-108
Sarcocystis fusiformis, calves infected with sporocysts from dogs, development of cysts

Sarcocystis fusiformis
Fayer, R.; and Johnson, A. J., 1975, J. Parasitol., v. 61 (5), 932-936
Sarcocystis fusiformis, calves (exper.), amprolium reduced severity of acute sarcocystosis (as measured by overall weight gains, packed cell volume, body temperature, postmortem and histological examinations, and serum enzyme levels)

Sarcocystis fusiformis
Fayer, R.; Johnson, A. J.; and Hildebrandt, P. K., 1976, J. Parasitol., v. 62 (1), 10-14
Sarcocystis fusiformis, narrow specificity of asexual stages for bovine intermediate host, wider but restricted host range of sexual stages for coyotes, dogs, foxes, and raccoons
bovine (nat. and exper.)
Canis familiaris (exper.)
Vulpes fulva (exper.)
Procyon lotor (exper.)
coyote

Sarcocystis fusiformis
Fayer, R.; Johnson, A. J.; and Lunde, M., 1976, J. Infect. Dis., v. 134 (6), 624-628
Sarcocystis fusiformis, field bred cows (exper.), clinical and pathological signs of infection, occurrence of abortions, no evidence of congenital transmission to calves or fetuses; Sarcocystis may play significant role in bovine abortions

Sarcocystis fusiformis
Fayer, R.; Mahrt, J. L.; and Johnson, A. J., 1973, J. Protozool., v. 20 (4), 509
Sarcocystis fusiformis, life cycle and pathogenic effects in calves (exper.)

Sarcocystis fusiformis Moule, 1887
Gadaev, A.; and Abidzhanov, A. A., 1976, Uzbek. Biol. Zhurnal (4), 57-59
brief description
[Bos taurus]: meat packing plants in Tashkent and Karshi, Uzbekistan

Sarcocystis fusiformis, Railliet, 1897, pro parte
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovicanis n. spec.

Sarcocystis fusiformis, Railliet, 1897, pro parte
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovifelis n. spec.

Sarcocystis fusiformis, Railliet, 1897, pro parte
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis bovihominis n. spec.

Sarcocystis fusiformis, illus.
Heydorn, A. O.; Mehlhorn, H.; and Gestrich, R., 1975, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 233 (1), 123-137
Sarcocystis fusiformis, calves (exper.), infected with oocysts and sporocysts of large form of Isospora bigemina from dogs, fine structure of cyst stages

Sarcocystis fusiformis
Johnson, A. J.; Hildebrandt, P. K.; and Fayer, R., 1975, Am. J. Vet. Research, v. 36 (7), 995-999
Sarcocystis fusiformis, calves fed sporocysts isolated from coyotes and dogs, clinicopathologic manifestations, macroscopic observations, histopathologic changes

Sarcocystis fusiformis Railliet 1897
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: presumably S. blanchardi Doflein 1901; S. siamensis von Linstow 1903; S. bubali Willey, Chalmers, and Marshall 1904; S. bubalis Dogiel 1916

Sarcocystis fusiformis Railliet 1897 of Babudieri (1932) and auctores, pro parte (non) S. fusiformis (Railliet 1897) Bernard and Bauche 1912
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Sarcocystis fusiformis Railliet 1897 of Babudieri (1932) and auctores, pro parte (non) S. fusiformis (Railliet 1897) Bernard and Bauche 1912
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hirsuta Moule 1888

Sarcocystis fusiformis Railliet 1897 of Babudieri (1932) and auctores, pro parte, (non) S. fusiformis Railliet 1897 Bernard and Bauche, 1912
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hominis (Railliet and Lucet 1891) comb. n.

Sarcocystis fusiformis
Mahrt, J. L.; and Fayer, R., 1975, J. Parasitol., v. 61 (5), 967-969
Sarcocystis fusiformis, calves (exper.), acute phase of infection, hematologic and serum enzyme changes: oligocythemic anemia, leukocytic shift to the left, elevation of serum SGOT, LDH, and CPK levels

Sarcocystis fusiformis
Markus, M. B.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 3 [Demonstration]
Sarcocystis fusiformis, attempted infection of chimpanzees and cats, one cat shed sporocysts, all animals sero-negative to indirect fluorescent antibody test

Sarcocystis fusiformis Railliet, 1897
Mehlhorn, H.; Heydorn, O.; and Gestrich, R., 1975, J. Protozool., v. 22 (3), 83A [Abstract]
calves infected with Isospora hominis sporocysts or I. bigemina large forms from dog or cat, development of Sarcocystis fusiformis cysts in musculature, each of 3 species of Isospora forms characteristic cysts, "Ainsi, il semble prouve que les kystes de Sarcocystis fusiformis Railliet, 1897 font partie des cycles de developpement d'au moins trois especes differentes de Coccidies."

Sarcocystis fusiformis
Sibalic, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 148-152
humans with high Sabin-Feldman dye test titers thought to have toxoplasmosis found apparently also infected with Sarcocystis spp., specimens of their blood were injected into mice and Citellus citellus and Sarcocystis organisms recovered as cystozoites in peritoneal exudate or cysts in musculature; further verification positive using Sarcocystis fusiformis antigen for indirect immunofluorescence on human blood

Sarcocystis fusiformis Railliet, 1897 (as used by numerous authors in later literature) pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora cruzi (Hasselmann, 1926) comb. nov.

Sarcocystis fusiformis Railliet, 1897 (as used by numerous authors in later literature) pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hirsuta (Moule, 1887) comb. nov.

Sarcocystis fusiformis Railliet, 1897 (as used by numerous authors in later literature) pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hominis (Railliet & Lucet, 1891) comb. nov.

Sarcocystis fusiformis, illus.
Zaman, V.; and Colley, F. C., 1972, Southeast Asian J. Trop. Med. and Pub. Health, v. 3 (4), 489-495
Sarcocystis fusiformis, fine structure, observation of asexual reproduction by endodyogeny
Bubalus bubalis (esophagus): Singapore

Sarcocystis fusiformis (bovifelis)
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
". . . according to International Nomenclature Rules S. fusiformis and S. tenella constitute a pro parte situation and the old names must be retained for two selected species. In this case we would prefer to denote the cat-cattle and cat-sheep species with the old names adding the new names in brackets: S. fusiformis (bovifelis) and S. tenella (ovifelis)."

Sarcocystis fusiformis (bovifelis) Heydorn, Gestrich, Mehlhorn & Rommel, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora cruzi (Hasselmann, 1926) comb. nov.

Sarcocystis gigantea (Railliet, 1886)
Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
"Balbiania gigantea by Railliet, is now generally recognized as being a Sarcocystis, and the name Sarcocystis gigantea (Railliet, 1886) should be applied to it."
Syn.: S. ovifelis Heydorn et al., 1975

Sarcocystis hemionilatrantis, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis hemionilatrantis Hudkins and Kistner, 1977
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
"proper name for the organism in mule deer"

Sarcocystis hemionilatrantis, illus.
Koller, L. D.; Kistner, T. P.; and Hudkins, G. G., 1977, Am. J. Vet. Research, v. 38 (8), 1205-1209
Sarcocystis hemionilatrantis in Odocoileus hemionus hemionus fawns (tissue and perivascular spaces in skeletal and cardiac muscle; macrophages) (exper.), macroscopic and histopathological lesions

Sarcocystis hirsuta, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review
Syn.: Sarcocystis bovifelis

Sarcocystis hirsuta, Moule, 1886
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis hirsuta Moule 1888
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Miescheria cruzi Hasselmann 1926 pro parte; Sarcocystis fusiformis Railliet 1897 of Babudieri (1932) and auctores, pro parte, (non) S. fusiformis (Railliet 1897) Bernard and Bauche 1912; Sarcocystis marcovi Vershinin 1975 pro parte; Sarcocystis bovifelis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Bos taurus
Felis catus
Felis silvestris

Sarcocystis hirsuta Moule, 1887
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora hirsuta (Moule, 1887) comb. nov.

Sarcocystis hominis [? n. comb.]
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Syn.: Sarcocystis bovihominis
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis hominis (Railliet and Lucet 1891) comb. n.
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Miescheria cruzi Hasselmann 1926 pro parte; Sarcocystis fusiformis Railliet 1897 of Babudieri (1932) and auctores, pro parte, (non) S. fusiformis Railliet 1897 Bernard and Bauche, 1912; Coccidium bigeminum var. hominis Railliet and Lucet 1891 pro parte; Lucetina hominis (Railliet and Lucet, 1891) Henry and Leblois, 1926 pro parte; Isospora hominis (Railliet and Lucet 1891) Wenyon 1923 pro parte; Sarcocystis bovihominis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Bos taurus
Homo sapiens
Macaca mulatta
Papio cynocephalus
?Pan troglodytes

Sarcocystis iturbei Vogelsang 1938
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
probably as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Sarcocystis kortei
Prosl, H., 1976, Ztschr. Parasitenk., v. 50 (2), 214
Rhesusaffe

Sarcocystis leporum, Crawley 1914
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis lindemanni, illus.
Jeffrey, H. C., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 17-29
Sarcocystis lindemanni, differential diagnosis from Toxoplasma gondii, 2 case reports of infection in man with critical evaluation of previously reported cases: Nepal

Sarcocystis marcovi Vershinin 1975 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis cruzi (Hasselmann, 1926) Wenyon, 1926

Sarcocystis marcovi Vershinin 1975 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis hirsuta Moule 1888

Sarcocystis marcovi Vershinin, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
"Vershinin (1975) erected a new species S[arcocystis] marcovi . . . . We believe Vershinin was dealing with a mixed infection with E[ndorimospora] hirsuta + E. cruzi."

Sarcocystis miescheri, Lankester, 1882, pro parte
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
as syn. of Sarcocystis suihominis n. spec.

Sarcocystis miescheri Lankester 1882
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis miescheriana (Kuhn 1865) Labbe 1899

Sarcocystis miescheriana
Arru, E.; and Cosseddu, A. M., 1976, Clin. Vet., Milano, v. 99 (7), 322-327
incidence in regularly slaughtered animals
swine: Sardinia

Sarcocystis miescheriana
Cironeanu, I., 1975, Rev. Crest. Animalelor, v. 25 (2), 81-82
Sarcocystis miescheriana, Trichinella spiralis, differential diagnosis of cysts

Sarcocystis meischeriana, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis miescheriana (Kuehn, 1865) Lankester, 1882
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis meischeriana Kuhn, 1865
Gadaev, A.; and Abidzhanov, A. A., 1976, Uzbek. Biol. Zhurnal (4), 57-59
brief description
[Sus scrofa]: meat packing plants in Tashkent and Karshi, Uzbekistan

Sarcocystis miescheriana
Greve, E., 1973, Norwegian J. Zool., v. 21 (4), 329 [Abstract]
Sarcocystis miescheriana in pigs, seasonal variation: slaughterhouse in Roskilde, Denmark

Sarcocystis miescheriana
Greve, E., 1975, Medlemsbl. Danske Dyrlaegeforen., v. 58 (18), 689-693
Sarcocystis miescheriana, swine, method of diagnosis at time of slaughter

Sarcocystis miescheriana, Kuehn, 1865, pro parte
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
as syn. of Sarcocystis suihominis n. spec.

Sarcocystis miescheriana (Kuhn 1865) Labbe 1899
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Synchytrium miescherianum Kuhn 1865; Sarcocystis miescheri Lankester 1882; Coccidium bigemina var. canis Railliet and Lucet 1891 pro parte; 'Isospora rivolta' sporocysts of Gassner (1940) and Levine and Ivens (1965) pro parte; 'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) and Heydorn, Mehlhorn, and Gestrich (1975) pro parte
Sus scrofa
Canis familiaris

Sarcocystis miescheriana
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Sus scrofa: Chile

Sarcocystis miescheriana (Kuhn, 1865) Lankester, 1882 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora miescheriana (Kuhn, 1865) comb. nov.

Sarcocystis miescheriana (Kuhn, 1865) Lankester, 1882 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora suihominis nom. nov. [i.e., n. sp.]

Sarcocystis miescheriana
Wojcik, A. R.; and Grzywinski, L., 1975, Medycyna Wet., v. 31 (10), 597-598
incidence in swine, economic losses: Torun slaughterhouse

Sarcocystis moulei Neveu-Lemaire, 1912
Gadaev, A.; and Abidzhanov, A. A., 1976, Uzbek. Biol. Zhurnal (4), 57-59
brief description
[Capra hircus]: meat packing plants in Tashkent and Karshi, Uzbekistan

Sarcocystis muris, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis muris (Blanchard, 1885) Labbe, 1899
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis muris (Blanchard 1885) Labbe 1899
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Miescheria muris Blanchard 1885; Coccidium bigeminum var. cati Railliet and Lucet 1891; Sarcocystis musculi Blanchard 1885 of Kalyakhin and Zasukhin 1975 lapsus calami
Mus musculus
Rattus norvegicus
Rattus rattus
Microtus arvalis
Microtus agrestis
Clethrionomys glareolus
Oryzomys goeldi
Felis catus

Sarcocystis muris, "similar to"
Powell, E. C.; and McCarley, J. B., 1975, J. Parasitol., v. 61 (5), 928-931
murine sarcocysts fed to cats produced isosporan infection, resulting oocysts fed to mice produced sarcocysts, sarcocysts from exper.-infected mice fed to other cats again produced Isospora felis-like infection

Sarcocystis muris, illus.
Ruiz, A.; and Frenkel, J. K., 1976, J. Infect. Dis., v. 133 (4), 409-418
Sarcocystis muris, obligatory 2-host life cycle (mouse-cat), development, pathology
Mus musculus (nat. and exper.) (liver, skeletal muscle): Costa Rica
cats (exper.) (intestine, feces)

Sarcocystis muris, illus.
Sheffield, H. G.; Frenkel, J. K.; and Ruiz, A., 1977, J. Parasitol., v. 63 (4), 629-641
Sarcocystis muris, mice, development of parasite, cyst wall, and parasitized muscle cells during course of infection and in correlation with potential infectivity of cyst organisms, ultrastructural study

Sarcocystis muris (Blanchard, 1885) Labbe, 1899
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora muris (Blanchard, 1885) comb. nov.

Sarcocystis musculi Blanchard 1885 of Kalyakhin and Zasukhin 1975 lapsus calami
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis muris (Blanchard 1885) Labbe 1899

Sarcocystis nesbitti
Prosl, H., 1976, Ztschr. Parasitenk., v. 50 (2), 214
Rhesusaffe

Sarcocystis orientalis sp. n., illus.
Zaman, V.; and Colley, F. C., 1975, Ztschr. Parasitenk., v. 47 (3), 169-185
Sarcocystis orientalis sp. n., life cycle, light and electron microscopy
Python reticulatus (small intestine, intestinal epithelium): Chinese butcher shop, Singapore
Rattus norvegicus (exper.) (blood, lungs, striated muscle)

Sarcocystis orientalis Zaman and Colley, 1975
Zaman, V., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (4), 603
Sarcocystis orientalis, redescription and revision of new species diagnosis

Sarcocystis orientalis Zaman and Colley, 1975
Zaman, V., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (1), 112
Sarcocystis orientalis, study of host range shows S. orientalis to be a highly pathogenic parasite of Rattus genus, not infective for common domestic and laboratory animals, possible use in biological control of rats
Rattus norvegicus (exper.)
Python reticulatus (caecal contents) (exper.)

Sarcocystis orientalis Zaman and Colley, 1975
Zaman, V.; and Colley, F. C., 1976, Ztschr. Parasitenk., v. 51 (1), 137
preoccupied by S. orientalis Machulskii and Miskaryan, 1958, renamed S. singaporensis sp. n. [i.e., nom. nov.]

Sarcocystis ovicanis n. spec.
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
Syns.: Sarcocystis tenella, Railliet, 1886, pro parte; Isospora bigemina, Stiles, 1891, Luhe, 1906, pro parte
Ovis aries
Canis familiaris

Sarcocystis ovicanis Heydorn et al., 1975
Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
as syn. of Sarcocystis tenella (Railliet, 1886)

Sarcocystis ovicanis, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis ovicanis, Heydorn, Gestrich, Mehlhorn and Rommel, 1975
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis ovicanis Heydorn et al., 1975, illus.
Heydorn, A.-O.; and Gestrich, R., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (1), 1-5
Sarcocystis ovicanis, developmental stages in lambs, lamb muscles infectious for experimental dogs

Sarcocystis ovicanis, illus.
Leek, R. G.; and Fayer, R., 1978, Cornell Vet., v. 68 (1), 108-123
Sarcocystis ovicanis-induced abortion in sheep (exper.), clinical signs, pathology, no evidence of intrauterine transmission

Sarcocystis ovicanis, illus.
Leek, R. G.; Fayer, R.; and Johnson, A. J., 1977, J. Parasitol., v. 63 (4), 642-650
Sarcocystis ovicanis, lambs (exper.) (cardiac and skeletal muscles, brain parenchyma, optic nerve, vascular endothelial cells), clinical data, gross and histopathologic findings; unsuccessful attempts to infect cats and simultaneous successful infection of dogs with parts of same naturally infected sheep tissue

Sarcocystis ovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Sarcocystis tenella Railliet 1886 pro parte; 'free sporocysts of Isospora rivolta' of Gassner (1940) and Levine and Ivens (1965) pro parte; 'large form of Isospora bigemina' of Mehlhorn, Heydorn, and Gestrich (1975) pro parte; '?Cryptosporidium' of Bearup (1954) pro parte; probably Hoareosporidium pellerdyi Pande, Bhatia, and Chauhan 1972
Ovis aries
Canis familiaris

Sarcocystis ovicanis
Mehlhorn, H.; and Heydorn, A. O., 1976, Ztschr. Parasitenk., v. 50 (2), 209-210
Sarcocystis ovicanis, S. ovifelis, cyst wall morphology

Sarcocystis ovicanis Heydorn et al., 1975, illus.
Mehlhorn, H.; Heydorn, A. O.; and Gestrich, R., 1975, Ztschr. Parasitenk., v. 48 (2), 83-93
Sarcocystis ovicanis, lambs infected with cysts from dogs which had been infected with sheep muscles infected with S. tenella, light and electron microscopy of cysts in lambs, cyst wall structure

Sarcocystis ovifelis n. spec.
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
Syn.: Sarcocystis tenella, Railliet, 1886, pro parte; Balbiania gigantea, Railliet, 1886; Isospora bigemina, Stiles, 1891, Luhe, 1906, pro parte
". . . according to International Nomenclature Rules S. fusiformis and S. tenella constitute a pro parte situation and the old names must be retained for two selected species. In this case we would prefer to denote the cat-cattle and cat-sheep species with the old names adding the new names in brackets: S. fusiformis (bovifelis) and S. tenella (ovifelis)."
Ovis aries
Felis catus

Sarcocystis ovifelis Heydorn et al., 1975
Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
as syn. of S. gigantea (Railliet, 1886)

Sarcocystis ovifelis (= S. tenella pro parte), illus.
D'Haese, J.; Mehlhorn, H.; and Peters, W., 1977, Internat. J. Parasitol., v. 7 (6), 505-518
Sarcocystis ovifelis, Besnoitia jellisoni, Eimeria falciformis, pellicular structures, fine structure, implications for mechanism of motility

Sarcocystis ovifelis
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
as syn. of Sarcocystis tenella

Sarcocystis ovifelis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis tenella Railliet 1886

Sarcocystis ovifelis
Mehlhorn, H.; and Heydorn, A. O., 1976, Ztschr. Parasitenk., v. 50 (2), 209-210
Sarcocystis ovicanis, S. ovifelis, cyst wall morphology

Sarcocystis porcifelis n. sp., illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis porcifelis, Dubey, 1976
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis porcihominis n. sp.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis porcihominis
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]
"Tadros and Laarman [1976] also proposed 2 new organisms: Endorimospora (Sarcocystis) suihominis from the pig and Isospora heydorni from the dog. I [1976] named them Sarcocystis porcihominis and Isospora wallacei respectively. Because Tadros and Laarman's paper was published a month earlier than mine, the names proposed by Tadros and Laarman have priority over Sarcocystis porcihominis and Isospora wallacei."

Sarcocystis siamensis von Linstow 1903
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis fusiformis Railliet 1897

Sarcocystis singaporensis sp. n. [i.e., nom. nov.]
Zaman, V.; and Colley, F. C., 1976, Ztschr. Parasitenk., v. 51 (1), 137
for Sarcocystis orientalis Zaman and Colley, 1975, preoccupied by S. orientalis Machulskii and Miskaryan, 1958

Sarcocystis suihominis n. sp., illus.
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
Sarcocystis suihominis n. sp., life cycle study; S. bovihominis failure to infect pigs proves that it is a different species from S. suihominis n. sp.
Syns.: Sarcocystis miescheriana, Kuehn, 1865, pro parte; Synchytrium miescherianum, Kuehn, 1865, pro parte; Gregarina miescheriana, Rivolta, 1878, pro parte; Sarcocystis miescheri, Lankester, 1882, pro parte; Miescheria utriculosa, Harz, 1886, pro parte; Isospora bigemina, Stiles 1891, Luehe 1906, pro parte; I. hominis, Railliet und Lucet, 1891, Wenyon 1923, pro parte
Sus scrofa domestica (nat. and exper.)
Homo sapiens (exper.)

Sarcocystis suihominis, illus.
Heydorn, A. O., 1977, Arch. Lebensmittel.-Hyg., v. 28 (1), 27-31
Sarcocystis bovihominis, S. bovicanis, exper. human infection from raw beef, minor symptoms; S. suihominis, exper. human infection from raw pork, severe symptoms, parasite pathogenic or toxic

Sarcocystis suihominis, illus.
Mehlhorn, H.; and Heydorn, A. O., 1977, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 239 (1), 124-139
Sarcocystis suihominis, pigs (exper.), cyst fine structure and development in muscle fibre, nervous and connective tissue cells, light and electron microscopy, concluded that development of S. suihominis is somewhat quicker than other sarcosporidia

Sarcocystis tenella
Arru, E.; and Cosseddu, A. M., 1976, Clin. Vet., Milano, v. 99 (7), 322-327
incidence in regularly slaughtered animals
sheep: Sardinia

Sarcocystis tenella
Aryeetey, M. E.; and Piekarski, G., 1976, Ztschr. Parasitenk., v. 50 (2), 109-124
Sarcocystis tenella-infected rats, antibodies detected by indirect immunofluorescence test; no cross reaction with Toxoplasma-infected mice; rats fed infected sausage showed negative reaction to Sarcocystis antigen

Sarcocystis tenella (Railliet, 1886), illus.
Ashford, R. W., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 29-34
synonymy, description, fox as definitive host
Vulpes vulpes (intestine, feces) (nat. and exper.): Wales; Exmoor

Sarcocystis tenella, illus.
Bergmann, V.; and Kinder, E., 1975, Monatsh. Vet.-Med., v. 30 (20), 772-774
Sarcocystis tenella, sheep, differences in cyst wall structure, electron microscopy, thin- and thick-walled microcysts, macrocysts, possibly two species involved

Sarcocystis tenella
Bordjoski, A.; et al., 1972, Acta Parasitol. Iugoslavica, v. 3 (2), 129-135
Sarcocystis tenella, improved method for preparation of antigen for indirect fluorescent antibody test

Sarcocystis tenella
Bordjoski, A.; and Conic, V., 1973, Acta Parasitol. Iugoslavica, v. 4 (2), 99-105
Sarcocystis tenella, preparation of antigens for complement fixation test and fluorescent antibody test, survey of humans, low number of low titre positive reactions not considered conclusive proof that sarcosporidiosis does not occur in humans

Sarcocystis tenella
Bordjochki, A.; Savin, Z.; and Sovitsch, M., 1977, Bull. Acad. Vet. France, v. 50 (1), 71-74
Sarcocystis tenella spores, partial or total loss of antigenic properties when heated at various temperatures (60°-90°C), guinea pigs immunized and then tested by complement fixation

Sarcocystis tenella, illus.
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (10), 1061-1078
Syn.: Sarcocystis ovifelis
Sarcocystis of domestic animals, feline and canine Coccidia, nomenclature, life cycles, pathogenicity, extensive review

Sarcocystis tenella, Railliet, 1886
Dubey, J. P., 1977, J. Am. Vet. Med. Ass., v. 170 (8), 778, 782 [Letter]

Sarcocystis tenella Railliet, 1886
Gadaev, A.; and Abidzhanov, A. A., 1976, Uzbek. Biol. Zhurnal (4), 57-59
brief description
[Ovis aries]: meat packing plants in Tashkent and Karshi, Uzbekistan

Sarcocystis tenella, illus.
Goksu, K., 1975, Vet. Fak. Dergisi, Istanbul Univ., v. 1 (1), 110-127
incidence in sheep (oesophagus, diaphragm, abdominal and intercostal muscles, tongue, heart), 1971-1975: Ankara and Zeytinburnu meat plants

Sarcocystis tenella, Railliet, 1886, pro parte
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis ovicanis n. spec.

Sarcocystis tenella, Railliet, 1886, pro parte
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
as syn. of Sarcocystis ovifelis n. spec.

Sarcocystis tenella
Kepka, O.; and Rezaeian, M., 1976, Ztschr. Parasitenk., v. 50 (2), 210
Frenkelia glareoli, Sarcocystis tenella, chemotaxonomic comparison, disc electrophoresis, serology, close but not identical species

Sarcocystis tenella Railliet 1886
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Syns.: Balbiania gigantea Railliet 1886; Sarcocystis ovifelis Heydorn, Gestrich, Mehlhorn, and Rommel 1975
Ovis aries
Felis catus

Sarcocystis tenella Railliet 1886 pro parte
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis ovicanis Heydorn, Gestrich, Mehlhorn, and Rommel 1975

Sarcocystis tenella
Ludvik, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Sarcocystis tenella, Toxoplasma gondii, Eimeria tenella, negative-stained preparations, ultrastructure

Sarcocystis tenella, illus.
Macchioni, G.; and Marconcini, A., 1974, Ann. Fac. Med. Vet.,Pisa, v. 26, 1973, 402-409
Sarcocystis tenella, life cycle in cats experimentally infected with cysts from sheep oesophagus, not infective to dogs, mice and chickens; incidence in sheep

Sarcocystis tenella
Mehlhorn, H.; Heydorn, A. O.; and Gestrich, R., 1975, Ztschr. Parasitenk., v. 48 (2), 83-93
Sarcocystis ovicanis, lambs infected with cysts from dogs which had been infected with sheep muscles infected with S. tenella, light and electron microscopy of cysts in lambs, cyst wall structure

Sarcocystis tenella
Mueller, B. E. G.; Mehlhorn, H.; and Scholtyseck, E., 1973, J. Protozool., v. 20 (4), 505
ultrastructure and origin of cyst wall in Toxoplasmatea

Sarcocystis tenella
Oberg, C.; Diaz, L.; and Valenzuela, G., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 99-102
Ovis aries: Chile

Sarcocystis tenella, illus.
Porchet-Hennere, E., 1975, J. Protozool., v. 22 (2), 214-220
Sarcocystis tenella endozoite, ultrastructure using negative staining techniques

Sarcocystis tenella, illus.
Scholtyseck, E.; Mehlhorn, H.; and Mueller, B.E.G., 1974, J. Protozool., v. 21 (2), 284-294
Sarcocystis tenella, Besnoitia jellisoni, Frenkelia sp., Toxoplasma gondii, nature of cyst, fine structure of cyst wall

Sarcocystis tenella Railliet, 1886 pro parte
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora ovicanis Heydorn, Gestrich, Mehlhorn & Rommel, 1975 [comb. nov.]

Sarcocystis tenella Railliet, 1886
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora tenella (Railliet, 1886) comb. nov.

Sarcocystis tenella (ovifelis)
Heydorn, A.-O., et al., 1975, Ztschr. Parasitenk., v. 48 (2), 73-82
". . . according to International Nomenclature Rules S. fusiformis and S. tenella constitute a pro parte situation and the old names must be retained for two selected species. In this case we would prefer to denote the cat-cattle and cat-sheep species with the old names adding the new names in brackets: S. fusiformis (bovifelis) and S. tenella (ovifelis)."

Sarcocystis tenella (ovifelis) Heydorn, Gestrich, Mehlhorn & Rommel, 1975
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora tenella (Railliet, 1886) comb. nov.

Sarcocisti, illus.
Cornaglia, E., 1976, Ann. Fac. Med. Vet. Torino, v. 28, 259-267
sarcosporidic cysts, ibex (heart), pathology

Sarcocysts
Cunningham, C. C., 1973, Vet. Rec., v. 92 (25), 684 [Letter]
foal (heart muscle)

Sarcocysts, sarcosporidial cysts
Kaliner, G.; Grootenhuis, J. G.; and Protz, D., 1974, J. Wildlife Dis., v. 10 (3), 237-238
Gazella granti: Kenya
G. thomsoni: Kenya
Aepycerus melampus: Kenya
Syncerus caffer: Kenya
Taurotragus oryx: Kenya
Connochaetes taurinus: Kenya
Kobus defassa: Kenya
Alcelaphus buselaphus: Kenya
Phacochoerus aethiopicus: Kenya
Equus burchelli: Kenya; Northern Tanzania
Giraffa camelopardalis: Kenya
Redunca redunca: Kenya
R. chanleri: Kenya
Ourebia ourebi: Kenya

Sarcosporidia
Chattopadhyay, S. K.; and Sharma, R. M., 1972, Indian J. Animal Sc., v. 42 (9), 705-710
sheep and goats from slaughterhouses, lesions in pericardium and heart, findings include Sarcosporidia, Multiceps, hydatid cyst, Cysticercus tenuicollis: India

Sarcosporidia
Meshkov, S., 1973, Vet. Med. Nauki, v. 10 (34), 73-81
survey of slaughtered sheep: Burgas abattoir

Sarcosporidia
Meshkov, S., 1975, Vet. Med. Nauki, v. 12 (5), 55-61
survey of slaughtered cattle: Burgas abattoir

Sarcosporidia [sp.], illus.
Bergmann, V.; and Kinder, E., 1976, Monatsch. Vet.-Med., v. 31 (20), 785-788
Sarcosporidia, sarcocysts, electron microscopy of wall structure, four types of cysts recorded
Wildschwein
Reh
(skeletel muscles of all)

Sarcosporidia [sp.], illus.
Janitschke, K.; Protz, D.; and Werner, H., 1976, Ztschr. Parasitenk., v. 48 (3-4), 215-219
Sarcosporidia [sp.], cysts from Gazella granti (muscle) fed to dogs and cats which shed oocysts and sporocysts (feces), morphological study

Sarcosporidian cysts
Cerna, Z., 1976, J. Protozool., v. 23 (4), 6A [Abstract]
coccidian oocysts isolated from Tyto alba caused heavy sarcosporidian infection in Mus musculus, asexual development of parasite

Sarcosporidiens
Cerna, Z.; and Senaud, J., 1976, J. Protozool., v. 23 (4), 6A-7A [Abstract]
Isospora buteonis oocysts caused sarcosporidian infection in mice (exper.), ultrastructure of asexual stages

Sarcosporidiosis
Arru, E.; Tanda, S.; and Cosseddu, A. M., 1977, Clin. Vet., Milano, v. 100 (5), 341-346
sarcospiridiosis, danger of raw meat for man and animal, importance of meat inspection

Sarcosporidiosis
Basson, P. A.; et al., 1970, Onderstepoort J. Vet. Research, v. 37 (1), 11-28
parasitic and other diseases of Syncerus caffer, some pathological findings, age of host
Syncerus caffer (oesophagus, pharyngeal and laryngeal musculature): Kruger National Park

Sarcosporidiosis
Markus, M. B., 1973, N. England J. Med., v. 289 (18), 980-981 [Letter]
human Toxoplasma gondii, no evidence for cross-reactions of routine serologic tests for toxoplasmosis with sarcosporidiosis and isosporosis

Sarcosporidiosis
Markus, M. B., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 415-416 [Letter]
human sarcosporidiosis and toxoplasmosis, problems in serological differential diagnosis

Sarcosporidiosis
Shukla, D. C.; and Victor, D. A., 1976, Indian Vet. J., v. 53 (11), 852-854
sarcosporidiosis, bovines, diagnosis, complement fixation effective, agglutination and gel diffusion were of no value

Sarcosporidiosis
Young, E.; et al., 1973, Research J. National Parks Republic South Africa (16), 195-198
Antidorcas marsupialis (cardiac musculature): Mountain Zebra National Park near Cradock, Cape Province

Sarcosporidium
Szazados, I.; and Takacs, J., 1976, Magy. Allat. Lapja, v. 98, v. 31 (10), 671-673
parasites of sheep, importance in meat inspection

Saurocytozoon Lainson & Shaw, 1969
Bennett, G. F.; et al., 1975, J. Protozool., v. 22 (1), 24-30
extensive remarks on taxonomic and nomenclatural status of Leucocytozoon, classification (Haemosporina, Leucocytozoidae), emended generic diagnosis, proposal that authorship be credited to Sambon 1908 and that L. majoris (Laveran, 1902) be designated as type species (L. danilewskyi being a nomen nudum), rejection of recent synonymy of Akiba and Saurocytozoon with Leucocytozoon

Saurocytozoon Lainson & Shaw, 1969
Hsu, C.-K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon Berestnev, 1904

Saurocytozoon tupinambi Lainson & Shaw, 1969
Hsu, C.-K.; Campbell, G. R.; and Levine, N. D., 1973, J. Protozool., v. 20 (2), 195-203
as syn. of Leucocytozoon (Saurocytozoon) tupinambi (Lainson & Shaw, 1969) nov.comb.

Sauromella Pienaar, 1954
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses

Sauroplasma du Toit, 1937
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses

Sauroplasma du Toit, 1937
Svahn, K., 1976, Norwegian J. Zool., v. 24 (1), 1-6
Theileriidae

Sauroplasma boreale sp. n., illus.
Svahn, K., 1976, Norwegian J. Zool., v. 24 (1), 1-6
Ixodes ricinus as possible transmitter of Sauroplasma boreale sp. n.
Lacerta agilis (blood): North shore of Lake Snogeholmssjon in Scania, southern Sweden; Jydelejet on the island of Mon, Denmark

Schaudinnella Nusbaum, 1903
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
Schaudinnellidae
diagnosis; checklist of species

Schaudinnella enchytraei (von Kolliker, 1848) Meier, 1956
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
synonymy

Schaudinnella henleae Nusbaum, 1903
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Schaudinnella enchytraei (von Kolliker, 1848) Meier, 1956

Schaudinnellidae Poche, 1913
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
diagnosis; checklist of genera and species of 6 aseptate gregarine families

Schellackia
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243
generic definition; differentiation of species from lizards and amphibians, includes: Schellackia bolivari; S. occidentalis; S. brygooi; S. landauae; S. balli; S. golvani

Schellackia balli n. sp., illus.
Le Bail, O.; and Landau, I., [1975], Ann. Parasitol., v. 49 (6), 1974, 663-668
life cycle
Bufo marinus (nat. and exper.) (sang, intestin grele): Maripassoula (Guyane francaise)

Schellackia balli, illus.
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243

Schellackia bolivari, illus.
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243

Schellackia brygooi, illus.
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243

Schellackia golvani n. sp., illus.
Rogier, E.; and Landau, I., 1975, Bull. Mus. National Hist. Nat., Paris, 3. s. (284), Zool. (194), 91-97
Anolis marmoratus (intestin): Guadeloupe

Schellackia landauae sp.nov., illus.
Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243
development, life cycle, exper. transmission
Polychrus marmoratus (blood (erythrocytes, lymphocytes, monocytes), liver, spleen, lungs, small intestine): Capanema, Para State, north Brazil
Culex pipiens fatigans (gut) (exper.)

Schizotrypanum cruzi
Fistein, B.; and Deer, A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 306 [Abstract]
Rhodnius prolixus, inheritance of ability to transmit Schizotrypanum cruzi

Schneideria schneiderae n. sp., illus.
da Cunha, A. B.; et al., 1975, Rev. Brasil. Biol., v. 35 (4), 693-697
Trichosia pubescens (intestinal caeca, midintestine): Taiacupeba, Mogi das Cruzes, State of Sao Paulo

Schultzellina mucronata Cepede, 1910
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
as syn. of Maupasella mucronata (Cepede, 1910) nov. comb. de Puytorac, 1954

Selenidioides sp.
Douglass, T. G., 1977, J. Electr. Micr., v. 26 (2), 177 [Abstract]
Selenidioides sp., function of anterior complex of merozoites (conoid, rhoptries, micronemes, subpellicular microtubules) in feeding and other activities, electron microscopy
Boccardia proboscidea (intestinal epithelium)

Selenidioides giganteum (Harant, 1931) Levine, 1971 pro parte (?)
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria maculata Ormieres, 1965

Selenidium annulata (Greeff, 1885) comb. nov.
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syns.: Polyrabdina annulata (Greeff, 1885) Mingazzini, 1893; Polyrhabdina annulata (Greeff, 1885) Labbe, 1899

Selenidium cirratuli (Lankester, 1866) Caullery & Mesnil, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Polyrhabdina cirratuli Mingazzini, 1893 of Labbe (1899) pro parte

Selenidium fallax MacGregor & Thomasson, 1965
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Polyrhabdina cirratuli (Lankester) of Labbe (1899) pro parte

Selenidium fauchaldi sp. n., illus.
Levine, N. D., 1974, J. Protozool., v. 21 (1), 8-9
Phragmatopoma californica (free in lumen of posterior intestine): intertidal zone of Pacific Ocean on Santa Catalina Island, California

Selenidium (gen.?) giganteum Harant, 1931 pro parte (?)
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
as syn. of Lankesteria maculata Ormieres, 1965

Selenidium hollandei
Metais, M.; and Schrevel, J., 1974, J. Protozool., v. 21 (3), 473 [Abstract]
Selenidium hollandei, mechanism of motility, role of cortical fibrillar structures

Selenidium sabellae (Lankester, 1863) Ray, 1930
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Polyrhabdina sabellae (Lankester, 1863) Labbe, 1899

Selenidium serpulae (Lankester, 1863) Caullery & Mesnil, 1899
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
synonymy

Selenidium spionis (von Kolliker, 1845) Ray, 1930
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Syn.: Polyrabdina spionis Mingazzini, 1871 pro parte

Selenidium spionis (von Kolliker, 1848) Ray, 1930
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
"Monocystis spionis (von Kolliker, 1848) Lankester, 1863. Syn. of Polyrhabdina spionis (von Kolliker, 1848) Mingazzini, 1891 pro parte and of Selenidium spionis (von Kolliker, 1848) Ray, 1930 pro parte."

Selenidium synaptae (Mingazzini, 1893) Levine, 1971
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Syn.: Esarabdina synaptae Mingazzini, 1893

Selenidium telepsavi (Stuart, 1871) comb. nov.
Levine, N. D., 1974, J. Protozool., v. 21 (1), 8-9
Syn.: Monocystis telepsavi Stuart, 1871

Semitrichodina
Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Trichodinidae

Serpentoplasma Pienaar, 1954
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses

Sicuophora macropharyngea (Bezzenberger, 1904) comb. nov., illus.
Albaret, J.-L., 1973, J. Protozool., v. 20 (1), 51-57
description
Syns.: Nyctotherus macropharyngeus Bezzenberger, 1904; Prosicuophora macropharyngeus (Bezzenberger, 1904) de Puytorac & Oktem, 1967
Rana cyanophlyctis (rectum): Afghanistan

Sicuophora xenopi de Puytorac & Grain, 1968, illus.
Albaret, J.-L., 1973, J. Protozool., v. 20 (1), 51-57
nuclear apparatus, polysaccharide elements, fibrillar system
Xenopus mulleri (rectum): Ippy, Republique Centrafricaine
X. fraseri (rectum): La Maboke, Republique Centrafricaine

Silvatoshellina
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Sinuolinea magna Yoshino & Noble 1973
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Coryphaenoides acrolepis: off Oregon, Mexico, northern, southern California, Oregon, Canada
C. pectoralis: northern California
(urinary bladder of all)

Sinuolinea rebae Tripathi 1948
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Coelorinchus coelorhincus (urinary bladder): off Italy
C. gladius (urinary bladder): off Hawaii
Malacocephalus laevis (urinary bladder): off South Africa
Trachonurus villosus (gall bladder): Gulf of Mexico
Trachyrincus sp. (gall bladder): off Ecuador

Sinuolinea triangulata Schulman 1966
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Coryphaenoides acrolepis (urinary bladder, kidney): off northern Mexico, southern, northern California, Oregon
C. armatus (urinary bladder, kidney): off northern Mexico, southern, central, northern California, Oregon
C. filifer (urinary bladder): off Oregon
C. leptolepis (urinary bladder): off northern California, Oregon
C. pectoralis (kidney): off northern California

Sleeping sickness, Rhodesian. See [Trypanosoma rhodesiense]

Sogdianella moshkovskii
Peirce, M. A., 1973, J. Protozool., v. 20 (5), 543-546
"On the available evidence. . .regarded as valid avian piroplasm. . ."

Sphaerocystis hydrophili Foerster, 1938
Baudoin, J.; and Maillard, Y. P., 1972, Protistologica, v. 8 (1), 53-63
as syn. of Enterocystis hydrophili n. comb.

Sphaerocystis hydrophili Foerster, 1938
Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
as syn. of Enterocystis hydrophili (Foerster, 1938) Baudoin and Maillard, 1972

Sphaeromyxa intermediata sp. n., illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Malacocephalus occidentalis: off Colombia (Caribbean Sea)
Nezumia cyrano: Caribbean Sea
Ventrifossa macropogon: off Venezuela (Caribbean Sea)
V. mucocephalus: Caribbean Sea

Sphaeromyxa lomi sp. n., illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Malacocephalus occidentalis (gall bladder): off Borneo

Sphaeromyxa maiyai sp.n., illus.
Morrison, N. D.; and Pratt, I., 1973, J. Protozool., v. 20 (2), 214-217
Microgadus proximus (gall bladder): bays and oceans to at least 40 fathoms off Newport, Oregon

Sphaeromyxa minuta Poljanskii 1955
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 93-96
Coelorinchus argentatus (gall bladder): off Philippine Islands

Sphaerospora divergens Thelohan, 1895
Moser, M.; and Noble, E. R., 1977, Ztschr. Parasitenk., v. 51 (2), 159-163
Coelorinchus coelorhincus carminatus (gall bladder): off Surinam

Sphaerospora elegans Thelohan, 1892
Campbell, A. D., 1974, Proc. Roy. Soc. Edinb., sect. B, Biol., v. 74, 347-364
Gasterosteus aculeatus: Loch Leven, Scotland

Spiriopsis adipophila (Arvy et Delage), illus.
Arvy, L.; and Sowa, R., 1976, Ann. Parasitol., v. 51 (1), 111-120
Ephemera danica
Ephemerella ignita
all from region de Cracovie, Pologne

Spiriopsis adipophila (Arvy et Delage 1966) Arvy and Peters, 1972, illus.
Desportes, I.; and Delage-Darchen, B., 1975, Compt. Rend. Acad. Sc., Paris, v. 281, s. D, Sc. Nat. (14), 1001-1002
Spiriopsis adipophila, micronemes and paraglycogen in cytoplasm, considered a sporozoan, possibly in family Barrouxiidae
Ephemera vulgata (cavite generale)

Spirobuetschliella Hovasse, 1950
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
Intoshellinidae

Spirobuetschliella clignyi (Cepede, 1922) Hovasse, 1950
de Puytorac, P., 1972, Protistologica, v. 8 (1), 5-42
synonymy
Potamoeeros triqueter

Spironucleus Lavier
Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Diplomonadina emend.

Spironucleus bocis (Hexamita bocis) Lavier
Brugerolle, G., 1976, J. Protozool., v. 23 (4), 5A [Abstract]
Box boops (intestin)

Spironucleus (Hexamita) muris, illus.
Kunstyr, I., 1977, Lab. Animals, v. 11 (3), 185-188
Spironucleus muris, description of mature and immature cysts
mice (exper.) (faeces, lower intestine)

Spironucleus (Hexamita) muris, illus.
Kunstyr, I.; Ammerpohl, E.; and Meyer, B., 1977, Lab. Animal Sc., v. 27 (5), part II, 782-788
Spironucleus muris, nude mouse (exper.) an excellent model for immunologic and pharmacologic studies; dimetridazole, metronidazole, tinidazole, and Acranil tested

Spironucleus salpae (Hexamita salpae) Lavier 1936
Brugerolle, G., 1976, J. Protozool., v. 23 (4), 5A [Abstract]
Box salpa (intestin posterieur)

Sporadina juli Frantzius, 1848
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora juli (Frantzius)

Sporadina oblongata (Hamm.) Frantzius, 1848
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stylocephalus oblongatus (Hammerschmidt)

Sporodinium
Loeblich, A. R., III, 1974, J. Protozool., v. 21 (3), 435 [Abstract]
organism isolated from plankton of Puget Sound, Washington, corresponds to organism described by Dogiel, 1906 (mistakenly considered by him to be synonymous with Gymnodinium lunula) and appears to be congeneric with Sporodinium pseudocalanii (parasite of copepod eggs); another organism isolated off San Diego, California, corresponds to Schuett's description of G. lunula and is a free-living autotroph

Sporozoa [sp.], illus.
Dissanaike, A. S.; Lim, B. L.; and Poopalachelvam, M., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 141-142 [Demonstration]
intracellular sporozoan parasite with 8 "spore-like" bodies discovered in spleen and liver smears and sections from Echinosorex gymnurus: Selangor/Gombak Forest Reserve

Sporozoa [sp.], illus.
Ferguson, H. W.; and Roberts, R. J., 1976, Prog. Exper. Tumor Research, v. 20, 212-216
Sporozoa [sp.] existing within lymphoma-like condition affecting 2-year-old Scophthalmus maximus being raised at a marine culture unit, causal relationship not established but possibly the neoplastic condition provided suitable altered environment for proliferation of the protozoan parasites: Scotland

Sporozoa [sp.] of undetermined taxonomic position, illus.
Lainson, R., 1972, J. Protozool., v. 19 (4), 582-586
description
Bos taurus (lung, liver): England

Sporozoa [sp.] of undetermined taxonomic position, illus.
Lainson, R., 1972, J. Protozool., v. 19 (4), 582-586
description
Dasypus novemcinctus (liver, brain capillaries, spleen): British Honduras

Steinella [sp.]
Ball, I. R.; and Khan, R. A., 1976, J. Fish. Biol., v. 8 (5), 419-426
Micropharynx parasitica (intestinal lumina): North Atlantic Ocean, off coast of Newfoundland

Steinina ovalis
Harry, O. G., 1973, J. Protozool., v. 20 (4), 526
4 species of gregarines in Tenebrio molitor, incidence, effect of seasonal changes in temperature and humidity on infection: British Isles

Steinina ovalis, illus.
Ruehl, H., 1976, Arch. Protistenk., v. 118 (4), 353-363
ultrastructure
Tenebrio molitor

Stempellia sp.
Hazard, E.; and Fukuda, T., 1974, J. Protozool., v. 21 (4), 497-504
Psorophora ferox (fat body, Malpighian tubules)

Stempellia sp.
Hazard, E.; and Fukuda, T., 1974, J. Protozool., v. 21 (4), 497-504
Psorophora horrida

Stempellia sp.
Hazard, E.; and Fukuda, T., 1974, J. Protozool., v. 21 (4), 497-504
Uranotaenia sapphirina

Stempellia lunata Hazard & Savage 1968, illus.
Hazard, E.; and Fukuda, T., 1974, J. Protozool., v. 21 (4), 497-504

Stempellia magna (Kudo 1920), illus.
Hazard, E.; and Fukuda, T., 1974, J. Protozool., v. 21 (4), 497-504

Stempellia milleri sp. n., illus.
Hazard, E.; and Fukuda, T., 1974, J. Protozool., v. 21 (4), 497-504
host specificity, symptomology in mosquito host
Culex pipiens quinquefasciatus (blood cells, adipose tissue) (nat. and exper.): Louisiana; Texas
C. p. fatigans: Bangkok, Thailand
C. p. pipiens (exper.)
C. salinarius (exper.)
C. territans (exper.)
C. tarsalis (exper.)

Stempellia rubtsovi Issi, 1968, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Stempellia simulii Maurand & Manier, 1968, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Stenocephalus juli: Schneider, 1875 [pro parte]
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora juli (Frantzius)

Stenocephalus juli: Schneider, 1875 pro parte
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora julimarginati (Leidy)

Stenoductus gen. n.
Ramachandran, P., 1976, Zool. Anz., Jena, v. 196 (1-2), 125-132
Monoductidae
tod: S. penneri sp. n.

Stenoductus penneri sp. n. (tod), illus.
Ramachandran, P., 1976, Zool. Anz., Jena, v. 196 (1-2), 125-132
Floridobolus penneri (midgut, hindgut): vicinity of Archbold Biological Station, Highlands County, Florida

Stenophora sp., illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Oxidus affinis: Coree du sud

Stenophora sp. 1, illus.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Oxydesmidae: Cote d'Ivoire

Stenophora sp. 2, illus.
Theodorides, J.; Desportes, I.; and Mateu, J., 1976, Ann. Parasitol., v. 51 (5), 499-512
Strongylosomidae: Cote d'Ivoire

Stenophora caudata sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Chromatoiulus projectus (intestine): Bialowieza, Poland

Stenophora hoshidei n. sp., illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Orthomorpha uncinata: Khao Yai, Thailande

Stenophora juli (Frantzius), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, redescription
Schizophyllum sabulosum (intestine): Poland

Stenophora juli (Frantzius): Labbe, 1899 pro parte
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora julimarginati (Leidy)

Stenophora julimarginati (Leidy), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Schizophyllum sabulosum (intestine): Poland

Stenophora julimarginati: Leger et Duboscq, 1904
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora julimarginati (Leidy)

Stenophora larvata (Leidy) Ellis, 1913 pro parte; Watson, 1916 pro parte
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stenophora julimarginati (Leidy)

Stenophora nematoides Leger et Duboscq, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
description
Strongylosoma pallipes (intestine): Poland

Stenophora orthomorphae sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Orthomorpha gracilis (intestine): Poznan, Poland

Stenophora poznanensis sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Orthomorpha gracilis (intestine): Poznan, Poland

Stenophora sarmatiuli sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Sarmatiulus vilnensis (intestine): Kornik, Poland

Stenophora schizophylli sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Schizophyllum sabulosum (intestine): Bialowieza, Poland

Stenophora strongylosomae sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Strongylosoma pallipes (intestine): Bialowieza, Poland

Stenophora thyropygi (Rodgi et Ball)
Theodorides, J.; and Desportes, I., [1975], Ann. Parasitol., v. 49 (6), 1974, 785-786
synonymy

Stenophora uncigeri sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Unciger foetidus (intestine): Siemianice, Poland

Stictospora provincialis Leger, illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Amphimallon solstitialis (intestine): Poland

Stylocephalus bahli Misra, illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Syn.: Stylocephalus japonicus Hoshide
Gonocephalum sp.: Khon Kaen, Thailande

Stylocephalus bahli Misra, illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
Syn.: Stylocephalus japonicus Hoshide
Gonocephalum coriaceum
Mesomorphus villiger
all from Coree du sud

Stylocephalus carabi sp. n., illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
Carabus glabratus (intestine): Bialowieza, Poland

Stylocephalus (Stylocephalus) hoffmannseggi sp. nov., illus.
Devdhar, M. J.; and Amoji, S. D., 1977, Arch. Protistenk., v. 119 (1-2), 75-85
Stylocephalus spp., morphological and life-cycle studies
Gonocephalus hoffmannseggi (gut): Dharwar, Karnataka state, India

Stylocephalus japonicus Hoshide
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
as syn. of Stylocephalus bahli Misra

Stylocephalus japonicus Hoshide
Theodorides, J.; Desportes, I.; and Jolivet, P., 1976, Ann. Parasitol., v. 51 (2), 161-173
as syn. of Stylocephalus bahli Misra

Stylocephalus (Xiphocephalus) karnatakaensis sp. nov., illus.
Devdhar, M. J.; and Amoji, S. D., 1977, Arch. Protistenk., v. 119 (1-2), 75-85
Stylocephalus spp., morphological and life-cycle studies
[lapsus p. 79 as S. karnatakanensis]
Gonocephalum hoffmannseggi (gut): Dharwar, Karnataka state, India

Stylocephalus karnatakanensis [lapsus for karnatakaensis]
Devdhar, M. J.; and Amoji, S. D., 1977, Arch. Protistenk., v. 119 (1-2), 79

Stylocephalus longicollis F. Stein, illus.
Desportes, I., 1975, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 17 (2), 215-228
Stylocephalus longicollis, scanning electron microscope study of different stages of life cycle
Blaps sulcata: Pyrenees-Orientales

Stylocephalus oblongatus (Hammerschmidt), illus.
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
synonymy, description
Opatrum sabulosum (intestine): Poland

Stylocephalus oblongatus (Hammerschmidt), illus.
Theodorides, J.; Desportes, I.; and Jolivet, P., 1975, Ann. Parasitol., v. 50 (2), 145-159
Pseudoblaps javana: Hua Hin and Phuket, Thailande

Stylocephalus (Xiphocephalus) serpentula sp. nov., illus.
Devdhar, M. J.; and Amoji, S. D., 1977, Arch. Protistenk., v. 119 (1-2), 75-85
Stylocephalus spp., morphological and life-cycle studies
Gonocephalum hoffmannseggi (gut): Dharwar, Karnataka State, India

Stylorhynchus oblongatus (Hamm.) Schneider, 1875
Lipa, J. J., 1967, Acta Protozool., v. 5 (8), 97-179
as syn. of Stylocephalus oblongatus (Hammerschmidt)

Sycia Leger, 1892
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Sycia cirratuli H. Hoshide, 1958
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Sycia inopinata Leger, 1892
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Sycia legeri Ganapati, 1946
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Sycia polydorae Ganapati, 1946
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Synchytrium mieschierianum, Kuehn, 1865, pro parte
Heydorn, A. O., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (11), 218-224
as syn. of Sarcocystis suihominis n. spec.

Synchytrium mieschierianum Kuhn 1865
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Sarcocystis miescheriana (Kuhn 1865) Labbe 1899

Synchytrium mieschierianum Kuhn, 1865 [pro parte]
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Endorimospora miescheriana (Kuhn, 1865) comb. nov.

Synchytrium miescherianum Kuhn, 1865 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Endorimospora suihominis nom. nov. [i.e., n. sp.]

Syncystis Cuenot, 1891
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Urospora Schneider, 1875

Syncystis holothuriae (Schneider, 1858) Cuenot, 1891
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Urospora holothuriae (Schneider, 1858) Tregouboff, 1918

Syncystis muelleri Cuenot, 1891
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Urospora muelleri (Cuenot, 1891) Cuenot, 1892, on p. 102; but as syn. of Urospora synaptae (Cuenot, 1891) Leger, 1892, on p. 106

Synophrya [sp.]
  Johnson, C. A., III; and Bradbury, P. C., 1976, J. Protozool., v. 23 (2), 252-256
    Synophrya in decapods, occurrence may be related to salinity, possibly important pathogen in post-larval crabs
    Sicyonia brevirostris
    Albunea sp.
    Ranila muricata
    Dromidia antillensis
    Persephona punctata
    Calappa flammea
    C. angusta
    Hepatus epheliticus
    Cycloes bairdii
    Ovalipes ocellatus
    O. quadulpensis
    Portunus sayi
    P. ordwayi
    P. spinicarpus
    Pilumnus sayi
    Micropanope sp.
    Palicus faxoni
    Euchirograpsus americanus
    Mithrax pleuracanthus
    M. forceps
    M. acuticornis
    Stenocionops furcata
    Parthenope fraterculus
    P. agona
    all from coastal North Carolina, U.S.A.

Synophrya hypertrophica Chatton & Lwoff 1926, illus.
  Johnson, C. A., III; and Bradbury, P. C., 1976, J. Protozool., v. 23 (2), 252-256
    Ovalipes ocellatus
    Macropipus sp.
    both from Roscoff, France

Telomyxa glugeiformis Leger et Hesse
Arvy, L.; and Sowa, R., 1976, Ann. Parasitol., v. 51 (1), 111-120
Ephemera danica: region de Cracovie, Pologne

Terebrospira
Bradbury, P. C., 1974, J. Protozool., v. 21 (3), 424 [Abstract]
Terebrospira, fine structure of feeding stage in Palaemonetes pugio

Terebrospira chattoni sp. n., illus.
Bradbury, P. C.; Clamp, J. C.; and Lyon, J. T., III, 1974, J. Protozool., v. 21 (5), 678-686
life cycle
Palaemonetes pugio (endocuticle) (nat. and exper.): South Creek near Pamlico Marine Station, Aurora, North Carolina

Terebrospira chattoni, illus.
Bradbury, P. C.; and Goyal, V., 1976, Tissue and Cell, v. 8 (4), 573-582
Terebrospira chattoni, fine structure during ingestion of exoskeleton of Palaemonetes pugio, demonstration of extracellular acid phosphatase and acid phosphatase bound to cell membrane

Tetrahymena
Cox, D.; and Miglietta, L., 1975, J. Protozool., v. 22 (3), 34A-35A [Abstract]
Tetrahymena killer parasite of mosquito colonies isolated from outbreaks in Louisiana, laboratory maintenance in vitro

Tetrahymena [sp.]
Grassmick, R. A., 1975, J. Protozool., v. 22 (3), 29A [Abstract]
Tetrahymena, 2 clones, host specificity: Aedes aegypti completely refractory, 2 spp. of Culex are susceptible, Anopheles quadrimaculatus susceptible to infection by one clone and refractory to infection with the other

Tetrahymena pyriformis, illus.
Seaman, G. R.; et al., 1972, J. Protozool., v. 19 (4), 644-647
Tetrahymena pyriformis, pathogenicity of hemocoelomic injections of various concentrations into adult female Periplaneta americana, recoverability of ciliates into axenic cultures from infected insects, morphologic changes of ciliates after recovery and reestablishment in axenic culturc

Tetrahymenina [sp.], illus.
Clark, T. B.; and Brandl, D. G., 1976, J. Invert. Path., v. 28 (3), 341-349
Tetrahymenina infection of Aedes sierrensis larvae (nat. and exper.), melanized spots on cuticle were sites of invasion or attempted invasion, invasion sites were capped by hemispherical membranes or cysts vacated by invading ciliates, factors such as host activity, host age, and cuticle thickness limited successful infection : near Kings River, Fresno County, California

Tetratrichomonas [sp.]
Nohynkova, E., 1974, J. Protozool., v. 21 (3), 457-458 [Abstract]
Haemopis sanguisuga (intestine): Czechoslovakia

Tetratrichomonas gallinarum (Martin & Robertson 1911) Honigberg 1963, illus.
Kulda, J.; Suchankova, E.; and Svoboda, S., 1974, Acta Vet. Brno, v. 43 (1), 53-64
Tetratrichomonas gallinarum, synonymy; axenic culture of 10 strains isolated from poultry, virulence determined by mouse assay did not reflect pathogenicity for turkey poults, results indicate T. gallinarum should not be considered serious pathogen; enterohepatitis caused by mixed infection with Histomonas meleagridis from contaminated culture
Meleagris gallopavo (nat. and exper.): Czechoslovakia
Gallus gallus: Czechoslovakia
Anas platyrhynchos f. dom.: Czechoslovakia
Anser anser f. dom.: Czechoslovakia
mice (exper.)

Thalicola
Theodorides, J.; and Desportes, I., 1975, J. Protozool., v. 22 (3), 85A-86A [Abstract]
gregarines of marine invertebrates, organization of epicyte

Theileria
Irvin, A. D.; et al., 1972, Vet. Rec., v. 91 (21), 513, 514-516, 517
theilerial piroplasms acquired by Rhipicephalus appendiculatus and R. simus (salivary glands) (exper.) fed on naturally infected Taurotragus oryx pattersonianus were not transmitted to cattle, role of eland in epidemiolgy of cattle theilerioses is questionable: Kenya

Theileria
Jadin, J. B.; et al., 1977, J. Protozool., v. 24 (4), 60A [Abstract]
relations between rickettsia and piroplasms

Theileria
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Theileria
Mehlhorn, H.; and Schein, E., 1976, J. Protozool., v. 23 (4), 14A [Abstract]
Theileria, sexual development in intestine and hemolymph of Hyalomma anatolicum excavatum

Theileria, resembling T. mutans
Miller, R. M.; Price, M. A.; and Kuttler, K. L., 1976, Southwest Entom., v. 1 (3), 107-110
Amblyomma americanum, A. cajennense, nymphs feeding on Anaplasma marginale and Theileria infected cattle, attempted transstadial transmission to splenectomized calves, unsuccessful

Theileria
Pal, M.; Verma, J. D.; and Dahiya, S. M., 1976, Indian J. Animal Research, v. 10 (2), 93-95
Theileria, cattle, higher incidence in young calves of exotic breed (Holstein Friesian) than adults: Satbari, Delhi

Theileria, illus.
Ristic, M., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 831-870
babesiosis, theileriosis, immunology, review

Theileria
Ristic, M., 1976, Vet. Parasitol., v. 2 (1), 31-47
intracellular blood protista: intraerythrocytic behavior, transfer, and circulatory clearance; survival and development within macrophages; persistence of organism in immunologically hostile host; immune responses and protection; serodiagnosis; vaccination

Theileria and/or Cytauxzoon [spp.]
Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
blood parasites of wild Bovidae, incidence, epizootiological importance
Syncerus caffer
Connochaetes taurinus
Damaliscus lunatus
Kobus leche
Aepyceros melampus
Hippotragus niger
Tragelaphus strepsiceros
Taurotragus oryx
(blood of all): all from Ngamiland, Botswana

Theileria sp., illus.
Grootenhuis, J. G.; et al., 1975, J. Wildlife Dis., v. 11 (1), 122-127
impala Theileria, morphology, transmission experiments, attempted transmission to splenectomized steer by inoculation and tick vector Rhipicephalus appendiculatus; no cross-reactivity with other theilerial parasites, indirect fluorescent antibody test
Aepyceros melampus (nat. and exper.): Machakos District, Kenya

Theileria [sp.] "eland Theileria"
Grootenhuis, J. G.; et al., 1975, J. Wildlife Dis., v. 11 (1), 122-127
impala Theileria, no cross-reactivity with other theilerial parasites, indirect fluorescent antibody test

Theileria sp., illus.
Grootenhuis, J. G.; et al., 1977, J. Parasitol., v. 63 (6), 1127-1129
Theileria sp. from naturally-infected eland transmitted transstadially by Rhipicephalus appendiculatus experimentally to a susceptible eland

Theileria [sp.]
Kuttler, K. L.; and Craig, T. M., 1975, Am. J. Vet. Research, v. 36 (3), 323-325
splenectomized calves inoculated with pooled blood samples from eastern Texas cattle developed dual infections of Anaplasma marginale and a Theileria resembling Theileria mutans, treatment with gloxazone and imidocarb selectively eliminated Anaplasma, pure Theileria infections only mildly pathogenic

Theileria species
Loehr, K.-F.; and Meyer, H., 1974, Tropenmed. u. Parasitol., v. 25 (3), 288-292
Theileria parva, T. mutans, capillary-tube agglutination and indirect fluorescent antibody tests on sera from Masai cattle (reacted with both species) and 5 game animals (reacted only with T. parva), intra erythrocytic "theileria organisms" found in all game animals but could not be needle-transmitted to splenectomized calves, "one theilerial species which was clearly of T. parva and not of T. mutans antigenicity", "cannot, however, be excluded that these piroplasms may not belong to members of the genus Cytauxzoon"
Connochaetes taurinus
Alcelaphus buselaphus
Aepyceros melampus
Gazella grantii
G. thomsonii
all from Kapiti Plains near Konza, Kenya

Theileria spp.
Morzaria, S. P.; et al., 1977, Research Vet. Sc., v. 22 (3), 330-333
Theileria mutans, comparison of British strain with East and South African strains and other Theileria spp. using indirect fluorescent antibody test, no cross-reactions were detected, concluded that British strain of T. mutans could be a distinct species from African strains of T. mutans

Theileria sp.
Stagg, D. A.; et al., 1976, Research Vet. Sc., v. 20 (2), 122-126
theilerial parasite or parasites (Theileria and/or Cytauxzoon), isolation from Taurotragus oryx in Kenya, establishment of five infected cell lines

Theileria [spp.]
Uilenberg, G.; et al., 1977, Austral. Vet. J., v. 53 (6), 271-273
Theileria mutans, African, serological and morphological comparisons with Theileria of cattle in Australia and Britain; results indicate that Australian and British theilerias were similar to each other but different from T. mutans

Theileria sp.
Uilenberg, G.; and Schreuder, B. E. C., 1976, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 29 (2), 119-126
presence in splenectomized sheep used in transmission and pathogenesis studies of Haematoxenus separatus

Theileria sp. (Githunguri), illus.
Young, A. S.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (2), 185-194
successful transmission of Theileria parasites from eland (Taurotragus oryx) to cattle by feeding Rhipicephalus appendiculatus and R. pulchellus on them; cattle recovered from infections produced antibodies to Theileria sp. (eland) and T. sp. (Githunguri) previously also recovered from cattle, indications that the 2 species may be related and possibly represent a new species infective for cattle

Theileria species (eland), illus.
Young, A. S.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (2), 185-194
successful transmission of Theileria parasites from eland (Taurotragus oryx) to cattle by feeding Rhipicephalus appendiculatus and R. pulchellus on them; cattle recovered from infections produced antibodies to Theileria sp. (eland) and T. sp. (Githunguri) previously also recovered from cattle, indications that the 2 species may be related and possibly represent a new species infective for cattle

Theileria [sp.], illus.
Young, A. S.; Irvin, A. D.; and Woodford, M. J., 1973, J. Wildlife Dis., v. 9 (1), 94-98
description
Syncerus caffer: Queen Elizabeth Park, Uganda

Theileria annulata
Bansal, G. C.; and Gaur, S. N. S., 1977, Pantnagar J. Research, v. 2 (2), 221-222
Theileria annulata, cross-bred calves (exper.), possibility of utilizing biochemical changes as diagnostic tool

Theileria annulata
Bansal, G. C.; Gaur, S. N. S.; and Shah, H. L., 1977, Pantnagar J. Research, v. 2 (2), 244-246
Theileria annulata, survey of cattle and buffaloes, low prevalence and lack of symptoms indicate resistance and carrier status; cross transmission experiments from calf with Hyalomma anatolicum as vector: successful only with calves, unsuccessful with guinea-pigs, rabbits and kids: Nainital Tarai

Theileria annulata Dschunskowsky & Luhs, 1904, illus.
Bhattacharyulu, Y.; Chaudhri, R. P.; and Gill, B. S., 1975, Ann. Parasitol., v. 50 (4), 397-408
Theileria annulata, endogenous development in Hyalomma anatolicum anatolicum larvae, nymphs, and adults

Theileria annulata
Dhar, S.; and Gautam, O. P., 1977, Indian J. Animal Sc., v. 47 (4), 169-172
Theileria annulata, total serum levels of bilirubin, calcium, sodium and potassium in calves (exper.), use in supportive therapy

Theileria annulata
Dhar, S.; and Gautam, O. P., 1977, Indian J. Animal Sc., v. 47 (7), 389-394
Theileria annulata, calves (nat. and exper.), complement-fixation and conglutinating-complement-absorption tests, more reliable in determining latent Theileria infection than blood smear examination

Theileria annulata
Dhar, S.; and Gautam, O. P., 1977, Indian J. Animal Sc., v. 47 (8), 458-462
Theileria annulata, cattle (nat. and exper.), capillary-tube-agglutination test

Theileria annulata
Dhar, S.; and Gautam, O. P., 1977, Indian J. Animal Sc., v. 47 (9), 566-570
Theileria annulata, cattle (nat. and exper.), haemagglutination-inhibition, indirect haemagglutination, and agar-gel-precipitation tests evaluated, haemagglutination-inhibition test most sensitive and more reliable than blood smear examination in predicting latent Theileria infection

Theileria annulata
Dhar, S.; and Gautam, O. P., 1977, Indian J. Animal Sc., v. 47 (11), 720-723
Theileria annulata, cattle (exper.), indirect fluorescent-antibody test, more reliable than blood smear examination for predicting latent Theileria infection

Theileria annulata
Dhar, S.; and Gautam, O. P., 1977, Indian Vet. J., v. 54 (1), 21-24
Theileria, cattle, species differentiation using complement-fixation test

Theileria annulata
Dipeolu, O. O.; and Ogunji, F. O., 1977, Lab. Animals, v. 11 (1), 39-40
Theileria annulata, transmission to a rabbit by larvae from eggs from engorged female tick Hyalomma rufipes from infected cattle, possibility of transovarial transmission

Theileria annulata
El Refaii, A. H.; and Michael, S. A., 1976, Brit. Vet. J., v. 132 (4), 363-368
Theileria annulata, clinical and subclinical infections in calves used for testing of rinderpest vaccine controlled by imidocarb dipropionate, enables vaccine testing to continue without complications previously attributable to T. annulata infection: Egypt

Theileria annulata
Gill, B. S.; et al., 1976, Nature (5584), v. 264, 355-356
Theileria annulata, calves, vaccination with schizonts grown in tissue culture, full immunity to severe tick (Hyalomma dromedarii) challenge

Theileria annulata
Gill, B. S.; et al., 1977, Ann. Recherches Vet., v. 8 (3), 285-292
Theileria annulata, cattle, successful immunization by infecting with one-tick stabilate (Hyalomma dromedarii) and simultaneous treatment with chlortetracycline

Theileria annulata
Gill, B. S.; Bhattacharyulu, Y.; and Kaur, D., 1976, Research Vet. Sc., v. 21 (2), 146-149
Theileria annulata, calves, tick-induced infections (Hyalomma anatolicum anatolicum or H. dromedarii), immunization using chlortetracycline prophylactically

Theileria annulata
Gill, B. S.; Bhattacharyulu, Y.; and Kaur, D., 1977, Ann. Parasitol., v. 52 (6), 597-608
Theileria annulata in Bos taurus x Bos indicus cross-bred calves, symptoms, sequence of appearance of parasite, pathology

Theileria annulata
Hashemi-Fesharki, R.; and Shad-Del, F., 1973, Am. J. Vet. Research, v. 34 (11), 1465-1467
Theileria annulata, vaccination of calves and milking cows with cell-suspension culture, immunity persisted for more than 1 year

Theileria annulata
Hashemi-Fesharki, R.; and Shad-Del, F., 1975, Arch. Inst. Razi (27), 67-70
Theileria annulata, cattle (exper.), Terramycin, administered during incubation period, at beginning of clinical syndrome, and at height of parasitaemia and fever, no therapeutic value

Theileria annulata
Hooshmand-Rad, P., 1976, Research Vet. Sc., v. 20 (3), 324-329
Theileria annulata, intact susceptible calves, premune calves following splenectomy, pathogenesis of anemia, role of erythrocytic forms and schizonts, involvement of autoimmune reaction is proposed

Theileria annulata
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Theileria annulata
Laiblin, C.; and Mueller, M., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (12), 234-237
Theileria annulata, steers, resochin did not prevent acute symptoms and death, but did prevent chronic disease in recovering animals; prophylactic treatment negative

Theileria annulata (Dschunkowsky, Luhs, 1904), illus.
Mehlhorn, H.; et al., 1975, Ztschr. Parasitenk., v. 48 (2), 137-150
Theileria annulata, developmental stages in intestine and haemolymph of Hyalomma anatolicum excavatum, microgamont-like stages, microgamete-like stages, ookinete-like stages, electron microscopy

Theileria annulata Dschunkowsky & Luhs, 1904, illus.
Mehlhorn, H.; and Schein, E., 1977, J. Protozool., v. 24 (2), 249-257
Theileria annulata in Hyalomma anatolicum excavatum, development of kinetes, electron microscopic studies

Theileria annulata
Morzaria, S. P.; et al., 1977, Research Vet. Sc., v. 22 (3), 330-333
Theileria mutans, comparison of British strain with East and South African strains and other Theileria spp. using indirect fluorescent antibody test, no cross-reactions were detected, concluded that British strain of T. mutans could be a distinct species from African strains of T. mutans

Theileria annulata
Pipano, E.; et al., 1977, Vet. Parasitol., v. 3 (1), 11-22
Theileria annulata, cattle, immunization using killed schizont vaccine, protection against blood challenge but not against tick-transmitted infection, high antibody levels not correlated with protection

Theileria annulata
Pipano, E.; Goldman, M.; and Kapuler, S., 1974, J. Protozool., v. 21 (3), 465 [Abstract]
Theileria annulata-infected lymphocytes, continuous line culture, globulin-producing cells

Theileria annulata
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Theileria annulata
Samish, M., 1977, Nature, London (5632), v. 270, 51-52
Theileria annulata, temperature of 37°C and relative humidity of 95% stimulates production of infective parasite forms in infected adult Hyalomma excavatum ticks without the need for a blood meal

Theileria annulata
Samish, M., 1977, J. Protozool., v. 24 (4), 67A-68A [Abstract]
Theileria annulata, transmission by Hyalomma excavatum, high temperature and humidity may cause uninfective particles in unfed ticks to change into infective ones

Theileria annulata
Samish, M.; and Pipano, E., 1977, J. Protozool., v. 24 (2), Suppl., 19A [Abstract]
Theileria annulata-infected Hyalomma detritum, storage at -79°C. of triturated tick suspensions, successful subsequent infection of calves

Theileria annulata
Samish, M.; Pipano, E.; and Tsafrir, N., 1975, J. Protozool., v. 22 (3), 73A [Abstract]
Theileria annulata, exper. transmission to cattle by Hyalomma detritum

Theileria annulata, illus.
Schein, E., 1975, Ztschr. Parasitenk., v. 47 (2), 165-167
Theileria annulata, microgametes, macrogametes, zygotes, ookinetes, sporonts, in midgut or haemolymph of Hyalomma anatolicum excavatum (exper.)

Theileria annulata
Schein, E., 1976, Ztschr. Parasitenk., v. 50 (2), 205-206
Theileria annulata, developmental stages in Hyalomma anatolicum excavatum

Theileria annulata, illus.
Schein, E.; Buescher, G.; and Friedhoff, K. T., 1975, Ztschr. Parasitenk., v. 48 (2), 123-136
Theileria annulata, developmental stages in gut of engorged Hyalomma anatolicum excavatum nymphs, microgamonts, microgametes, macrogamonts, macrogametes, zygotes, kinetes, light microscopy

Theileria annulata (Dschunkowsky, Luhs, 1904), illus.
Schein, E.; Mehlhorn, H.; and Warnecke, M., 1977, Tropenmed. u. Parasitol., v. 28 (3), 349-360
Theileria annulata, fine structural study of erythrocytic stages with 2 forms (comma-shaped stage and spherical stage) found within the erythrocytes

Theileria annulata
Sharma, L. D.; et al., 1977, Indian Vet. J., v. 54 (12), 979-983
Theileria annulata, virulent strain, splenectomized calves (exper.), amodiaquine hydrochloride and imidocarb dipropionate reduced parasitema and mortality, and controlled pyrexia, dyspnea and tachycardia (criteria for drug success)

Theileria annulata
Sharma, L. D.; Sabir, M.; and Bhattacharyya, N. K., 1976, Indian J. Physiol. and Pharmacol., v. 20 (2), 69-70
Theileria annulata, bull calves (exper.), diethylcarbamazine citrate, good results

Theileria annulata
Singh, D. K.; Jagdish, S.; and Gautam, O.,P., 1977, Research Vet. Sc., v. 23 (3), 391-392
Theileria annulata, cattle, cell-mediated immunity demonstrated using leucocyte migration inhibition test and delayed skin hypersensitivity reaction

Theileria annulata
Srivastava, P. S.; and Sharma, N. N., 1975, Indian J. Animal Research, v. 9 (1), 8-10
Theileria annulata premune calves, oral administration of prednisolone, immunosuppressive effect, clinical relapses; need for caution in use of corticosteroids in enzootic areas; relapse induction useful for detection of carriers

Theileria annulata
Srivastava, P. S.; and Sharma, N. N., 1976, Indian J. Animal Research, v. 10 (1), 21-26
Hyalomma anatolicum anatolicum, laboratory rearing and maintenance on rabbits and cattle, maintenance of Theileria annulata in ticks for transmission experiments

Theileria annulata
Srivastava, P. S.; and Sharma, N. N., 1976, Acta Vet. Beograd, v. 26 (6), 315-321
Theileria annulata-infected calves, splenectomy of T. annulata premune calves provoked only mild relapses, severe disease symptoms manifested in calves splenectomized during febrile reaction period

Theileria annulata
Srivastava, P. S.; and Sharma, N. N., 1976, Internat. J. Radiation Biol., v. 29 (2), 159-168
Hyalomma anatolicum, effects of ^{60}Co irradiation on development; 1000 R to ticks infected with Theileria annulata failed to prevent transmission to calves

Theileria annulata (Dschunowsky & Luhs, 1904)
Srivastava, P. S.; and Sharma, N. N., 1976, Pantnagar J. Research, v. 1 (1), 70-72
Theileria annulata, cross-bred calves, infectivity and immunogenicity of infected washed bovine erythrocytes, calves highly susceptible to tick challenge (infected Hyalomma anatolicum)

Theileria annulata, illus.
Srivastava, P. S.; and Sharma, N. N., 1976, Pantnagar J. Research, v. 1 (2), 83-88
Theileria annulata, calves (exper.), morphological features and characteristics, Hyalomma anatolicum anatolicum-transmitted virulent strain

Theileria annulata, illus.
Srivastava, P. S.; and Sharma, N. N., 1976, Pantnagar J. Research, v. 1 (2), 147-150
Theileria annulata, crossbred calves (exper.), lesions, histopathological changes of cerebral tissues

Theileria annulata
Srivastava, P.S.; and Sharma, N.N., 1977, Vet. Parasitol., v.3 (1), 23-31
Theileria annulata, calves, immunoprophylaxis using schizonts attenuated by cobalt-60 irradiation in bovine lymphocytes

Theileria annulata
Srivastava, P. S.; and Sharma, N. N., 1977, Vet. Parasitol., v. 3 (2), 183-188
Theileria annulata in salivary gland suspensions of Hyalomma anatolicum, attenuation by cobalt-60 irradiation, further investigations needed to test suitability of technique for immunoprophylaxis

Theileria annulata
Tsafrir, N.; Pipano, E.; and Hadani, A., 1973, J. Protozool., v. 20 (4), 534
Hyalomma detritum was able to transmit Theileria annulata regardless of mode of infection of calves on which fed during preimaginal stage (natural tick transmission vs. blood inoculation from acutely or chronically infected calves)

Theileria barnetti
Schreuder, B.E.C.; Uilenberg, G.; and Tondeur, W., 1977, Tropenmed. u. Parasitol., v. 28 (3), 367-371
"It therefore appears to be possible that T. barnetti is a synonym for T. mutans." Theileriidae, reinfection and retransmission experiments in cattle and African buffalo

Theileria cervi
Durham, K. A.; Corstvet, R. E.; and Hair, J. A., 1976, J. Parasitol., v. 62 (6), 1000-1002
fluorescent antibody technique suitable for identification of Theileria cervi in salivary glands or oral secretions of Amblyomma americanum (laboratory infected and field collected), high infection rates of field collected ticks indicate potentially a very efficient vector: eastern Oklahoma

Theileria dispar
Angelovski, T.; and Iliev, A., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 17-20
Theileria dispar, cattle, occurrence of H[yalomma] excavatum and H[yalomma] detritum on most sick animals: Macedonia

Theileria gorgonis
Burridge, M. J.; and Kimber, C. D., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 186-191
Theileria spp. antigens tested against sera from cattle inoculated with wildebeest blood containing Theileria gorgonis or cattle recovered from infection with T. parva, T. lawrencei, or T. mutans, indirect fluorescent antibody test, results show antigenic distinctness of T. gorgonis with only slight degree of cross-reaction with other Theileria spp.

Theileria gorgonis
Grootenhuis, J. G.; et al., 1975, J. Wildlife Dis., v. 11 (1), 122-127
impala Theileria, no cross-reactivity with other theilerial parasites, indirect fluorescent antibody test

Theileria gorgonis
Morzaria, S. P.; et al., 1977, Research Vet. Sc., v. 22 (3), 330-333
Theileria mutans, comparison of British strain with East and South African strains and other Theileria spp. using indirect fluorescent antibody test, no cross-reactions were detected, concluded that British strain of T. mutans could be a distinct species from African strains of T. mutans

Theileria gorgonis
Purnell, R. E.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 181-185
Theileria gorgonis exacerbation in 4 splenectomized Connochaetes taurinus, failure to transmit to cattle by blood passage or via Rhipicephalus appendiculatus ticks despite establishment of parasite in ticks; exacerbated Anaplasma marginale infection in C. taurinus was successfully blood-passaged to calves; results indicate that blue wildebeest is probably not a reservoir for T. gorgonis but may be for A. marginale: Tanzania; Kenya

Theileria gorgonis, illus.
Smith, K.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (3), 293-300
Theileria gorgonis, Anaplasma marginale, intra-erythrocytic stages isolated from splenectomized Connochaetes taurinus, fine structure

T[heileria] hirci (Levine, 1973), illus.
Gautam, O. P.; et al., 1975, Haryana Vet., v. 14 (2), 94-100
T[heileria] hirci, morphology, sheep, case history, pathology, diagnosis, steclin, berenil, nivaquine, fair results: Hissar, transferred to Kameri

Theileria hirci
Hawa, N.; Latif, B. M. A.; and Bakir, F. A., 1976, Trop. Animal Health and Prod., v. 8 (2), 97-101
Theileria hirci, sheep (nat. and exper.), diagnosis, indirect fluorescent antibody test, serological response of host to cell culture schizont antigen

Theileria lawrencei
Burridge, M. J.; and Kimber, C. D., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 186-191
Theileria spp. antigens tested against sera from cattle inoculated with wildebeest blood containing Theileria gorgonis or cattle recovered from infection with T. parva, T. lawrencei, or T. mutans, indirect fluorescent antibody test, results show antigenic distinctness of T. gorgonis with only slight degree of cross-reaction with other Theileria spp.

Theileria lawrencei (Serengeti)
Grootenhuis, J. G.; et al., 1975, J. Wildlife Dis., v. 11 (1), 122-127
impala Theileria, no cross-reactivity with other theilerial parasites, indirect fluorescent antibody test

Theileria lawrencei
Lawrence, J. A., 1977, Vet. Rec., v. 100 (22), 470-471
Theileria parva (Muguga) and T. lawrencei from Rhodesia serologically indistinguishable, indirect immunofluorescent test

Theileria lawrencei
Morzaria, S. P.; et al., 1977, Research Vet. Sc., v. 22 (3), 330-333
Theileria mutans, comparison of British strain with East and South African strains and other Theileria spp. using indirect fluorescent antibody test, no cross-reactions were detected, concluded that British strain of T. mutans could be a distinct species from African strains of T. mutans

Theileria lawrencei
Purnell, R. E.; et al., 1974, J. Comp. Path., v. 84 (4), 533-537
Theileria lawrencei Serengeti strain, comparative infectivity for cattle of stabilates derived from adult vs. nymphal Rhipicephalus appendiculatus

Theileria lawrencei
Radley, D. E.; et al., 1975, Vet. Parasitol., v. 1 (1), 35-41
Bos taurus, chemoprophylactic (oxytetracycline) immunization against Theileria parva (Muguga) or each of 5 recently-isolated theilerial strains (4 T. parva and 1 T. lawrencei), response to homologous and heterologous challenge

Theileria lawrencei
Radley, D. E.; et al., 1975, Vet. Parasitol., v. 1 (1), 43-50
Bos taurus immunized either by chemoprophylaxis (oxytetracycline) or sub-lethal infection against various strains of Theileria lawrencei or T. parva, response to homologous and heterologous challenge

Theileria lawrencei
Radley, D. E.; et al., 1975, Vet. Parasitol., v. 1 (1), 51-60
Bos taurus, chemoprophylactic (oxytetracycline) immunization using a combination of theilerial strains (Theileria parva and T. lawrencei), response to challenge better than animals immunized with only 1 or 2 strains

Theileria lawrencei, illus.
Stagg, D. A.; et al., 1974, Research Vet. Sc., v. 16 (1), 125-127
establishment of Theileria lawrencei-infected lymphoblastoid cell lines from a Syncerus caffer calf obtained from an enzootic area in Kenya, use of bovine embryo spleen feeder layers

Theileria lawrencei
Wagner, G. G.; and Duffus, W. P. H., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 97-107
anti-lymphocyte antibody response in cattle inoculated with Theileria parva- or T. lawrencei-infected lymphoblastoid cell lines, apparently unrelated to specific antibody response to parasite itself,does not appear to either interfere with or enhance development of subsequent immunity to challenge

Theileria lawrencei
Young, A. S.; et al., 1977, J. Parasitol., v. 63 (5), 903-907
Theileria lawrencei-infected Rhipicephalus appendiculatus, experimental field population established in a paddock and maintained by Syncerus caffer, transmission to cattle introduced into paddock, the lethal T. lawrencei challenge established in the paddock may be suitable for testing the efficacy of T. lawrencei immunization procedures for cattle

Theileria lawrencei
Young, A. S.; et al., 1977, Vet. Parasitol., v. 3 (4), 283-290
cattle immunized with Theileria lawrencei stabilates from Rhipicephalus appendiculatus either by fortuitous recovery or by chemoprophylaxis with oxytetracycline, survival without signs of clinical disease after prolonged natural challenge to T. lawrencei derived from Syncerus caffer

Theileria lawrencei
Young, A. S.; and Purnell, R. E., 1973, Trop. Animal Health and Prod., v. 5 (3), 146-152
tick (Rhipicephalus appendiculatus) transmission of Theileria lawrencei (Serengeti) from buffalo to cattle and between intact cattle (Bos taurus), difficult to produce infective stabilates

Theileria mutans
Brocklesby, D. W.; Morzaria, S. P.; and Harradine, D. L., 1975, Tropenmed. u. Parasitol., v. 26 (3), 295-302
Theileria mutans in cattle (exper.), experimental transmission by Haemaphysalis punctata, only transstadial transmission occurred, and after 7 passages there was no evidence of increased virulence

Theileria mutans
Burridge, M. J.; and Kimber, C. D., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 186-191
Theileria spp. antigens tested against sera from cattle inoculated with wildebeest blood containing Theileria gorgonis or cattle recovered from infection with T. parva, T. lawrencei, or T. mutans, indirect fluorescent antibody test, results show antigenic distinctness of T. gorgonis with only slight degree of cross-reaction with other Theileria spp.

Theileria mutans
Dhar, S.; and Gautam, O. P., 1977, Indian Vet. J., v. 54 (1), 21-24
Theileria, cattle, species differentiation using complement-fixation test

Theileria mutans (Aitong)
Grootenhuis, J. G.; et al., 1975, J. Wildlife Dis., v. 11 (1), 122-127
impala Theileria, no cross-reactivity with other theilerial parasites, indirect fluorescent antibody test

Theileria mutans, illus.
Kimber, C. D.; and Young, A. S., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 1-10
Theileria mutans (Aitong), cattle infected by cyclical and mechanical transmission, response to infection studied using indirect fluorescent antibody technique with piroplasm and schizont antigens, indistinguishable reactions with different strains of T. mutans but easily distinguishable reactions from other Theileria spp.

Theileria mutans
Kuttler, K. L.; and Craig, T. M., 1975, Am. J. Vet. Research, v. 36 (3), 323-325
splenectomized calves inoculated with pooled blood samples from eastern Texas cattle developed dual infections of Anaplasma marginale and a Theileria resembling Theileria mutans, treatment with gloxazone and imidocarb selectively eliminated Anaplasma, pure Theileria infections only mildly pathogenic

Theileria mutans
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Theileria mutans
Loehr, K.-F.; and Meyer, H., 1974, Tropenmed. u. Parasitol., v. 25 (3), 288-292
Theileria parva, T. mutans, capillary-tube agglutination and indirect fluorescent antibody tests on sera from Masai cattle (reacted with both species) and 5 game animals (reacted only with T. parva), intra erythrocytic "theileria organisms" found in all game animals but could not be needle-transmitted to splenectomized calves, "one theilerial species which was clearly of T. parva and not of T. mutans antigenicity", "cannot, however, be excluded that these piroplasms may not belong to members of the genus Cytauxzoon": Kapiti Plains near Konza, Kenya

Theileria mutans
Mehlitz, D.; and Ehret, R., 1974, Tropenmed. u. Parasitol., v. 25 (1), 3-10
Anaplasma, Babesia, Theileria, cattle, serological survey, capillary-tube agglutination test, indirect fluorescent antibody test, and complement fixation test: Botswana

Theileria mutans
Mohammed, A. N., 1976, Tijdschr. Diergeneesk., v. 101 (8), 413-420
Friesian cattle (blood), first finding, experimental transmission by H[aemaphysalis] punctata: Netherlands

Theileria mutans
Mohammed, A. N.; and Leeflang, P., 1975, Samaru Agric. Newsletter, v. 17 (2), 75-77
cattle (blood): Shika, Nigeria

Theileria mutans
Morzaria, S. P.; et al., 1977, Research Vet. Sc., v. 22 (3), 330-333
Theileria mutans, comparison of British strain with East and South African strains and other Theileria spp. using indirect fluorescent antibody test, no cross-reactions were detected, concluded that British strain of T. mutans could be a distinct species from African strains of T. mutans

Theileria mutans
Morzaria, S. P.; Brocklesby, D. W.; and Harradine, D. L., 1977, Vet. Rec., v. 100 (23), 484-487
Babesia major, Theileria mutans, cattle (exper.), indirect fluorescent antibody test found to be species specific, possible use in field survey of cattle in Britain

Theileria mutans
Mpangala, C.; Uilenberg, G.; and Schreuder, B. E. C., 1976, Tropenmed. u. Parasitol., v. 27 (2), 192-196
serological characterization of Haematoxenus veliferus from cattle using the indirect fluorescent antibody test, no cross-reactions with either Theileria parva or T. mutans

Theileria mutans (Aitong)
Purnell, R. E.; et al., 1975, J. Parasitol., v. 61 (4), 725-729
comparison of development of Theileria mutans in adult Amblyomma variegatum with T. parva in Rhipicephalus appendiculatus

Theileria mutans
Purnell, R. E.; Brocklesby, D. W.; and Harradine, D. L., 1975, Vet. Parasitol., v. 1 (2), 199-202
Babesia divergens, B. major, Theileria mutans, successful cryopreservation using either dimethyl sulphoxide or glycerol as cryoprotectants

Theileria mutans
van Rensburg, I. B. J., 1976, J. South African Vet. Ass., v. 47 (2), 137-141
Theileria mutans, bovine cerebral theileriosis, 5 case reports, splenic infarction: South Africa

Theileria mutans
Robson, J.; et al., 1977, Trop. Animal Health and Prod., v. 9 (4), 219-231
East Coast fever immunization of Zebu cattle with 3 isolates of T. parva, natural challenge of T. parva and T. mutans, some protection: Uganda

Theileria mutans
Rodriguez, O. N.; Espaine, L.; and Rivas, A., 1975, Rev. Cubana Cien. Vet., v. 5 [on cover], v. 6 [on title page] (1-2), 145-153
Theileria mutans, incidence in cattle with or without Anaplasma marginale, Babesia bigemina and B. argentina natural infections: Cuba

Theileria mutans
Schreuder, B. E. C.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (1), 26-34
Theileria parva, immunization of cattle using an infection and treatment method; attempted immunization against pathogenic T. mutans using an apathogenic strain

Theileria mutans
Schreuder, B. E. C.; and Uilenberg, G., 1976, Tropenmed. u. Parasitol., v. 27 (4), 422-426
technique for infecting ticks (Amblyomma variegatum and Rhipicephalus appendiculatus) with cattle Theileriidae by injecting bovine blood with high parasitemia through cuticle into fully engorged nymphs, after moulting ticks transmitted parasite to susceptible calf

Theileria mutans
Schreuder, B.E.C.; Uilenberg, G.; and Tondeur, W., 1977, Tropenmed. u. Parasitol., v. 28 (3), 367-371
"It therefore appears to be possible that T. barnetti is a synonym for T. mutans." Theileriidae, reinfection and retransmission experiments in cattle and African buffalo

Theileria mutans
Trees, A. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 287 [Demonstration]
blood from healthy indigenous Nigerian cattle inoculated into splenectomized calf, calf became infected with Babesia bigemina, B. bovis, Anaplasma marginale, Borrelia theileri, Theileria mutans, Eperythrozoon teganodes, E. wenyoni ; isolation of pure stabilate of B. bigemina

Theileria mutans
Uilenberg, G.; et al., 1976, Tropenmed. u. Parasitol., v. 27 (3), 329-336
Theileria parva, cattle immunized by infection and treatment method using 3 strains of T. parva survived exposure to natural tick infestation; all controls died from T. parva infections, 3 immunized cattle died from T. mutans or Babesia bigemina infection but none from T. parva

Theileria mutans (Theiler 1906)
Uilenberg, G.; et al., 1977, Austral. Vet. J., v. 53 (6), 271-273
Theileria mutans, African, serological and morphological comparisons with Theileria of cattle in Australia and Britain; results indicate that Australian and British theilerias were similar to each other but different from T. mutans

Theileria mutans (Theiler, 1906)
Uilenberg, G.; Robson, J.; and Pedersen, V., 1974, Tropenmed. u. Parasitol., v. 25 (2), 207-216
Amblyomma variegatum, successful transstadial transmission of Theileria mutans, doubtful transmission of Theileria parva; Rhipicephalus appendiculatus, successful transmission of Theileria parva but transmission of Theileria mutans needs confirmation

Theileria mutans
Uilenberg, G.; and Schreuder, B. E. C., 1976, Tropenmed. u. Parasitol., v. 27 (1), 106-111
successful transstadial transmission of Haematoxenus veliferus by Amblyomma variegatum in exper. cattle; Theileria mutans and rickettsial organism transmitted at the same time

Theileria mutans
Uilenberg, G.; Schreuder, B. E. C.; and Mpangala, C., 1976, Tropenmed. u. Parasitol., v. 27 (3), 323-328
Theileria mutans, successful experimental transmission to cattle by Amblyomma variegatum; attempted transmission using Rhipicephalus appendiculatus unsuccessful in 7 trials, suggests possibility that T. mutans in East Africa is different species from that of South Africa

Theileria ovis
Leeflang, P.; and Ilemobade, A. A., 1977, Trop. Animal Health and Prod., v. 9 (4), 211-218
diseases known to be or possibly tick-borne, domestic animals, research summary, 1966-1976: northern Nigeria

Theileria ovis Rodhain, 1916
Neitz, W. O., 1972, Onderstepoort J. Vet. Research, v. 39 (2), 83-85
Theileria ovis, experimental transmission by Rhipicephalus evertsi mimeticus and R. bursa to splenectomized sheep; reaction of sheep to infestation

Theileria parva
Allsopp, B. A.; et al., 1977, J. Gen. Microbiol., v. 100 (2), 319-328
Theileria parva, purification and characterization of precipitating antigens from schizont and piroplasm stages, results indicate that precipitating antigens are not involved in immune mechanism responsible for immunity to East Coast fever

Theileria parva
Brown, C. G. D.; et al., 1973, Nature (5420), v. 245, 101-103
Theileria parva, infection and transformation of bovine lymphoid cells in vitro using both in vitro tick feed material and ground tick supernatant (Rhipicephalus appendiculatus)

Theileria parva
Brown, C. G. D.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (3), 342-348
Theileria parva, immunization of cattle against East Coast fever by inoculation of a tick-derived (Rhipicephalus appendiculatus) stabilate of T. parva infective particles and the intramuscular injection at daily intervals of N-pyrrolidinomethyl tetracycline

Theileria parva
Burdin, M. L.; and Boarer, C. D. H., 1972, Vet. Rec. (4894), v. 90 (11), 299-302
Theileria parva, Bos indicus, B. taurus and mixed breeds, glucose 6 phosphate dehydrogenase levels, haemoglobin types, relation to resistance: East Africa

Theileria parva
Burridge, M. J.; and Kimber, C. D., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 186-191
Theileria spp. antigens tested against sera from cattle inoculated with wildebeest blood containing Theileria gorgonis or cattle recovered from infection with T. parva, T. lawrencei, or T. mutans, indirect fluorescent antibody test, results show antigenic distinctness of T. gorgonis with only slight degree of cross-reaction with other Theileria spp.

Theileria parva
Burridge, M. J.; and Kimber, C. D., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 305-308
Theileria parva, cattle, levels of antibodies in colostrum of dams recovered from exper. East Coast Fever and in the sera of their calves, indirect fluorescent antibody studies

Theileria parva
Cunningham, M. P., 1977, Advances Exper. Med. and Biol., v. 93, 189-207
Theileria parva, cattle, immunization, review: stabilates, tissue culture, investigation of quantum of infection hypothesis, irradiation of protozoites harvested from ticks, infection and treatment with drugs in tetracycline series, duration of immunity, cross-immunity, field trials

Theileria parva
Cunningham, M. P.; et al., 1973, Isotopes and Radiation Parasitol. III, 145-154
Theileria parva harvested from infected Rhipicephalus appendiculatus, relationship between number of infective particles inoculated into cattle and severity of ensuing infection; effects of irradiation at various doses on suspensions of infective particles of high or low concentration subsequently inoculated into cattle

Theileria parva
Cunningham, M. P.; et al., 1973, J. Protozool., v. 20 (2), 298-300
Theileria parva infective particles harvested from Rhipicephalus appendiculatus using an in vitro feeding technique and subjected to cobalt irradiation, increasing doses of irradiation destroyed increasing numbers of IP's, no evidence that IP's which survived were attenuated, appears unlikely that vaccination against East Coast fever could be achieved using these methods

Theileria parva
Danskin, D.; and Wilde, J. K. H., 1976, Nature, London (5558), v. 261, 311-312
Theileria parva, development to ultimate intraerythrocytic stage in vitro

Theileria parva
Danskin, D.; and Wilde, J. K. H., 1976, Trop. Animal Health and Prod., v. 8 (3), 175-185
cultivation of Theileria parva-infected lymphoid cells, addition of calf lymph and bovine red blood cells caused stimulation of lymphoid cells and parasites, formation of developmental stages

Theileria parva, illus.
DeMartini, J. C.; and Moulton, J. E., 1973, J. Comp. Path., v. 83 (3), 281-298
Theileria parva, cattle (exper.), origin of parasitized lymphoblasts, routes of their metastasis, ultrastructural features of affected lymph nodes and parasitized cells

Theileria parva
DeMartini, J. C.; and Moulton, J. E., 1973, J. Comp. Path., v. 83 (3), 299-306
Theileria parva, calves (exper.), lymph samples collected from tracheal vs. thoracic lymph ducts at various stages of East Coast fever, comparison of prevalence of parasitized cells, number of mitoses, and size distribution of lymphocytes

Theileria parva
Duffus, W. P. H.; and Wagner, G. G., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 85-95
Theileria parva, cattle, rosette-forming cell technique as an indicator of cell-mediated immune response

Theileria parva (Muguga)
Grootenhuis, J. G.; et al., 1975, J. Wildlife Dis., v. 11 (1), 122-127
impala Theileria, no cross-reactivity with other theilerial parasites, indirect fluorescent antibody test

Theileria parva, illus.
Irvin, A. D., 1976, Vet. Rec., v. 98 (18), 351-356
fusion of Theileria parva-infected bovine lymphoid cells and hamster cells, detection of bovine/hamster heterokaryons by presence of H thymidine labelled nuclei and unlabelled nuclei in the same cell, autoradiography, possible use as laboratory model for East Coast fever, review

Theileria parva
Irvin, A. D.; et al., 1973, Isotopes and Radiation Parasitol. III, 155-159
Theileria parva-macroschizont-infected bovine lymphoid cells inoculated into whole-body-irradiated mice, tumors containing macroschizont-infected lymphoid cells developed at inoculation site, no evidence that parasites had invaded mouse cells nor that mouse cells were involved in tumor formation, parasitized cells from tumors successfully transferred back to tissue culture and into other irradiated mice

Theileria parva
Irvin, A. D.; et al., 1975, Nature (5511), v. 255, 713-714
Theileria parva-infected lymphoid cells, growth in irradiated athymic mice (extensive infiltration causing malignant neoplastic condition), comparison with bovine lymphosarcoma cells (discrete circumscribed tumors with no evidence of metastasis)

Theileria parva, illus.
Irvin, A. D.; et al., 1975, Research Vet. Sc., v. 19 (2), 142-151
Theileria parva-infected hybrid cells formed by fusion of hamster or mouse cells with parasitized bovine lymphoid cells using inactivated Sendai virus

Theileria parva
Irvin, A. D.; et al., 1976, J. Comp. Path., v. 86 (1), 51-57
Theileria parva-infected bovine lymphoid cells grown in mice, immunization of cattle with cells passaged in mice

Theileria parva
Irvin, A. D.; et al., 1977, Vet. Parasitol., v. 3 (2), 141-160
Theileria parva-infected lymphoid cells grown in culture inoculated by different routes into neonatal and adult immunosuppressed Swiss mice and into irradiated and non-irradiated athymic (nude) mice, successful establishment and passage in irradiated nude mice indicate potential usefulness of this model

Theileria parva
Irvin, A. D.; Brown, C. G. D.; and Crawford, J. G., 1972, Research Vet. Sc., v. 13 (6), 589-590
unsuccessful attempts to grow Theileria parva-infected bovine lymphoid cells in a variety of laboratory animal systems (normal mice, newborn mice, hamster cheek pouches, anterior chamber of rabbits' eyes)

Theileria parva
Irvin, A. D.; and Stagg, D. A., 1976, Parasitology, v. 73 (2), xix [Abstract]
Theileria parva in bovine lymphoid cells, purine and pyrimidine metabolism, possibility of exploiting defect in purine metabolism in therapy with folate antagonists

Theileria parva
Irvin, A. D.; and Stagg, D. A., 1977, Exper. Parasitol., v. 41 (1), 172-185
Theileria parva-infected bovine lymphoid cells, apparent defect in purine metabolism suggests that folate antagonists such as aminopterin and methotrexate may have application in therapy of East Coast fever

Theileria parva
Irvin, A. D.; Stagg, D. A.; and Kanhai, G. K., 1976, Research Vet. Sc., v. 21 (2), 197-204
fusion of Theileria parva-infected bovine lymphoid cells with Ehrlich ascites tumor cells, attempts to increase fusion percentage and harvest of heterokaryons

Theileria parva
Joyner, L. P., 1971, Symposia Brit. Soc. Parasitol., v. 9, 1-37
laboratory maintenance of tick-borne Babesia and Theileria, extensive review: collection of field material and infection of laboratory animals; management and preparation of experimental animals; cyclical maintenance; mechanical transmission; tissue culture; low-temperature preservation

Theileria parva
Kimber, C. D.; Purnell, R. E.; and Sellwood, S. A., 1973, Research Vet. Sc., v. 14 (1), 126-127
Theileria parva, detection in salivary glands of Rhipicephalus appendiculatus using fluorescent antibody techniques

Theileria parva
Kimeto, B. A., 1976, Am. J. Vet. Research, v. 37 (4), 443-447
Theileria parva, cattle, sequence of ultrastructural changes of platelets during fatal course of East Coast fever

Theileria parva
Lawrence, J. A., 1977, Vet. Rec., v. 100 (22), 470-471
Theileria parva (Muguga) and T. lawrencei from Rhodesia serologically indistinguishable, indirect immunofluorescent test

Theileria parva
Loehr, K.-F.; and Meyer, H., 1974, Tropenmed. u. Parasitol., v. 25 (3), 288-292
Theileria parva, T. mutans, capillary-tube agglutination and indirect fluorescent antibody tests on sera from Masai cattle (reacted with both species) and 5 game animals (reacted only with T. parva), intra erythrocytic "theileria organisms" found in all game animals but could not be needle-transmitted to splenectomized calves, "one theilerial species which was clearly of T. parva and not of T. mutans antigenicity", "cannot, however, be excluded that these piroplasms may not belong to members of the genus Cytauxzoon"
Connochaetes taurinus
Alcelaphus buselaphus
Aepyceros melampus
Gazella grantii
G. thomsonii
all from Kapiti Plains near Konza, Kenya

Theileria parva
Loehr, K. F.; Ross, J. P. J.; and Meyer, H., 1974, Tropenmed. u. Parasitol., v. 25 (2), 217-226
sera of 1505 game animals of 19 different species screened for antibodies to Anaplasma marginale, Babesia bigemina, and Theileria parva, capillary tube agglutination and indirect fluorescent antibody tests, antibodies more prevalent in sera of antelopes grazing in vicinity of non-dipped cattle than in areas where cattle are either dipped regularly or are not present at all, need for studies on transmission of these organisms from game to cattle and vice versa: Kenya; Tanzania; Uganda; Zambia

Theileria parva
McCall, H., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 286 [Demonstration]
Rhipicephalus appendiculatus, stimulation of acid phosphatase activity in salivary glands from Theileria parva infected and non-infected unfed ticks, parasite maturation not induced

Theileria parva
McHardy, N.; Haigh, A. J. B.; and Dolan, T. T., 1976, Nature, London (5562), v. 261, 698-699
Theileria parva, drug screening in vitro, only Menoctone showed appreciable activity, subsequent tests in cattle artificially infected with ECF confirmed activity of Menoctone against the disease, mode of action not clear but seems to act against schizont or earlier stages and probably along a widely occurring biochemical pathway

Theileria parva
Malmquist, W. A.; and Brown, C. G. D., 1974, Research Vet. Sc., v. 16 (1), 134-135
establishment of Theileria parva-infected lymphoblastoid cell lines from cattle by using homologous feeder layers of bovine embryonic spleen cells

Theileria parva
Martins, M., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 286 [Demonstration]
Rhipicephalus appendiculatus, histochemical studies of salivary glands of non-infected and Theileria parva-infected ticks

Theileria parva, illus.
Mehlhorn, H.; and Schein, E., 1976, Tropenmed. u. Parasitol., v. 27 (2), 182-191
electron microscopic study of developmental stages of Theileria parva in the intestine of the tick Hyalomma anatolicum

Theileria parva
Mehlitz, D.; and Ehret, R., 1974, Tropenmed. u. Parasitol., v. 25 (1), 3-10
Anaplasma, Babesia, Theileria, cattle, serological survey, capillary-tube agglutination test, indirect fluorescent antibody test, and complement fixation test: Botswana

Theileria parva
Morzaria, S. P.; et al., 1977, Research Vet. Sc., v. 22 (3), 330-333
Theileria mutans, comparison of British strain with East and South African strains and other Theileria spp. using indirect fluorescent antibody test, no cross-reactions were detected, concluded that British strain of T. mutans could be a distinct species from African strains of T. mutans

Theileria parva
Mpangala, C.; Uilenberg, G.; and Schreuder, B. E. C., 1976, Tropenmed. u. Parasitol., v. 27 (2), 192-196
serological characterization of Haematoxenus veliferus from cattle using the indirect fluorescent antibody test, no cross-reactions with either Theileria parva or T. mutans

Theileria parva
Mugera, G. M.; Bitakaramire, P. K.; and Munyua, W. K., 1973, Isotopes and Radiation Parasitol. III, 129-137
cattle, immunization against East Coast fever using Theileria parva-infected Rhipicephalus appendiculatus irradiated at graded doses, comparison of immunity produced by attachment of ticks vs. inoculation of tick tissue, circulating antibodies demonstrated by capillary agglutination test

Theileria parva (Theiler, 1904), illus.
Mugera, G. M.; and Munyua, W. K., 1973, Bull. Epizoot. Dis. Africa, v. 21 (1), 51-66
Theileria parva, developmental stages in tick and bovine tissues, electron microscopy

T[heileria] parva
Mugera, G. M.; Mworia, G. M.; and Munyua, W. K., 1973, Bull. Epizoot. Dis. Africa, v. 21 (4), 501-505
T[heileria] parva, calves (exper.), transmission by Rhipicephalus appendiculatus, reduction of parasites and, in some cases, complete recovery and immunity after early treatment with chlortetracycline hydrachloride

Theileria parva
Muhammed, S. I.; Lauerman, L. H., jr; and Johnson, L. W., 1975, Am. J. Vet. Research, v. 36 (4), Part 1, 399-402
Theileria parva, cattle experimentally infected with standardized suspensions of infected Rhipicephalus appendiculatus, treatment with immune serum or concentrated immune globulins, no effect on establishment of infection nor clinical and hematologic changes, immunity seen in cattle recovered from East Coast fever is therefore probably cell-mediated

Theileria parva
Muhammed, S. I.; Wagner, G. G.; and Lauerman, L. H., jr., 1974, Immunology, v. 27 (6), 1033-1037
Theileria parva, cattle, leucocyte migration inhibition as model for demonstration of sensitized cells in East Coast fever

Theileria parva
Munyua, W. K.; et al., 1973, Bull. Epizoot. Dis. Africa, v. 21 (1), 39-49
Theileria parva, pathological lesions in exper. infected cattle

Theileria parva
Munyua, W. K.; Mugera, G. M.; and Bitakaramire, P. K., 1973, Bull. Epizoot. Dis. Africa, v. 21 (1), 75-85
cattle, pathogenesis and pathology of East Coast fever induced by gamma irradiated Theileria parva infected Rhipicephalus appendiculatus, immunogenesis

Theileria parva
Oteng, A. K., 1973, Bull. Epizoot. Dis. Africa, v. 21 (4), 449-459
Uganda strain of Theileria parva, cattle (exper.), symptoms, fever, haematology, parasitaemia, pathology, transmission by blood and Rhipicephalus appendiculatus, no cross-immunity between strains of T. parva

Theileria parva
Oteng, A. K., 1976, Pathophysiol. Parasit. Infect., 211-219
Theileria parva in Bos taurus and B. indicus, fever plays significant and critical role in course and pathogenesis of East Coast Fever

Theileria parva
Purnell, R. E.; et al., 1973, Isotopes and Radiation Parasitol. III, 139-144
Rhipicephalus appendiculatus (unfed nymphs; engorged nymphs; moulting nymphs; unfed adults), effects of irradiation assessed by evaluating subsequent performance of ticks when fed on rabbits (mortality, attachment, feeding, mating, egg-laying); effects of irradiation on Theileria parva in salivary glands of adult ticks

Theileria parva (Muguga)
Purnell, R. E.; et al., 1975, J. Parasitol., v. 61 (4), 725-729
comparison of development of Theileria mutans in adult Amblyomma variegatum with T. parva in Rhipicephalus appendiculatus

Theileria parva
Purnell, R. E.; et al., 1975, Trop. Animal Health and Prod., v. 7 (3), 133-137
Theileria parva-infected ticks (Rhipicephalus appendiculatus), establishment of an experimental field population for field challenge of cattle (Bos taurus) immunized against East Coast fever

Theileria parva
Purnell, R. E.; Radley, D. E.; and Cunningham, M. P., 1974, Vet. Rec., v. 94 (13), 278
sheep and goats not susceptible to experimental infection

Theileria parva
Purnell, R. E.; Sansom, B. F.; and Sellwood, S. A., 1973, Isotopes and Radiation Parasitol. III, 161-163
Theileria parva infective particles harvested from Rhipicephalus appendiculatus, possible technique for quantitative assay by labelling with ^{3}H-thymidine

Theileria parva (Muguga)
Radley, D. E.; et al., 1975, Vet. Parasitol., v. 1 (1), 35-41
Bos taurus, chemoprophylactic (oxytetracycline) immunization against Theileria parva (Muguga) or each of 5 recently-isolated theilerial strains (4 T. parva and 1 T. lawrencei), response to homologous and heterologous challenge

Theileria parva
Radley, D. E.; et al., 1975, Vet. Parasitol., v. 1 (1), 43-50
Bos taurus immunized either by chemoprophylaxis (oxytetracycline) or sub-lethal infection against various strains of Theileria lawrencei or T. parva, response to homologous and heterologous challenge

Theileria parva
Radley, D. E.; et al., 1975, Vet. Parasitol., v. 1 (1), 51-60
Bos taurus, chemoprophylactic (oxytetracycline) immunization using a combination of theilerial strains (Theileria parva and T. lawrencei), response to challenge better than animals immunized with only 1 or 2 strains

Theileria parva
Robson, J.; et al., 1977, Trop. Animal Health and Prod., v. 9 (4), 219-231
East Coast fever immunization of Zebu cattle with 3 isolates of T. parva, natural challenge of T. parva and T. mutans, some protection: Uganda

Theileria parva
Schein, E.; and Warnecke, M., 1976, Ztschr. Parasitenk., v. 50 (2), 211
Theileria parva, developmental stages in intestine of Rhipicephalus appendiculatus and Hyalomma a. excavatum

Theileria parva, illus.
Schein, E.; Warnecke, M.; and Kirmse, P., 1977, Parasitology, v. 75 (3), 309-316
Theileria parva, life cycle in gut of Rhipicephalus appendiculatus, morphology of developmental stages

Theileria parva
Schreuder, B. E. C.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (1), 26-34
Theileria parva, immunization of cattle using an infection and treatment method; attempted immunization against pathogenic T. mutans using an apathogenic strain

Theileria parva
Schreuder, B. E. C.; and Uilenberg, G., 1976, Tropenmed. u. Parasitol., v. 27 (4), 422-426
technique for infecting ticks (Amblyomma variegatum and Rhipicephalus appendiculatus) with cattle Theileriidae by injecting bovine blood with high parasitemia through cuticle into fully engorged nymphs, after moulting ticks transmitted parasite to susceptible calf

Theileria parva, illus.
Smith, K.; Irvin, A. D.; and Stagg, D. A., 1976, Research Vet. Sc., v. 21 (2), 205-214
heterokaryons derived from Theileria parva-infected lymphoblasts and Ehrlich ascites tumor cells, ultrastructure of contained parasites

Theileria parva
Uilenberg, G.; et al., 1976, Tropenmed. u. Parasitol., v. 27 (3), 329-336
Theileria parva, cattle immunized by infection and treatment method using 3 strains of T. parva survived exposure to natural tick infestation; all controls died from T. parva infections, 3 immunized cattle died from T. mutans or Babesia bigemina infection but none from T. parva

Theileria parva (Theiler, 1904)
Uilenberg, G.; Robson, J.; and Pedersen, V., 1974, Tropenmed. u. Parasitol., v. 25 (2), 207-216
Amblyomma variegatum, successful transstadial transmission of Theileria mutans, doubtful transmission of Theileria parva; Rhipicephalus appendiculatus, successful transmission of Theileria parva but transmission of Theileria mutans needs confirmation

Theileria parva
Uilenberg, G.; and Schreuder, B. E. C., 1977, Tropenmed. u. Parasitol., v. 28 (2), 181-184
Rhipicephalus evertsi, successful transmission of Theileria parva from engorged nymphs to cross-bred steers (exper.) confirms the potential of R. evertsi as vector of East Coast fever; similar trials with Amblyomma variegatum were unsuccessful

Theileria parva
Wagner, G. G.; et al., 1975, Research Vet. Sc., v. 19 (2), 209-211
Theileria parva, diminished antibody response to rinderpest vaccination in cattle undergoing severe East Coast fever reactions

Theileria parva
Wagner, G. G.; and Duffus, W. P. H., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 97-107
anti-lymphocyte antibody response in cattle inoculated with Theileria parva- or T. lawrencei-infected lymphoblastoid cell lines, apparently unrelated to specific antibody response to parasite itself, does not appear to either interfere with or enhance development of subsequent immunity to challenge

Theileria parva lawrencei Neitz, 1955 [? n. rank]
Schreuder, B.E.C.; Uilenberg, G.; and Tondeur, W., 1977, Tropenmed. u. Parasitol., v. 28 (3), 367-371
Theileriidae, reinfection and retransmission experiments in cattle and African buffalo "We consider T. lawrencei Neitz, 1955 and T. parva (Theiler, 1904) to be biologically different strains of the same species.... For the sake of facility we prefer to consider them as subspecies of T. parva."

Theileria parva parva
Schreuder, B.E.C.; Uilenberg, G.; and Tondeur, W., 1977, Tropenmed. u. Parasitol., v. 28 (3), 367-371
Theileriidae, reinfection and retransmission experiments in cattle and African buffalo "We consider T. lawrencei Neitz, 1955 and T. parva (Theiler, 1904) to be biologically different strains of the same species.... For the sake of facility we prefer to consider them as subspecies of T. parva."

Theileria sergenti
Takahashi, K.; et al., 1976, Brit. Vet. J., v. 132 (1), 112-117
Theileria sergenti, cattle (exper.), antibody levels, parasitaemia, packed cell volume, indirect fluorescent antibody test on whole serum, IgG, and IgM, possible role of humoral antibody detected by IFA test in inhibition of parasitaemia

Theileria sergenti
Takahashi, K.; Yamashita, S.; and Shimizu, Y., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (5), 295-301
Theileria sergenti, cattle, indirect fluorescent antibody test; relationship of varying antibody titer to course of infection and clinical signs; severe infection in splenectomized calves with no antibody developed; resistance to challenge infection in previously infected calves; recrudescence of infection in calves given corticosteroids; possible humoral factor in immune mechanism and cell-mediated immunity relationship to relapse

Theileria tragelaphi or Cytauxzoon sp., illus.
Bigalke, R. D.; Keep, M. E.; and Schoeman, J. H., 1972, Onderstepoort J. Vet. Research, v. 39 (4), 225-227
Tragelaphus scriptus (blood): Hluhluwe Game Reserve, Natal, Zululand

Theilerial piroplasms, illus.
Irvin, A. D.; et al., 1972, Vet. Rec., v. 91 (21), 513, 514-516, 517
theilerial piroplasms acquired by Rhipicephalus appendiculatus and R. simus (salivary glands) (exper.) fed on naturally infected Taurotragus oryx pattersonianus were not transmitted to cattle, role of eland in epidemiolgy of cattle theilerioses is questionable: Kenya

Theileriasis
Anjaria, J. V.; Jhala, V. M.; and Shersingh, 1976, Gujarat Agric. Univ. Research J., v. 1 (2), 119-124
theileriasis, crossbred and exotic cattle, chloroquine phosphate; clinico-pharmacological approach

Theileriosis
Bazargani, T. T., 1976, J. Vet. Fac. Univ. Tehran, v. 31 (3-4), 1975-1976, 87-92
theileriosis, clinical symptoms, 10-day-old calf, possible congenital infection: Iran

Theileriasis
Burridge, M. J., 1975, J. Wildlife Dis., v. 11 (1), 68-75
bovine theileriasis, role of wild mammals in epidemiology, review: East Africa

Theileriasis
Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
African buffalo as potential reservoir of bovine theileriosis in Botswana

Theileriasis
Rao, G. S.; Surendran, N. S.; and Choudary, K. V., 1973, Indian Vet. J., v. 50 (1), 104-105
theileriasis, exotic and cross-bred cows, treatment, best results using Berenil coupled with Achromycin: Andhra Pradesh

Theileriasis
Soltys, M. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 309-322
immunization against protozoan diseases of animals, review

Theileriasis
Uilenberg, G., 1976, World Animal Rev. (17), 8-15
babesiasis, theileriasis, anaplasmosis, epizootiology of tick-borne diseases, review

Thelohanellus pyriformis, illus.
Gattaponi, P., 1972, Parassitologia, v. 14 (2-3), 311-316
Thelohanellus pyriformis, infection of tench, Tinca tinca, vegetative stages most commonly on gills, description on gills, acid mucopolysaccharides in polar cyst walls, no appreciable damage, seasonal distribution irregular, limnological data not correlated with variation in localization and dimensions: Lake Trasimeno

Thelohanellus pyriformis Thelohan
Gattaponi, P., 1972, Atti Soc. Ital. Sc. Vet., v. 26, 512-517
Tinca tinca: Lake Trasimeno

Thelohania
Codreanu-Balcescu, D.; Codreanu, R.; and Porchet-Hennere, E., 1975, J. Protozool., v. 22 (3), 78A [Abstract]
Thelohania, Trichoduboscqia epeori, mitosis, ultrastructural aspects

Thelohania [sp.]
Desportes, I.; Ginsburger Vogel, T.; and Zerbib, C., 1976, J. Protozool., v. 23 (4), 8A-9A [Abstract]
Orchestia gammarellus (muscles)

Thelohania sp.
Fahy, E., 1975, Entom. Gaz., v. 26 (2), 146-148
recurrent epizootic, symptoms
Rhithrogena semicolorata: Co. Mayo, Ireland

Thelohania [sp.]
Jakowska, S., 1964, An. 2. Cong. Latino-Am. Zool., v. 1, 265-273
Lophius americanus (celulas nervosas): Aquario de New York

Thelohania sp.
Martins, R. R.; and Perondini, A. L. P., 1977, J. Invert. Path., v. 30 (3), 422-428
Thelohania sp.-infected Rhynchosciara angelae (exper.), nuclear hypertrophy and RNA synthesis in striated parietal muscle

Thelohania sp., illus.
Vernick, S. H.; Sprague, V.; and Krause, D., 1977, J. Protozool., v. 24 (1), 94-99
Thelohania sp., Golgi apparatus, ultrastructural and functional aspects
Pandalus jordani: Queen Charlotte Sound, B. C.

Thelohania sp., illus.
Wilson, G. G., 1975, Canad. J. Zool., v. 53 (12), 1799-1802
description
Choristoneura fumiferana: Ontario, Canada [and/or] northern Minnesota, U.S.A.

Thelohania anomala Sen, 1941
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania anomala (Sen 1941)

Thelohania apodemi Doby, Jeannes et Rault, 1963, illus.
Doby, J. M.; Rault, B.; and Barker, R., 1975, Compt. Rend. Soc. Biol., Paris, v. 169 (4), 1053-1056
Thelohania apodemi, intense autofluorescence in presence of ultraviolet radiation, probably due to presence of chitin in spore wall

Thelohania bracteata Strickland
Ebsary, B. A.; and Bennett, G. F., 1975, Canad. J. Zool., v. 53 (8), 1058-1062
Simulium corbis
Simulium latipes
Simulium venustum
Simulium vittatum
all from insular Newfoundland

Thelohania bracteata (Strickland, 1913), illus.
Liu, T. P.; and Davies, D. M., 1973, J. Protozool., v. 20 (5), 622-630
Thelohania bracteata, spore envelope during development, ultrastructural architecture and organization, freeze-etching technique

Thelohania bracteata (Strickland, 1913) Debaisieux & Gastaldi, 1919, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Thelohania ceccaldii [n. sp.]
Vivares, C. P.; and Tuzet, O., 1974, J. Protozool., v. 21 (3), 476 [Abstract]
Thelohania naenadis in Carcinus mediterraneus and C. maenas, T. octospora in Palaemon serratus, T. ceccaldii in Processa edulis, comparative ultrastructure of development

Thelohania ceccaldii n. sp., illus.
Vivares, C. P., 1975, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 17 (2), 141-178
Thelohania maenadis, T. octospora, T. ceccaldii, comparative fine structure
Processa edulis edulis (muscles squelettiques): Marseille

Thelohania columbaczense Weiser, 1960, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Thelohania contejeani Henneguy, 1892, illus.
Quilter, C. G., 1976, N. Zealand J. Marine and Freshwater Research, v. 10 (1), 225-231
Thelohania contejeani, description, infection rate, results suggest that infection does not greatly affect normal locomotor activity of host, but does reduce total muscular power available
Paranephrops zealandicus (musculature): Leith Stream, Dunedin, New Zealand

Thelohania diazoma Kramer, 1965
Fowler, J. L.; and Reeves, E. L., 1974, J. Protozool., v. 21 (4), 538-542
microsporidan isolate previously considered to represent mixed infection of Nosema necatrix and Thelohania diazoma determined to be one species with spore dimorphism, electrophoretic protein profile of two spore types, temperature effects on ratio of mono- to octospores, recommended that name Nosema necatrix be retained for this isolate

Thelohania duorara
Iversen, E. S.; and Kelly, J. F., 1976, J. Invert. Path., v. 27 (3), 407-408
Penaeus duorarum: Biscayne Bay

Thelohania ephestiae Mattes 1927
Weiser, J., 1977, J. Protozool., v. 24 (2), Suppl., 17A [Abstract]
Thelohania ephestiae, occurrence of two types of sporogony

Thelohania fibrata (Strickland)
Boemare, N.; and Maurand, J., 1976, Bull. Soc. Zool. France, v. 101 (3), 377-385
Thelohania fibrata, Pleistophora debaisieuxi, respiratory metabolism of healthy and infected Tetisimulium bezzii larvae compared

Thelohania fibrata Strickland
Ebsary, B. A.; and Bennett, G. F., 1975, Canad. J. Zool., v. 53 (8), 1058-1062
Simulium latipes
Simulium tuberosum
Simulium venustum
Simulium vittatum
all from insular Newfoundland

Thelohania fibrata (Strickland, 1913) Debaisieux & Gastaldi, 1919, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Thelohania herediteria
Bulnheim, H. P., 1977, Biol. Zentralbl., v. 96 (1), 61-78
Thelohania herediteria and Octosporea effeminans in Gammarus d. duebeni (ovary) (nat. and exper.), feminized hosts, androgenic gland development inhibited, partially suppressed endocrine function: Gebiet der Elbmundung (Graben des Deichvorlandes bei Muggendorf)

Thelohania illinoisensis Kudo 1921
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania illinoisensis (Kudo 1921)

Thelohania illinoisensis Kudo (in part), 1922
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania anophelis (Kudo 1924) [n. comb.]

Thelohania indica Kudo, 1929
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania indica (Kudo 1929)

Thelohania legeri Kudo (in part), 1924 [et auct.]
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania anophelis (Kudo 1924) [n. comb.]

Thelohania legeri Kudo (in part), 1924 [et auct.]
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania illinoisensis (Kudo 1921)

Thelohania legeri Hesse, 1904
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania legeri (Hesse 1904)

Thelohania legeri Chapman et al., 1966
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
as conditional synonym of Parathelohania obesa (Kudo 1924)

Thelohania maenadis Perez, 1904, illus.
Vivares, C. P., 1975, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 17 (2), 141-178
Thelohania maenadis, T. octospora, T. ceccaldii, comparative fine structure
Carcinus mediterraneus: l'etang de Lapalme (entre Narbonne et Perpignan)
C. maenas: Bassin d'Arcachon (hemolymphe of all)

Thelohania maenadis
Vivares, C. P.; Bouix, G.; and Manier, J.-F., 1977, J. Protozool., v. 24 (1), 83-94
Carcinus mediterraneus: etangs cotiers de la region du Languedoc mediterraneen

Thelohania maenadis
Vivares, C. P.; Loubes, C.; and Bouix, G., 1975, J. Protozool., v. 22 (3), 45A-46A [Abstract]
Thelohania maenadis, cytochemistry

Thelohania maenadis
Vivares, C. P.; Loubes, C.; and Bouix, G., 1975, J. Protozool., v. 22 (3), 86A [Abstract]
Thelohania maenadis, cytochemistry

Thelohania maenadis Perez, 1904, illus.
Vivares, C. P.; Loubes, C.; and Bouix, G., 1976, Ann. Parasitol., v. 51 (1), 1-14
Thelohania maenadis, Nosema pulvis, cytochemistry
Carcinus mediterraneus: divers etangs littoraux languedociens et catalans

Thelohania maenadis
Vivares, C. P.; Trellu, J.; and Ceccaldi, H. J., 1977, Experientia, v. 33 (10), 1311-1312
Thelohania maenadis-infected vs. uninfected Carcinus mediterraneus, electrophoregrams (proteingram and zymogram) of hemolymph proteins, parasitized crabs show increase in chymotrypsin activity and variations in zones of esterase activity
Carcinus mediterraneus (musculature): etang de Vic, pres de Montpellier, France

Thelohania maenadis
Vivares, C. P.; and Tuzet, O., 1974, J. Protozool., v. 21 (3), 476 [Abstract]
Thelohania naenadis in Carcinus mediterraneus and C. maenas, T. octospora in Palaemon serratus, T. ceccaldii in Processa edulis, comparative ultrastructure of development

Thelohania obesa Kudo, 1924
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania obesa (Kudo 1924)

Thelohania obesa Weiser (in part), 1961
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania indica (Kudo 1929)

Thelohania obesa Weiser (in part), 1961
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull.(1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania obscura (Kudo 1929)

Thelohania obscura Kudo, 1929
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania obscura (Kudo 1929)

Thelohania octospora Henneguy, 1892, illus.
Vivares, C. P., 1975, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 17 (2), 141-178
Thelohania maenadis, T. octospora, T. ceccaldii, comparative fine structure
Palaemon serratus (musculature): La Rochelle

Thelohania octospora
Vivares, C. P.; and Tuzet, O., 1974, J. Protozool., v. 21 (3), 476 [Abstract]
Thelohania naenadis in Carcinus mediterraneus and C. maenas, T. octospora in Palaemon serratus, T. ceccaldii in Processa edulis, comparative ultrastructure of development

Thelohania pallipedis n. sp., illus.
Vivares, C. P., 1975, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 17 (2), 141-178
"les microsporidies ...considerees par les auteurs comme appartenant a l'espece T. contejeani, presentent des difference notables, ce qui m'autorise a attribuer au parasite decrit par Cossins et Bowler (1974) un nom specifique"

Thelohania penaei
Iversen, E. S.; and Kelly, J. F., 1976, J. Invert. Path., v. 27 (3), 407-408
successful transmission of Thelohania penaei by feeding infected trout feces to pink shrimp
Penaeus duorarum (nat. and exper.): Biscayne Bay
Cynoscion nebulosus (exper.)

Thelohania periculosa Kellen and Wills, 1962
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania periculosa (Kellen and Wills 1962)

Thelohania solenopsae n. sp., illus.
Knell, J. D.; Allen, G. E.; and Hazard, E. I., 1977, J. Invert. Path., v. 29 (2), 192-200
light and electron microscope study
Solenopsis invicta (fat body, ovaries): University campus, Cuiaba, Mato Grosso, Brazil

Thelohania varians (Leger, 1897) Debaisieux, 1919, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Thelohaniidae [sp.]
Lainson, R.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 381 [Demonstration]
morphology
Lutzomyia, anduzei group (midgut): Monte Dourado area of North Brazil

Toddia
de Sousa, M. A.; and Weigl, D. R., 1975, J. Protozool., v. 22 (3), 36A [Abstract]
evidence that Toddia is not a protozoan but probably a virus

Toddia [sp.]
Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses
Natrix rhombifera: Mexico
Oxybelis aeneus: Honduras

Toddia [sp.]
de Sousa, M. A.; de Biasi, P.; and Pessoa, S. B., 1973, J. Protozool., v. 20 (4), 530
Bothrops moojeni
B. pradoi
B. jararaca
Chironius flavolineatus
(blood of all): all from Brazil

Toddia sp., illus.
Sullivan, J. S.; and Sullivan, J. J., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (1), 126-128
Leptobrachium hasselti (blood): Peninsular Malaysia

Toxoglugea missiroli Weiser, 1961
Hazard, E. I.; and Anthony, D. W., 1974, Tech. Bull. (1505) U. S. Dept. Agric., 1-26
as syn. of Parathelohania legeri (Hesse 1904)

Toxoplasma
Akao, S., 1977, J. Electr. Micr., v. 26 (3), 266
Toxoplasma, localization of perioxidase activity in mitochondria and dense granules, biochemical characterization

Toxoplasma
Beier, T. V.; Siim, I. K.; and Hutchison, W. M., 1977, Tsitologiia, v. 19 (12), 1362-1368
Eimeriidae, Isosporinae

Toxoplasma, illus.
Berard-Badier, M.; et al., 1968, Path. Biol., v. 16 (19-20), 829-835
acute congenital toxoplasmosis in guinea pigs (exper.), placental lesions varied with Toxoplasma strains used, foetus not affected if placenta lesions were absent

Toxoplasma
Box, E. D., 1975, J. Protozool., v. 22 (2), 165-169
Isospora serini, description of exogenous stages, asexual stages in macrophages are indistinguishable from parasites previously called avian Toxoplasma, Atoxoplasma, and Lankesterella

Toxoplasma
Bozdech, V., 1976, Ang. Parasitol., v. 17 (2), 70-75
toxoplasmin skin test, humans, percentage of incidence, higher frequency in males, in females with increasing age: Kaduna, Nigeria

Toxoplasma
Dale, D. C.; and Petersdorf, R. G., 1973, Med. Clin. North Am., v. 57 (5), 1277-1287
parasitic infections in the immuno-suppressed steroid-treated patient, review

Toxoplasma, illus.
Dobos-Kovacs, M.; and Kardevan, A., 1976, Magy. Allat. Lapja, v. 98, v. 31 (1), 53-61
Toxoplasma, dogs, case histories and histopathology of generalized infection, tachyzoites, concurrent with distemper, generalized form of toxoplasmosis may be regarded as secondary infection associated mainly with distemper, reduced resistance making its manifestation possible

Toxoplasma, illus.
Dominguez, A.; and Giron, J. J., 1976, Acta Cytol., v. 20 (3), 269-271
Toxoplasma cysts in human vaginal smears, need for accurate differential diagnosis from cancer, possible source of congenital toxoplasmosis and/or abortions as well as individual reinfection

Toxoplasma, illus.
Donev, A., 1975, Vet. Med. Nauki, v. 12 (10), 65-69
Toxoplasma, isolation of strains, digestion of diaphragm musculature, sheep, cattle, pigs

Toxoplasma
Droller, M. J.; and Remington, J. S., 1975, Cellular Immunol., v. 19 (2), 349-355
Toxoplasma-infected mice, correlation between decrease in adenyl cyclase activity in lymphocytes and macrophages and resistance to tumor growth, data suggest that production of cyclic AMP by lymphocytes is inhibited with activation of certain cell-mediated immune functions

Toxoplasma
Dubey, J. P., 1976, J. Am. Vet. Med. Ass., v. 169 (11), 1197-1199
Toxoplasma not isolated from bovine tissues administered to cats and mice; cattle serum examined for antibodies; absence of Toxoplasma infection in cattle: Ohio abattoir

Toxoplasma
Dymowska, Z.; Migdalska, Z.; and Kraus, A., 1971, Med. Dosw. i Mikrobiol., v. 23 (2), 167-173
Toxoplasma, immunized rabbits (exper.), evaluation of immune and cellular reactions using percent pattern of hemogram

Toxoplasma Nicolle and Manceaux 1909
Frenkel, J. K., 1977, J. Parasitol., v. 63 (4), 611-628
Sarcocystidae, Toxoplasmatinae
diagnosis, life cycle pattern

Toxoplasma, illus.
van Furth, R.; and Jones, T. C., 1975, Infect. and Immun., v. 12 (4), 888-890
mouse peritoneal macrophages that have ingested killed Toxoplasma, hydrocortisone had no effect on phagosome-lysosome interaction, no change in intracellular microbicidal activity in macrophages treated with glucocorticosteroids

Toxoplasma
Hagiwara, T.; Katsube, Y.; and Hanaki, T., 1976, Japan. J. Vet. Sc., v. 38 (5), 517-520
Toxoplasma, swine, dye and hemagglutination tests evaluated for detection of latent infection

Toxoplasma
Hans, J. C., 1975, Ann. Med. Vet., v. 119 (7), 429-433
Toxoplasma, incidence in pigs, cattle, sheep, Sabin-Feldman dye test, higher incidence in sheep: Belgian slaughterhouse

Toxoplasma
Ito, S.; et al., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 229-234
Toxoplasma, strains from cats infected with Isospora bigemina, oocysticidal effect of disinfectants, alcohols and other chemicals, only Lomasept, Neo Kurehasol, ethanol, methanol, peracetic acid, ammonia water, ammonium sulfide, and formalin were effective

Toxoplasma
Ito, S.; and Tsunoda, K., 1975, Japan Agric. Research Quart., v. 9 (4), 221-224
piglets inoculated with 2 strains of Toxoplasma oocysts: natural transfer to uninfected piglets in same pen, exper. transfer to mice from infected visceral organs of piglets; piglets nat. and exper. infected with toxoplasmosis by feeding on leaf-moulds contaminated with cat feces, treatment with 2-sulfamoyl-4,4'-diaminodiphenyl-sulfone (SDDS)

Toxoplasma
Janitschke, K., 1976, Ztschr. Parasitenk., v. 50 (2), 208-209
Toxoplasma, transmission of oocysts from cats to rabbits

Toxoplasma
Katsube, Y.; et al., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 245-252
Toxoplasma, latent infection in swine grouped according to presence or absence of gross lesions of lymph nodes and liver, comparison of site of localization and virulence of isolated strains for mice

Toxoplasma
Landau, I., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 403-407
analysis of analogy between Toxoplasma and Hepatozoon life cycles; importance and role of cyst formation in the Coccidia

Toxoplasma Nicolle and Manceaux 1909
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
Eimeriidae
diagnosis

Toxoplasma Nicolle & Manceaux, 1909
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
diagnosis, taxonomic revision, assigned to Eimeriidae

Toxoplasma
Long, P. L.; and Speer, C. A., 1977, Symposia Brit. Soc. Parasitol., v. 15, 1-26
coccidia, invasion of host cells, symposium presentation: general considerations; excystation; invasion of cells by Eimeria in vivo; host reactions to invasion by Eimeria; invasion of cells by Eimeria sporozoites in vitro; factors affecting invasion of cells by sporozoites; invasion of cultured cells by merozoites; invasion of cells by Toxoplasma

Toxoplasma
Lunde, M. N.; and Fayer, R., 1977, J. Parasitol., v. 63 (2), 222-225
Sarcocystis, soluble antigen prepared from zoites obtained by pepsin digestion techniques, indirect hemagglutination test and agar gel diffusion test in cattle, possible use in diagnosis, antigen did not cross-react with sera of Toxoplasma-positive humans and did not react with sera from Sarcocystis-infected dogs

Toxoplasma
Murosaku, A., 1976, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 29 (5), 263-267
Toxoplasma antibody survey, cats, indirect hemagglutination test, medication with 2-sulfamoyl-4,4'-diaminodiphenylsulfone, no negative reactions changed to positive, no increase in antibody titer in positive cats

Toxoplasma
Peychl, L.; Zastera, M.; and Dostal, V., 1976, J. Hyg., Epidemiol., Microbiol. and Immunol., v. 20 (2), 248-251
Toxoplasma, human nodular, pathology, clinical course, immunity, 6-year study of 93 subjects: Czechoslovakia

Toxoplasma
Rakovec, R.; and Senk, L., 1975, Zborn. Bioteh. Fak. Univ. Ljubljani, v. 12 (1), 165-171
Isospora bigemina, sporulated oocysts from cats infecting mice, resulting in cysts in brain similar to Toxoplasma, structure; negative results in reinfecting other mice with brain cysts; negative Sabin-Feldman test results in all mice

Toxoplasma
Rasin, K., 1973, Vet. Med., Praha, v. 46, v. 18 (10), 619-624
Toxoplasma, incidence in Ondatra shown by complement fixation test and isolation, importance of muskrats as reservoir hosts and migratory carriers: Czechoslovakia

Toxoplasma
Rasin, K.; and Kovarik, K., 1973, Vet. Med., Praha, v. 46, v. 18 (10), 625-631
Toxoplasma, incidence in guinea pigs shown by complement fixation test, incidence by sex; isolation of 31 strains, low pathogenicity for mice

Toxoplasma
Rifaat, M. A.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 118-120
toxoplasmosis, immunoserological survey conducted by randomly sampling the populations of governorates in Egypt

Toxoplasma
Ruiz, A.; and Frenkel, J. K., 1977, J. Parasitol., v. 63 (5), 931-932
Toxoplasma isolated from cat feces deposited in false attics of homes, survival of oocysts under these conditions in field experiments, possible role in transmission: Costa Rica

Toxoplasma
Saari, M.; et al., 1976, Arch. Ophth., Chicago, v. 94 (9), 1485-1488
toxoplasmic chlorioretinitis accompanying acute acquired systemic infection, case report of man probably infected through saliva or nasal secretions of infected cow, treated with pyrimethamine in conjunction with sulfadiazine and prednisone: Finland

Toxoplasma
Sasaki, S.; et al., 1976, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 29 (2), 77-82
Toxoplasma, collective outbreak on swine breeding farm, sulfamonomethoxine

Toxoplasma cysts
Seamon, P. J.; et al., 1977, Vet. Rec., v. 101 (16), 324-325
Toxoplasma cysts, mice (exper.), latex agglutination test as a rapid diagnostic method, application to diagnosis of ovine abortion

Toxoplasma
Shettigara, P. T.; Choi, N. W.; and Abu-Zeid, H. A. H., 1976, Am. J. Epidemiol., v. 104 (3), 340 [Abstract]
prevalence survey of prenatal sera for presence of Toxoplasma antibodies and correlation of findings with ethnic origins, socioeconomic status and climatic differences: Manitoba Province

Toxoplasma
Shirahata, T.; Shimizu, K.; and Suzuki, N., 1976, Ztschr. Parasitenk., v. 49 (1), 11-23
Toxoplasma, effects of immune lymphocyte products and serum antibody on parasite multiplication in murine peritoneal macrophages

Toxoplasma
Sogandares-Bernal, F.; et al., 1975, J. Parasitol., v. 61 (5), 965-966
higher prevalence of Toxoplasma antibodies in dairy vs. range cattle, possible danger of human infection by drinking unpasteurized milk or milk from "Certified" herds: Bitterroot Valley, Montana

Toxoplasma
Stagno, S.; et al., 1977, Pediatrics, Am. Acad. Pediat., v. 59 (5), 669-678
human congenital Toxoplasma, auditory and visual defects resulting from symptomatic and subclinical infections, need for early screening of suspected cases in order to prevent chorioretinitis and ameliorate brain damage

Toxoplasma
Supperer, R.; and Leibetseder, J., 1977, Wien. Tierarztl. Monatsschr., v. 64 (6-7), 188-189
Sarcocystis, Toxoplasma, raw meat and slaughtering offals should be cooked to prevent transmission of parasites to dog and cat

Toxoplasma
  Suzuki, M.; Tsunematsu, Y.; and Waki, S., 1973, J. Protozool., v. 20 (4), 528
    Toxoplasma, mechanism of Sabin-Feldman dye test, importance of complement

Toxoplasma
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    Sarcocystis and related coccidian parasites, general review with discussion of biological aspects of life cycles and new proposal for classification

Toxoplasma
  Waldeland, H., 1977, Acta Vet. Scand., v. 18 (2), 227-236
    Toxoplasma, four breeding flocks of sheep, long-term (3-6 years) epidemiological studies, abnormal reproductive performance, increased prevalence with age, higher incidence in lowland pastures than in mountain pastures, lower incidence in summer than in winter: Norway

Toxoplasma
  Werner, H., 1968, Bol. Chileno Parasitol., v. 23 (3-4), 95-98
    Toxoplasma, strains of varying virulence, mouse mortality after oral vs. intraperitoneal infection, variation in dye-test titers in relation to cyclic development of parasite

Toxoplasma
  Werner, H., 1976, Ztschr. Parasitenk., v. 50 (2), 176-177
    Toxoplasma, mice, rabbits, congenital infection despite immunity after reinfection during pregnancy

Toxoplasma [sp.]
  Altan, Y.; Heydorn, A. O.; and Janitschke, K., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (22), 433-435
    Toxoplasma oocysts, horses (exper.), infectivity limited

Toxoplasma [sp.]
  Burtscher, H.; and Meingassner, J. G., 1976, Ztschr. Parasitenk., v. 50 (2), 220
    Chinchilla laniger

Toxoplasma sp.
  Guterbock, W. M.; and Levine, N. D., 1977, J. Am. Vet. Med. Ass., v. 170 (12), 1411-1413
    cats (feces): east central Illinois

Toxoplasma sp. Shortt, 1950
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    "nomen nudum"

Toxoplasma sp. auctores from various birds
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    mostly Atoxoplasma spp.

Toxoplasma sp. Plimmer, 1916
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    as syn. of Toxoplasma colubri Tibaldi, 1921

Toxoplasma spp.
  Owen, D.; and Chessum, B. S., 1977, Vet. Rec., v. 101 (20), 402-404
    Toxoplasma spp., sheep, goat, Sabin Feldman dye test, no seasonal fluctuations demonstrated in antibody titres

Toxoplasma alencari (da Costa & Pereira, 1971) comb. nov.
  Levine, N. D.; and Nye, R. R., 1976, J. Protozool., v. 23 (4), 488-490
    Syn.: Lankesterella alencari da Costa & Pereira, 1971

Toxoplasma alencari (da Costa & Pereira, 1971) Levine & Nye, 1976
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    diagnosis
    Syn.: Lankesterella alencari da Costa & Pereira, 1971

Toxoplasma avium Marullaz, 1913
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    as syn. of Atoxoplasma paddae (Aragao, 1911) Laird, 1959

Toxoplasma avium Marullaz, 1913 pro parte
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Isospora lacazei (Labbe, 1893)

Toxoplasma avium Marullaz, 1913 [pro parte]
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Isospora serini (Aragao, 1933) Box, 1975

Toxoplasma brumpti Coutelen, 1932
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    diagnosis

Toxoplasma butasturis de Mello, 1935
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    as syn. of Lankesterella butasturis (de Mello, 1935) comb. nov.

Toxoplasma canis Mello 1910
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma canis Mello, 1910
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma canis Mello, 1910
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    as syn. of Isospora gondii (Nicolle & Manceaux, 1908) comb. nov.

Toxoplasma caviae Carini and Migliano 1916
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma caviae Carini & Migliano, 1916
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma caviae Carini & Migliano, 1916
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora gondii (Nicolle & Manceaux, 1908) comb. nov.

Toxoplasma colubri Tibaldi, 1921
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
synonymy, diagnosis

Toxoplasma columbae Yakimoff and Kohl-Yakimoff 1912
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma columbae Yakimoff & Kohl-Yakimoff, 1912
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma crocidurae Galli-Valero 1933
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma crocidurae Galli-Valero, 1933
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma cuniculi (Splendore 1908)
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma cuniculi Splendore, 1908
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma cuniculi Splendore, 1910
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora gondii (Nicolle & Manceaux, 1908) comb. nov.

Toxoplasma francae (de Mello 1915) Wenyon 1926
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma francae (de Mello, 1915) Wenyon, 1926
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma fulicae de Mello 1935
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma fulicae de Mello, 1935
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma gallinarum Hepding 1939
Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma gallinarum Hepding, 1939
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma glareoli Erhardova, 1955
Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
as syn. of Frenkelia glareoli (Erhardova, 1955) Tadros, Bird & Ellis, 1972

Toxoplasma gondii, illus.
Aikawa, M.; et al., 1977, Am. J. Path. (420), v. 87 (2), 285-296
Toxoplasma gondii tachyzoites, ultrastructural study of penetration of host cell by parasite

Toxoplasma gondii
Akinshina, G. T.; and Zalkind, S. Ia., 1975, Biul. Eksper. Biol. i Med., v. 80 (10), 122-125
Toxoplasma gondii-infected pig embryonic kidney cells, host cell response to infection (morphological changes, absence of severe or advanced cytotoxic effect, preservation of high mitotic activity); persistence of infection

Toxoplasma gondii
Altichieri, M.; et al., 1975, Minerva Pediat., v. 27 (11), 656-657
Toxoplasma gondii, seroimmunologic survey of newborn infants and their mothers for evidence of infection

Toxoplasma gondii, illus.
Alvarez San Cristobal, A.; and Sanmarti Roset, M., 1976, Acta Cytol., v. 20 (4), 285-286
Toxoplasma gondii cysts in human vaginal and cervical smears, clinical case: Barcelona, Spain

Toxoplasma gondii
do Amaral, V.; Santos, S. M.; and Reboucas, M. M., 1975, Biologico, S. Paulo, v. 41 (4), 105-107
Toxoplasma gondii, prevalence of antibodies in pigs, hemagglutination test: States of Sao Paulo and Rio Grande do Sul, Brazil

Toxoplasma gondii
Anderson, S. E.; Bautista, S.; and Remington, J. S., 1976, J. Immunol., v. 117 (2), 381-387
intracellular Toxoplasma gondii infection in human monocyte-derived macrophages in vitro can be inhibited or killed by soluble lymphocyte products contained in supernatants prepared from either antigen or mitogen-stimulated lymphocytes (homologous antigen more active than heterologous antigen or mitogen)

Toxoplasma gondii
Anderson, S. E., jr.; Bautista, S. C.; and Remington, J. S., 1976, Clin. and Exper. Immunol., v. 26 (3), 375-380
specific antibody-dependent killing of Toxoplasma gondii by normal macrophages

Toxoplasma gondii
Aparicio Garrido, J.; et al., 1972, Med. Trop., Madrid, v. 48 (1-2), 11-23
sheep, cattle and pigs slaughtered at local abbatoir surveyed for serologic evidence of Toxoplasma gondii infection, possible correlation of human infections transmitted through consumption of meat animals and comparisons with similar studies done in other parts of world: Madrid

Toxoplasma gondii, illus.
Aparicio Garrido, J.; et al., 1972, Med. Trop., Madrid, v. 48 (1-2), 24-39
serological and fecal survey of cats for evidence of Toxoplasma gondii infections: Madrid

Toxoplasma gondii
Apt, W., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 65-68
Toxoplasma gondii, criteria for treating humans with pyrimethamine and sulfa drugs, pharmacologic action, treatment of toxic side effects with folinic acid, recommendation for prophylactic treatment of newborn infants exposed to infection

Toxoplasma gondii
Apt, W.; et al., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 37-41
Toxoplasma gondii, human, diagnosis, establishment of criteria for interpretation of toxoplasmin skin test and comparison with results of hemagglutination test

Toxoplasma gondii
Apt, W.; Niedmann, G.; and Atias, A., 1968, Bol. Chileno Parasitol., v. 23 (3-4), 102-107
known or suspected Toxoplasma gondii infections in pregnant women, treatment during pregnancy with pyrimethamine combined with sulfa drugs, favorable results in selected clinical trials

Toxoplasma gondii
Arambulo, P. V. III; Cabrera, B. D.; and Alge, M. H., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 9-11
Toxoplasma gondii, serologic survey of market-age pigs for evidence of toxoplasmosis using the indirect hemagglutination test: Philippine Islands

Toxoplasma gondii
Araujo, F. G.; et al., 1976, Infect. and Immun., v. 13 (5), 1528-1530
Toxoplasma gondii, 7 inbred and outbred strains of mice, susceptibility to infection with trophozoites at 2 different doses, striking differences in susceptibility and changes of susceptibility with dosage change, may be due to genetic factors

Toxoplasma gondii
Araujo, F. G.; and Nascimento, E., 1977, J. Parasitol., v. 63 (6), 1120-1121
parasitemia by Trypanosoma cruzi lower in mice chronically infected with Toxoplasma gondii than in controls, mortality rate similar

Toxoplasma gondii
Araujo, F. G.; and Remington, J. S., 1975, J. Immunol., v. 115 (2), 335-338
Toxoplasma gondii, newborn rabbits, passively administered IgG significantly suppresses IgM antibody response, results may explain absence of IgM antibody response in congenitally infected human infants by suppressive effect of maternally-transmitted IgG

Toxoplasma gondii
Arnaudov, D.; et al., 1976, Vet. Med., Praha, v. 49, v. 21 (6), 375-384
Toxoplasma gondii, sheep, immunoepizootiological study by hemagglutination, indirect fluorescence, and microprecipitation reaction in agar gel; higher incidence in aborting ewes and in sheep in montane regions: Bulgaria; Czechoslovakia

Toxoplasma gondii
Arribada, A.; et al., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 22-32
survey of patients with cardiomyopathies established Toxoplasma gondii as frequent etiologic agent with mixed infection of toxoplasmosis and Chagas disease discovered in one person

Toxoplasma gondii
Atias M., A., 1970, Bol. Chileno Parasitol., v. 25 (3-4), 123-126
Toxoplasma gondii, human, review of congenital and acquired forms, diagnosis and medical management

Toxoplasma gondii
Atias, A.; and Saavedra, R., 1966, Bol. Chileno Parasitol., v. 21 (2), 30-34
Toxoplasma gondii, seroimmunologic survey for presence of infection in women with history of spontaneous and repeated abortions, frequency and serologic activity higher in these women than in controls

Toxoplasma gondii, illus.
Attwood, H. D.; Woolley, P. A.; and Rickard, M. D., 1975, J. Wildlife Dis., v. 11 (4), 543-551
Toxoplasma gondii, prevalence in dasyurid marsupials obtained from the field or maintained or reared in the laboratory, puzzling absence of infection in Antechinus stuartii
Antechinomys spenceri
Antechinus apicalis
A. macdonnellensis
Dasycercus cristicauda
Dasyuroides byrnei
Sminthopsis crassicaudata
S. larapinta
S. leucopus
all from Australia

Toxoplasma gondii, illus
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Toxoplasma gondii
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persistence of viable Toxoplasma gondii oocysts in soil up to 18 months in Kansas and 1 year in Costa Rica, experimentally buried infected cat feces; Musca fly, Armadillium sp., earthworms acted as transport hosts with earthworm possibly also vector as food for birds; seasonal distribution, effects of weather on infectivity

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Mus musculus (exper.)

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Mus musculus
(brains of all): all from Iran

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Toxoplasma gondii, clinical and differential diagnostic review of infection in humans

T[oxoplasma] gondii
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T[oxoplasma] gondii, dog, encephalitis with generalized necrotizing myopathy of masticatory muscles

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Toxoplasma gondii, human acute and chronic infections, chromosome abnormalities in lymphocytes, possibly produced by parasitic infection

Toxoplasma gondii
Mishra, G. S.; and Gonzalez, J. P., 1975, Arch. Inst. Pasteur Tunis, v. 52 (1-2), 71-87
cats fed raw rat liver passed oocysts in feces 72 hours after feeding, none thereafter; cats fed cooked rat liver passed no oocysts
Rattus norvegicus (visceres): Tunis, Tunisia

Toxoplasma gondii (RH and Beverley strains)
Miyata, A., 1973, Nettai Igaku (Trop. Med.), v. 15 (4), 204-213
Entamoeba histolytica, Toxoplasma gondii, Plasmodium berghei, Trypanosoma gambiense, preservation by freezing, evaluation of precooling and cryo-protective solutions, length of survival; T. gondii (Beverley strain) unable to survive freezing

Toxoplasma gondii, illus.
Miyata, A., 1976, Nettai Igaku (Trop. Med.), v. 18 (3), 113-124
Toxoplasma gondii, RH and Beverley strains, attempts to change virulence using antitoxoplasma drugs and steroids, variations in mouse (exper.) survival time

Toxoplasma gondii
Montenegro, D.; Lira, V.; and Azoubel, E., 1972, Rev. Microbiol., Sao Paulo, v. 3 (4), 177-178
Toxoplasma gondii antibodies in sera of blood bank donor detected by indirect immunofluorescence: Recife, PE, Brazil

Toxoplasma gondii
Motomura, I., 1967, Nettai Igaku (Trop. Med.), v. 9 (4), 201-225
Toxoplasma gondii, mice (exper.), survival comparisons of Beverley and RH strains under various physico-chemical conditions (osmotic pressure, pH, temperature, moisture, irradiation)

Toxoplasma gondii
Motomura, I., 1967, Nettai Igaku (Trop. Med.), v. 9 (4), 226-243
Toxoplasma gondii, cysts and proliferative forms, in vitro survival in different concentrations of disinfectants or on incubation with digestive enzymes or bovine bile

Toxoplasma gondii
Motomura, I., 1967, Nettai Igaku (Trop. Med.), v. 9 (4), 244-255
Toxoplasma gondii, mice (exper.), infectivity of Beverley cysts and RH proliferative forms, oral and percutaneous inoculations

Toxoplasma gondii
Motomura, I.; and Jo, K., 1970, Nettai Igaku (Trop. Med.), v. 12 (2), 41-50
Toxoplasma gondii, cyst distribution and development within brain of infected mice, statistics of numbers of toxoplasmas within a single brain cyst

Toxoplasma gondii
Mueller, B. E. G.; Mehlhorn, H.; and Scholtyseck, E., 1973, J. Protozool., v. 20 (4), 505
ultrastructure and origin of cyst wall in Toxoplasmatea

Toxoplasma gondii
Mueller, F.; and Klein, P., 1975, Med. Microbiol. and Immunol., v. 162 (1), 55-61
possible immunologic differentiation between immunoglobulin M, immunofluorescent and complement fixing antibodies, role in serodiagnosis of human Toxoplasma gondii

Toxoplasma gondii
Munday, B. L., 1973, Med. J. Australia, v. 1 (24), 1183-1185
survey of human Toxoplasma gondii infection, probable significant factor in eye and lymphatic disease in Tasmania

Toxoplasma gondii
Munday, B. L., 1973, Med. J. Australia, v. 2 (11), 564
potential sources of Toxoplasma gondii infection for pregnant women and suggested control measures

Toxoplasma gondii
Munday, B. L.; et al., 1975, Research Vet. Sc., v. 18 (2), 218-219
Toxoplasma gondii, sera from aborted or newborn lambs with congenital infections fractionated on Sephadex G-200, sera and fractions tested for antibodies using indirect fluorescent antibody test, most of antibody found in IgG fraction

Toxoplasma gondii
Nakabayashi, T.; et al., 1967, Nettai Igaku (Trop. Med.), v. 9 (2), 111-125
Toxoplasma gondii, mice (exper.), survival of parasites at refrigerating and cold storage room temperatures of slaughterhouse

Toxoplasma gondii
Nakabayashi, T.; et al., 1969, Nettai Igaku (Trop. Med.), v. 11 (1), 16-26
detection in pigs suspected to be infected with Toxoplasma gondii using hemagglutination test with the mouse inoculation method or fluorescent antibody test

Toxoplasma gondii, illus.
Nakabayashi, T.; and Motomura, I., 1968, Nettai Igaku (Trop. Med.), v. 10 (2), 72-80
Toxoplasma gondii, separation of cysts from infected mouse brain using multi-layered centrifugation with gum arabic solution

Toxoplasma gondii
Nation, P. N.; and Allen, J. R., 1976, Canad. Vet. J., v. 17 (12), 308-310
survey for prevalence in cattle, sheep, cats, indirect hemagglutination test, antibody levels for sheep and cattle nil, 3.4% for cats: Saskatchewan

Toxoplasma gondii
Nejmi, S.; and Alami, S., 1973, Maroc Med. (572), v. 53, 561-568
Toxoplasma gondii, epidemiologic survey of Marocaine population using immunofluorescence: Maroc

Toxoplasma gondii
Nguyen, B. T.; and Stadtsbaeder, S., 1975, Path. Europ., v. 10 (4), 307-315
Toxoplasma gondii in culture, trimethoprim alone or with sulfamethoxazole as synergist (cotrimoxazole) resulted in inhibition of intracellular multiplication, prolonged therapy resulted in eradication of organisms from cell monolayers; similar trials with spiramycin ineffective

Toxoplasma gondii
Nguyen, B. T.; and Stadtsbaeder, S., 1976, Infect. and Immun., v. 13 (3), 884-889
spontaneous interaction in vitro between lymphocytes and syngeneic peritoneal macrophages of normal as well as Toxoplasma gondii-immune mice

Toxoplasma gondii
Nozais, J. P.; et al., 1975, Medecine Trop., v. 35 (5), 413-417
Toxoplasma gondii, epidemiologic survey of disease prevalence using indirect immunofluorescence test, infection at early age, probably through ingestion of contaminated soil as a result of poor hygiene habits: Ivory Coast

Toxoplasma gondii
Olisa, E. G.; et al., 1977, Exper. Parasitol., v. 41 (2), 307-313
Toxoplasma gondii RH strain, differences in survival time of four strains of mice each infected with various doses

Toxoplasma gondii
Omata, Y.; and Suzuki, N., 1975, Research Bull. Obihiro Univ., s. I, v. 9 (3), 473-482
Toxoplasma gondii-infected rats, subpopulations of T- and B-lymphocytes in spleen, thymus, and peripheral blood, compared with healthy rats

Toxoplasma gondii
Omata, Y.; and Suzuki, N., 1975, Research Bull. Obihiro Univ., s. I, v. 9 (2), 265-272 (35-42)
Toxoplasma gondii, humoral antibody responses, appearance of I_gM and I_gG antibodies in adult female rats and their infants; transmission of infection from chronically infected rats to their fetuses and infants, no congenital toxoplasmosis detected

Toxoplasma gondii
Omland, T.; Tønjum, A.; and Frentzel-Beyme, B. R., 1977, Tropenmed. u. Parasitol., v. 28 (3), 372-376
Toxoplasma gondii, prevalence survey for presence of antibodies to toxoplasmosis in native Liberians using the Sabin-Feldman dye test, differing habitats

Toxoplasma gondii
Ourth, D. D.; Lunde, M. N.; and Watson, R. R., 1976, Ztschr. Immunitaetsforsch., v. 151 (3), 254-262
Toxoplasma gondii, guinea pigs, demonstration of delayed hypersensitivity by macrophage migration inhibition, skin-testing, and lymphocyte transformation

Toxoplasma gondii
Overdulve, J. P., 1976, Trop. and Geogr. Med., v. 28 (4), 363 [Abstract]
Toxoplasma gondii, light and electron microscopy of development in small intestine of cat

Toxoplasma gondii
Ozcan, K., 1975, Mikrobiyol. Bul., v. 9 (4), 281-290
Toxoplasma gondii, indirect fluorescent antibody technique and Sabin-Feldman dye test compared for diagnosis in patients with eye diseases: Turkey

Toxoplasma gondii
Palick, P.; Micek, E.; and Kerka, J., 1975, Veterinarstvi, v. 25 (10), 457-458
Toxoplasma gondii, human incidence determined by complement fixation reaction and microprecipitation in agar gel; incidence in domestic animals determined by above methods plus isolation in mice: Czechoslovakia

Toxoplasma gondii
Partono, F.; and Cross, J. H., 1975, Southeast Asian J. Trop. Med. and Pub. Health, v. 6 (4), 472-476
Toxoplasma gondii, comparative survey of Indonesian and Chinese medical students for prevalence of antibodies to toxoplasmosis: Jakarta, Indonesia

Toxoplasma gondii
Partono, F.; and Cross, J. H., 1976, Trop. and Geogr. Med., v. 28 (1), 63-64
Toxoplasma gondii antibody titers in mother and 18-month old child with blindness and microphthalmia, probable prenatal infection: Indonesia

Toxoplasma gondii
Pasmanik, S.; and Atias, A., 1966, Bol. Chileno Parasitol., v. 21 (1), 7-14
human ocular Toxoplasma gondii, pathology, diagnosis (presence of rosetta-like macular chorioretinitis or by seroimmunologic testing)

Toxoplasma gondi
Pav, J.; and Zajicek, D., 1974, Veterinarstvi, v. 24 (11), 517-520
Tetrao urogallus: CSSR

Toxoplasma gondii, illus.
Pelster, B., 1975, Ztschr. Parasitenk., v. 48 (2), 95-110
Toxoplasma gondii, immunized mice, superinfection not causing death, cellular immune reactions, mouse exudate containing high percentage of lymphocytes; peritoneal exudate cells incubated in vitro with trophozoites cause their death by lysis

Toxoplasma gondii
Pelster, B., 1976, Ztschr. Parasitenk., v. 50 (2), 175-176
Toxoplasma gondii, infected mice, effect of antithymus serum

Toxoplasma gondii, illus.
Pelster, B.; Piekarski, G.; and Suzuki, N., 1976, Ztschr. Parasitenk., v. 49 (2), 113-125
Toxoplasma gondii, mice, virulent and avirulent strains, histopathological changes in thymus

Toxoplasma gondii, illus.
Pestre-Alexandre, M.; and Nicolas, J. A., 1972, Medecine et Malad. Infect., v. 1 (12), special no., 557-564
Toxoplasma gondii, morphology, life cycle, epidemiology, domestic cat as intermediate host in transmission to man

Toxoplasma gondii
Pettersen, E. K., 1977, Acta Path. et Microbiol. Scand., v. 85B (1), 95-102
Toxoplasma gondii, avirulent strain made virulent by mouse passage and then attenuated by storage, comparison of cyst abilities in mice and rabbits, rabbits infected with attenuated parasites survived challenge with virulent parasites

Toxoplasma gondii
Pfefferkorn, E. R.; and Pfefferkorn, L. C., 1976, Exper. Parasitol., v. 39 (3), 365-376
Toxoplasma gondii, chemical mutagenesis, selection of temperature-sensitive mutants, less virulent for mice than wild type RH strain

Toxoplasma gondii, illus.
Pfefferkorn, E. R.; and Pfefferkorn, L. C., 1976, J. Parasitol., v. 62 (6), 993-999
Toxoplasma gondii, adenine arabinoside inhibits DNA synthesis, more inhibitory to parasite than to cultured human fibroblast host cells, selection of single-step resistant mutant; cytosine arabinoside notably more inhibitory to cultured human cells than to T. gondii

Toxoplasma gondii, illus.
Pfefferkorn, E. R.; and Pfefferkorn, L. C., 1977, Exper. Parasitol., v. 41 (1), 95-104
Toxoplasma gondii, growth in Lesch-Nyhan cells which are incapable of incorporating hypoxanthine or guanine as a technique for specific labeling of nucleic acids of intracellular parasites vs. their host cells, autoradiography

Toxoplasma gondii, illus.
Pfefferkorn, E. R.; and Pfefferkorn, L. C., 1977, Exper. Parasitol., v. 42 (1), 44-55
Toxoplasma gondii, inhibition of growth and RNA and DNA synthesis by 5-fluorodeoxyuridine (FUDR), isolation and partial characterization of FUDR-resistant mutant also resistant to fluorouracil and fluorouridine, examination of possible mechanisms of resistance yielded new insights into pyrimidine salvage pathways of the parasite

Toxoplasma gondii, illus.
Pfefferkorn, E. R.; and Pfefferkorn, L. C., 1977, J. Protozool., v. 24 (3), 449-453
Toxoplasma gondii, specific labelling of intracellular parasites with radioactive uracil, not significantly incorporated into nucleic acids of human fibroblast host cells

Toxoplasma gondii
Pfefferkorn, E. R.; Pfefferkorn, L. C.; and Colby, E. D., 1977, J. Parasitol., v. 63 (1), 158-159
Toxoplasma gondii, development of gametes and oocysts in cats fed cysts derived from cloned trophozoites, results suggest that genome of each T. gondii contains genetic potential for production of both macro- and micro-gametes

Toxoplasma gondii
Plank, G.; et al., 1974, Orvosi Hetilap, v. 115 (51), 3055-3056
prenatal Toxoplasma gondii infection in human resulting in icterus similar to Rh incompatibility and death, case report: Hungary

Toxoplasma gondii, illus.
Plant, J. W.; Richardson, N.; and Moyle, G. G., 1974, Austral. Vet. J., v. 50 (1), 19-21
Toxoplasma gondii, sheep, contamination of grain by cat feces containing oocysts, examination of cats inhabiting feed shed revealed oocysts

Toxoplasma gondii, illus.
Poitrat, C., 1973, Medecine Infant., v. 80 (3), 345-365
Toxoplasma gondii, general review of life cycle, morphology, diagnosis, epidemiology and clinical management

Toxoplasma gondii
Poletti, T.; et al., 1970, Minerva Med., v. 61 (48), 2696-2701
acquired lymphatic Toxoplasma gondii in young adults, frequent cause of diagnostic difficulties, case reports: Italy

Toxoplasma gondii, illus.
Premuzic-Lampic, M.; and Paskvalin, L. J., 1975, Medicina, Rijeka, v. 12 (1), 24-28
review of 3 cases of human acquired lymphatic toxoplasmosis, clinical aspects, hematologic diagnosis: Yugoslavia

Toxoplasma gondii
Raisanen, S., 1976, Norwegian J. Zool., v. 24 (4), 458 [Abstract]
Toxoplasma gondii trophozoites, survival in vitro, best preservation media are mouse ascitic fluid and rabbit serum

Toxoplasma gondii
Raisanen, S.; and Koivukangas, J., 1976, Norwegian J. Zool., v. 24 (4), 458-459 [Abstract]
Toxoplasma gondii, infectivity of trophozoites in body secretion examined by penetration through intact mucous membranes of mice, greater contamination hazard than previously known, easiest route of infection is intranasal

Toxoplasma gondii
Raisanen, S.; and Saari, M., 1976, Med. Biol., v. 54 (2), 152-155
effect of preservation time and changes in osmotic pressure on infectivity and survival of Toxoplasma gondii trophozoites

Toxoplasma gondii
Raizman, R. E.; and Neva, F. A., 1975, J. Infect. Dis., v. 132 (1), 44-48
Toxoplasma gondii, mice and rabbits (exper.), detection and characterization of circulating antigen

Toxoplasma gondii, illus.
Rao, N. A.; and Font, R. L., 1977, Arch. Ophth., Chicago, v. 95 (2), 273-277
Toxoplasma gondii causing retinochoroiditis, formalin-fixed human eye tissue, electron microscopic study of cysts, immunofluorescent-antibody technique in diagnosis when routine histologic preparation fails to reveal parasites

Toxoplasma gondii, illus.
Rasin, K., 1973, Vet. Med., Praha, v. 46, v. 18 (10), 593-618
Toxoplasma gondii (14 strains), Isospora felis, cats, natural and exper. infections, chronology of oocyst elimination; mixed infections; no cross immunity

Toxoplasma gondii
Reikvam, A., 1976, Acta Path. and Microbiol. Scand., v. 84C (2), 124-130
Toxoplasma gondii, mice, mechanism(s) of initiation and regulation of macrophage proliferation and activation, relationship to lymphocyte stimulation

Toxoplasma gondii
Reikvam, A.; and Lorentzen-Styr, A. M., 1976, Nature, London (5560), v. 261, 508-509
Toxoplasma gondii, mice infected with Beverley strain and reinfected with RH strain, 14-18 months after challenge Toxoplasma with full virulence of RH strain could be isolated from some mice, in brains of others only immunizing (Beverley) strain could be found, implications for human infections

Toxoplasma gondii
Remington, J. S.; Krahenbuhl, J. L.; and Mendenhall, J. W., 1972, Infect. and Immun., v. 6 (5), 829-834
activated macrophages from mice which were chronically infected with Toxoplasma gondii or Besnoitia jellisoni or which had received Freund complete adjuvant had enhanced capacity to kill intracellular Toxoplasma

Toxoplasma gondii
Remington, J. S.; Miller, M. J.; and Brownlee, I., 1968, Pediatrics, Am. Acad. Pediat., v. 41 (6), 1082-1091
Toxoplasma gondii, diagnosis of congenital infection in newborn infants using the fluorescent antibody test to detect IgM antibodies (in absence of placental leak) in newborn serum

Toxoplasma gondii
van Renterghem, L.; and van Nimmen, L., 1976, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 235 (4), 559-565
Toxoplasma gondii, indirect immunofluorescence test, polar fluorescence, frequency higher in adults than in children, nonspecificity, polar factor shown to be nonspecific IgM

Toxoplasma gondii
Ricciardi, I. D.; Sandoval, E. F. D.; and Mayrink, W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 516-517
Toxoplasma gondii, human prevalence survey, distribution by age: Brazil

Toxoplasma gondii
Riemann, H. P.; et al., 1974, J. Wildlife Dis., v. 10 (4), 471-477
Toxoplasma gondii, pallas cats, indirect hemagglutination test, pathological findings in kittens
Felis manul: imported to California zoo

Toxoplasma gondii
Riemann, H. P.; et al., 1975, J. Wildlife Dis., v. 11 (2), 272-276
Toxoplasma gondii, prevalence of antibodies among carnivores (Lynx rufus, Procyon lotor, Taxidea taxus, Canis latrans), cysts found in brain tissues of one Taxidea taxus: northern California (Napa County; Humboldt County)

Toxoplasma gondii
Riemann, H. P.; et al., 1975, J. Wildlife Dis., v. 11 (4), 529-533
Toxoplasma gondii, survey of antibodies in free-living mammals provides evidence that it is being maintained in wildlife: Kenya; Tanzania; Uganda; Zambia

Toxoplasma gondii
Riemann, H. P.; et al., 1977, J. Am. Vet. Med. Ass., v. 171 (12), 1260-1264
Toxoplasma gondii, prevalence of Toxoplasma antibodies in sheep, higher incidence in breeding ewes than in lambs sent to market, management and environmental factors: California; Nevada; Idaho; Oregon

Toxoplasma gondii
Rifaat, M. A.; et al., 1976, J. Trop. Med. and Hyg., v. 79 (3), 67-70
seroepidemiolgoic survey of stray cats for Toxoplasma gondii antibodies and role in epidemiology of human infection, no cross-immunity with Isospora spp. in relationship study, high percentage of cats surveyed shedding oocysts in feces: Cairo, Egypt

Toxoplasma gondii
Roberts, C. O.; Chaparas, S. D.; and McLaughlin, D., 1976, J. Protozool., v. 23 (2), 34A [Abstract]
Toxoplasma gondii, mammalian and avian cell cultures tested to determine ability to support growth and plaque induction, inhibition of plaque formation by lincocin, sulfisoxazole, cleocin, and aureomycin and by anti-toxoplasma antibody

Toxoplasma gondii, illus.
Roberts, C. O.; Chaparas, S. D.; and McLaughlin, D., 1976, Tr. Am. Micr. Soc., v. 95 (3), 470-482
Toxoplasma gondii, plaque assay technique used to (1) select best tissue culture system for studying progress of infection, (2) assess effect of drugs and antibiotics, (3) observe effect of immune sera on tissue culture infection, and (4) investigate cellular hypersensitivity by in vivo and in vitro systems

Toxoplasma gondii
Robinson, R. A., 1973, N. Zealand Med. J. (489), v. 77, 97-101
Toxoplasma gondii, potential role of cats in epidemiology of human infections

Toxoplasma gondii
Rommel, M.; and v. Seyerl, F., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (20), 398-399
Hammondia hammondi, isolation from cat feces, exper. mouse infection, epidemiological significance for distinguishing from Toxoplasma gondii: Germany

Toxoplasma gondii
Rose, N. R.; Milisauskas, V.; and Zeff, G., 1975, Immunol. Commun., v. 4 (1), 1-16
antigenic and enzymatic changes in infected (including by Toxoplasma gondii) and transformed human diploid cells

Toxoplasma gondii
Rossi, G. A.; Brancaccio, M.; and Gatti, A., 1974, Atti Soc. Ital. Sc. Vet., v. 28, 840-845
Toxoplasma gondii, isolation from dogs with severe pneumoencephalitis, case histories, severity possibly related to distemper vaccination or winter climate

Toxoplasma gondii
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fatal infection in premature infant, Toxoplasma gondii isolated from brain tissue at autopsy, case report, serological evidence of toxoplasmosis in both parents: Australia

Toxoplasma gondii
Ruitenberg, E. J.; and van Knapen, F., 1977, J. Infect. Dis., v. 136, suppl., S267-S273
application of the enzyme-linked immunosorbent assay to the detection of human and animal helmintic and protozoal infections, advantages of assay for seroepidemiology, discussion of performance of assay in tubes and microplates

Toxoplasma gondii
Ruskin, J.; and Remington, J. S., 1971, J. Reticuloendothel. Soc., v. 9 (5), 465-479
mice immunized with Toxoplasma vaccine and adjuvant are protected against challenge with Listeria

Toxoplasma gondii
Ryning, F. W.; and Remington, J. S., 1977, Infect. and Immun., v. 18 (3), 746-753
Toxoplasma gondii, role for activated macrophage as effector in resistance of lung to infection

Toxoplasma gondii, illus.
Ryu, E.; et al., 1975, Taiwan J. Vet. Med. and Animal Husb. (26), 7-15
Toxoplasma gondii, swine, survey, diagnosis by intradermal skin test, hemagglutination test, dye test, isolation: Taiwan

Toxoplasma gondii
Saari, K. M.; and Raisanen, S. A., 1977, Lancet, London (8047), v. 2, 1077 [Letter]
Toxoplasma gondii, studies show that trophozoites are capable of transmitting infections in acute stages of toxoplasmosis

Toxoplasma gondii
Saathoff, M.; and Dogba, C., 1974, Tropenmed. u. Parasitol., v. 25 (4), 401-404
Toxoplasma gondii, prevalence of infection in juveniles and adults determined by Sabin Feldman dye test: south Togo

Toxoplasma gondii
Sakurai, Y.; et al., 1976, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 29 (6), 316-319
Toxoplasma gondii, survey on hemagglutination antibody in cats, breed or sex not significant in rate of positive reaction; high positive rate in cats from vicinity of piggery or slaughterhouse: Tohoku district

Toxoplasma gondii
Salazar, E.; et al., 1972, Arch. Inst. Cardiol. Mexico, v. 42 (6), 840-849
human Toxoplasma gondii resulting in clinical and electrocardiographic evidence of cardiomyopathy, case reports

Toxoplasma gondii
Samir Yassin, M. A.; et al., 1975, Riv. Parassitol., Roma, v. 36 (2-3), 81-88
observations on parasitemia in experimental animals
starling (blood, brain, spleen)
pigs (blood)
cats (blood, brain)
polecat
(all exper.)

Toxoplasma gondii
Schaal, E.; and Kleikamp, I., 1976, Berl. u. Munchen. Tierarztl. Wchnschr., v. 89 (17), 341-344
Toxoplasma gondii, incidence in pigs surveyed by Sabin Feldman test: Westphalia

Toxoplasma gondii
Schenk, M. A. M.; et al., 1976, Arq. Escola Vet. Univ. Fed. Minas Gerais, v. 28 (1), 33-35
frequency of antibodies in armadillos (Dasypus novemcinctus): Minas Gerais, Brazil

Toxoplasma gondii
Schenk, M. A. M.; Lima, J. D.; and Viana, F. C., 1976, Arq. Escola Vet. Minas Gerais, v. 28 (3), 261-266
Toxoplasma gondii, frequency in swine, diagnosis, methylene blue dye test (Sabin-Feldman) compared with indirect fluorescent antibody test: Belo Horizonte, Minas Gerais

Toxoplasma gondii
Schenone, H.; et al., 1974, Bol. Chileno Parasitol., v. 29 (3-4), 109-111
Toxoplasma gondii, preliminary studies suggest that congenital toxoplasmosis not frequent cause of mental deficiency: Chile

Toxoplasma gondii
Schierz, G.; and von Busch, K., 1976, Munchen. Med. Wchnschr., v. 118 (26), 839-842
differentiation between human clinical toxoplasmosis and Toxoplasma gondii infection, comparison of Sabin-Feldman dye test, indirect immunofluorescence to detect IgM and IgG antibodies, and complement fixation test

Toxoplasma gondii, illus.
Scholtyseck, E.; Mehlhorn, H.; and Mueller, B.E.G., 1974, J. Protozool., v. 21 (2), 284-294
Sarcocystis tenella, Besnoitia jellisoni, Frenkelia sp., Toxoplasma gondii, nature of cyst, fine structure of cyst wall

Toxoplasma gondii
Seah, S. K. K., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 76-77 [Letter]
Toxoplasma gondii, pretreatment and treatment with vitamin C offers some protective effect in experimental infection in mice

Toxoplasma gondii
Seguela, J. P.; et al., 1975, Medecine et Malad. Infect., v. 5 (11), 546-548
epidemiologic survey of Toxoplasma gondii among native populations in French Guiana

Toxoplasma gondii, illus.
Senaud, J., 1972, Bull. Inst. Pasteur, Paris, v. 70 (1), 3-27
extensive review of life cycle, morphology and development of Toxoplasma gondii

Toxoplasma gondii
Senet, J. M.; and Robert, R., 1974, Medecine et Malad. Infect., v. 4 (1), 21-22
Toxoplasma gondii, use of direct and indirect hemagglutination and immunofluorescence in seroimmunologic diagnosis, comparison with Sabin-Feldman dye test

Toxoplasma gondii
Sengbusch, H. G.; and Sengbusch, L. A., 1976, Am. J. Epidemiol., v. 103 (6), 595-597
comparative survey for prevalence of Toxoplasma gondii antibodies in veterinary personnel and selected population with no exposure to cats; hygiene, type of association, duration of exposure and infectivity of cats as considerations for disease transmission

Toxoplasma gondii, illus.
Sethi, K. K.; et al., 1973, Nature, London, N. Biol. (129), v. 243, 255-256
Toxoplasma gondii, multiplication in enucleated L cells proves that nucleo-cytoplasmic integrity is not required for intracytoplasmic multiplication of this parasite

Toxoplasma gondii
Sethi, K. K.; et al., 1975, J. Immunol., v. 115 (4), 1151-1158
Toxoplasma gondii, immunity induced in vitro in non-immune mouse macrophages with specifically immune lymphocytes

Toxoplasma gondii, illus.
Sethi, K. K.; et al., 1977, J. Parasitol., v. 63 (6), 1076-1080
Toxoplasma gondii, lectin-binding sites demonstrated on cell wall of brain cysts but not on surface membrane of trophozoites

Toxoplasma gondii
Shafer, N., 1975, N. York State J. Med., v. 75 (7), 1049-1061
Toxoplasma gondii in humans, extensive clinical review of current medical progress

Toxoplasma gondii
Sharma, H. K.; and Vashishta, M. S., 1976, Livestock Advis., v. 1 (8), 25-26
zoonosis, brief review

Toxoplasma gondii, illus.
Sheffield, H. G.; and Melton, M. L., 1975, J. Parasitol., v. 61 (4), 704-712
Toxoplasma gondii, fine structure and multiplication in cell cultures, effects of pyrimethamine and sulfadiazine singly and in combination

Toxoplasma gondii
Sheffield, H. G.; and Melton, M. L., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 379-383
Toxoplasma gondii, intestinal development in cats, effect of combined pyrimethamine and sulfadiazine treatment, prolonged prepatent period, shortened patent period, inhibition of oocyst excretion, infection induced immunity to oocyst-producing reinfection in both control and treated cats

Toxoplasma gondii
Sheffield, H. G.; Melton, M. L.; and Neva, F. A., 1976, Proc. Helminth. Soc. Washington, v. 43 (2), 217-225
Hammondia hammondi, artificially excysted sporozoites, development in cell culture, ultrastructure, endodyogeny only type division observed, characteristics to differentiate from Toxoplasma gondii

Toxoplasma gondii
Shin, Y. H.; Glass, L.; and Evans, H. E., 1976, Pediat. Ann., v. 5 (2), 106-113
Toxoplasma gondii, congenital toxoplasmosis as possible cause of birth defects and teratogenic changes, clinical review

Toxoplasma gondii
Shirahata, T.; and Shimizu, K., 1975, Research Bull. Obihiro Univ., s. I, v. 9 (2), 257-264 (27-34)
Toxoplasma gondii, mice (exper.), humoral antibody responses, indirect fluorescent antibody test, complement fixation test, Sabin-Feldman dye test; effect of serum antibody on viability in vitro

Toxoplasma gondii
Shirahata, T.; Shimizu, K.; and Suzuki, N., 1975, Japan. J. Vet. Sc., Tokyo, v. 37 (3), 235-243
Toxoplasma gondii, in vitro, lymphocyte-mediated immunity, results suggest resistance to and microbicidal activity upon Toxoplasma parasites may be conferred to macrophages by culture supernatant from immune lymphocytes reacting with Toxoplasma antigen

Toxoplasma gondii
Sibalic, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 148-152
humans with high Sabin-Feldman dye test titers thought to have toxoplasmosis found apparently also infected with Sarcocystis spp., specimens of their blood were injected into mice and Citellus citellus and Sarcocystis organisms recovered as cystozoites in peritoneal exudate or cysts in musculature; further verification positive using Sarcocystis fusiformis antigen for indirect immunofluorescence on human blood

Toxoplasma gondii
Sibalic, D., 1975, Acta Parasitol. Iugoslavica, v. 6 (1), 7-11
human, isolation in patients confirms parasitemia in initial as well as chronic phase besides persistence of specific toxoplasma antibodies

Toxoplasma gondii
Sibalic, D.; et al., 1973, Acta Parasitol. Iugoslavica, v. 4 (2), 107-117
Toxoplasma gondii in humans with acquired infection, clinical signs in relation to titre in modified Sabin-Feldman test, isolation of parasites

Toxoplasma gondii
Sibalic, D.; and Radovic, M., 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 21-27
Toxoplasma gondii, antibody titers among workers in a meat packing plant in relation to their contact with animals, higher titers among those in contact with animals, much higher incidence of new infections among those in contact with animals: Yugoslavia

Toxoplasma gondii
Sibalic, D.; and Radovic, M.; 1970, Acta Parasitol. Iugoslavica, v. 1 (1-2), 29-32
Toxoplasma gondii, patients serologically and clinically suspect for infection, inoculation of material from lymph node puncture and of saliva produced infection in mice

Toxoplasma gondii
Sibalic, D.; Radovic, M.; and Martinovic, M., 1972, Acta Parasitol. Iugoslavica, v. 3 (2), 89-96
Toxoplasma gondii, incidence of infection in various occupational groups: Yugoslavia

Toxoplasma gondii
Sibalic, D.; Radovic, M.; and Martinovic, M., 1975, Acta Parasitol. Iugoslavica, v. 6 (1), 13-18
Toxoplasma gondii, human, survival in blood (stored at +4°C), viability tested by inoculation into white mice

Toxoplasma gondii Nicolle et Manceaux 1909
Sima, O.; and Rasin, K., 1973, Vet. Med., Praha, v. 46, v. 18 (10), 633-640
Toxoplasma gondii, incidence in domestic slaughter rabbits measured by complement fixation test; higher incidence and higher titres in small-scale than in large-scale breeding farms: southern Moravia

Toxoplasma gondii
Singh, M.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 88 [Demonstration]
Cavia cobaya (brain): Singapore

Toxoplasma gondii, illus.
Singh, M.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 292-296
isolation from a naturally infected population of guinea pigs (Cavia cobaya) (brain) reared for experimental purposes: Singapore

Toxoplasma gondii
Somoza, M. J.; Monteverde, D. A.; and Morganfeld, M. C., 1972, Prensa Med. Argent., v. 59 (2), 65-70
Toxoplasma gondii, human acquired toxoplasmic lymphadinitis with neurologic symptoms, review of clinical findings in 47 patients: Argentina

Toxoplasma gondii
Stadtsbaeder, S.; and Nguyen, B. T., 1977, Ann. Immunol., v. 128C (1-2), 149-150
Toxoplasma gondii, susceptibility of nude mice to infection was same as controls, vaccination of nude mice conferred incomplete immunity

Toxoplasma gondii
Southern, P. M., jr., 1972, Obst. and Gynec., v. 39 (1), 45-47
Toxoplasma gondii, no evidence to support etiologic relationship between human toxoplasmosis and fetal wastage

Toxoplasma gondii
Stadtsbaeder, S.; Nguyen, B. T.; and Calvin-Preval, M. C., 1975, Ann. Immunol., v. 126C (4), 461-474
Toxoplasma gondii, mice, immunization with living parasites concomitant to cotrimoxazol treatment, phagocytosis/penetration and intracellular multiplication of Toxoplasma in normal or immune macrophages in the absence or presence of specific antibodies

Toxoplasma gondii
Stagno, S.; and Saavedra, P., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 122-125
combined antigen of free trophozoites of Toxoplasma gondii and cultured forms of Trypanosoma cruzi used to simultaneously diagnose both diseases by the indirect immunofluorescence test

Toxoplasma gondii
Stagno, S.; Saavedra, P.; and Thiermann, E., 1970, Bol. Chileno Parasitol., v. 25 (3-4), 102-105
Toxoplasma gondii, human, diagnosis, indirect immunofluorescence found comparable to Sabin-Feldman dye test

Toxoplasma gondii
Stagno, S.; and Thiermann, E., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 9-15
Toxoplasma gondii, use of indirect immunofluorescence test in place of Sabin-Feldman dye test in diagnosis

Toxoplasma gondii
Stagno, S.; and Thiermann, E., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 16-21
Toxoplasma gondii, prevalence survey of children 6 years old and under for presence of antibodies, importance of consideration of acquired toxoplasmosis in differential diagnosis of childhood diseases: Santiago, Chile

Toxoplasma gondii, illus.
Stahl, W.; et al., 1976, Exper. Parasitol., v. 39 (1), 135-142
Toxoplasma gondii, disease patterns in methotrexate-treated mice, examination of several time- and dose-dependent drug regimes, immunosuppressive effect when administered early in infection, potential use as experimental model for clinical toxoplasmosis

Toxoplasma gondii
Starzyk, J.; et al., 1977, Arzneimittel-Forsch., v. 27 (6), 1198-1199
Trichomonas vaginalis, Toxoplasma gondii, in vitro, lethal effect of ethanol extract of propolis

Toxoplasma gondii
Stempien, R.; Fabianowski, J.; and Wardecka, J., 1976, Polski Tygod. Lekar., v. 31 (43), 1847-1849
Toxoplasma gondii, humans, diagnosis and clinical management of lymph node infections

Toxoplasma gondii, illus.
Steriu, D.; and Dunareanu, G., 1976, Arch. Roumaines Path. Exper. et Microbiol., v. 35 (3), 213-222
Toxoplasma gondii endozoids, fine structure, mechanism of endo- and exocytosis, fragmentation of rhoptries resulting in formation of round bodies which give rise to new rhoptries in course of formation of daughter cells

Toxoplasma gondii, illus.
Stoll, L.; and Kraft, B., 1976, Deutsche Tierarztl. Wchnschr., v. 83 (4), 137-140
Toxoplasma gondii, swine, diagnosis, indirect fluorescent antibody test for lymph nodes (details of technique described) more sensitive than examination of blood samples by Sabin-Feldman test

Toxoplasma gondii
Stolte, F.; Stolte, J. B.; and The Lan Hing, 1972, Nederl. Tijdschr. Geneesk., v. 116 (34), 1511-1517
Toxoplasma gondii in humans, case reports of subacute lymphatic infections: Netherlands

Toxoplasma gondii, illus.
Stray-Pedersen, B.; and Lorentzen-Styr, A.M., 1977, Am. J. Obst. and Gynec., v. 128 (7), 716-721
Toxoplasma gondii in pregnant women, possible role of uterine infections in sporadic and habitual abortions, comparative study of controls and women with suspected infections using seroimmunologic methods and endometrial biopsies: Oslo, Norway

Toxoplasma gondii
Strickland, G. T.; and Sayles, P. C., 1977, Infect. and Immun., v. 15 (1), 184-190
Toxoplasma gondii, mice,depressed antibody responses to thymus-dependent antigen (sheep erythrocytes), results show that immunodepression associated with Toxoplasma infection is complicated and they provide no definitive explanation for the mechanism

Toxoplasma gondii, illus.
Sulzer, A. J.; et al., 1974, J. Protozool., v. 21 (5), 710-714
Toxoplasma gondii, comparative study of morphology by freeze-etch replication vs. thin sectioning technique

Toxoplasma gondii
Suzuki, M.; and Tsunematsu, Y., 1974, Ann. Trop. Med. and Parasitol., v. 68 (1), 33-39
Toxoplasma gondi high, low, and moderate virulence strains, comparison of susceptibility of Meriones unguiculatus vs. mice, illness and death rates, proliferation of parasites in various organs as revealed by fluorescent antibody technique, development of dye test antibodies, M. unguiculatus as good model host

Toxoplasma gondii
Suzuki, N.; et al., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 35-40
Toxoplasma gondii, mice and rats (exper.), blood changes; rats (exper.), antibody titers by dye test and indirect immunofluorescence, erythrocytes showed positive Coombs' reaction suggesting presence of auto-immune acquired hemolytic process

Toxoplasma gondii
Szabo, T., 1974, Mount Sinai J. Med., v. 41 (6), 765-773
human Toxoplasma gondii, current status in the United States, clinical review (pathology, transmission, epidemiology, diagnosis, prophylaxis, temporal variations)

Toxoplasma gondii
Szeri, I.; and Csoka, R., 1976, Acta Microbiol. Acad. Scient. Hungar., v. 23 (3), 235-237
Toxoplasma gondii, acute infections in mice (exper.), spleen, thymus and lymphocyte changes

Toxoplasma gondii Nicolle & Manceaux, 1909
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
as syn. of Isospora gondii (Nicolle & Manceaux, 1908) comb. nov.

Toxoplasma gondii
Tan, D. S. K.; and Zaman, V., 1973, Med. J. Malaysia, v. 27 (3), 188-191
statistical survey for prevalence of Toxoplasma gondii antibodies in inhabitants of West Malaysia

Toxoplasma gondii, illus.
Terragna, A., 1975, Paediatrician, v. 4 (2-3), 138-154
Toxoplasma gondii in children, present perspectives, extensive review (biology, epidemiology, life cycle, clinical aspects, diagnosis, fine structure, therapy)

Toxoplasma gondii
Terragna, A.; et al., 1970, Minerva Med., v. 61 (23), 1129-1133
Toxoplasma gondii in mice (exper.), laboratory trials using sulfamethoxypyrazine

Toxoplasma gondii, illus.
Terragna, A.; et al., 1975, Pathologica (969-970), v. 67, 329-335
Toxoplasma gondii, diagnosis of human lymphatic toxoplasmosis by ultrastructural demonstration of Toxoplasma in the lymph node

Toxoplasma gondii, illus.
Theologides, A.; and Lee, J. C., 1970, Minnesota Med., v. 53 (6), 615-619
concomitant systemic Toxoplasma gondii, Pneumocystis carinii pneumonitis and cytomegalic inclusion disease in woman undergoing steroid therapy for chronic leukemia, case report: Minnesota

Toxoplasma gondii, illus.
Theologides, A.; Pflueger, O. H., jr.; and Kennedy, B. J., 1969, Minnesota Med., v. 52 (5), 737-741
Pneumocystis carinii pneumonitis and Toxoplasma gondii of brain and myocardium in man being treated with steroids for chronic leukemia, case report: Minnesota

Toxoplasma gondii
Thiermann, E.; Apt, W.; and Niedmann, G., 1966, Bol. Chileno Parasitol., v. 21 (3), 82-88
Toxoplasma gondii in humans, review and assessment of current diagnostic methods, seroimmunologic and direct microscopic

Toxoplasma gondii
Thomaidis, T.; et al., 1977, Arch. Dis. Childhood, v. 52 (5), 403-407
Toxoplasma gondii, survey of relationship between toxoplasmosis and chronic lymphadenopathy in children, need for inclusion in differential diagnosis of lymphoglandular enlargement: Greece

Toxoplasma gondii
Timofeev, B. A.; and Akulov, A. V., 1975, Sel'skokhoz. Biol., v. 10 (2), 257-263
Toxoplasma gondii, life cycle, oocysts isolated from cats and alimentarily introduced into calves, sheep, and rabbits caused mild to severe infections, recommended removal from animal houses of toxoplasma positive cats

Toxoplasma gondii
Timoney, J. F., 1976, Irish Vet. J., v. 30 (3), 41-42
Toxoplasma gondii, healthy dogs in animal shelter, incidence as shown by positive indirect hemagglutination test: Dublin

Toxoplasma gondii
Tizard, I. R.; Billett, J. B.; and Ramsden, R. O., 1976, J. Wildlife Dis., v. 12 (3), 322-325
Toxoplasma gondii, antibody prevalence in wild mammals, apparent correlation with host consumption of rodents, possible importance of carnivorism as a route of transmission vs. oocyst-derived infection: Ontario, Canada

Toxoplasma gondii
Tizard, I. R.; Carrington, M.; and Lai, C. H., 1977, Canad. Vet. J., v. 18 (10), 274-277
Toxoplasma gondii, goats, prevalence of anti-Toxoplasma antibodies, Sabin-Feldman dye test, no significant clinical problem in goats surveyed, epidemiological aspects discussed, public health hazard in relation to goat's milk: southern Ontario

Toxoplasma gondii
Tizard, I. R.; Chauhan, S. S.; and Lai, C. H., 1977, J. Hyg., Cambridge, v. 78 (2), 275-282
Toxoplasma gondii, epidemiologic survey, prevalence of antibodies in human serum, inverse relationship between urban size and infection prevalence: Province of Ontario, Canada

Toxoplasma gondii
Tizard, I. R.; Fish, N. A.; and Quinn, J. P., 1976, J. Hyg., Cambridge, v. 77 (1), 11-21
extensive survey of human serum for presence of antibodies to Toxoplasma gondii and observations on probable epidemiology: Canada

Toxoplasma gondii, illus.
Todd, K. S., jr., 1972, Illinois Vet., v. 15 (1), 14-16
Toxoplasma gondii, cats, historical review; public health importance and suggestions for preventing human infection

Toxoplasma gondii, illus.
de la Torre Rendon, F. E.; and Gorraez de la Torre, M. T., 1974, Patologia, v. 12 (1), 15-39
Toxoplasma gondii encephalitis with central nervous system lymphomatous infiltrates in young girl being treated with immunosuppressive drugs for Hodgkin's disease, diagnosis at postmortem examination, case report, extensive bibliography: Mexico

Toxoplasma gondii, illus.
Tsai, C. S.; and Cross, J. H., 1970, Southeast Asian J. Trop. Med. and Pub. Health, v. 1 (4), 556-557 [Demonstration]
Toxoplasma gondii, re-confirmation of coccidian-like cycle in the intestinal tract of the domestic cat (exper.)

Toxoplasma gondii
Turner, G. V. S., 1976, J. South African Vet. Ass., v. 47 (3), 227-231
epidemiology and public health significance, life cycle, review

Toxoplasma gondii
Uhlikova, M.; and Huebner, J., 1974, J. Protozool., v. 21 (3), 458-459 [Abstract]
Toxoplasma gondii, Toxocara cati, concomitant incidence of infection in 8 children

Toxoplasma gondii
Valkoun, A.; and Jira, J., 1975, Ceskoslov. Epidemiol., Mikrobiol., Imunol., v. 24 (5), 289-292
Toxoplasma gondii, cultivation in human embryonic lung cell cultures, preparation of pure suspensions of zoites free from host cell contamination by filtration through paper filters

Toxoplasma gondii
Valkoun, A.; and Jira, J., 1975, J. Protozool., v. 22 (3), 69A [Abstract]
Toxoplasma gondii, use of lyophilized antigen for indirect fluorescent antibody test

Toxoplasma gondii
Valkoun, A.; and Jira, J., 1977, J. Protozool., v. 24 (2), Suppl., 16A-17A [Abstract]
Toxoplasma gondii, purified antigen for skin test

Toxoplasma gondii
Vallat, M.; et al., 1976, Nouv. Presse Med., v. 5 (38), 2543-2544 [Letter]
Toxoplasma gondii, 2 case reports of acquired toxoplasmosis with ocular symptoms in women suffering from Hodgkin's disease, implications for defective immunologic responses: France

Toxoplasma gondii, illus.
Van Pelt, R. W.; and Dieterich, R. A., 1973, J. Wildlife Dis., v. 9 (3), 258-261
Toxoplasma gondii in Phoca vitulina richardii (liver), complicating generalized staphyloccal infection, clinical course with fatal outcome, pathologic findings: captured at Cold Bay, Alaska, held in captivity until death 21 days later

Toxoplasma gondii
Vethanyagam, A.; and Bryceson, A. D. M., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 524-525
Toxoplasma gondii in man presenting and treated as hepatitis with mixed Giardia lamblia infection, later diagnosis of active acquired toxoplasmosis and treatment with pyrimethamine and sulfadimidine resolved condition: Great Britain

Toxoplasma gondii, illus.
Villegas-Gonzalez, J.; Fastag de Shor, A.; and Villegas-Silva, R., 1976, Bol. Med. Hosp. Inf., v. 33 (1), 137-149
Toxoplasma gondii, human congenital infections, pathology

Toxoplasma gondii, illus.
Vivier, E.; and Petitprez, A., 1972, Protistologica, v. 8 (2), 199-221
Toxoplasma gondii, ultrastructure, cytochemistry, with special reference to apical complex and cytoplasmic organelles

Toxoplasma gondii, illus.
Vrablic, J., 1977, Biologia, Bratislava, s. B, Zool. (1), v. 32 (2), 89-99
Toxoplasma gondii, schizogony and gametogony in small intestine of cats (exper.)

Toxoplasma gondii, illus.
Vrablic, J., 1977, Biologia, Bratislava, s. B, Zool. (4), v. 32 (11), 841-850
Toxoplasma gondii, intestine of cat, intraepithelial stages of asexual and sexual multiplication; macrogamogony and oogony

Toxoplasma gondii, illus.
Vrablic, J., 1977, Biologia, Bratislava, s. B, Zool. (3), v. 32 (8), 565-573
Toxoplasma gondii, microgamogony, cats

Toxoplasma gondii
Vrablic, J., 1977, J. Protozool., v. 24 (2), Suppl., 17A [Abstract]
gamonts, some morphological and biological characteristics, comparison with schizonts

Toxoplasma gondii
Waldeland, H., 1976, Norwegian J. Zool., v. 24 (4), 459 [Abstract]
prevalence in sheep in four regions of Norway

Toxoplasma gondii
Waldeland, H., 1976, Acta Vet. Scand., v. 17 (4), 403-411
Toxoplasma gondii, sheep muscular tissue, incidence correlated with dye test titres and haemoglobin type (higher in type B than A or AB), possible genetic influence on infection, epidemiology, meat inspection: southern Norway

Toxoplasma gondii
Waldeland, H., 1976, Acta Vet. Scand., v. 17 (4), 412-425
Toxoplasma gondii, sheep, importance as a cause of reproductive loss, main cause of abortion: Norway

Toxoplasma gondii
Waldeland, H., 1976, Acta Vet. Scand., v. 17 (4), 426-431
Toxoplasma gondii, sheep, Sabin and Feldman dye test, high correlation between microtiter system and conventional method, advantages of microtiter system

Toxoplasma gondii
Waldeland, H., 1976, Acta Vet. Scand., v. 17 (4), 432-440
Toxoplasma gondii, prevalence in lambs, comparison with ewes, variations in epidemiological patterns, no significant differences in geographical distribution: southern Norway

Toxoplasma gondii
Waldeland, H., 1977, Acta Vet. Scand., v. 18 (1), 91-97
Toxoplasma gondii, sheep flocks, reproductive loss in specific age groups, apparently no transmission between sheep, probably transmitted by silage

Toxoplasma gondii
Waldeland, H., 1977, Acta Vet. Scand., v. 18 (2), 237-247
Toxoplasma gondii, sheep, antibody formation, dye test titres higher in ewes that had aborted; course of titre levels in young lambs; titre not influenced by listeric encephalitis; higher titres in sheep with hemoglobin type B

Toxoplasma gondii
Waldeland, H., 1977, Acta Vet. Scand., v. 18 (2), 248-256
Toxoplasma gondii, sheep experimentally infected with sheep strain and RH strain, no significant changes in blood values, T. gondii cysts recovered from meat samples of lambs inoculated with sheep strain, but not from lambs inoculated with RH strain

Toxoplasma gondii
Wallace, G. D., 1976, Am. J. Trop. Med. and Hyg., v. 25 (1), 48-53
Toxoplasma gondii, human, prevalence of dye-test antibody in relation to host age and ethnic group, generally high except for people of Japanese ancestry living in Hawaii and for aborigines and ethnic Chinese living in Taiwan: Pacific Islands

Toxoplasma gondii
Weinberg, J. B.; and Hibbs, J. B., jr., 1977, Nature, London (5625), v. 269, 245-247
endocytosis of red blood cells or haemoglobin by activated macrophages from Toxoplasma gondii or BCG-infected mice inhibits tumoricidal effect

Toxoplasma gondii
Wellensiek, H. J.; et al., 1976, Ztschr. Immunitaetsforsch., v. 152 (2), 123 [Abstract]
Toxoplasma gondii, Sabin-Feldman dye test, immunocytolysis caused by properdin-dependent alternate pathway activation of human complement

Toxoplasma gondii, illus.
Werner, H., 1968, Bol. Chileno Parasitol., v. 23 (3-4), 98-101
Toxoplasma gondii, human latent uterine toxoplasma infections with resulting congenital transmission, prospects for diagnosis, need for antitoxoplasma treatment of women who have had habitual abortions or suspected infections

Toxoplasma gondii
Werner, H.; and Catar, G., 1974, Tropenmed. u. Parasitol., v. 25 (3), 313-317
Toxoplasma gondii, mice, SDDS in combination with pyrimethamine or sulfamonomethoxine, possible to prevent formation of cysts but not to eliminate cysts from chronic infections

Toxoplasma gondii, illus.
Werner, H.; and Egger, I., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 174-180
Toxoplasma gondii, mice, latent or chronic infection protects against fatality from challenge with virulent strain, host may harbor cysts of all strains inoculated irrespective of whether homologous or heterologous strain was used for challenges

Toxoplasma gondii, illus.
Werner, H.; and Janitschke, K., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 57-64
Toxoplasma gondii, morphological variations in developmental stages, details of sexual forms and oocysts, life cycle completed only in domestic cat, partial common antigenicity demonstrated between Toxoplasma and Isospora

Toxoplasma gondii
Wildfuehr, G.; and Wildfuehr, W., 1975, Toxoplasmose. Ratgeber fur Artzte und Tierarzte, 320 pp., illus.
Toxoplasma gondii, guidebook for physicians and veterinarians, morphology, life cycle, diagnosis, epidemiology, pathology, prenatal infection, veterinary aspects, extensive review

Toxoplasma gondii
Williams, D. M.; Krick, J. A.; and Remington, J. S., 1976, Am. Rev. Resp. Dis., v. 114 (3), 593-627
parasitic pulmonary infections in the compromised host, immunopathologic aspects, clinical manifestations, diagnosis and treatment, review

Toxoplasma gondii, illus.
Wilson, H. R.; and Fairchild, D. G., 1977, Stain Tech., v. 52 (2), 105-111
modified Whipf's polychrome staining to demonstrate protozoans and tissue components in cutaneous lesions of infected animals

Toxoplasma gondii
Wing, E. J.; et al., 1977, Nature, London (5621), v. 268, 642-644
macrophages activated by infection with Toxoplasma gondii or Trichinella spiralis, ability to inhibit tumor cell DNA synthesis and to inhibit intracellular multiplication of Toxoplasma gondii, results show that macrophages characterized as activated by one criterion may not satisfy other criteria of activation and that differences in functional capacity depend on the method used to activate the macrophages

Toxoplasma gondii
Wing, E. J.; and Remington, J. S., 1977, Cellular Immunol., v. 30 (1), 108-121
Toxoplasma gondii, mice, studies on regulation of lymphocyte reactivity by normal and activated macrophages

Toxoplasma gondii
Wing, E. J.; and Remington, J. S., 1977, West. J. Med., San Francisco, v. 126 (1), 14-31
role of cell-mediated immunity (and enhancement by levamisole) in host defense against human parasitic diseases, extensive review

Toxoplasma gondii
Wosu, N. J.; et al., 1977, Lab. Animal Sc., v. 27 (2), 210-216
Encephalitozoon cuniculi, rabbits (exper.), diagnosis by immunofluorescence and intradermal test, no cross reactions between E. cuniculi and experimentally induced Toxoplasma gondii, Eimeria perforans, or E. stiedai

Toxoplasma gondii
Wu, Y. H.; et al., 1975, Taiwan J. Vet. Med. and Animal Husb. (27), 19-24
pigs (brain, diaphragm) and pork meat, detection survey, skin test, hemagglutination test, isolation method: Kaohsiung and Pingtung areas, Taiwan

Toxoplasma gondii, illus.
Wyers, M.; and Marchand, A., 1977, Rec. Med. Vet., v. 153 (1), 9-13
stages of life cycle, review, practical applications

Toxoplasma gondii, illus.
  Wyers, M.; and Marchand, A., 1977, Rec. Med. Vet., v. 153 (3), 169-174
    Toxoplasma gondii, summary of clinical signs of human toxoplasmosis and means of transmission to people; review of risks of cat owners and role of veterinary surgeons in control of human infection

Toxoplasma gondii, illus.
  Yoshii, Z.; Tokunaga, J.; and Tawara, J., 1976, Atlas of scanning electron microscopy in microbiology, 233 pp.
    Trichomonas vaginalis, Toxoplasma gondii, atlas of scanning electron microscopy

Toxoplasma gondii, illus.
  Yoshizumi, M. O., 1976, Arch. Path. and Lab. Med., v. 100 (9), 487-490
    light and electron microscopic study of pathology of Toxoplasma gondii retinitis lesions in exper. rabbit

Toxoplasma gondii, illus.
  Zaman, V., 1968, Med. J. Malaya, v. 22 (3), 195-197
    Toxoplasma gondii invasion of sarcoma mouse ascites tumor, increased ascites caused decreased infection

Toxoplasma gondii
  Zaman, V., 1972, Southeast Asian J. Trop. Med. and Pub. Health, v. 3 (4), 623
    unsuccessful attempted infection of Felis bengalensis

Toxoplasma gondii, illus.
  Zaman, V.; and Colley, F. C., 1970, Southeast Asian J. Trop. Med. and Pub. Health, v. 1 (4), 457-464
    Toxoplasma gondii, light microscopic study of endogenous stages

Toxoplasma gondii
  Zaman, V.; and Colley, F. C., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (3), 408 [Demonstration]
    Toxoplasma gondii, ultrastructural study of penetration of the host cell by parasite

Toxoplasma gondii
  Zardi, O.; et al., 1969, Acta Parasitol. Polon., v. 16 (1-19), 1968-1969, 1-8
    Toxoplasma gondii, survey, incidence, chickens, dye test and immunofluorescence, seven strains isolated by inoculation and serial passage in mice: Lazio, vicinity of Rome

Toxoplasma gondii
  Zigas, V., 1976, Papua N. Guinea Med. J., v. 19 (4), 225-230
    Toxoplasma gondii in humans, epidemiologic survey for prevalence of antibodies, possible correlation with ingestion of raw meat rather than presence of cats: New Britain, Papua New Guinea

Toxoplasma gondii, illus.
  Zimmermann, W. J., 1976, Pub. Health Rep., U.S. Pub. Health and Mar. Hosp. Serv., v. 91 (6), 526-532
    Toxoplasma gondii, no definitive relationships could be determined between epidemiologic characteristics examined and the presence of antibodies to toxoplasmosis in survey of veterinary college staff and students at the Iowa State University

Toxoplasma gondii
  Zukovic, M.; et al., 1973, Acta Parasitol. Iugoslavica, v. 4 (1), 43-45
    Isospora bigemina, small form from dogs given to mice orally or peritoneally, single or repeated dose; toxoplasmosis did not develop; no identity with Toxoplasma gondii found

Toxoplasma gondii, illus.
  van der Zypen, E.; and Piekarski, G., 1968, Bol. Chileno Parasitol., v. 23 (3-4), 90-94
    Toxoplasma gondii, ultrastructure of endodyogeny

Toxoplasma hammondi (Frenkel 1974) Levine and Nye (in press) [?n. comb.]
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    Syn.: Hammondia hammondi Frenkel 1974
    Mus musculus
    Rattus norvegicus
    Mesocricetus auratus
    Felis catus

Toxoplasma hammondi (Frenkel & Dubey, 1975) comb. nov.
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    diagnosis
    Syn.: Hammondia hammondi Frenkel & Dubey, 1975

Toxoplasma hominis Wolf, Cowen, and Paige 1939
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma hominis Wolf, Cowen & Paige, 1939
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 36-41
    as syn. of Toxoplasma gondii (Nicolle & Manceaux, 1908) Nicolle & Manceaux, 1909

Toxoplasma laidlawi Coutelen 1932
  Levine, N. D., 1977, J. Parasitol., v. 63 (1), 36-51
    as syn. of Toxoplasma gondii (Nicolle and Manceaux 1908) Nicolle and Manceaux 1909

Toxoplasma laidlawi Coutelen, 1932
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Mugil cephalus (gills)
Liza ramada (gills)
Siganus luridus
S. rivulatus
all from Elat, Gulf of Aqaba, Red Sea

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A. dieffenbachii
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Bos taurus: Chile

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Rattus norvegicus (caecum): Tunis, Tunisia

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Mus musculus
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Trichomonas muris, illus.
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Tsvetkova, A.; Iankov, N.; and Dorevska, K., 1975, Vet. Med. Nauk., v. 12 (10), 83-87
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Trichomonas vaginalis
Basile, F., 1970, Minerva Ginec., v. 22 (11), 584-587
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human parasitic gynecologic diseases, clinical aspects, epidemiology, sexual transmission, review

Trichomonas vaginalis
Tatra, G., 1972, Munchen. Med. Wchnschr., v. 114 (16), 773-776
human Trichomonas vaginalis causing vulvo-vaginal infections, dequalinium chloride in combination with metronidazole, good results

Trichomonas vaginalis
Teisanu, E.; and Popescu-Tomus, D., 1971, Oto-Rino-Laringol., v. 16 (4), 265-269
Trichomonas vaginalis, human sinusitis resulting from parasitic infection, associated asthma, case reports: Romania

Trichomonas vaginalis Donne, 1836, illus.
Toyos Aparicio, J. M., 1974, An. Fac. Vet. Leon, v. 20, 35-90
35 strains of Trichomonas vaginalis, taxonomy, morphology, culture, biochemistry, diagnosis, pathology, comparison of pathogenicity in natural and experimental infections, mice, no relationship observed between pathogeny and clinical picture

Trichomonas vaginalis
Trzeciak, B., 1975, Polski Tygod. Lekar., v. 30 (28), 1187-1188
women, incidence survey results show only 7.6% infection rate in Poland

Trichomonas vaginalis
Valent, M.; et al., 1977, J. Protozool., v. 24 (2), Suppl., 16A [Abstract]
Trichomonas vaginalis, failure to isolate strain with in vitro resistance to metronidazole

Trichomonas vaginalis
Villani, L.; and Moneta, E., 1970, Minerva Ginec., v. 22 (18), 880-886
Trichomonas vaginalis, human vaginitis associated with fungal and bacterial infections, good results with mysteclin-V, case statistics: Roma

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Trichomonas vaginalis, Toxoplasma gondii, atlas of scanning electron microscopy

Trichomonas vaginalis
Zajac, W.; and Malyszko, E., 1977, Przegl. Dermat., v. 64 (2), 185-189
Trichomonas vaginalis, analysis of incidence of human infection of urogenital tract in the absence of venereal diseases: Poland

Trichomonas vaginalis
Zaremba, A., 1976, Przegl. Dermat., v. 63 (4), 513-517
Trichomonas vaginalis, humans, problems of transmission in asymptomatic trichomoniasis

Trichomonas vaginalis
Zaremba, A.; and Szarmach, H., 1977, Przegl. Dermat., v. 64 (4), 461-463
Trichomonas vaginalis, diagnosis of asymptomatic infections in males, comparison of several diagnostic methods, positive findings increased significantly after prostatic massage

Trichomonas vaginalis
Zawadzki, J., 1977, Przegl. Lek., v. 34 (4), 445-447
Trichomonas vaginalis, canesten used alone or simultaneously with metronidazole in women with vaginitis, less complications after gynecological surgery if treated prophylactically prior to surgery

Trichomoniasis
Aure, J. C.; and Tveter, L., 1969, Tidsskr. Norske Laegefor., v. 89 (23), 1792-1794
human trichomoniasis colpitis, clinical trials using atricana not as successful as trials with metronidazole and nifuratel

Trichomoniasis
Baron, A., 1970, Minerva Ginec., v. 22 (4), 269-271
human vaginal trichomoniasis, good results with macmiror: Poland

Trichomoniasis
Cavaglieri, D.; Falsetti, L.; and Caputo, G., 1974, Minerva Ginec., v. 26 (5), 303-305
human vaginal trichomoniasis, drug trials comparing nimorazole and metronidazole in varying doses

Trichomoniasis
Celaya Barrera, R., 1976, Medicina, Mexico (1214), an. 57, v. 56, 336-338
human vaginal trichomoniasis, clinical trials comparing efficacy of tinidazole and nitrimidazine: Mexico

Trichomoniasis
Counio, 1975, Marseille Med., v. 112 (4), 233-237
human vaginitis, good treatment results in combined use of hormones and trichomonicides: France

Trichomoniasis
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human vaginal trichomoniasis, single-dose treatment with tinidazole

Trichomoniasis
Dykers, J. R., jr., 1976, N. England J. Med., v. 295 (7), 395-396 [Letter]
human vaginal trichomoniasis, single dose therapy with metronidazole, follow-up report of 2 women with recurring infections: North Carolina

Trichomoniasis
Epureanu, P.; and Lesnic, C., 1970, Presse Therm. et Climat., v. 107 (2), 112-114
human trichomoniasis, chronic cervicitis and cervico-colpitis treated with sulfurous mineral water baths, inflammatory symptoms improved but there was no actual parasiticidal effects from the baths: Calimanesti, Roumanie

Trichomoniasis
Gualandri, V.; and Galeazzi, L., 1971, Minerva Ginec., v. 23 (3), 148-150
human vaginal trichomoniasis, statistical survey of occurrence in conjunction with genital neoplasms

Trichomoniasis
Gwiezdzinski, Z.; Pekowski, M.; and Kozlowski, J., 1971, Polski Tygod. Lekar., v. 26 (17), 642-643
trichomoniasis in young man resulting in erosive balanoposthitis, successful cure with metronidazole, differential diagnosis from atypical syphilitic lesions

Trichomoniasis
Hauser, K. W., 1976, Prakt. Tierarzt , v. 57, Sondernummer, 20-21
trichomoniasis of pigeons, successful treatment with Carnidazol

Trichomoniasis
Jenny, J., 1977, Praxis, Bern, v. 66 (8), 247-250
human vaginal trichomoniasis, clinical aspects, diagnosis using microscopic examination of fresh vaginal smears, treatment with fasigyn given orally

Trichomoniasis
Kazmier, F. J., 1976, Mayo Clin. Proc., v. 51 (12), 782-784
trichomoniasis, woman on warfarin anticoagulant therapy, metronidazole treatment of parasitic infection resulted in drug interactions with increased prothrombin times and internal hemorrhaging

Trichomoniasis
Kern, A., 1974, Paediat. u. Grenzgeb., v. 13 (2-3), 147-162
urogenital trichomoniasis in children and adolescents, diagnosis and management

Trichomoniasis
Kessler, H. J.; Schulz, V.; and Wagener, B., 1976, Arzneimittel-Forsch., v. 26 (7), 1371-1375
distribution, absorption and elimination of trichomonocidal drug ZK 25 095 tested in normal subjects

Trichomoniasis
Komaroff, A. L.; Cohen, A. B.; and McCue, J., 1976, J. Am. Med. Ass., v. 235 (19), 2081-2082
possible carcinogenicity and mutagenicity of metronidazole, discussion of continued use to treat human trichomoniasis

Trichomoniasis
Kulichkin, P. N., 1975, Vestnik Sel'skokhoz. Nauki Kazakhstana (9), 110-111
trichomoniasis, bovine, microscopic examination of vaginal or prepucial washings at farm most expedient method of diagnosis, laboratory culture useful for cases not discovered by microscopic method

Trichomoniasis
Laudisio, V.; Lirosi, G.; and Guarascio, A., 1970, Minerva Ginec., v. 22 (21), 1059-1063
human vaginal trichomoniasis, relationship between hormonal levels and predisposition to vaginal infections

Trichomoniasis
Mahony, J. D. H.; Harris, J. R. W.; and Farrer, C. J., 1975, Brit. J. Clin. Pract., v. 29 (3), 71-72
human vaginal trichomoniasis, comparative clinical trials using nimorazole and metronidazole: England

Trichomoniasis
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human vaginitis, clinical trials using metronidazole tablets with nystatin pesseries, comparison with nitrofuratel tablets and pesseries

Trichomoniasis
Perju, A.; and Ciuca, T., 1971, Pediatria, Bucuresti, v. 20 (6), 561-564
urogenital trichomoniasis in young girls, treatment with metronidazole or flagyl: Romania

Trichomoniasis
Sackman Muriel, C., 1977, Gac. Vet., Buenos Aires (317), v. 39, 46-53
bovine trichomoniasis, dimetridazole, bulls: provincias de Buenos Aires, Santa Fe, Cordoba y San Luis

Trichomoniasis
Underhill, R. A.; and Peck, J. E., 1974, Brit. J. Clin. Pract., v. 28 (4), 134-136
human trichomonal vaginitis, metronidazole, single-dose treatment compared with 7-day therapy: England

Trichomoniasis
Voogd, C. E.; van der Stel, J. J.; and Jacobs, J. J. J. A. A., 1974, Mutation Research, v. 26 (6), 483-490
possible mutagenic action of metronidazole and nimorazole used to treat human trichomoniasis and dimetridazole and ronidazole used prophylactically against Histomonas meleagridis in poultry

Trichorhynchus pulcher
Marques, A.; Ormieres, R.; and Puissegur, C., 1977, J. Protozool., v. 24 (4), 61A [Abstract]
ultrastructure
Scutigera coleoptrata

Tricystis leuckarti (Mingazzini, 1893) Ormieres, 1965
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Syn.: Lankesteria leuckarti (Mingazzini, 1893) Labbe, 1899

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Brugerolle, G., 1975, J. Protozool., v. 22 (3), 468-475
Diplomonadina emend.

Tripartiella
Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Trichodinidae

Tritrichomonas augusta
Honigberg, B. M., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 469-550
immunology of trichomonads, review

Tritrichomonas augusta
Juillerat, F., 1975, Proc. Indiana Acad. Sc., v. 84, 1974, 131-132
trichomonad costae type A and B, molecular weight differences in polypeptides, usefulness of technique for biochemical studies of evolution of trichomonads

Tritrichomonas augusta
Rainis, K. G.; and Buttrey, B. W., 1976, J. Protozool., v. 23 (2), 12A [Abstract]
Tritrichomonas augusta, formation of pseudocysts in relation to temperature

Tritrichomonas augusta
Samuels, R.; and Pratt, M., 1975, J. Protozool., v. 22 (3), 21A [Abstract]
Tritrichomonas augusta, β-galactosidases, partial characterization

Tritrichomonas augusta
Toma, M. J.; and Concannon, J. N., 1973, J. Protozool., v. 20 (4), 501-502
Tritrichomonas augusta, culture in serum-free medium, maintained with weekly transfer for over five months

Tritrichomonas caviae, illus.
Brugerolle, G., 1972, Protistologica, v. 8 (3), 353-363
trichomonads, ultrastructural and cytochemical characterization of two types of cytoplasmic granules

Tritrichomonas eberthi
Leibovitz, L., 1973, Am. J. Vet. Research, v. 34 (8), 1053-1061
Protozoa contributing to pathogenesis of necrotic enteritis in commercial white Pekin breeder ducks; necrotic enteritis also observed in other wild and domestic Anseriformes

Tritrichomonas equi, illus.
Humphrey, W. J., 1977, Mod. Vet. Pract., v. 58 (4), 365-367
horses, protozoal colitis, diagnosis

Tritrichomonas foetus
Budilova, M.; and Kulda, J., 1977, J. Protozool., v. 24 (4), 49A [Abstract]
Tritrichomonas foetus, enhancement of virulence (infectivity + mortality) for mice by ferric ammonium citrate, indicates accessibility of iron in form utilizable by parasite as critical factor in infection

T[ritrichomonas] foetus
Cerkasov, J.; et al., 1974, J. Protozool., v. 21 (3), 454-455 [Abstract]
T[ritrichomonas] foetus, ATP forming systems in hydrogenosomes

Tritrichomonas foetus
Cerkasov, J.; Cerkasovova, A.; and Dohnalova, M., 1977, J. Protozool., v. 24 (4), 49A [Abstract]
Tritrichomonas foetus, malate dehydrogenases

Tritrichomonas foetus
Cerkasovova, A.; et al., 1973, J. Protozool., v. 20 (4), 535
Tritrichomonas foetus, large granule fraction, biochemical characterization, importance in energy-producing metabolism

Tritrichomonas foetus
Christensen, H. R.; Clark, B. L.; and Parsonson, I. M., 1977, Austral. Vet. J., v. 53 (3), 132-134
Tritrichomonas foetus, incidence in young replacement bulls following introduction into an infected herd, implications for control of trichomoniasis

Tritrichomonas foetus
De Carli, G. A.; and Guerrero, J., 1976, Rev. Latinoam. Microbiol., v. 18 (3), 167-171
Tritrichomonas suis and T. foetus, antigenic comparison by gel immunodiffusion; closely related antigenic structure found

Tritrichomonas foetus
Dohnalova, M.; and Kulda, J., 1975, J. Protozool., v. 22 (3), 61A [Abstract]
Tritrichomonas foetus, 19 strains, pathogenicity assayed on laboratory mice inoculated subcutaneously or intraperitoneally

Tritrichomonas foetus
Dohnalova, M.; and Kulda, J., 1977, J. Protozool., v. 24 (4), 50A [Abstract]
Tritrichomonas foetus, meagre respiration with erythritol as substrate, no stimulatory effect of erythritol on growth in vitro, results indicated that confinement of T. foetus to lumen of bovine uterus is not related to presence of erythritol at this site (in contrast to Brucella abortus)

Tritrichomonas foetus
Honigberg, B. M., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 469-550
immunology of trichomonads, review

Tritrichomonas foetus
Honigberg, B. M.; and Mohn, F. A., 1973, J. Protozool., v. 20 (1), 146-150
Trichomonas vaginalis, T. gallinae, Tritrichomonas foetus, method for isolation of purified, highly polymerized native DNA

Tritrichomonas foetus
Juillerat, F., 1975, Proc. Indiana Acad. Sc., v. 84, 1974, 131-132
trichomonad costae type A and B, molecular weight differences in polypeptides, usefulness of technique for biochemical studies of evolution of trichomonads

Tritrichomonas foetus
Kovac, L.; Cerkasov, J.; and Cerkasovova, A., 1977, J. Protozool., v. 24 (4), 51A [Abstract]
Tritrichomonas foetus cells, osmotic and permeability properties

Tritrichomonas foetus
Kralova, J.; and Kulda, J., 1974, J. Protozool., v. 21 (3), 456 [Abstract]
Tritrichomonas foetus, course of mitosis in culture

Tritrichomonas foetus
Kulda, J.; and Budilova, M., 1977, J. Protozool., v. 24 (4), 51A [Abstract]
Tritrichomonas foetus, multiplication of parasites in mouse peritoneal cavity stimulated by ferric ammonium citrate, indicates importance of trichomonad iron metabolism in host-parasite relationships

Tritrichomonas foetus
Kulda, J.; Budilova, M.; and Dohnalova, M., 1977, J. Protozool., v. 24 (2), Suppl., 15A-16A [Abstract]
Tritrichomonas foetus, enhancement of virulence to mice by parenteral administration of ferric-ammonium citrate

Tritrichomonas foetus
Kulda, J.; Kralova, J.; and Vavra, J., 1974, J. Protozool., v. 21 (3), 456-457 [Abstract]
Tritrichomonas foetus, ultrastructural 3,3'-diaminobenzidine-positive component of hydrogenosomes, considered to be site of oxidative (possibly peroxidative) reaction without catalase involvement

Tritrichomonas foetus
Kulda, J.; Kralova, J.; and Vavra, J., 1977, J. Protozool., v. 24 (4), 51A-52A [Abstract]
Tritrichomonas foetus, hydrogenosomes, ultrastructure and 3,3'-diaminobenzidine cytochemistry

Tritrichomonas foetus
Kulda, J.; and Zavadova, H., 1975, J. Protozool., v. 22 (3), 65A-66A [Abstract]
Tritrichomonas foetus, T. suis, high neuraminidase activity, none in T[richomonas] vaginalis, possible pathogenetic significance in Tritrichomonas foetus

Tritrichomonas foetus
Lindmark, D. G.; and Mueller, M., 1973, J. Protozool., v. 20 (4), 500
Tritrichomonas foetus, Monocercomonas sp., subcellular distribution of flavins, results in agreement with suggestion of flavin-mediated electron transport

Tritrichomonas foetus
Lindmark, D. G.; and Mueller, M., 1974, J. Protozool., v. 21 (3), 436 [Abstract]
affinity of metronidazole to reducing enzyme system(s) in Trichomonas vaginalis and Tritrichomonas foetus homogenates, inhibitory action of metronidazole on these enzymes due to competition for low redox potential electrons

Tritrichomonas foetus
Lindmark, D. G.; and Mueller, M.; 1975, J. Protozool., v. 22 (3), 13A [Abstract]
Tritrichomonas foetus, acetate formation from pyruvate by hydrogenosomes

Tritrichomonas foetus
McLoughlin, D. K.; and Chute, M. B., 1975, Proc. Helminth. Soc. Washington, v. 42 (2), 183-184
Trypanosoma theileri in peripheral blood and in bloody vaginal washings of Hereford cows being used for exper. treatment studies of Tritrichomonas foetus infections (dimetridazole), transitory urogenital disturbance

Tritrichomonas foetus
Meingassner, J. G.; and Mieth, H., 1976, Experientia, v. 32 (2), 183-184
Tritrichomonas foetus, induction of metronidazole resistance, marked cross-resistance to other 5-nitroimidazole-derivatives in vitro and in vivo

Tritrichomonas foetus
Miyata, A., 1973, Nettai Igaku (Trop. Med.), v. 15 (3), 141-153
Trichomonas vaginalis, Tritrichomonas foetus, preservation by freezing, effects of precooling and of cryo-protective glycerol, dimethyl sulfoxide or salt concentration on storage

Tritrichomonas foetus
Mueller, M.; and Lindmark, D. G., 1976, Antimicrob. Agents and Chemotherapy, v. 9 (4), 696-700
Tritrichomonas foetus, Trichomonas vaginalis, Entamoeba invadens, uptake of metronidazole under anaerobic vs. aerobic conditions, effect on viability of T. foetus

Tritrichomonas foetus (Riedmuller, 1928)
Santos, S. M.; do Amaral, V.; and Reboucas, M. M., 1975, Biologico, S. Paulo, v. 41 (8), 234-237
cattle, found in preputial, vaginal and stomach fluid of aborted foetuses; geographic distribution: State of Sao Paulo, Brazil

Tritrichomonas foetus
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Tritrichomonas foetus
Wosu, L. O., 1977, Vet. Microbiol., v. 2 (1), 89-93
Tritrichomonas foetus, simplified medium for cultivation, rapid technique for obtaining pure cultures without sub-cultivation

Tritrichomonas foetus var. belfast
Turnbull, P. A., 1977, Austral. Vet. J., v. 53 (6), 274-279
Bos (genitalia): 3 Western Australian abattoirs

Tritrichomonas foetus var. belfast
Wosu, L. O., 1977, Austral. Vet. J., v. 53 (7), 340-341
Tritrichomonas foetus, serotyping of isolates from a bull and infected cows showed that strain in bull had apparently changed from var. brisbane to var. belfast over a four-month period, techniques in serotyping of strains, possible cause of change in serotype, possible mixed infection initially, with predominant strain later eliminated

Tritrichomonas foetus var. brisbane
Clark, B. L.; Dufty, J. H.; and Parsonson, I. M., 1977, Austral. Vet. J., v. 53 (4), 170-172
Tritrichomonas foetus var. brisbane, passive transmission from infected to non-infected cows during mating with non-infected bulls; application of T. foetus var. brisbane to vulva failed to cause genital infection in cows, suggests that parasite is unable to migrate to vagina

Tritrichomonas foetus var. brisbane
Parsonson, I. M.; Clark, B. L.; and Dufty, J. H., 1976, J. Comp. Path., v. 86 (1), 59-66
Tritrichomonas foetus var. brisbane, pathogenesis and pathology in virgin heifers mated to an infected bull

Tritrichomonas foetus var. brisbane
Wosu, L. O., 1977, Austral. Vet. J., v. 53 (7), 340-341
Tritrichomonas foetus, serotyping of isolates from a bull and infected cows showed that strain in bull had apparently changed from var. brisbane to var. belfast over a four-month period, techniques in serotyping of strains, possible cause of change in serotype, possible mixed infection initially, with predominant strain later eliminated

Tritrichomonas lacertae, illus.
Brugerolle, G., 1972, Protistologica, v. 8 (3), 353-363
trichomonads, ultrastructural and cytochemical characterization of two types of cytoplasmic granules

Tritrichomonas muris, illus.
Grewal, M. S., 1975, Research Bull. Panjab Univ., v. 23 (3-4), 1972, 119-124
Rattus rattus: Chandigarh, India

Tritrichomonas muris
Wilhelm, W. E.; and Pope, D. C., 1974, J. Protozool., v. 21 (3), 434-435 [Abstract]
Tritrichomonas muris, infections of laboratory colony of Mesocricetus auratus (caecum, small intestine, colon), hamsters fed high protein diet became trichomonad free, trichomonad-free hamsters showed higher mortality, weight loss, and fur thinning suggesting possible mutualistic relationship, successful transfaunation to laboratory and wild Mus musculus, reinfection of trichomonad-free Mesocricetus auratus, parasite morphology

Tritrichomonas suis
De Carli, G. A.; and Guerrero, J., 1976, Rev. Latinoam. Microbiol., v. 18 (3), 167-171
Tritrichomonas suis and T. foetus, antigenic comparison by gel immunodiffusion; closely related antigenic structure found

Tritrichomonas suis
Kulda, J.; and Zavadova, H., 1975, J. Protozool., v. 22 (3), 65A-66A [Abstract]
Tritrichomonas foetus, T. suis, high neuraminidase activity, none in T[richomonas] vaginalis, possible pathogenetic significance in Tritrichomonas foetus

Tritrichomonas suis, illus.
Marr, J. L., 1976, Proc. Indiana Acad. Sc., v. 85, 1975, 411-417
Tritrichomonas suis, in vitro culture in chick chorio-allantoic fluid resulted in lower numbers and smaller individuals than in commercial C.P.L.M. medium; difference perhaps based on different carbohydrate utilization

Troglodytella abrassarti Brumpt and Joyeux 1921
File, S. K.; McGrew, W. C.; and Tutin, C. E. G., 1976, J. Parasitol., v. 62 (2), 259-261
Pan troglodytes schweinfurthii (feces): Gombe National Park, Tanzania

Trypanophis grobbeni "flagellate" form, illus.
Cachon, J.; Cachon, M.; and Charnier, M., 1972, Protistologica, v. 8 (2), 223-236
Trypanophis grobbeni, ultrastructure, forms previously thought to be different life cycle stages are probably 2 separate parasite species, the "flagellate" form being Bodonidae and the "gregarinian" form probably Sporozoa
Chelophyes (oleocyte)
Lensia (oleocyte)
Diphyes (oleocyte)
Muggia (oleocyte)
Halistema (canaus endodermique des bractees)
all from Villefranche

Trypanophis grobbeni "gregarinian" form, probably Sporozoa [sp.], illus.
Cachon, J.; Cachon, M.; and Charnier, M., 1972, Protistologica, v. 8 (2), 223-236
Trypanophis grobbeni, ultrastructure, forms previously thought to be different life cycle stages are probably 2 separate parasite species, the "flagellate" form being Bodonidae and the "gregarinian" form probably Sporozoa
Chelophyes (cavite gastro-vasculaire, mesoglee)
Lensia (cavite gastro-vasculaire, mesoglee)
all from Villefranche

Trypanoplasma
Lom, J., 1973, J. Protozool., v. 20 (4), 537
Trypanosoma and Trypanoplasma from freshwater fishes, cross-infection experiments reveal low degree of host specificity, stress increases susceptibility to infection

Trypanoplasma sp. nov.
Nigrelli, R. F.; Pokorny, K. S.; and Ruggieri, G. D., 1975, J. Protozool., v. 22 (3), 43A [Abstract]
leech as apparent vector
Opsanus tau (blood)
Piscicola funduli
all from Long Island Sound

Trypanoplasma borelli
Lom, J.; and Nohynkova, E., 1977, J. Protozool., v. 24 (4), 52A [Abstract]
Trypanoplasma borelli bloodstream phase, surface coat, presence of carbohydrates

Trypanosoma
Capron, A.; et al., 1977, Ann. Immunol., v. 128C (1-2), 541-556
impairment of immune response in parasitic infections characterized by high prevalence of autoantibodies and by immunosuppression, review discussing malaria, trypanosomiasis, trichinosis, and schistosomiasis, with some original material on the last

Trypanosoma
Desowitz, R. S., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 551-596
immunology of African trypanosomes, review

Trypanosoma
Doyle, J. J., 1977, Advances Exper. Med. and Biol., v. 93, 31-63
salivarian trypanosomes, antigenic variation, review: life cycle, variant antigens, antigens other than variant-specified, antigens and protection, spectrum of variant types; effect of cyclical transmission on antigenic variation and its relevance to epidemiology; mechanisms of antigenic variation

Trypanosoma
Greiner, E. C.; et al., 1975, Canad. J. Zool., v. 53 (12), 1762-1787
avian hematozoa, prevalence with reference to distribution by geographic region, by host family, by vertical stratification of nesting sites and by feeding behavior of known vectors: North America north of Mexico
[Checklist includes 388 bird species and contains both published and unpublished records. For records from specific hosts, see entries in Supplement 22, Part 7, Hosts.]

Trypanosoma
Jordan, A. M., 1976, Vet. Parasitol., v. 2 (1), 143-152
Glossina spp. as vectors of trypanosomes, factors affecting infection rates in tsetse flies, possible immune responses, effects of vector on characteristics of strains of trypanosomes, review

Trypanosoma
Kallinikova, V. D., 1977, Molec. and Cell. Biochem., v. 14 (1-3), Feb. 4, 55-58
Kinetoplastida, kinetoplast genetic system, implications for concept of cell differentiation and phylogeny, relationship to phenotype of these flagellates

Trypanosoma
Lom, J., 1973, J. Protozool., v. 20 (4), 537
Trypanosoma and Trypanoplasma from freshwater fishes, cross-infection experiments reveal low degree of host specificity, stress increases susceptibility to infection

Trypanosoma
Mauel, J.; and Behin, R., 1974, Transplant. Rev., v. 19, 121-146
cell-mediated and humoral immunity to protozoan infections (leishmaniasis, malaria, trypanosomiasis), review

Trypanosoma
Risby, E. L., 1976, J. Protozool., v. 23 (2), 20A [Abstract]
Crithidia fasciculata, Blastocrithidia culicis, Trypanosoma, phosphohexose isomerase, comparison of characteristics

Trypanosoma
Simpson, L., 1973, J. Protozool., v. 20 (1), 2-8
salivarian and stercorarian trypanosomes, kinetoplast DNA, structure and function, review

Trypanosoma
Toure, S. M., 1973, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 26 (4), 113a-122a
use of trypanocides in veterinary medicine, review

Trypanosoma
Trigg, P. I.; and Gutteridge, W. E., 1977, Symposia Brit. Soc. Parasitol., v. 15, 57-81
parasitic Protozoa, morphological, biochemical, and physiological changes occurring during life cycles, symposium presentation

Trypanosoma [sp.]
Ashford, R. W.; et al., 1976, J. Wildlife Dis., v. 12 (3), 409-426
Ardeola ralloides
Actophilornis africana
Streptopelia semitorquata
S. decipiens
Turtur tympanistria
Halcyon leucocephala
Merops lafresnayii
Tockus alboterminatus
Riparia riparia
R. paludicola
Hirundo abyssinica
Motacilla alba
Pycnonotus barbatus
Chlorocichla flavicollis
Dryoscopus gambensis
Tchagra minuta
Cercomela familiaris
Cossypha semirufa
Turdus pelios
Turdoides leucopygius
T. rubiginosus
Acrocephalus arundinaceus
A. gracilirostris
Sylvia nisoria
Phylloscopus trochilus
P. collybita
Cisticola erythrops
C. brachyptera
Prinia subflava
Sylvietta brachyura
Platysteira cyanea
Terpsiphone viridis
Nectarinia venusta
Zosterops abyssinica
Z. senegalensis
Serinus mozambicus
S. citrinelloides
Hypochera chalybeata
Amadina fasciata
Pytelia phoenicoptera
Estrilda paludicola
E. astrild
Lagonosticta larvata
L. senegala
Lonchura cucullata
Ploceus luteolus
P. taeniopterus
P. cucullatus
P. rubiginosus
P. ocularis
P. nigricollis
Quelea erythrops
Euplectes hordeaceus
Plocepasser mahali
Passer griseus
P. eminibey
Creatophora cinerea
Dicrurus adsimilis
all from Ethiopia

Trypanosoma [sp.]
Bennett, G. F.; Okia, N. O.; and Cameron, M. F., 1974, J. Wildlife Dis., v. 10 (4), 458-465
survey, avian hematozoa, seasonal prevalence
Ceryle rudis
Ispidina picta
Tympanistria tympanistria
Tchagra australis
Ploceus aurantius
P. cucullatus
P. nigerrimus
Pycnonotus virens
Zosterops senegalensis
(blood of all): all from Uganda

Trypanosoma sp.
Colley, F. C.; et al., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 84-85 [Demonstration]
chickens (peripheral blood): Gombak, Selangor, Malaysia

Trypanosoma sp.
Davies, R. B.; and Clark, G. G., 1974, J. Wildlife Dis., v. 10 (1), 63-65
Cervus canadensis: Red Rock Wildlife Area, New Mexico (captured in Jackson Hole, Wyoming)
Hybomitra laticornis: Gila National Forest, New Mexico

Trypanosom[a] (Megatrypanum) [sp.]
Garnham, P. C. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 386 [Demonstration]
Miopithecus talapoin (blood): Rio Muni

Trypanosoma sp. Indon., illus.
Heywood, P.; Weinman, D.; and Lipman, M., 1974, J. Protozool., v. 21 (2), 232-238
Trypanosoma cyclops maintained in blood agar medium, ultrastructure, structure of pigment bodies, compared with T. cyclops grown in hemoglobin-free medium and a non-pigmented Trypanosoma sp.
Macaca ira: Indonesia

Trypanosoma spp.
Lanham, S. M.; and Scott, C. M., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 274 [Demonstration]
importance of controls in electrophoretic zymograms as shown by pyruvate kinase and adenylate kinase of trypanosomes on thin-layer starch gel

T[rypanosoma] (Trypanozoon) strains
Mehlitz, D., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 86 [Letter]
at least 3 of 4 strains of trypanosomes of the subgenus Trypanozoon isolated from domestic pigs were serum-resistant in the blood incubation infectivity test (possibly or probably T. b[rucei] gambiense, at any rate potentially dangerous to man): Liberia

Trypanosoma [spp.]
Misra, K. K.; and Choudhury, A., 1976, J. Protozool., v. 23 (2), 32A-33A [Abstract]
Bandicota bengalensis bengalensis
B. indica norvegicus

Trypanosoma sp., illus.
Miyata, A.; and Tsukamato, M., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 113-130
Callosciurus juvencus: Palawan Island, the Philippines

Trypanosoma sp.
Morton, J. K.; and Kingston, N., 1976, J. Wildlife Dis., v. 12 (2), 233-236
Cervus canadensis
Odocoileus hemionus
O. virginianus
O. hemionus columbianus
(blood of all): all from Wyoming

Trypanosoma [sp.]
Padmavathi, P.; et al., 1977, Indian Vet. J., v. 54 (12), 1035
sheep (blood): Rajendranagar

Trypanosoma sp., illus.
Peirce, M. A.; and Bevan, B. J., 1977, Vet. Rec., v. 100 (14), 282-283
Psittinus cyanurus (blood): imported to London from Indonesia

Trypanosoma sp., illus.
Ponnampalam, J. T.; and Seow, C. L., 1976, Kajian Vet., v. 8 (2), 83-84
Cygnus olor (blood): National Zoo, Kuala Lumpur, imported from Holland

Trypanosoma sp. Dorney, 1967
Riedel, D., 1975, J. Protozool., v. 22 (1), 8-17
as syn. of Trypanosoma (Herpetosoma) tamiasi sp. n.

Trypanosoma species
Ruitenberg, E. J.; and van Knapen, F., 1977, J. Infect. Dis., v. 136, suppl., S267-S273
application of the enzyme-linked immunosorbent assay to the detection of human and animal helmintic and protozoal infections, advantages of assay for seroepidemiology, discussion of performance of assay in tubes and microplates

Trypanosoma (Herpetosoma) sp., resembling T. (H.) lewisi
Shrivastava, K. K.; and Shrivastava, G. P., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 3-4 [Demonstration]
humans (blood): Village Parsda, India

Trypanosoma (Herpetosoma) sp., resembling T. (H.) lewisi, illus.
Shrivastava, K. K.; and Shrivastava, G. P., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 143-144
humans: India

Trypanosoma (Megatrypanum) [sp.], illus.
Stuht, J. N., 1975, J. Wildlife Dis., v. 11 (2), 256-262
Trypanosoma [sp.], morphology, higher incidence in older deer
Odocoileus virginianus
Cervus canadensis
all from Michigan

Trypanosoma sp., unidentified C-shaped trypanosome
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Rana erythraea: Malayan Peninsula

Trypanosoma sp., similar to T. borelli
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Rana erythraea
Bufo melanostictus
all from Malayan Peninsula

Trypanosoma sp., resembling T. chattoni
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Rana erythraea
R. hosei
Bufo melanostictus
Rana limnocharis
R. macrodon
all from Malayan Peninsula

Trypanosoma [sp.]
Telford, S. R., jr., 1977, Internat. J. Parasitol., v. 7 (4), 299-314
saurian malaria and other haematozoa, distribution and zoogeography, taxonomic problems, incidence by locality and season, implications for vector searches, mixed infections, comparative effectiveness of initial vs. repeated examinations of blood smears for detection: Middle America
Anolis lemurinus: Belize
Corytophanes cristatus: Belize
Thecadactylus rapicaudus: Panama
Gonatodes albogularis: Panama
Anolis capito: Panama
A. limifrons: Panama
A. poecilopus: Panama
A. biporcatus: Panama
A. frenatus: Panama
A. tropidogaster: Panama
Mabuya mabouya: Panama

Trypanosoma sp., lewisi-like, illus.
Wiger, R., 1971, Norwegian J. Zool., v. 19 (1), 83-87
Trypanosoma sp. from Lemmus lemmus, description, incidence, age of host; unable to experimentally infect Microtus oeconomus: More-Romsdal county, Norway

Trypanosoma [sp.]
Wiger, R., 1973, Norwegian J. Zool., v. 21 (4), 325 [Abstract]
Trypanosoma [sp.] and Grahamella [sp.], morphophysiological responses of naturally infected Lemmus lemmus and Clethrionomys glareolus: Norway

Trypanosoma sp., illus.
Wiger, R., 1973, J. Wildlife Dis., v. 9 (2), 125-128
Lemmus lemmus (blood): Norway

Trypanosoma acanthobramae n. sp.
Warsi, A. A.; and Fattohy, Z. I., 1976, Current Sc., Bangalore, v. 45 (23), 838-839 [Letter]
Acanthobrama marmid (blood): River Tigris, Mosul (Iraq)

Trypanosoma acomys
El-On, J.; and Greenblatt, C. L., 1973, J. Protozool., v. 20 (4), 531
Trypanosoma lewisi, T. acomys, growth in vitro at 37°C. in culture media with mammalian cells, rats partially protected against T. lewisi after receiving 6 injections of concentrated culture supernatant fluid

Trypanosoma acomys, illus.
El-On, J.; and Greenblatt, C. L., 1977, Exper. Parasitol., v. 41 (1), 31-42
Trypanosoma lewisi, T. acomys, T. cruzi, method for cultivation with mammalian tissue, results not good with T. equiperdum and T. congolense

Trypanosoma avium
Bennett, G. F.; et al., 1975, J. Wildlife Dis., v. 11 (2), 280-289
hematozoa of Anatidae, prevalence survey
Anas rubripes rubripes
Anas discors
all from Maritime provinces of Canada

Trypanosoma avium
Bennett, G. F.; and Blancou, J., 1974, J. Wildlife Dis., v. 10 (3), 239-240
Acridotheres tristis
Falco newtoni
(in blood of all): all from Madagascar

Trypanosoma avium
Bennett, G. F.; and Herman, C. M., 1976, J. Wildlife Dis., v. 12 (1), 59-65
(blood of all)
Anthropetes collaris: Tanzania
Cyanomitra olivacea: Tanzania
Numida meleagris: Tanzania
Oriolus auratus: Zaire
Hyphanturgis ocularis: Tanzania
Pycnonotus xanthopygus: Tanzania
Turdus olivaceus: Kenya

Trypanosoma avium
Stabler, R. M.; Kitzmiller, N. J.; and Braun, C. E., 1977, J. Wildlife Management, v. 41 (1), 128-130
Columba fasciata (blood): Colorado; Mexico

Trypanosoma avium Danilewsky, 1885
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Coracias garrulus]
[Merops apiaster]
[Upupa epops]
all from north-east Azerbaidzhan

Trypanosoma baigulensis n. sp., illus.
Pandey, K. C.; and Pandey, A. K., 1976, Indian J. Zoot., v. 15 (1), 1974, 15-18
Trypanosoma baigulensis, blood pathology, piscine hosts
Cirrhina reba
Osteobrama cotio
(blood of all): all from Baigul Dam, district Nainital, India

T[rypanosoma] (Herpetosoma) blanchardi
Hommel, M.; and Miltgen, F., 1973, J. Protozool., v. 20 (4), 527
Trypanosoma blanchardi, T. rabinowitschae, adaptation to mice by passage through cultures and then sarcomatous mice, strong cross immunity between T. musculi and mouse-adapted strains; adaptation of T. lewisi to mice using same method was not possible; this new culture system was used to cultivate T. brucei

Trypanosoma blanchardi
Hommel, M.; and Robertson, E., 1976, Experientia, v. 32 (4), 464-466
Trypanosoma blanchardi, in vitro system for study of attachment of trypanosomes to plastic (polystyrene flasks) by means of hemidesmosomes, significance of this type of attachment unknown, reproduces natural stage in life cycle

Trypanosoma bouffardi, illus.
Molyneux, D. H., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 223-228
Trypanosoma bouffardi in Lagonosticta senegala senegala (nat. and exper.), Glossina morsitans submorsitans (exper.), G. tachinoides (exper.), G. palpalis (exper.), morphology, course of infection and distribution in Glossina spp., development abortive and infection not established, avian trypanosomes as possible source of error in determining infection rate of tsetses with pathogenic trypanosomes

Trypanosoma bouffardi
Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 294 [Abstract]
Trypanosoma bouffardi development in Glossina suggests that Foster's T. grayi may have been an avian trypanosome instead, need for awareness of avian trypanosomes in Glossina spp.

Trypanosoma bouffardi, illus.
Molyneux, D. H.; and Robertson, E., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 369-377
Trypanosoma bouffardi bloodstream forms, ultrastructure

Trypanosom[a] brucei
Adam, K. M. G.; and Dipeolu, O. O., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 293 [Abstract]
Trypanosom[a] brucei, T. congolense, cyclical development, comparison of strains

Trypanosoma brucei
Aiyedun, B. A.; and Amodu, A. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 67-71
trypanosomiasis, albino rats (exper.), humans, diagnosis, capillary-tube precipitation test (known as capillary-tube agglutination test), rapid and specific technique that could be used in field survey work

Trypanosoma brucei
Al Chalabi, K.; and Gutteridge, W. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146 [Abstract]
Trypanosoma spp., Crithidia fasciculata, C. oncopelti, thymidine phosphorylase activity, salvage pathways, inhibition of activity by suramin and ethidium

Trypanosoma brucei
Al Chalabi, K.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), xv-xvi [Abstract]
6 spp. of Trypanosomatidae, thymidylate synthetase activity, ranks as potential target for chemotherapeutic attack since it is markedly more sensitive to suramin inhibition than isofunctional mammalian and bacterial enzymes

Trypanosoma brucei
Al Chalabi, K.; and Gutteridge, W. E., 1977, Parasitology, v. 74 (3), 299-312
deoxythymidylate catabolism in some trypanosomatids

Trypanosoma brucei
Alexander, J.; and Phillips, R. S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 273 [Demonstration]
Trypanosoma brucei in mice suppresses growth of cutaneous nodules caused by Leishmania spp., subcurative treatment of T. brucei with Berenil allowed nodule to recommence growth but when trypanosome infection relapsed the lesion again stopped growing, possible mechanism, no evidence of cross-immunity

Trypanosoma brucei subgroup
Allsopp, B. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 270-271 [Abstract]
Trypanosoma brucei subgroup, 4S (surface) antigen, separation of components

Trypanosoma brucei
Anosa, V.; Jennings, F. W.; and Urquhart, G. M., 1977, J. Comp. Path., v. 87 (4), 569-579
Trypanosoma brucei, mice, anemia slightly increased in splenectomized mice compared to intact mice; spleen not essential for red cell destruction

T[rypanosoma] brucei
Barker, D. C.; and Brack, C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 269 [Abstract]
Trypanosoma brucei, T. mega, Crithidia fasciculata, dark-field electron microscopy of kinetoplast DNA

Trypanosoma brucei
Barry, D., 1977, J. Protozool., v. 24 (2), Suppl., 5A [Abstract]
Trypanosoma brucei, loss of variant antigen by parasites transforming in vitro and in the tsetse fly

Trypanosoma brucei
Barry, J. D.; and Vickerman, K., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 271-272 [Abstract]
Trypanosoma brucei in vitro, changes in surface antigens of clone populations during transformation from bloodstream to procyclic stage

Trypanosoma brucei
Barry, J. D.; and Vickerman, K., 1977, Parasitology, v. 75 (2), xxx [Abstract]
Trypanosoma brucei, effects of antibodies on surface, "capping"

Trypanosoma brucei
Barry, J. D.; and Vickerman, K., 1977, J. Protozool., v. 24 (4), 42A [Abstract]
Trypanosoma brucei, short stumpy forms (SSF) of a particular parasitaemic peak are of same variable antigen type as their long slender predecessors, SSF appear more resistant to host antibodies possibly as a result of binding host serum components

Trypanosoma brucei
Boreham, P. F. L.; and Facer, C. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 279 [Abstract]
Trypanosoma brucei, rabbits, marked increases in concentration of fibrinogen degradation products

T[rypanosoma] (T[rypanozoon]) brucei
Boreham, P. F. L.; and Facer, C. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 153 [Abstract]
T[rypanosoma] brucei, rabbits (exper.), changes in plasma fibrinogen concentration during infection, possible relationships to disease pathologic features

Trypanosoma brucei
Boreham, P. F. L.; and Facer, C. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 274 [Abstract]
Trypanosoma brucei, experimental rabbits, changes in blood viscosity

Trypanosoma brucei
Boreham, P. F. L.; and Wright, I. G., 1976, Brit. J. Pharmacol., v. 58 (1), 137-139
hypotension in rabbits infected with Trypanosoma brucei probably result of immune complex formation of trypanosomes with antibody

Trypanosoma brucei
Boreham, P. F. L.; and Wright, I. G., 1976, Parasitology, v. 73 (2), xxxi [Abstract]
Trypanosoma brucei, rabbits, hypotension, possibly caused by trypanosome-antibody complexes and mediated by kallikrein

T[rypanosoma] brucei
Boulton, F. E.; Jenkins, G. C.; and Lloyd, M. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 153-154 [Abstract]
T[rypanosoma] brucei-infected rabbits, changes in clotting mechanism during infections and possible role of disseminated intravascular coagulation in causing hemolysis and anemia; increased production of clotting proteins

Trypanosoma brucei
Bridgen, P. J.; Cross, G. A. M.; and Bridgen, J., 1976, Nature, London (5578), v. 263, 613-614
Trypanosoma brucei, N-terminal amino acid sequences of variant-specific surface antigens

Trypanosoma brucei, illus.
Brown, L. A.; and Losos, G. J., 1977, Research Vet. Sc., v. 23 (2), 196-203
Trypanosoma congolense and T. brucei, comparative pathology in both bled and non-bled albino rats (exper.): parasitemia, packed cell volumes, weight of spleen and lymph, histology of thymus, spleen, lymph nodes, and bone marrow

Trypanosoma brucei
Buengener, W., 1975, Tropenmed. u. Parasitol., v. 26 (3), 281-284
Trypanosoma musculi in inbred mice (exper.), parasite multiplication and course of infection in single infection, after pre-infection with T. congolense, or with added infection of T. brucei

Trypanosoma (Trypanozoon) brucei
Buengener, W.; and Mehlitz, D., 1976, Tropenmed. u. Parasitol., v. 27 (4), 405-410
Trypanosoma brucei, experimentally infected Capra ibex s. hircus (Herzmuskel, Haut, Skelettmuskulatur, Niere, Fett- und Bindegewebe), severe infection resulting in death

Trypanosoma brucei
Burnett, J. K., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 254-255 [Abstract]
Trypanosoma brucei, characterization of kinetoplast DNA

Trypanosoma brucei
Buys, J.; and Ruitenberg, E. J., 1977, Trop. and Geogr. Med., v. 29 (2), 205 [Abstract]
Trypanosoma brucei, T. rhodesiense, comparative application of enzyme-linked immunosorbent assay (ELISA) and immunofluorescence for serodiagnosis of human trypanosomiasis; ELISA represents good alternate method particularly suited for mass screening purposes; cross-reaction only in one person in whom antibodies to Leishmania were detected

Trypanosoma (Trypanozoon) brucei
Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
blood parasites of wild Bovidae, incidence, epizootiological importance
Syncerus caffer
Tragelaphus strepsiceros
(blood of all): all from Ngamiland, Botswana

Trypanosoma brucei
Clarkson, M. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 272 [Abstract]
Trypanosoma brucei and T. congolense in mice, immunoglobulin changes during infection

Trypanosoma brucei
Clarkson, M. J., 1976, Parasitology, v. 73 (2), viii [Abstract]
Trypanosoma brucei, differential serum IgM response in different strains of mice, immunosuppression to sheep red blood cells demonstrated in mice with both high and low IgM concentrations

Trypanosoma brucei
Clarkson, M. J., 1976, Vet. Parasitol., v. 2 (1), 9-29
trypanosomes, immunological problems, review: antigens; host reactions to infection; immunological state of host

Trypanosoma brucei
Clarkson, M. J.; Choudhry, M.; and Cullingham, T. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 2 [Demonstration]
Trypanosoma brucei-infected mice (exper.), changes in immunoglobulins

Trypanosom[a] brucei
Clarkson, M. J.; and McCabe, W. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 12 [Demonstration]
Trypanosoma spp., relative susceptibility of mice to infection by buccal, gastric, or intraperitoneal routes, possible significance of oral transmission under natural conditions

Trypanosoma brucei
Cook, R. M., 1976, Parasitology, v. 73 (2), viii [Abstract]
Trypanosoma brucei, effect of specific antigen-antibody complexes on attachment to mouse peritoneal macrophages

Trypanosoma brucei
Corsini, A. C.; et al., 1977, Clin. and Exper. Immunol., v. 29 (1), 122-131
Trypanosoma brucei, mice, immunodepressing effect of infection can be attributed to clonal exhaustion of B-cell potential caused by undefined blastogenic stimulus from parasites which may operate at least in part by generation of suppressive T cells and macrophages

Trypanosoma brucei
Cross, G. A. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 255-256 [Abstract]
Trypanosoma brucei, T. rhodesiense, development of defined media for growth

Trypanosoma brucei
Cross, G. A. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 261 [Abstract]
Trypanosoma brucei, identification and purification of class of soluble surface proteins, differences in coat proteins isolated from closely related clones, possible significance in antigenic variation

Trypanosoma brucei
Cross, G. A. M., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, Pt. 2), 240-244
Trypanosoma brucei, variant surface antigens, summary of structure and possible functions, assessment of prospects for immunoprophylaxis and improved diagnosis of African trypanosomiasis, workshop report

Trypanosoma brucei
Cross, G. A. M.; and Klein, R. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 266-267 [Abstract]
Trypanosoma brucei, utilization of amino acids and glucose during growth in culture, effects on respiration

Trypanosoma brucei
Cunningham, I.; and Latif, B. M. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 270 [Abstract]
Trypanosoma brucei, T. congolense, antigenic properties during transformation from blood to culture forms

Trypanosoma brucei
Damper, D.; and Patton, C. L., 1976, Biochem. Pharmacol., v. 25 (3), 271-276
Trypanosoma brucei, pentamidine transport, kinetics, specificity (competitive inhibition by other trypanocidal aromatic diamidines), structural specificity in relationship to varying drug affinities for transport site and possible role in drug cross-resistance

Trypanosoma brucei
Damper, D. W.; and Patton, C. L., 1976, J. Protozool., v. 23 (2), 16A [Abstract]
Trypanosoma brucei, pentamidine transport system, structural specificity

Trypanosoma brucei
Dar, F. K.; et al., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 21-29
techniques used to study serotypes of salivarian trypanosomes being circulated by tsetse flies in 4 separate geographical areas, methods used to isolate and serologically type the trypanosomes are described and discussed in detail: Uganda and Kenya, East Africa

Trypanosoma brucei
Duxbury, R. E.; et al., 1973, Isotopes and Radiation Parasitol. III, 179-180
Trypanosoma rhodesiense, mice, rats, cattle, rhesus monkeys; T. congolense, mice, dogs, cattle; T. brucei, cattle: attempted immunization with irradiated trypanosomes, brief communication

Trypanosoma brucei
Eardley, D. D.; and Jayawardena, A. N., 1977, J. Immunol., v. 119 (3), 1029-1033
Trypanosoma brucei, T cells from infected mice can mediate suppression of in vitro primary antibody response, adherent cells are also suppressive, B cells are not suppressive but neither are they normally responsive

Trypanosoma brucei
Eggitt, M. J., 1977, J. Protozool., v. 24 (4), 42A [Abstract]
synthesis of variable and common antigens in rabbit reticulocyte cell-free system directed by Trypanosoma brucei RNA

Trypanosoma brucei
Eggitt, M. J.; Tappenden, L.; and Brown, K. N., 1977, Parasitology, v. 75 (2), 133-141
Trypanosoma brucei, RNA with messenger activity extracted from both blood and culture forms of parasite and translated into reticulocyte cell-free system, system used to investigate synthesis of polypeptides and especially of the variant antigen

Trypanosoma brucei
Elce, B. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 162 [Abstract]
Trypanosoma congolense, T. brucei, T. rhodesiense, development in Glossina morsitans and small laboratory mammals

Trypanosoma brucei
Ellis, D. S.; and Ormerod, W. E., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 276 [Abstract]
occult visceral forms, morphology

Trypanosoma brucei
Emslie, V. W.; and Kershaw, W. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 7 [Demonstration]
mice infected with Trypanosoma brucei, treated with antrypol and then infected with Litomosoides carinii, decreased host resistance to subsequent infection and to relapse from first infection, few visible immunologic changes observed

Trypanosoma brucei
Evans, D. A.; and Brown, R. C., 1972, J. Protozool., v. 19 (4), 686-690
Trypanosoma brucei culture forms, utilization of glucose and proline

Trypanosoma brucei
Evans, D. A.; and Brown, R. C., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 258 [Abstract]
Trypanosoma brucei, inhibition of cyanide-insensitive respiration of both bloodstream and culture forms by m-chlorobenzhydroxamic acid

Trypanosoma (Trypanozoon) brucei
Facer, C. A., 1976, J. Comp. Path., v. 86 (3), 393-408
Trypanosoma brucei, rabbits (exper.), blood hyperviscosity primarily determined by plasma concentration of macroglobulin and fibrinogen, pathology

Trypanosoma brucei
Fairlamb, A. H.; and Bowman, I. B. R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 275 [Demonstration]
Trypanosoma brucei, cell disruption and subcellular fractionation by differential centrifugation, ultrastructural changes in surface coat and pellicular membrane

Trypanosoma brucei
Fairlamb, A. H.; and Bowman, I. B. R., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 268 [Abstract]
Trypanosoma brucei, studies on glycerophosphate oxidase, mechanism of action, suramin a potential inhibitor

Trypanosoma brucei
Fairlamb, A. H.; and Bowman, I. B. R., 1977, Exper. Parasitol., v. 43 (2), 353-361
Trypanosoma brucei, effect of suramin and other trypanocidal compounds on sn-glycerol-3-phosphate oxidase, results suggest this enzyme is one of principal sites of action of suramin in vivo

Trypanosoma brucei
Fairlamb, A. H.; and Bowman, I. B. R., 1977, Internat. J. Biochem., v. 8 (9), 659-668
Trypanosoma brucei, particulate sn-glycerol-3-phosphate oxidase, isolation and characterization

Trypanosoma brucei
Fairlamb, A. H.; and Bowman, I. B. R., 1977, Internat. J. Biochem., v. 8 (9), 669-675
Trypanosoma brucei, particulate sn-glycerol-3-phosphate oxidase, inhibitor studies

Trypanosoma brucei
Fairlamb, A. H.; Opperdoes, F. R.; and Borst, P., 1977, Nature, London (5591), v. 265, 270-271
Trypanosoma brucei, effect of salicylhydroxamic acid + glycerol on motility and pyruvate production, shows how knowledge of trypanosome metabolism has led to new methods for screening drugs

T[rypanosoma] (T[rypanozoon]) brucei
Fearnhead, R. W.; Jenkins, G. C.; and Kawasaki, K., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 155 [Abstract]
T[rypanosoma] brucei-infected rabbits and mice, tissue samples from various organs show trypanosomes with extra cellular membranous coats origin of which is unknown

Trypanosoma brucei
Fregene, A. O.; et al., 1975, J. Parasitol., v. 61 (6), 1070-1073
comparative responses of radioattenuated Trypanosoma brucei and T. congolense in rats: radiation sensitivity with respect to infectivity; immunogenicity

Trypanosom[a] brucei subgroup
Geigy, R.; Kauffmann, M.; and Jenni, L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 284-286 [Abstract]
lion
spotted hyaena
Alcelaphus buselaphus
defassa waterbuck
warthog
all from Serengeti

T[rypanosoma] brucei-complex
Gibson, W., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 386 [Demonstration]
T[rypanosoma] brucei-complex, isoenzyme patterns

Trypanosoma (Trypanozoon) brucei subgroup
Goedbloed, E.; et al., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 31-43
Trypanosoma brucei subgroup, antigenic types of 38 strains isolated from more than 73,000 Glossina spp. from 4 geographically separate areas, intra- and inter-area and intra- and inter-strain comparisons, direct agglutination test used primarily: Uganda and Kenya, East Africa

Trypanosoma brucei
Goldstein, P. D.; and Patton, C. L., 1976, J. Protozool., v. 23 (2), 16A [Abstract]
Trypanosoma brucei, pentamidine transport in relation to antigenic variation and drug resistance

Trypanosoma brucei
Goodwin, L. G.; and Guy, M. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 12 [Demonstration]
Trypanosoma brucei, rabbits, serum and tissue fluid changes caused by infection

T[rypanosoma] brucei
Goodwin, L. G.; and Guy, M. W., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 156 [Abstract]
T[rypanosoma] brucei-infected rabbits, connective tissue pathology, antitrypanosomal antibody penetration of tissue fluid

Trypanosoma brucei
Goodwin, L. G.; and Tierney, E. D., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21 [Demonstration]
trypanosome suspension (Trypanosoma brucei), titration technique on plastic microtest plates (Nunclon) to determine trypanocidal drug activity in body fluids and to detect contribution of humoral immunity to chemotherapeutic activity

Trypanosoma (Trypanozoon) brucei
Goodwin, L. G.; and Tierney, E. D., 1977, Parasitology, v. 74 (1), 33-45
Trypanosoma brucei, T. evansi, bioassay technique for titration of trypanocidal activity in microtest plates using very small samples, use to study activity of plasma and tissue fluid from normal and infected rabbits treated with curative drugs, activity higher in samples from infected rabbits due to participation of immune response

Trypanosoma brucei
Guy, M. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 429 [Demonstration]
Trypanosoma brucei-infected rabbits, changes in total lipid, cholesterol and tryglyceride levels in serum and tissue fluid

Trypanosoma brucei
Harmsen, R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 364-373
Trypanosoma brucei, low survival rate in Glossina pallidipes interpreted as in part result of establishment barrier which is less active in young vs. older flies, peritrophic membrane appears unlikely to be establishment barrier, postulated adjustment period for trypanosomes in flies supported by evidence on temperature sensitivity of parasite enzymes

Trypanosoma brucei
Haston, W. S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146 [Abstract]
Trypanosoma brucei, gradient diver technique used to measure oxygen uptake of culture forms for the purpose of studying metabolism of developmental stages

Trypanosoma brucei
Haston, W., 1975, J. Protozool., v. 22 (3), 52A [Abstract]
Trypanosoma brucei, substrate utilization by transforming midgut forms, decreased glucose oxidation, increased proline oxidation, suggests that trypanosomes have adapted their metabolism to utilize the most available substrate in their mammalian vs. insect hosts

Trypanosoma brucei
Hawking, F., 1971, Tr. Roy. Soc. Trop. Med. and Hyg., v. 65 (5), 672-675
Trypanosoma brucei, attempted culture at human blood temperature (37 C) in presence of tissue cultures, survival and multiplication for 24 hours with diminishing numbers up to 10 days

Trypanosom[a] brucei
Hawking, F., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 262 [Abstract]
trypanocidal substances in human serum found to be macroglobulins

Trypanosoma brucei
Hawking, F., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 517-527
blood incubation infectivity test for differentiating Trypanosoma brucei and T. rhodesiense, modifications of technique and of interpretation

Trypanosoma brucei
Hawking, F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 504-512
Trypanosoma brucei, T. rhodesiense, T. equiperdum, polymorphic strains, sensitivity to human plasma, composition and detailed analysis of typical strains using α_2-macroglobulin test

Trypanosoma brucei
Hawking, F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 513-520
Trypanosoma gambiense, T. rhodesiense, T. brucei, 60 polymorphic strains from East and West Africa, in vivo test of sensitivity or resistance to human plasma, application to infectivity for man

Trypanosoma brucei
Hendow, H. T.; Storey, D. M.; and Kershaw, W. E., 1976, Parasitology, v. 73 (2), xiii [Abstract]
Litomosoides carinii and Trypanosoma brucei, hooded rats, effect of combined infections (in some of which trypanosomiasis was cured with Berenil and in some allowed to run its course)

Trypanosoma brucei
Herbert, W. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 272 [Abstract]
Trypanosoma brucei, growth inhibition of antigenic types between 2 strains, possible factor in control of sequence in which antigenic variants appear during infection

Trypanosom[a] brucei
Herbert, W. J.; and Inglis, M. D., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 268 [Abstract]
Trypanosom[a] brucei, mice, immunization by administration of syngeneic red blood cells which had been exposed to plasma taken from infected animals

Trypanosoma brucei
Herbert, W. J.; and Lumsden, W. H. R., 1976, Exper. Parasitol., v. 40 (3), 427-431
Trypanosoma brucei, rapid method for estimating host's parasitemia by matching microscopic appearance of wet film of infected blood with one of a series of 8 pictures of microscope fields

Trypanosoma brucei subspp.
Herbert, W. J.; Mucklow, M. G.; and Lennox, B., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 4 [Demonstration]
Trypanosoma brucei subspp., primary cause of death in acute murine infection characterized by convulsions is hypoglycemia with associated undetermined pathology

Trypanosoma (Trypanozoon) brucei
Hicks, B. R.; and Fethney, A. P., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 282 [Demonstration]
appraisal of matching technique for estimation of trypanosome parasitaemias

Trypanosoma brucei, illus.
Hirumi, H.; Doyle, J. J.; and Hirumi, K., 1977, Science (4293), v. 196, 992-994
Trypanosoma brucei, long-term propagation of animal-infective bloodstream forms in vitro, organisms retained morphological and biochemical characteristics and displayed variant-antigen on their surfaces

Trypanosoma brucei
Hoare, C. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 421-422 [Letter]
comment on use of term "Trypanosoma brucei-subgroup"

Trypanosoma brucei, illus.
Hogan, J. C., jr.; and Patton, C. L., 1976, J. Protozool., v. 23 (2), 205-215
Trypanosoma brucei bloodstream forms from intact or lethally irradiated rats, freeze-cleaved and thin sectioned preparations, variation in intramembrane components, immunological implications

T[rypanosoma] brucei
Holmes, P. H.; and Jennings, F. W., 1976, Pathophysiol. Parasit. Infect., 199-210
T[rypanosoma] congolense, T. brucei, rabbits infected and subsequently treated with diminazene aceturate, haematological indices, changes in circulating red cell volume and plasma volume, changes in erythropoiesis, red cell survival studies, dramatic effects of treatment on anemia, similar preliminary results with T. congolense in Zebu calves

Trypanosoma brucei
Hommel, M.; and Miltgen, F., 1973, J. Protozool., v. 20 (4), 527
Trypanosoma blanchardi, T. rabinowitschae, adaptation to mice by passage through cultures and then sarcomatous mice, strong cross immunity between T. musculi and mouse-adapted strains; adaptation of T. lewisi to mice using same method was not possible; this new culture system was used to cultivate T. brucei

Trypanosoma brucei
Honigberg, B. M.; et al., 1975, J. Protozool., v. 22 (3), 23A [Abstract]
Trypanosoma brucei, bloodstream, culture, midgut, and proventricular forms, antigenic analysis by quantitative fluorescent antibody method

Trypanosoma brucei
Honigberg, B. M.; et al., 1975, J. Protozool., v. 22 (3), 23A-24A [Abstract]
Trypanosoma brucei, bloodstream and salivary gland forms, antigenic analysis by quantitative fluorescent antibody method

Trypanosoma brucei
Honigberg, B. M.; et al., 1976, Exper. Parasitol., v. 39 (3), 496-522
Trypanosoma brucei brucei, comparative antigenic analysis of different developmental stages (bloodstream, vector, and culture forms) by quantitative fluorescent antibody methods

Trypanosoma brucei
Hudson, K. M.; Byner, C.; and Terry, R. J., 1976, Parasitology, v. 73 (2), ix [Abstract]
Trypanosoma brucei in S42 mice, data which support idea of link between immunodepression and raised levels of IgM characteristic of infection

Trypanosoma brucei
Ikede, B. O.; et al., 1977, Trop. Animal Health and Prod., v. 9 (2), 93-98
Trypanosoma brucei in Equus asini (exper.), clinical, haematological and pathological studies

Trypanosoma brucei
Ikede, B. O.; and Losos, G. J., 1972, Bull. Epizoot. Dis. Africa, v. 20 (3), 221-228
Trypanosoma brucei, dog, case report, diffuse non-suppurative meningo-encephalomyelitis associated with presence of large numbers of trypanosomes in brain, meninges and choroid plexus: Nigeria (imported from Britain)

Trypanosoma brucei
Ikede, B. O.; and Losos, G. J., 1975, J. Comp. Path., v. 85 (1), 23-31
Trypanosoma brucei, sheep (exper.), clinical signs

Trypanosoma brucei
Ikede, B. O.; and Losos, G. J., 1975, J. Comp. Path., v. 85 (1), 33-36
Trypanosoma brucei, sheep (exper.), cerebrospinal fluid changes, presence of trypanosomes in CSF in terminal stages

Trypanosoma brucei
Ikede, B. O.; and Losos, G. J., 1975, J. Comp. Path., v. 85 (1), 37-44
Trypanosoma brucei, sheep (exper.), hypophysial and other endocrine lesions

Trypanosoma brucei
Isoun, M. J.; and Isoun, T. T., 1974, Tropenmed. u. Parasitol., v. 25 (3), 283-287
Trypanosoma vivax, T. brucei, in vitro cultivation, improved survival and multiplication in MEM 199 medium buffered by HEPES in comparison to bicarbonate, importance of strict control of pH

T[rypanosoma] brucei
Itazi, O. K. A.; and Enyaru, J. C., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 263 [Abstract]
T[rypanosoma] brucei, rabbits, proteins excreted in urine are very similar to normal serum proteins

Trypanosom[a] brucei
Itazi, O. K. A.; and Njogu, A. R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 262 [Abstract]
Trypanosom[a] brucei, sugar nucleotides in antigen synthesis

Trypanosoma brucei
Jadin, J. M.; Lambert, P. H.; and Comtesse, A., 1974, J. Protozool., v. 21 (3), 472 [Abstract]
Trypanosoma brucei, mice, acute infection, histopathology

Trypanosoma brucei
James, D. M., 1976, Internat. J. Parasitol., v. 6 (2), 179-182
Trypanosoma brucei, T. congolense, induction of protective immunity in rodents receiving living trypanosomes pretreated with isometamidium chloride or diminazene aceturate tetrahydrate

Trypanosoma brucei
Jayawardena, A. N.; and Waksman, B. H., 1977, Nature (5594), v. 265, 539-541
spleen cells from Trypanosoma brucei-infected normal or nude mice, DNA synthetic response to T-cell and B-cell mitogens and allogeneic cells, ability of these spleen cells to influence responses of normal cells, evidence that thymus-dependent suppressor cells are involved in immunological hyporesponsiveness observed in trypanosomiasis

Trypanosoma brucei
Jenkins, G. C.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 437-438 [Demonstration]
Trypanosoma brucei, rabbits, haematological observations

T[rypanosoma] (T[rypanozoon]) brucei
Jenkins, G. C.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 154 [Abstract]
T[rypanosoma] brucei-infected rabbits, anemia associated with infection due to hemolytic mechanism either of immunologic or microangiopathic nature

T[rypanosoma] (T[rypanozoon]) brucei
Jenkins, G. C.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 154-155 [Abstract]
T[rypanosoma] brucei-infected rabbits, spleen possibly main destructive agent for red cells during infection and for sequestration of platelets

Trypanosoma brucei
Jennings, F. W.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 277 [Abstract]
Trypanosoma brucei, mice, findings of increased catabolism of plasma proteins may be contributory factor in immunosuppressive effects associated with trypanosomiasis

T[rypanosoma] brucei
Jennings, F. W.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 151 [Abstract]
T[rypanosoma] brucei-infected mice, immunosuppression, assessment of lymphoid cell response to antigenic stimulation by incorporation of iododeoxyuridine into lymph nodes; immunosuppression apparently specifically associated with presence of living trypanosomes as immunity partially restored if sensitization occurred same day as drug treatment

Trypanosoma brucei
Jennings, F. W.; et al., 1974, Research Vet. Sc., v. 16 (1), 70-76
Trypanosoma brucei, rats, mice, description of anemia, results indicative of hemolytic origin

Trypanosoma brucei
Jennings, F. W.; Whitelaw, D. D.; and Urquhart, G. M., 1977, Parasitology, v. 75 (2), 143-153
Trypanosoma brucei, mice, relationship between duration of infection and efficacy of chemotherapy, discovery of type of relapsing infection which in some respects resembles drug resistance but is apparently due to another cause since infections were always completely susceptible to chemotherapy administered at early stage of infection

T[rypanosoma] brucei
Jennings, F. W.; Whitelaw, D. D.; and Urquhart, G. M., 1977, Vet. Rec., v. 100 (17), 367
T[rypanosoma] brucei, mice (exper.), diminazene aceturate, relapse when administered later than 14 days post-infection, results suggest that trypanosomes are in insusceptible stage of development or in an inaccessible body location, not drug resistance

Trypanosoma brucei
Johnson, J. G.; and Cross, G. A. M., 1977, J. Protozool., v. 24 (4), 587-591
Trypanosoma brucei, carbohydrate composition of variant-specific surface antigen glycoproteins

Trypanosoma brucei
Klein, R. A.; and Cross, G. A. M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 267 [Abstract]
Trypanosoma brucei, culture form S42, L-threonine significant and possibly preferred source of fatty acid carbon units

Trypanosoma brucei
Klein, R. A.; and Linstead, D. J., 1976, Biochem. Soc. Tr., v. 4 (1), 48-50
Trypanosoma brucei, culture forms, threonine as preferred source of 2-carbon units for lipid synthesis

Trypanosoma brucei
Klein, R. A.; and Miller, P. G. G., 1977, Parasitology, v. 75 (2), xxvii [Abstract]
Trypanosoma brucei, aspects of intermediary metabolism, acetyl CoA production and utilization

Trypanosoma brucei
Kushimo, J. B., 1976, Immunology, v. 30 (5), 635-639
IgM and IgG antibody responses to complexes of denatured Trypanosoma brucei DNA and methylated bovine serum albumin, compared to exclusive IgM response to calf thymus DNA-MBSA complexes, rabbits

Trypanosoma brucei group, illus.
Lamy, L. H., 1971, Bull. Inst. Pasteur, Paris, v. 69 (4), 401-420
Leishmania donovani, Trypanosoma cruzi, T. brucei group, comparisons of life cycle, morphology, reproduction and development, metabolism, extensive review

Trypanosoma brucei, illus.
Langreth, S. G.; and Balber, A. E., 1975, J. Protozool., v. 22 (1), 40-53
Trypanosoma brucei, long slender and short stumpy bloodstream forms, culture forms, use of ferritin to study uptake and intracellular movement of protein, and of acid phosphatase to localize digestive activity cytochemically

Trypanosoma brucei
Le Ray, D.; van Meirvenne, N.; and Jadin, J. B., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 273-274 [Abstract]
Trypanosoma brucei, bloodstream and culture forms, common and variable antigens, immunoelectrophoretic characterization

Trypanosoma brucei
Linstead, D. J.; Klein, R. A.; and Cross, G. A. M., 1977, J. Gen. Microbiol., v. 101 (2), 243-251
Trypanosoma brucei, T. brucei brucei, threonine catabolism, inhibition of threonine dehydrogenase by wide range of agents

T[rypanosoma] brucei
Linstead, D. J.; Klein, R. A.; and Eldridge, M. V., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 267-268 [Abstract]
Trypanosoma mega, T. brucei, Crithidia fasciculata, presence of 3 enzymes capable of carbon dioxide fixation in culture as source of carbon for metabolism

Trypanosoma brucei
Lipschik, G.; et al., 1976, J. Protozool., v. 23 (2), 30A [Abstract]
trypanosomatids, polyamines and Mg^{++} content

Trypanosoma brucei
Luckins, A. G., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 337-344
Trypanosoma vivax, T. brucei, T. congolense-infected Tragelaphus scriptus, serum immunoglobulin levels and relative proportions of serum proteins, compared with Zebu cattle

Trypanosoma brucei
Luckins, A. G.; and Mehlitz, D., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 479-480
Trypanosoma brucei, T. vivax, T. congolense, cattle (exper.), single or mixed infections, serum immunoglobulin levels

Trypanosoma (Trypanozoon) brucei Plimmer & Bradford, 1899
Lumsden, W. H. R.; and Herbert, W. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 205-208
Trypanosoma brucei, pedigrees and inter-relationships of antigenic types

Trypanosoma brucei
Lumsden, W. H. R.; Kimber, C. D.; and Strange, M., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 421-424
Trypanosoma brucei, detection of low parasitemia in mice using a miniature anion-exchanger/centrifugation technique

Trypanosoma brucei
MacKenzie, A. R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 269 [Abstract]
Trypanosoma brucei, rabbits, anti-liver and anti-Wassermann response can occur in young animals not possessing the antibodies before infection, but production of Forssman (heterophile) antibody appears to require presence of antibody at time of infection

Trypanosoma brucei
MacKenzie, A. R.; Boreham, P. F. L.; and Facer, C. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 268 [Abstract]
Trypanosoma congolense, T. brucei, cattle, anti-liver, anti-Wassermann, and anti-fibrinogen antibodies, human serum from sleeping sickness patients also showed raised levels of these autoantibodies, trypanosome-infected lions and hyaenas did not

Trypanosoma brucei
Mansfield, J. M.; Craig, S. A.; and Stelzer, G. T., 1976, Infect. and Immun., v. 14 (4), 976-981
Trypanosoma brucei, T. congolense, mitogenic effects of trypanosome extracts in vitro for lymphocytes from normal rabbits, possible relationship to immunological dysfunctions occurring in chronic African trypanosomiasis

Trypanosoma (Trypanozoon) brucei
Mehlitz, D., 1975, Tropenmed. u. Parasitol., v. 26 (3), 265-275
Trypanosoma spp. of Nannomonas, Duttonella, and Trypanozoon subgenera, comparison of indirect fluorescent antibody test and complement fixation test as means of differentiating subgenera by serology and persistence of antibodies

Trypanosom[a] brucei
van Meirvenne, N.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 274-275 [Abstract]
Trypanosom[a] brucei, bloodstream and culture forms, common and variable antigens, immunogenic properties studied by trypanolysis (combined with neutralization and absorption) and immunoelectrophoretic analysis

Trypanosoma brucei
Miller, P. G. G.; Linstead, D. J.; and Klein, R. A., 1976, Parasitology, v. 73 (2), xvi [Abstract]
5 spp. of trypanosomatids, enzymes of acetyl CoA metabolism, possible routes for synthesis and utilization of acetyl CoA

Trypanosoma brucei
Minoprio, J. L., 1972, Prensa Med. Argent., v. 59 (8), 293-308
human Chagas disease, pathologic findings, comparison with characteristics of other Trypanosoma spp. and Leishmania spp., extensive review

Trypanosoma (Trypanozoon) brucei-subgroup
Moloo, S. K.; and Kutuza, S. B., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 403-409
Trypanosoma brucei-subgroup isolated from Glossina swynnertoni, indicative of vector role of local tsetse flies in endemic areas of Tanzania

Trypanosoma brucei
Moloo, S. K.; Losos, G. J.; and Kutuza, S. B., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 287 [Abstract]
Trypanosoma brucei, transmission to cats and dogs by feeding on infected goats, suggested that oral mode of transmission might be operative in areas where carnivores are frequently found infected

Trypanosoma brucei
Moloo, S. K.; Losos, G. J.; and Kutuza, S. B., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 331-334
Trypanosoma brucei, successful transmission to cats and dogs by feeding on infected goats, results suggest that oral mode of transmission might be operative in areas where wild carnivores have high rate of trypanosome infection

Trypanosoma brucei
Moulton, J. E.; and Sollod, A. E., 1976, Am. J. Vet. Research, v. 37 (7), 791-802
Trypanosoma brucei, calves (exper.), clinical changes, parasitemia, antibody titration (indirect fluorescent antibody technique), IgG and IgM, histopathology

Trypanosoma brucei
Murray, M.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 276-277 [Abstract]
Trypanosoma brucei, pathology in rats: progressive alteration in immunological apparatus leading to immunosuppressed state to other antigens; anemia; specific organ damage and failure

Trypanosoma brucei, illus.
Murray, M.; et al., 1974, Research Vet. Sc., v. 16 (1), 77-84
Trypanosoma brucei, 3 aspects of pathology in rats: progressive alteration in immunological apparatus of lymph nodes, spleen, and thymus, increase in activity of mononuclear phagocytic system; haemopoietic system changes, haemolytic anemia; specific organ damage (heart most markedly affected)

Trypanosoma brucei
Murray, M.; Lambert, P. H.; and Morrison, W. I., 1975, Medecine et Malad. Infect., v. 5 (12), special no., 638-641
Trypanosoma brucei-infected mice develop proliferative immune complex glomerulonephritis

Trypanosoma brucei
Murray, M.; Murray, P. K.; and McIntyre, W. I. M., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 325-326
African trypanosomiasis, light microscopic examination of buffy coat zone of a microhaematocrit capillary tube expressed onto a slide found to be more reliable than standard methods for diagnosing infections in cattle; method also allowed differentiation of species

Trypanosoma brucei
Murray, M.; and Urquhart, G. M., 1977, Advances Exper. Med. and Biol., v. 93, 209-241
African trypanosomiasis, immunoprophylaxis, review: cattle and sheep, laboratory animals; chemotherapeutic agents and immunity, nature of protective antigen, effector mechanisms in protection, manipulation of host resistance

Trypanosom[a] brucei
Murray, P. K.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 267 [Abstract]
Trypanosom[a] brucei, mice, inability to produce anti-sheep erythrocyte IgM, results suggest immunological defect induced by trypanosome infection may be associated with B cells

Trypanosoma brucei
Murray, P. K.; et al., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 133-150
Trypanosoma brucei in rodents, pathology of immune system, role of mononuclear phagocytic system in immunosuppression, humoral and T cell responsiveness, effect of infection on Nippostrongylus brasiliensis infection, recovery of immune response after trypanocidal therapy, review

Trypanosoma brucei
Musoke, A. J.; and Barbet, A. F., 1977, Nature, London (5636), v. 270, 438-440
Trypanosoma brucei, direct activation of human complement via classical pathway by isolated variant-specific surface antigen of parasite, possible role in pathogenesis

Trypanosom[a] brucei subgroup
Mwambu, P. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 286-287 [Abstract]
Trypanosom[a] rhodesiense, importance of cattle as natural reservoir hosts; T. brucei subgroup isolated from cattle, resistance to Ethidium, sensitivity to Berenil: Tanzania; Uganda

Trypanosoma brucei subgroups
Mwambu, P. M.; and Mayende, J. S. P., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 276-277 [Abstract]
Trypanosoma brucei subgroup isolates from cattle, game animals and tsetse-flies, resistance to ethidium

Trypanosoma brucei sspp.
Newton, B. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146-147 [Abstract]
Trypanosoma brucei sspp., amplification of kinetoplast DNA probably occurs in culture forms, much larger amounts of DNA in culture forms than in bloodstream forms; no significant differences found in T. lewisi forms

Trypanosoma brucei
Njogu, A. R.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 147-148 [Abstract]
Trypanosoma brucei subgroup, evidence that 4S (surface) antigens located on outer surface of cell membrane

Trypanosoma brucei
Opperdoes, F. R., 1977, Trop. and Geogr. Med., v. 29 (3), 315 [Abstract]
Trypanosoma brucei, glycerol-3-phosphate oxidase located in mitochondria and not in microbodies

Trypanosoma brucei
Opperdoes, F. R.; et al., 1977, European J. Biochem., v. 76 (1), 29-39
Trypanosoma brucei bloodstream form, subcellular fractionation, localization of glycerol-3-phosphate oxidase in mitochondrion and particulate NAD$^+$-linked glycerol-3-phosphate dehydrogenase in microbodies, supplementary studies with Crithidia luciliae

Trypanosoma brucei
Opperdoes, F. R.; and Borst, P., 1977, FEBS Letters, v. 80 (2), 360-364
Trypanosoma brucei, localization of 9 glycolytic enzymes in microbody-like organelle (the glycosome)

Trypanosoma brucei
Opperdoes, F. R.; Borst, P.; and Fonck, K., 1976, FEBS Letters, v. 62 (2), 169-172
Trypanosoma brucei, inhibition of glycerol-3-phosphate oxidase by salicylhydroxamic acid but motility and ATP production not drastically affected nor course of infection in rats, evidence for deficiencies in present ideas on trypanosome glycolysis, implications for chemotherapy

Trypanosoma brucei
Opperdoes, F. R.; Borst, P.; and Spits, H., 1977, European J. Biochem., v. 76 (1), 21-28
Trypanosoma brucei bloodstream form, screening for presence of enzymes that could be used as specific organelle markers in cell fractionation studies

Trypanosoma brucei
Ormerod, W. E., 1976, Science, v. 191 (4229), 815-821
possible ecological effects of trypanosomiasis control, increased cattle production leading to overgrazing and possibly to climatic change and drought: Africa

Trypanosoma brucei
Ormerod, W. E.; and Venkatesan, S., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 161 [Abstract]
Trypanosoma brucei, working hypothesis on the life cycle in mammalian hosts

Trypanosoma (Trypanozoon) brucei, illus.
Otieno, L. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (6), 856-869
Trypanosoma brucei, effect of irradiation or cyclophosphamide on course of infection in heat-stressed mice, reduced pathogenic effects in spite of elevated parasitemia, tempting to suggest that direct effect of temperature on trypanosomes was responsible for lowered pathogenicity, many bizarre morphological forms

Trypanosoma (Trypanozoon) brucei
Otieno, L. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (6), 886-887 [Letter]
Trypanosoma brucei in haemolymph of young Glossina morsitans (exper.)

Trypanosoma (Trypanozoon) brucei
Otieno, L. H.; Darji, N.; and Onyango, P., 1976, Acta Trop., v. 33 (2), 143-150
Trypanosoma brucei, development to infective stage in Glossina morsitans following inoculation of bloodstream forms into haemocoel, observations suggest possibility of cyclical transmission other than by the classically prescribed route of migration in tsetses; unusual modes of multiplication observed in haemocoel trypanosomes

Trypanosoma brucei
Paris, J.; Wilson, A. J.; and Gray, A. R., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 45-51
Trypanosoma brucei, 11 stabilates (prepared from isolates collected from cattle and tsetse flies in three areas of East Africa), serologic comparison of predominant variant antigens by direct agglutination tests, more sensitive in detection of antigenic relationships than comparisons founded on basic strain antigens

Trypanosoma brucei
Perie, N. M.; Tinnemans-Anggawidjaja, T.; and Zwart, D., 1975, Tropenmed. und Parasitol., v. 26 (4), 399-404
Trypanosoma spp. in experimentally infected domestic animals, sandwich immunofluorescent complement fixation test compared with indirect fluorescent antibody test for use in detecting binding of complement to antigen-antibody complex

Trypanosoma brucei
Politzar, H., 1974, Tropenmed. u. Parasitol., v. 25 (1), 22-27
Trypanosoma vivax alone or with T. brucei and T. congolense, cattle (exper.), homologous and heterologous antibody responses determined by indirect fluorescent antibody test

Trypanosoma brucei, illus.
Poltera, A. A.; Cox, J. N.; and Owor, R., 1977, East African Med. J., v. 54 (9), 497-499
human African trypanosomiasis, cardiac valvulitis observed in man with proven infection, possible relationship of trypanosomiasis with cardiomyopathies of unknown origins; Trypanosoma brucei-infected mice (exper.) also had valvulitis with parasites present in the lesions

Trypanosoma brucei
Raether, W.; Loewe, H.; and Seidenath, H., 1974, Tropenmed. u. Parasitol., v. 25 (3), 273-282
Trypanosoma brucei, mice, testing for trypanocidal effect of two cleavage products of Berenil and the possibility of their in vivo recoupling if administered in combination, compared with Berenil

T[rypanosoma] brucei
Raether, W.; and Seidenath, H., 1974, Tropenmed. u. Parasitol., v. 25 (1), 28-41
T[rypanosoma] rhodesiense, T[rypanosoma] brucei, influence of different physiological solutions and a blood substitute on motility and infectivity after deep-freezing in liquid nitrogen

Trypanosoma brucei
Rafael, A. G., 1975, An. Serv. Vet., Mocambique (20-21), 1972-1973, 331-350
Trypanosoma spp., pathology, economic importance, treatment, vector control, extensive review: Africa

Trypanosoma brucei
Reisner, A. H.; and Westwood, N. H., 1977, Austral. J. Exper. Biol. and Med. Sc., v. 55 (1), 19-25
Trypanosoma brucei and Paramecium aurelia soluble surface proteins compared, considerable degree of similarity, suggests that Paramecium aurelia system can serve as model for phenomenon of phase transformation (relapse syndrome) which occurs in trypanosomiasis

Trypanosoma brucei
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma (Trypanozoon) brucei
Rickman, L. R.; and Robson, J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 166-167 [Letter]
Trypanosoma brucei, observations on identification of strains isolated from non-human hosts

Trypanosoma brucei
Ridley, D. S.; and Hedge, E. C., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (6), 522-525
microfilariae of various spp., immunofluorescent reactions involving sheath, cuticle, and cytoplasm, relevance to immuno-evasive mechanisms: (1) microfilariae failed to adsorb non-specific immunoglobulins in contrast to other helminth larvae and non-blood protozoa; (2) sheath of Wuchereria bancrofti and Loa loa adsorbed specific A and B blood group antigens; (3) low titer reaction between microfilarial cytoplasm (L. loa and W. bancrofti) and host serum

Trypanosoma (Trypanozoon) brucei group
Riordan, K., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 11-20
Trypanosoma spp., survey of long-term variations in infection rates of Glossina spp. vectors found along major trade cattle route in southern Nigeria

Trypanosoma brucei
Risby, E. L., 1976, J. Protozool., v. 23 (2), 27A [Abstract]
Trypanosoma brucei, T. b. gambiense, T. b. rhodesiense, trypanocidal effects of methylglyoxal bis (guanylhydrazone) derivatives

Trypanosoma brucei
Roberts, C. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 275 [Abstract]
trypanosomiasis in ruminants, alterations in lipid metabolism

Trypanosoma brucei
Roberts, C. J.; and Gray, A. R., 1973, Trop. Animal Health and Prod., v. 5 (4), 220-233
Trypanosoma vivax, T. congolense, T. brucei, N'dama, Muturu, and Zebu cattle (exper.), comparison of host resistance, pathology; N'dama cattle demonstrated more resistance than other breeds

Trypanosoma brucei
Ruitenberg, E. J.; and Buys, J., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 31-36
Trypanosoma brucei, enzyme-linked immunosorbent assay (ELISA) for serodiagnosis of human African sleeping sickness, comparison tests with immunofluorescence technique showed good results in rabbits (exper.) and serum from infected humans, cross-reactions only in person with Leishmania antibodies, possible application to epidemiologic surveys

Trypanosoma brucei
Schindler, R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 290 [Abstract]
Trypanosoma spp., cattle, attempted serologic differential diagnosis gave equivocal results

Trypanosoma brucei
Sless, F.; et al., 1975, J. Protozool., v. 22 (3), 56A-57A [Abstract]
Trypanosoma brucei, mice treated with cyclophosphamide 2 days after infection developed high non-relapsing parasitaemia, mice treated 3 days before infection developed significantly lower parasitaemia than controls apparently due to increase in delayed hypersensitivity

Trypanosoma brucei
Slots, J. M. M.; et al., 1977, Exper. Parasitol., v. 43 (1), 211-219
Trypanosoma brucei, Trypanosoma vivax, antigen-antibody complexes as cause of platelet serotonin release in vitro and in vivo

T[rypanosoma] brucei
Smith, D. S.; Njogu, A. R.; and Cayer, M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 162-163 [Abstract]
T[rypanosoma] brucei blood stream forms, freeze-fracture and ultrastructural observations, flagellar attachment, surface cell membrane

Trypanosoma brucei
Soltys, M. A.; Thompson, S. M. R.; and Woo, P. T. K., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 399-402
Trypanosoma brucei, rats, guinea pigs, exper. transmission through skin and intact mucous membranes of conjunctiva, mouth, rectum, vagina; T. congolense, exper. transmission through same routes all failed

Trypanosoma brucei
Soothill, J. F.; Smith, M. D.; and Morgan, A. G., 1975, Symposia Brit. Soc. Parasitol., v. 13, 59-68
association of parasites with nephrotic syndrome, genetically and environmentally determined host variation may be the immunodeficiency underlying proneness to chronic soluble complex disease, extensive review with emphasis on Schistosoma spp., Plasmodium malariae, and some preliminary experiments with Trypanosoma brucei in mice

Trypanosoma brucei
Ssenyonga, G. S. Z., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 272 [Abstract]
Trypanosoma brucei and T. congolense in rodents, distribution in tissues and body fluids

Trypanosoma brucei
Steiger, R. F., 1976, J. Protozool., v. 23 (2), 33A-34A [Abstract]
Trypanosoma brucei, new culture medium

Trypanosoma brucei
Steiger, R.; and Jenni, L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 293-294 [Abstract]
Trypanosoma brucei-infected salivary glands of Glossina morsitans, tracer experiments, transport, uptake, and deposition of labeled amino acids

Trypanosoma brucei
Stevens, D. R.; and Moulton, J. E., 1977, J. Comp. Path., v. 87 (1), 109-118
Trypanosoma equiperdum and T. brucei in Peromyscus maniculatus (exper.), meningoencephalitis, model for human trypanosomal encephalitis

Trypanosoma brucei
Taylor, D. W.; and Cross, G. A. M., 1976, Parasitology, v. 73 (2), vii-viii [Abstract]
Trypanosoma brucei, development of in vitro system using fully defined medium HX 28 in order to facilitate studies into control of synthesis of variant-specific antigens of surface coat of bloodstream forms

Trypanosoma brucei
Taylor, D. W.; and Cross, G. A. M., 1977, Parasitology, v. 74 (1), 47-60
Trypanosoma brucei in vitro, variant-specific surface antigen, isolation, partial characterization, incorporation of [^{35}S]L-methionine, rate of synthesis

Trypanosoma brucei
Tetley, L.; and Vickerman, K., 1977, J. Protozool., v. 24 (4), 45A [Abstract]
Trypanosoma brucei, transformation from pleomorphic bloodstream forms to procyclic culture forms, freeze-fracture studies

Trypanosoma brucei
Toure, S. M., 1973, Bull. Epizoot. Dis. Africa, v. 21 (1), 1-3
Trypanosoma vivax, exper. goats and cattle; T. brucei, exper. cattle, intravenous Isometamidium injections, drug efficacy

Trypanosoma brucei
Toure, S. M., 1973, Medecine Afrique Noire, v. 20 (11), 855-862
trypanocides of veterinary interest and their indications for use, review

Trypanosoma brucei
Toure, S. M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 30 (1), 1-10
trypanosomiasis, domestic animals, diagnostic methods, review

Trypanosoma brucei
Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 1977, 1-15
17. Seminar on Trypanosomiasis (biochemistry and relevance to control; relevance of laboratory studies to tsetse control; field diagnosis; problems of land use and tsetse control)

Trypanosoma brucei subgroup
Trigg, P. I.; and Gutteridge, W. E., 1977, Symposia Brit. Soc. Parasitol., v. 15, 57-81
parasitic Protozoa, morphological, biochemical, and physiological changes occurring during life cycles, symposium presentation

Trypanosoma brucei
Urquhart, G. M.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 528-535
Trypanosoma brucei-infected rats, failure of humoral and immediate-type responses to superimposed Nippostrongylus brasiliensis infection; T. brucei-infected mice, cell-mediated immunity as measured by oxazolone sensitization still occurred to significant extent although less than in uninfected mice

Trypanosoma brucei
Vasquez, R.; Cattan, A.; and Herbert, W. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 436 [Demonstration]
Trypanosoma brucei, production of antibodies cytophilic for macrophages in infected mice

Trypanosoma brucei
Venkatesan, S.; and Bird, R. G., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 276 [Abstract]
heterogeneity of "Type I granules"

Trypanosoma brucei
Venkatesan, S.; and Ormerod, W. E., 1973, J. Protozool., v. 20 (4), 529
Trypanosoma brucei, long-thin vs. short-stumpy forms, differences in lipids and in lysosomal enzymes, short-stumpy considered to be degenerate form as result of action of hydrolytic enzymes released at lyso-phagosomal junction, thus disease remission occurs as result of action of lysosomal enzymes rather than antibody activity

Trypanosoma brucei
Voorheis, H. P., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 256-257 [Abstract]
Trypanosoma brucei, active site of N_1 amino acid transport carrier

Trypanosoma brucei
Voorheis, H. P., 1977, Biochem. J., v. 164 (1), 15-25
Trypanosoma brucei, changes in kinetic behaviour of threonine transport elicited by variation in hydrogen ion concentration

Trypanosom[a] brucei
Walker, P. J.; and Opiyo, E. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 281-282 [Abstract]
Trypanosom[a] brucei, T. rhodesiense, mice, effect of parasite burden on curative dose of trypanocides, possible source of variation in results of in vivo drug tests

Trypanosoma brucei
Wallace, L. E.; White, R. G.; and Herbert, W. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 279 [Demonstration]
Trypanosoma brucei in Gallus domesticus (exper.), massive segregation of B-cells in germinal centres of spleen

Trypanosoma brucei
Wiesenhuetter, E., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 60-66
trypanosomiasis, Zebu cattle, diagnosis, comparison of indirect fluorescent antibody test with microscopic examination, IFA proved to be reliable and rapid diagnostic aid useful for epizootiological studies: Tanzania

Trypanosom[a] brucei
Wilson, A. J.; Dar, F. K.; and Paris, J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 287-288 [Abstract]
trypanosomiasis in a herd of cattle maintained under chemotherapeutic cover in area of high tsetse fly challenge: fly infection rates; comparison of diagnostic methods; differential infection rates with 3 Trypanosom[a] spp.; frequency of Berenil requirement; body weight gains of calves

Trypanosoma brucei
Winkelmann, E.; et al., 1977, Arzneimittel-Forsch., v. 27, (12), 2251-2263
Entamoeba histolytica, Trichomonas foetus, Trypanosoma spp., in golden hamsters and mice (both exper.), laboratory trials with more than 135 new 2-methyl-5-nitroimidazoles substituted in the 1-position and 1-methyl-5-nitroimidazoles substituted in the 2-position, comparison of activity with that of metronidazole and tinidazole

Trypanosoma brucei
Woo, P. T. K.; and Rogers, D. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 319-326
Trypanosoma spp., statistical study of sensitivity of haematocrit centrifuge technique for detecting trypanosomes in blood, evaluation of relationship of probability of detection of trypanosome to trypanosome density

Trypanosoma brucei
Woo, P.; and Soltys, M. A., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 465-469
Trypanosoma brucei, T. rhodesiense, effect of suramin on blood and tissue forms

Trypanosoma brucei
Yesufu, H. M.; and Mshelbwala, A. S., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 307-312
prevalence of trypanosome species in cattle and tsetse flies along a trade cattle route
Zebu cattle
Glossina morsitans
all from southwestern Nigeria

Trypanosoma brucei
Zahalsky, A. C.; and Weinberg, R. L., 1976, J. Parasitol., v. 62 (1), 15-19
Trypanosoma brucei, inbred rats, humoral response to monomorphic strain during infection, during Berenil cure, after cure, and after rechallenge following drug-induced immunity; contribution of Berenil prophylaxis during refractory period, presence or absence of trypanosome-agglutinating antibodies, class of protective antibodies formed

Trypanosoma brucei brucei
Afchain, D.; et al., 1976, Path. Biol., v. 24 (9), 615-617
Trypanosoma brucei brucei, pure variant specific antigen obtained by immunochemical purification by affinity chromatography

Trypanosoma brucei brucei
Bagster, I. A.; and Parr, C. W., 1973, Nature (5415), v. 244, 364-366
trypanosome identification by electrophoresis of soluble enzymes

Trypanosoma (Trypanozoon) brucei brucei
Baker, J. R.; and McConnell, E., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 153-154 [Letter]
Trypanosoma brucei ssp., response of 2 strains to blood incubation infectivity test, Gambela IV from Glossina tachinoides identified as T. b. brucei, Gambela V from Tragelaphus scriptus identified as T. b. rhodesiense, however Gambela V was not infective to a human volunteer: near village of Pinybago on Gilo River, Illubabor Province, Ethiopia

Trypanosoma brucei brucei
Baker, J. R.; and Taylor, A. E. R., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 471-485
Trypanosoma brucei brucei- and T. brucei rhodesiense-infected Pan troglodytes (exper.), course of infection, serologic relationships between trypanosome species and strains, blood changes, cerebrospinal fluid changes, post-mortem observations

Trypanosoma b. brucei
Barry, D., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 273 [Demonstration]
Trypanosoma b. brucei, surface antigenic changes during transformation in culture, progressive loss of surface coat and progressive acquisition of antigenically universal culture form surface

Trypanosoma brucei brucei
Boehringer, S.; and Hecker, H., 1974, J. Protozool., v. 21 (5), 694-698
Trypanosoma brucei brucei, T. b. rhodesiense, pleomorphic strains, quantitative differences in organelle composition during transformation from slender to stumpy blood forms, compared with monomorphic T. b. brucei

Trypanosoma brucei brucei, illus.
Boehringer, S.; and Hecker, H., 1975, J. Protozool., v. 22 (4), 463-467
Trypanosoma brucei brucei developmental stages in Glossina morsitans, quantitative ultrastructure, morphometric analysis, compared with previously published results on bloodstream forms, results discussed with reference to changes in structure and in oxidative metabolism

Trypanosoma brucei brucei
Clarkson, A. B., jr.; and Brohn, F. H., 1976, Science (4261), v. 194, 204-206
Trypanosoma brucei brucei, T. brucei rhodesiense, mice, salicyl hydroxamic acid + glycerol block glucose catabolism of the parasite and it is rapidly destroyed but parasitemia recurs after treatment, with modification may be new approach to trypanosome chemotherapy

Trypanosoma brucei brucei
Cunningham, I., 1977, J. Protozool., v. 24 (2), 325-329
new culture medium for maintenance of Glossina morsitans morsitans tissues and growth of trypanosomatids, medium based on amino-acid composition of tsetse hemolymph and contains fetal bovine serum

Trypanosoma brucei brucei, illus.
Cunningham, I.; and Honigberg, B. M., 1977, Science (4310), v. 197, 1279-1282
Trypanosoma brucei brucei, reacquisition of infectivity for mice in cultures grown in presence of tsetse fly salivary gland explants in medium based on amino acid composition of Glossina hemolymph and containing fetal bovine serum

Trypanosoma brucei brucei
Damper, D.; and Patton, C. L., 1976, J. Protozool., v. 23 (2), 349-356
Trypanosoma brucei brucei, T. b. rhodesiense, pentamidine transport and sensitivity, concluded that reduced drug uptake is primary mechanism of pentamidine resistance in trypanosomes

Trypanosoma brucei brucei
Evans, D. A.; Brightman, C. J.; and Holland, M. F., 1977, Lancet, London (8041), v. 2, 769 [Letter]
Trypanosoma brucei brucei and T. b. rhodesiense infections in mice (exper.) successfully treated with salicylhydroxamic acid if combined with glycerol and administered before patent infection established; T. vivax infections of different durations all successfully cured

Trypanosoma brucei brucei
Evans, D. A.; and Brown, R. C., 1973, J. Protozool., v. 20 (1), 157-160
Trypanosoma brucei bloodstream and culture forms, inhibition of cyanide-insensitive respiration by m-chlorobenzhydroxamic acid, cyanide-sensitive respiration of culture forms not affected, possible importance as trypanocidal drug, implications for mechanism of terminal respiration

T[rypanosoma] b. brucei
Freeman, J. C.; Hudson, K. M.; and Byner, C., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 149 [Abstract]
trypanosome-infected mice, immunosuppression with reduction of IgG producing cells, variations in response with administration of trypanocides prior and subsequent to antigen administration

Trypanosoma brucei brucei, illus.
Fruit, J.; et al., 1977, Parasitology, v. 74 (2), 185-190
Trypanosoma brucei brucei, antigenic analysis of variant-specific component, localization on surface coat using labelled specific antibodies

Trypanosom[a] (Trypanozoon) brucei brucei
Godfrey, D. G.; and Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 260 [Abstract]
bloodstream trypanosomes of 4 different subgenera, relative activities of alanine vs. aspartate aminotransferases were characteristic of each subgenus

Trypanosoma (Trypanozoon) brucei brucei
Godfrey, D. G.; and Kilgour, V., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 219-224
Trypanosoma brucei brucei, T. b. rhodesiense, T. b. gambiense, enzyme electrophoresis used to differentiate between subspecies, potentially useful in search for animal reservoirs or in identifying foci of human infections

Trypanosoma brucei brucei
Haston, W., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 275 [Demonstration]
Trypanosoma brucei brucei, metabolic changes accompanying transformation from bloodstream forms to vector forms, studies using microrespirometric technique to measure oxygen consumption

Trypanosoma brucei brucei
Hawking, F., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 271 [Abstract]
human plasma resistance test for measurement of resistance of polymorphic trypanosomes to human plasma, differentiation of potentially infective Trypanosoma brucei rhodesiense from non-infective T. b. brucei

T[rypanosoma] brucei brucei
Hudson, K. M.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 273 [Abstract]
T[rypanosoma] brucei brucei in mice (exper.), depressed immune responsiveness on challenge with heterologous antigens

Trypanosoma brucei brucei
Hudson, K. M.; et al., 1976, Nature (5583), v. 264, 256-258
Trypanosoma brucei brucei, mice, link between immunodepression, high IgM levels and evasion of immune response

Trypanosoma brucei brucei
Hudson, K. M.; and Byner, C., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 265 [Abstract]
Trypanosoma brucei brucei, mice, changes in lymphoid organ architecture

Trypanosoma brucei brucei
van den Ingh, T., 1977, Tijdschr. Diergeneesk., v. 102 (20), 1210-1214 [Abstract of thesis]
Trypanosoma brucei brucei, rabbit, 3 distinguishable stages during course of infection, immunological response, pathomorphological changes

Trypanosoma brucei brucei
Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 11 [Demonstration]
Trypanosoma brucei brucei, T. vivax, T. congolense, isoenzymes of alanine aminotransferase as possible specific characters

Trypanosoma brucei brucei
Kilgour, V.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 270 [Abstract]
Trypanosoma brucei gambiense, distinctive isoenzyme patterns, attempted differentiation from T. b. rhodesiense and T. b. brucei by electrophoresis on thin-layer starch-gel

Trypanosoma brucei brucei
Kilgour, V.; and Godfrey, D. G., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 258 [Abstract]
Trypanosoma brucei brucei in mice, influence of dietary pyridoxine

Trypanosoma brucei brucei
Leeflang, P.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 412 [Letter]
Trypanosoma spp., concentration and differentiation of species using a hypotonic lysis method

Trypanosoma brucei brucei
Linstead, D. J.; Klein, R. A.; and Cross, G. A. M., 1977, J. Gen. Microbiol., v. 101 (2), 243-251
Trypanosoma brucei, T. brucei brucei, threonine catabolism, inhibition of threonine dehydrogenase by wide range of agents

T[rypanosoma] b. brucei
Longstaffe, J. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 150 [Abstract]
T[rypanosoma] b. brucei-infected guinea pigs, immunodepression in trypanosomiasis, attempts to characterize response of thymus-dependent lymphocytes

Trypanosom[a] brucei brucei
Longstaffe, J. A.; Freeman, J.; and Hudson, K. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 264-265 [Abstract]
Trypanosom[a] brucei brucei, mice, some thymus-dependent and thymus-independent immune responses, results support postulation of either impaired T cell viability or impaired T-B cell interaction

Trypanosoma brucei brucei
Molyneux, D. H., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 391-397
Trypanosoma brucei gambiense, technique of intratesticular inoculation of rabbits used for isolation of strains from patients with Gambian trypanosomiasis, behavior compared with T. b. brucei as a means of distinguishing the two subspecies

Trypanosoma (Trypanozoon) brucei brucei
Opperdoes, F. R.; et al., 1976, Exper. Parasitol., v. 40 (2), 198-205
Trypanosoma brucei brucei, salicylhydroxamic acid (SHAM), inhibition of trypanosomal respiration in vitro, determination of toxicity for rats, determination of plasma levels in rats, no significant therapeutic effect in vivo, discussion of possibility that SHAM could still be trypanocidal if used in conjunction with another inhibitor of glycolysis

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Trypanosoma brucei brucei, T. evansi, oligomycin sensitivity of mitochondrial ATPase as a simple biochemical marker for presence of functional kinetoplast DNA and mitochondrial protein-synthesizing system and, hence, the ability to develop functional mitochondria and to multiply in insects

Trypanosoma (Trypanozoon) brucei brucei
Rickman, L. R., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 262-264
Trypanosoma brucei, attempted subspecific identification of isolates from non-human hosts using the blood incubation infectivity test (BIIT)

Trypanosoma brucei brucei
Robson, J.; and Rickman, L. R., 1973, Trop. Animal Health and Prod., v. 5 (3), 187-191
Trypanosoma brucei subgroups, identified by blood incubation infectivity test, non-human hosts: Lambwe Valley, South Nyanza, Kenya

Trypanosoma brucei brucei
Seed, J. R., 1974, J. Protozool., v. 21 (5), 639-646
African trypanosomes, antigens and antigenic variability, review including some new work on antigens demonstrated in Trypanosoma brucei gambiense extracts that are antigenically identical to rat serum proteins

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carbohydrates and phospholipids in surface coat of Trypanosoma brucei brucei, variations from most types of mammalian cells

Trypanosoma brucei brucei
Shakespeare, P. G.; and Trigg, P. I., 1974, J. Protozool., v. 21 (3), 452 [Abstract]
Trypanosoma b. brucei, T. congolense, T. lewisi, bloodstream forms, glucose utilization

Trypanosoma brucei brucei
Taylor, A. E. R.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 433 [Demonstration]
Trypanosoma brucei brucei, T.vivax, column separation and washing in various media, surface coat and internal morphology, agglutinability, infectivity, motility

Trypanosoma brucei brucei
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

Trypanosoma brucei brucei
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trypanosomiasis, cattle, evaluation of indirect fluorescent antibody test in diagnosis and epizootiological surveys: Senegal

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Ackerman, S. B.; and Seed, J. R., 1976, Clin. and Exper. Immunol., v. 25 (1), 152-158
Trypanosoma brucei gambiense in Microtus montanus, immunodepression of both humoral and cell-mediated immune responses, previously established specific antibody levels not affected by infection, specific antibody-producing capabilities of suramin-cured hosts were comparable to uninfected controls

Trypanosoma brucei gambiense
Ackerman, S. B.; and Seed, J. R., 1976, Infect. and Immun., v. 13 (2), 388-391
Trypanosoma brucei gambiense-infected Microtus montanus, increased susceptibility to Ehrlich's tumor growth, implications of trypanosome-induced immunosuppression toward susceptibility to neoplastic growth

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Afchain, D.; et al., 1975, Medecine Afrique Noire, v. 22 (5), 351-360
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Bogucki, M. S.; and Seed, J. R., 1976, J. Protozool., v. 23 (2), 17A [Abstract]
Trypanosoma brucei gambiense, host antigens obtained from extracts of well-washed rat-harvested parasites identified as immunoglobulins

Trypanosoma brucei gambiense
Cunningham, I., 1977, J. Protozool., v. 24 (2), 325-329
new culture medium for maintenance of Glossina morsitans morsitans tissues and growth of trypanosomatids, medium based on amino-acid composition of tsetse hemolymph and contains fetal bovine serum

Trypanosoma brucei gambiense
Fromentin, H., 1974, J. Protozool., v. 21 (3), 470 [Abstract]
Trypanosoma brucei gambiense strain M'Bala Victor, ability to protect against 2 heterologous strains (Eliane and Huguette), mice

Trypanosoma (Trypanozoon) brucei gambiense
Godfrey, D. G.; and Kilgour, V., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 219-224
Trypanosoma brucei brucei, T. b. rhodesiense, T. b. gambiense, enzyme electrophoresis used to differentiate between subspecies, potentially useful in search for animal reservoirs or in identifying foci of human infections

Trypanosoma brucei gambiense
Greenwood, B. M.; and Whittle, H. C., 1976, Clin. and Exper. Immunol., v. 24 (1), 133-138
Trypanosoma brucei gambiense, humans, evidence that complement activation plays a role in pathogenesis of Gambian sleeping sickness: low levels of complement components C3, C4, and factor B in sera, high serum IgM levels, possible formation of large molecular weight IgM complexes, increase in serum C3 levels after melarsoprol treatment

Trypanosoma brucei gambiense
Kilgour, V.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 270 [Abstract]
Trypanosoma brucei gambiense, distinctive isoenzyme patterns, attempted differentiation from T. b. rhodesiense and T. b. brucei by electrophoresis on thin-layer starch-gel

Trypanosoma brucei gambiense
Mahmoud, A. A. F.; and Warren, K. S., 1976, J. Infect. Dis., v. 133 (4), 487-491
human African trypanosomiasis, diagnosis and management, epidemiology, travelers to Africa, review

T[rypanosoma] b[rucei] gambiense
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at least 3 of 4 strains of trypanosomes of the subgenus Trypanozoon isolated from domestic pigs were serum-resistant in the blood incubation infectivity test (possibly or probably T. b[rucei] gambiense, at any rate potentially dangerous to man): Liberia

Trypanosoma (Trypanozoon) brucei gambiense
Molyneux, D. H., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 391-397
Trypanosoma brucei gambiense, technique of intratesticular inoculation of rabbits used for isolation of strains from patients with Gambian trypanosomiasis, behavior compared with T. b. brucei as a means of distinguishing the two subspecies

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Trypanosoma brucei gambiense-infected Microtus montanus, free serum amino acids, alteration in molar ratios and diurnal variation, possible role in neuropsychiatric syndromes of African trypanosomiasis

Trypanosoma brucei gambiense
Newport, G. R.; and Page, C. R. III, 1977, J. Parasitol., v. 63 (6), 1060-1065
Trypanosoma brucei gambiense-infected Microtus montanus, free amino acids in brain, liver, and skeletal muscle tissues, relatively specific alteration of free tyrosine concentrations suggests alteration in host metabolism of this amino acid and/or parasite utilization, possible relationship to neurological syndromes associated with infection

Trypanosoma brucei gambiense
Oudart, J. L.; et al., 1976, Medecine Afrique Noire, v. 23 (1), 25-29
Trypanosoma brucei gambiense, electrophoresis of spinal fluid proteins in African blacks, comparison of normal standards with findings in trypanosomiasis and with standards in Europeans: West Africa

Trypanosoma brucei gambiense
Oudart, J. L.; Diallo, B.; and Rolez, S., 1976, Medecine Afrique Noire, v. 23 (1), 31-37
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Trypanosoma (Trypanozoon) brucei gambiense
Rickman, L. R., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 262-264
Trypanosoma brucei, attempted subspecific identification of isolates from non-human hosts using the blood incubation infectivity test (BIIT)

Trypanosoma brucei gambiense
Risby, E. L., 1976, J. Protozool., v. 23 (2), 27A [Abstract]
Trypanosoma brucei, T. b. gambiense, T. b. rhodesiense, trypanocidal effects of methylglyoxal bis (guanylhydrazone) derivatives

Trypanosoma brucei gambiense
Ristic, M.; and Smith, R. D., 1974, Proc. 6. Internat. Conf. World Ass. Adv. Vet. Parasitol. (Vienna, Austria, Sept. 18-20, 1973), 41-63
zoonoses caused by hemoprotozoa, review: trypanosomiasis, malaria, babesiosis

Trypanosoma brucei gambiense
Seed, J. R., 1974, J. Protozool., v. 21 (5), 639-646
African trypanosomes, antigens and antigenic variability, review including some new work on antigens demonstrated in Trypanosoma brucei gambiense extracts that are antigenically identical to rat serum proteins

Trypanosoma brucei gambiense
Seed, J. R., 1977, Internat. J. Parasitol., v. 7 (1), 55-60
Trypanosoma brucei gambiense, relative efficiency of IgM and IgG-type antibodies in presence and absence of complement compared on weight basis by both agglutination and in vitro protection tests, ability to passively transfer immunity in vivo also compared, hypothesized that IgM is responsible for relapse phenomena observed in blood while IgG is more active in extravascular locations

Trypanosoma brucei gambiense
Seed, J. R.; et al., 1976, Am. Midland Naturalist, v. 96 (2), 379-390
Trypanosoma brucei gambiense in Microtus montanus (exper.), role of spleen in host immunity; effect of infection (enlarged spleen, increase in adrenal weights, decrease in size of gonads), possible means of distinguishing parasite stress from other forms of natural population stress; possible relationship to reproductive potential and population density

Trypanosoma brucei gambiense
Seed, J. R.; and Varney, J. R., 1976, Exper. Parasitol., v. 40 (2), 238-249
Trypanosoma brucei gambiense-infected rabbits, changes in diurnal rhythm of body temperature, preliminary results on diurnal rhythms of water consumption, fecal and urine output, return of normal temperature curve following chemotherapy(hydrocortisone + pentamidine isethionate)

Trypanosoma brucei gambiense
Stibbs, H. H.; and Seed, J. R., 1973, J. Protozool., v. 20 (4), 503
Trypanosoma brucei gambiense-infected Microtus montanus, elevated serum tyrosine transaminase levels, high levels of this enzyme found in sonicated preparations of parasite itself

Trypanosoma brucei gambiense
Stibbs, H. H.; and Seed, J. R., 1976, Exper. Parasitol., v. 39 (1), 1-6
Trypanosoma brucei gambiense, chronically infected Microtus montanus, elevated serum and hepatic tyrosine aminotransferase, high serum levels may result from lysis of parasites (possibly due to agglutination by antibody) containing high levels of enzyme, implications for catecholamine metabolism and consequently for pathologic behavioral syndrome

Trypanosoma brucei gambiense
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

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Van Wettere, P., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 294-295 [Abstract]
Glossina palpalis palpalis, seasonal fluctuations of "Apparent Density" in endemic zone of T[rypanosoma] brucei gambiense sleeping sickness: Zaire

Trypanosoma brucei rhodesiense
Bagster, I. A.; and Parr, C. W., 1973, Nature (5415), v. 244, 364-366
trypanosome identification by electrophoresis of soluble enzymes

Trypanosoma (Trypanozoon) brucei rhodesiense
Baker, J. R.; and McConnell, E., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 153-154 [Letter]
Trypanosoma brucei ssp., response of 2 strains to blood incubation infectivity test, Gambela IV from Glossina tachinoides identified as T. b. brucei, Gambela V from Tragelaphus scriptus identified as T. b. rhodesiense, however Gambela V was not infective to a human volunteer: near village of Pinybago on Gilo River, Illubabor Province, Ethiopia

Trypanosoma brucei rhodesiense
Baker, J. R.; and Taylor, A. E. R., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 471-485
Trypanosoma brucei brucei- and T. brucei rhodesiense-infected Pan troglodytes (exper.), course of infection, serologic relationships between trypanosome species and strains, blood changes, cerebrospinal fluid changes, post-mortem observations

Trypanosoma brucei rhodesiense
Boehringer, S.; and Hecker, H., 1974, J. Protozool., v. 21 (5), 694-698
Trypanosoma brucei brucei, T. b. rhodesiense, pleomorphic strains, quantitative differences in organelle composition during transformation from slender to stumpy blood forms, compared with monomorphic T. b. brucei

Trypanosoma brucei rhodesiense
Cunningham, I., 1977, J. Protozool., v. 24 (2), 325-329
new culture medium for maintenance of Glossina morsitans morsitans tissues and growth of trypanosomatids, medium based on amino-acid composition of tsetse hemolymph and contains fetal bovine serum

Trypanosoma brucei rhodesiense
Dallas, A. B. C., 1976, J. Trop. Med. and Hyg., v. 79 (8), 182-188
Trypanosoma brucei rhodesiense, Plasmodium berghei yoelii, mice, mutual potentiation of concurrent infections, possibly caused by immunosuppression

Trypanosoma brucei rhodesiense
Damper, D.; and Patton, C. L., 1976, J. Protozool., v. 23 (2), 349-356
Trypanosoma brucei brucei, T. b. rhodesiense, pentamidine transport and sensitivity, concluded that reduced drug uptake is primary mechanism of pentamidine resistance in trypanosomes

Trypanosoma brucei rhodesiense
Ellis, D. S.; and Evans, D. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 20 [Demonstration]
Trypanosoma brucei rhodesiense, electron micrograph demonstration of passage of parasite through the anterior mid-gut of Glossina morsitans morsitans vectors

Trypanosoma brucei rhodesiense, illus.
Ellis, D. S.; and Evans, D. A., 1977, Nature (5614), v. 267, 834-835
Trypanosoma brucei rhodesiense, passage through peritrophic membrane of Glossina morsitans morsitans

Trypanosoma brucei rhodesiense
Ellis, D. S.; and Evans, D. A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 380 [Demonstration]
Trypanosoma brucei rhodesiense, electron micrographs of passage of Trypanosoma through the peritrophic membrane of Glossina morsitans in central region of midgut

Trypanosoma brucei rhodesiense
Ellis, D. S.; and Evans, D. A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 385 [Demonstration]
Trypanosoma brucei rhodesiense (Botswana strain 180), electron micrographs of "cyst-like bodies" found when grown in cultures, possible reproductive function

Trypanosoma (Trypanozoon) brucei rhodesiense
Ellis, D. S.; Ormerod, W. E.; and Lumsden, W. H. R., 1976, Acta Trop., v. 33 (2), 151-168
Trypanosoma brucei rhodesiense, origin and structure of 2 types of filaments (one associated with stumpy trypanosomes and the other with slender trypanosomes)

Trypanosoma brucei rhodesiense
Evans, D. A.; Brightman, C. J.; and Holland, M. F., 1977, Lancet, London (8041), v. 2, 769 [Letter]
Trypanosoma brucei brucei and T. b. rhodesiense infections in mice (exper.) successfully treated with salicylhydroxamic acid if combined with glycerol and administered before patent infection established; T. vivax infections of different durations all successfully cured

Trypanosoma brucei rhodesiense
Evans, D. A.; and Brown, R. C., 1973, J. Protozool., v. 20 (1), 157-160
Trypanosoma brucei bloodstream and culture forms, inhibition of cyanide-insensitive respiration by m-chlorobenzhydroxamic acid, cyanide-sensitive respiration of culture forms not affected, possible importance as trypanocidal drug, implications for mechanism of terminal respiration

T[rypanosoma] brucei rhodesiense
Gibson, W., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 386 [Demonstration]
T[rypanosoma] brucei-complex, isoenzyme patterns

Trypanosoma (Trypanozoon) brucei rhodesiense
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Trypanosoma brucei brucei, T. b. rhodesiense, T. b. gambiense, enzyme electrophoresis used to differentiate between subspecies, potentially useful in search for animal reservoirs or in identifying foci of human infections

Trypanosoma brucei rhodesiense
Hawking, F., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 271 [Abstract]
human plasma resistance test for measurement of resistance of polymorphic trypanosomes to human plasma, differentiation of potentially infective Trypanosoma brucei rhodesiense from non-infective T. b. brucei

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Jones, I. G.; Lowenthal, M. N.; and Buyst, H., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 388-395
Trypanosoma brucei rhodesiense, electrocardiographic abnormalities in infected humans

Trypanosoma brucei rhodesiense
Kilgour, V.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 270 [Abstract]
Trypanosoma brucei gambiense, distinctive isoenzyme patterns, attempted differentiation from T. b. rhodesiense and T. b. brucei by electrophoresis on thin-layer starch-gel

Trypanosoma brucei rhodesiense
Mahmoud, A. A. F.; and Warren, K. S., 1976, J. Infect. Dis., v. 133 (4), 487-491
human African trypanosomiasis, diagnosis and management, epidemiology, travelers to Africa, review

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Trypanosoma brucei, T. b. gambiense, T. b. rhodesiense, trypanocidal effects of methylglyoxal bis (guanylhydrazone) derivatives

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zoonoses caused by hemoprotozoa, review: trypanosomiasis, malaria, babesiosis

Trypanosoma brucei rhodesiense
Robson, J.; and Rickman, L. R., 1973, Trop. Animal Health and Prod., v. 5 (3), 187-191
Trypanosoma brucei subgroups, identified by blood incubation infectivity test, non-human hosts: Lambwe Valley, South Nyanza, Kenya

Trypanosoma brucei rhodesiense
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African trypanosomes, antigens and antigenic variability, review including some new work on antigens demonstrated in Trypanosoma brucei gambiense extracts that are antigenically identical to rat serum proteins

Trypanosoma brucei rhodesiense
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

Trypanosoma brucei rhodesiense, illus.
Venkatesan, S.; Bird, R. G.; and Ormerod, W. E., 1977, Internat. J. Parasitol., v. 7 (2), 139-147
Trypanosoma brucei rhodesiense, intracellular enzymes and their localization in slender and stumpy forms, possible relation to lipid content

Trypanosoma brucei rhodesiense
Venkatesan, S.; and Ormerod, W. E., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 12-13 [Demonstration]
Trypanosoma brucei rhodesiense, antigenicity and stilbamidine resistance

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Venkatesan, S.; and Ormerod, W. E., 1976, Comp. Biochem. and Physiol., v. 53 (4B), 481-487
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Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Bufo boreas (blood): central California

Trypanosoma bufophlebotomi Ayala 1970, illus.
Werner, J. K.; and Walewski, K., 1976, J. Parasitol., v. 62 (1), 20-25
brief description
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Kingston, N.; and Morton, J. K., 1975, Proc. Helminth. Soc. Washington, v. 42 (2), 179-181
Trypanosoma cervi in Cervus canadensis (spleen, liver), recovery of multiplication stages

Trypanosoma cervi
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Trypanosom[a] congolense
Adam, K. M. G.; and Dipeolu, O. O., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 293 [Abstract]
Trypanosom[a] brucei, T. congolense, cyclical development, comparison of strains

Trypanosoma congolense
Aiyedun, B. A.; and Amodu, A. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 67-71
trypanosomiasis, albino rats (exper.), humans, diagnosis, capillary-tube precipitation test (known as capillary-tube agglutination test), rapid and specific technique that could be used in field survey work

Trypanosoma congolense
Al Chalabi, K.; and Gutteridge, W. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146 [Abstract]
Trypanosoma spp., Crithidia fasciculata, C. oncopelti, thymidine phosphorylase activity, salvage pathways, inhibition of activity by suramin and ethidium

Trypanosoma congolense
Al Chalabi, K.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), xv-xvi [Abstract]
6 spp. of Trypanosomatidae, thymidylate synthetase activity, ranks as potential target for chemotherapeutic attack since it is markedly more sensitive to suramin inhibition than isofunctional mammalian and bacterial enzymes

Trypanosoma congolense
Al Chalabi, K.; and Gutteridge, W. E., 1977, Parasitology, v. 74 (3), 299-312
deoxythymidylate catabolism in some trypanosomatids

Trypanosoma congolense
Bagster, I. A.; and Parr, C. W., 1973, Nature (5415), v. 244, 364-366
trypanosome identification by electrophoresis of soluble enzymes

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Brown, L. A.; and Losos, G. J., 1977, Research Vet. Sc., v. 23 (2), 196-203
Trypanosoma congolense and T. brucei, comparative pathology in both bled and non-bled albino rats (exper.): parasitemia, packed cell volumes, weight of spleen and lymph, histology of thymus, spleen, lymph nodes, and bone marrow

Trypanosoma congolense
Buengener, W., 1975, Tropenmed. u. Parasitol., v. 26 (3), 281-284
Trypanosoma musculi in inbred mice (exper.), parasite multiplication and course of infection in single infection, after pre-infection with T. congolense, or with added infection of T. brucei

Trypanosoma (Nannomonas) congolense
Buengener, W.; and Mehlitz, D., 1976, Tropenmed. u. Parasitol., v. 27 (4), 405-410
experimentally infected Capra ibex s. hircus, few clinical signs of infection

Trypanosoma congolense, illus.
Buengener, W.; and Mueller, G., 1976, Tropenmed. u. Parasitol., v. 27 (3), 370-371
trypanosome aggregation in capillaries of host organs, 2 possible explanations for the phenomena

Trypanosoma (Nannomonas) congolense
Carmichael, I. H.; and Hobday, E., 1975, Onderstepoort J. Vet. Research, v. 42 (2), 55-62
blood parasites of wild Bovidae, incidence, epizootiological importance
Syncerus caffer
Kobus leche
Aepyceros melampus
Tragelaphus strepsiceros
(blood of all): all from Ngamiland, Botswana

Trypanosoma congolense
Clarkson, M. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 272 [Abstract]
Trypanosoma brucei and T. congolense in mice, immunoglobulin changes during infection

Trypanosoma congolense
Clarkson, M. J., 1976, Vet. Parasitol., v. 2 (1), 9-29
trypanosomes, immunological problems, review: antigens; host reactions to infection; immunological state of host

Trypanosom[a] congolense
Clarkson, M. J.; and McCabe, W. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 12 [Demonstration]
Trypanosoma spp., relative susceptibility of mice to infection by buccal, gastric, or intraperitoneal routes, possible significance of oral transmission under natural conditions

Trypanosoma congolense
Cunningham, I., 1977, J. Protozool., v. 24 (2), 325-329
new culture medium for maintenance of Glossina morsitans morsitans tissues and growth of trypanosomatids, medium based on amino-acid composition of tsetse hemolymph and contains fetal bovine serum

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Cunningham, I.; and Latif, B. M. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 270 [Abstract]
Trypanosoma brucei, T. congolense, antigenic properties during transformation from blood to culture forms

Trypanosoma congolense
Dar, F. K.; et al., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 21-29
techniques used to study serotypes of salivarian trypanosomes being circulated by tsetse flies in 4 separate geographical areas, methods used to isolate and serologically type the trypanosomes are described and discussed in detail: Uganda and Kenya, East Africa

Trypanosoma congolense
Diffley, P.; and Honigberg, B. M., 1977, J. Parasitol., v. 63 (4), 599-606
Trypanosoma congolense, presence, host specificity, and time of accretion of rat plasma components on parasite surface, quantitative indirect fluorescent antibody analysis

Trypanosoma (Nannomonas) congolense Broden
Diffley, P.; Honigberg, B. M.; and Mohn, F. A., 1976, J. Parasitol., v. 62 (1), 136-137
Trypanosoma congolense, improved method of cryopreservation in liquid nitrogen

Trypanosoma congolense
Duxbury, R. E.; et al., 1973, Isotopes and Radiation Parasitol. III, 179-180
Trypanosoma rhodesiense, mice, rats, cattle, rhesus monkeys; T. congolense, mice, dogs, cattle; T. brucei, cattle: attempted immunization with irradiated trypanosomes, brief communication

Trypanosoma congolense
Elce, B. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 162 [Abstract]
Trypanosoma congolense, T. brucei, T. rhodesiense, development in Glossina morsitans and small laboratory mammals

Trypanosoma congolense
El-On, J.; and Greenblatt, C. L., 1977, Exper. Parasitol., v. 41 (1), 31-42
Trypanosoma lewisi, T. acomys, T. cruzi, method for cultivation with mammalian tissue, results not good with T. equiperdum and T. congolense

Trypanosoma congolense
Franco, B.; and Halevy, S., 1977, J. Protozool., v. 24 (2), Suppl., 18A [Abstract]
Trypanosoma congolense-infected mice, decrease in activity of glucose-6-phosphatase in liver, changes in amounts of liver fatty acids, survival time of some mice extended by barbiturate treatment

Trypanosoma congolense
Fregene, A. O.; et al., 1975, J. Parasitol., v. 61 (6), 1070-1073
comparative responses of radioattenuated Trypanosoma brucei and T. congolense in rats: radiation sensitivity with respect to infectivity; immunogenicity

Trypanosom[a] (Nannomonas) congolense
Godfrey, D. G.; and Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 260 [Abstract]
bloodstream trypanosomes of 4 different subgenera, relative activities of alanine vs. aspartate aminotransferases were characteristic of each subgenus

Trypanosoma congolense
Goose, J., 1977, Parasitology, v. 75 (2), xxxv [Abstract]
Fasciola hepatica-infected rats, depressed immune response to sheep red blood cells, greater numbers of peritoneal macrophages and increased phagocytosis, increased resistance to infection with Trypanosoma congolense and Nippostrongylus brasiliensis not serum-transferable, concluded that Fasciola hepatica stimulates certain T-cell populations as well as reticulo-endothelial system

Trypanosoma congolense
Hawking, F., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 170
Trypanosoma congolense, periodicity in rodents (exper.)

Trypanosoma congolense
Hawking, F.; Wilson, A. J.; and Paris, J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 289 [Letter]
Trypanosoma congolense, effect of dexamethasone upon infection in calves, results suggest that anemia was not due to auto-immunization but probably due to direct toxic action of trypanosomes on blood

T[rypanosoma] congolense
Holmes, P. H.; and Jennings, F. W., 1976, Pathophysiol. Parasit. Infect., 199-210
T[rypanosoma] congolense, T. brucei, rabbits infected and subsequently treated with diminazene aceturate, haematological indices, changes in circulating red cell volume and plasma volume, changes in erythropoiesis, red cell survival studies, dramatic effects of treatment on anemia, similar preliminary results with T. congolense in Zebu calves

Trypanosoma congolense
Holmes, P. H.; and Mamo, E., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 274 [Abstract]
Trypanosoma congolense in experimentally infected zebu cattle, dynamics of development of trypanosomal anemia

Trypanosoma congolense
James, D. M., 1976, Internat. J. Parasitol., v. 6 (2), 179-182
Trypanosoma brucei, T. congolense, induction of protective immunity in rodents receiving living trypanosomes pretreated with isometamidium chloride or diminazene aceturate tetrahydrate

Trypanosoma congolense
Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 11 [Demonstration]
Trypanosoma brucei brucei, T. vivax, T. congolense, isoenzymes of alanine aminotransferase as possible specific characters

Trypanosoma congolense
Kobayashi, A.; Soltys, M. A.; and Woo, P.T.K., 1976, Ann. Trop. Med. and Parasitol., v. 70 (1), 53-58
Trypanosoma congolense, sheep, diagnosis, comparison of various parasitological techniques (wet mount preparation; hematocrit centrifuge technique; mouse inoculation test) with various serological techniques (immunolysis test; indirect fluorescent antibody test; complement fixation test; immunoconglutination test), effect of treatment with quinapyramine dimethosulphate on diagnosis

Trypanosoma congolense
Kobayashi, A.; and Tizard, I. R., 1976, Tropenmed. u. Parasitol., v. 27 (4), 411-417
Trypanosoma congolense-infected calves (exper.), changes in levels of immunoglobulins and complement fixing antibody titres during course of infection

Trypanosoma congolense
Kobayashi, A.; Tizard, I. R.; and Woo, P. T. K., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 401-406
Trypanosoma congolense, calves, anemia proved to be of immunological origin, antigen-antibody-complement complexes deposited on surface of erythrocytes results in their immune elimination and leads to clinical anemia

Trypanosoma congolense
Lanham, S. M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 270 [Abstract]
trypanosomes, enzymatic identification of precipitating antigens

Trypanosoma congolense
Leeflang, P.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 412 [Letter]
Trypanosoma spp., concentration and differentiation of species using a hypotonic lysis method

Trypanosoma (Nannomonas) congolense
Luckins, A. G., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 272-273 [Abstract]
Trypanosoma congolense, T. vivax, cattle, elevated IgM levels: Uganda

Trypanosoma congolense
Luckins, A. G., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 148-149 [Abstract]
Trypanosoma congolense, T. vivax in zebu cattle, increased levels of IgM in presence of infection

Trypanosoma congolense
Luckins, A. G., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 337-344
Trypanosoma vivax, T. brucei, T. congolense-infected Tragelaphus scriptus, serum immunoglobulin levels and relative proportions of serum proteins, compared with Zebu cattle

Trypanosoma congolense
Luckins, A. G., 1976, Ann. Trop. Med. and Parasitol., v. 70 (2), 133-145
Trypanosoma congolense, T. vivax, zebu cattle (exper.), serum immunoglobulin levels during infection and after administration of diminazene aceturate

Trypanosoma congolense
Luckins, A. G.; and Mehlitz, D., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 479-480
Trypanosoma brucei, T. vivax, T. congolense, cattle (exper.), single or mixed infections, serum immunoglobulin levels

Trypanosoma congolense
MacKenzie, A. R.; Boreham, P. F. L.; and Facer, C. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 268 [Abstract]
Trypanosoma congolense, T. brucei, cattle, anti-liver, anti-Wassermann, and anti-fibrinogen antibodies, human serum from sleeping sickness patients also showed raised levels of these autoantibodies, trypanosome-infected lions and hyaenas did not

Trypanosoma congolense
Mahmoud, M. M.; and Elmalik, K. H., 1977, Trop. Animal Health and Prod., v. 9 (3), 167-170
Trypanosoma congolense, goats and calves, experimental infection compared, goats developed chronic form of trypanosomiasis, may be a possible reservoir host

Trypanosoma congolense
Mamo, E.; and Holmes, P. H., 1975, Research Vet. Sc., v. 18 (1), 105-106
Trypanosoma congolense, chronically infected Ethiopian Zebu cattle, erythrokinetics, anemia due principally to massive loss of red cells from circulation

Trypanosoma congolense
Mansfield, J. M.; Craig, S. A.; and Stelzer, G. T., 1976, Infect. and Immun., v. 14 (4), 976-981
Trypanosoma brucei, T. congolense, mitogenic effects of trypanosome extracts in vitro for lymphocytes from normal rabbits, possible relationship to immunological dysfunctions occurring in chronic African trypanosomiasis

Trypanosoma congolense
Mansfield, J. M.; and Kreier, J. P., 1972, Infect. and Immun., v. 5 (5), 648-656
Trypanosoma congolense, rabbits (exper.), complement-fixing and precipitating autoantibodies to normal allogeneic and autologous tissues, passive transfer of autoantibody to normal rabbits did not produce observable pathology, cell-mediated autoimmunity was not shown

Trypanosoma congolense
Mansfield, J. M.; and Kreier, J. P., 1972, Infect. and Immun., v. 6 (1), 62-67
Trypanosoma congolense, rabbits (exper.), Arthus-type immediate hypersensitivity reactions demonstrated, no cell-mediated hypersensitivity reactions observed, role of immediate-type skin reaction in pathology of infection and possible use in diagnosis

Trypanosoma congolense
Maxie, M. G.; and Losos, G. J., 1977, Vet. Parasitol., v. 3 (4), 277-281
Trypanosoma congolense, cattle, release of parasites from microcirculation after injection of berenil, suggested that berenil acts by making trypanosomes available to host defenses such as the macrophage system

Trypanosoma congolense
Maxie, M. G.; Losos, G. J.; and Tabel, H., 1976, Pathophysiol. Parasit. Infect., 183-198
Trypanosoma congolense, T. vivax, cattle, development of macrocytic normochromic anemia, leukopenia, and persistent thrombocytopenia, significant differences in response of cattle to the two species

Trypanosoma (Nannomonas) congolense
Mehlitz, D., 1975, Tropenmed. u. Parasitol., v. 26 (3), 265-275
Trypanosoma spp. of Nannomonas, Duttonella, and Trypanozoon subgenera, comparison of indirect fluorescent antibody test and complement fixation test as means of differentiating subgenera by serology and persistence of antibodies

Trypanosoma congolense-type
Moloo, S. K., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 205-211
Trypanosoma vivax-type, T. congolense-type, infection rates in Glossina swynnertoni, relationship with types of mammals used as food sources: Serengeti National Park, Tanzania

Trypanosom[a] congolense-type
Moloo, S. K., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 286 [Abstract]
source of food as important factor determining trypanosome infection rate in natural population of Glossina swynnertoni: Ikoma-Serengeti area, Tanzania

Trypanosoma congolense
Murray, M.; Murray, P. K.; and McIntyre, W. I. M., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 325-326
African trypanosomiasis, light microscopic examination of buffy coat zone of a microhaematocrit capillary tube expressed onto a slide found to be more reliable than standard methods for diagnosing infections in cattle; method also allowed differentiation of species

Trypanosoma congolense
Murray, M.; and Urquhart, G. M., 1977, Advances Exper. Med. and Biol., v. 93, 209-241
African trypanosomiasis, immunoprophylaxis, review: cattle and sheep, laboratory animals; chemotherapeutic agents and immunity, nature of protective antigen, effector mechanisms in protection, manipulation of host resistance

Trypanosoma congolense
Nielsen, K.; and Sheppard, J., 1977, Experientia, v. 33 (6), 769-771
Trypanosoma congolense, T. lewisi, activation of complement, possible roles in pathology of disease

Trypanosoma congolense
Oduye, O. O.; and Dipeolu, O. O., 1976, J. Small Animal Practice, v. 17 (5), 331-337
blood parasites of dogs, single and mixed infections, correlation between incidence and rainfall, degree of parasitaemia, infectivity rate within age groups, no significant difference in host susceptibility of local and exotic breeds to infection: Ibadan, Nigeria

Trypanosoma congolense
Ormerod, W. E., 1976, Science, v. 191 (4229), 815-821
possible ecological effects of trypanosomiasis control, increased cattle production leading to overgrazing and possibly to climatic change and drought: Africa

Trypanosoma congolense, illus.
Perie, N. M.; Tinnemans-Anggawidjaja, T.; and Zwart, D., 1975, Tropenmed. und Parasitol., v. 26 (4), 399-404
Trypanosoma spp. in experimentally infected domestic animals, sandwich immunofluorescent complement fixation test compared with indirect fluorescent antibody test for use in detecting binding of complement to antigen-antibody complex

Trypanosoma congolense
Politzar, H., 1974, Tropenmed. u. Parasitol., v. 25 (1), 22-27
Trypanosoma vivax alone or with T. brucei and T. congolense, cattle (exper.), homologous and heterologous antibody responses determined by indirect fluorescent antibody test

Trypanosoma congolense
Rafael, A. G., 1975, An. Serv. Vet., Mocambique (20-21), 1972-1973, 331-350
Trypanosoma spp., pathology, economic importance, treatment, vector control, extensive review: Africa

Trypanosoma congolense
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma (Nannomonas) congolense group
Riordan, K., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 11-20
Trypanosoma spp., survey of long-term variations in infection rates of Glossina spp. vectors found along major trade cattle route in southern Nigeria

Trypanosoma congolense
Roberts, C. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 11 [Demonstration]
Trypanosoma congolense, T. vivax, cattle, sheep, serum lipid levels, significance of increase in plasma volume in trypanosome infections

Trypanosom[a] congolense
Roberts, C. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 279-280 [Abstract]
Trypanosom[a] vivax, T. congolense, ruminants, serum lipid levels

Trypanosoma congolense
Roberts, C. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 155 [Abstract]
Trypanosoma congolense and T. vivax-infected sheep, changes in lipid levels during infection, effects of infection on metabolic processes of serum lipoproteins

Trypanosoma congolense
Roberts, C. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 275 [Abstract]
trypanosomiasis in ruminants, alterations in lipid metabolism

Trypanosoma congolense
Roberts, C. J.; and Gray, A. R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 278-279 [Abstract]
Trypanosoma congolense isolates from wild game animals, pattern of infectivity for rats may be different from previously described patterns

Trypanosoma congolense
Roberts, C. J.; and Gray, A. R., 1973, Trop. Animal Health and Prod., v. 5 (4), 220-233
Trypanosoma vivax, T. congolense, T. brucei, N'dama, Muturu, and Zebu cattle (exper.), comparison of host resistance, pathology; N'dama cattle demonstrated more resistance than other breeds

Trypanosoma congolense
Sadun, E. H.; et al., 1972, Parassitologia, v. 14 (1), 25-28
Trypanosoma congolense, irradiated forms immunizing mice, dogs and cattle; T. rhodesiense, irradiated forms immunizing mice and rats; irradiated human strain of T. rhodesiense immunizing Macaca mulatta

Trypanosom[a] congolense
Saror, D. I., 1976, Vet. Rec., v. 98 (10), 196
Trypanosom[a] vivax, Trypanosom[a] congolense, plasma copper levels determined in exper. infected cattle, neither iron nor copper deficiencies are significant in the pathogenesis of bovine trypanosomiasis

Trypanosoma congolense
Schindler, R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 290 [Abstract]
Trypanosoma spp., cattle, attempted serologic differential diagnosis gave equivocal results

Trypanosoma congolense
Scott, J. M.; et al., 1977, Trop. Animal Health and Prod., v. 9 (3), 159-165
Trypanosoma congolense, cattle (exper.), some immunosuppression after vaccination with foot-and-mouth disease vaccine and clostridial vaccine, but protection against these diseases not impeded

Trypanosoma congolense
Shakespeare, P. G.; and Trigg, P. I., 1974, J. Protozool., v. 21 (3), 452 [Abstract]
Trypanosoma b. brucei, T. congolense, T. lewisi, bloodstream forms, glucose utilization

Trypanosoma congolense
Sheppard, J.; et al., 1977, Canad. Fed. Biol. Soc., Programme and Proc. 20. Ann. Meet., v. 20, 120 [Abstract]
Trypanosoma congolense, T. lewisi, complement activating factors, one of which may be a hexose polymer

Trypanosoma congolense
Soltys, M. A.; Thompson, S. M. R.; and Woo, P. T. K., 1973, Ann. Trop. Med. and Parasitol., v. 67 (4), 399-402
Trypanosoma brucei, rats, guinea pigs, exper. transmission through skin and intact mucous membranes of conjunctiva, mouth, rectum, vagina; T. congolense, exper. transmission through same routes all failed

Trypanosoma congolense
Ssenyonga, G. S. Z., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 272 [Abstract]
Trypanosoma brucei and T. congolense in rodents, distribution in tissues and body fluids

Trypanosoma congolense, illus.
Steiger, R. F.; et al., 1977, J. Parasitol., v. 63 (5), 861-867
Trypanosoma congolense, partial cyclic development in Glossina cell system, oxygen consumption

Trypanosoma congolense
Tabel, H.; Losos, G. J.; and Maxie, M. G., 1977, Canad. Fed. Biol. Soc., Programme and Proc. 20. Ann. Meet., v. 20, 118 [Abstract]
Trypanosoma vivax, T. congolense, cattle, determination of hemolytic complement activity and complement component C3 levels; impairment of complement system perhaps related to immunosuppression and susceptibility to secondary infection

Trypanosoma congolense
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

Trypanosoma congolense
Tizard, I. R.; et al., 1977, Canad. Fed. Biol. Soc., Programme and Proc. 20. Ann. Meet., v. 20, 4 [Abstract]
Trypanosoma congolense, autolysing in vitro, generation of hemolytic and cytotoxic fatty acids by endogenous phospholipase A, proposed as major pathogenic mechanism

Trypanosoma congolense
Tizard, I. R.; et al., 1977, Experientia, v. 33 (7), 901-902
Trypanosoma congolense, hemolytic activity is due to presence of free fatty acids generated by action of phospholipase A on endogenous phosphatidyl choline, some lysolecithin also contributes to lytic activity, T. lewisi is devoid of phospholipase A and does not generate free fatty acids and is therefore non-hemolytic

Trypanosoma congolense
Tizard, I. R.; and Holmes, W. L., 1976, Experientia, v. 32 (12), 1533-1534
Trypanosoma congolense, generation of toxic activity considered to be of potential significance in pathogenesis (phospholipase-like activity capable of mediating lysis of both nucleated cells and erythrocytes as well as acute inflammatory response on intradermal inoculation), Trypanosoma lewisi produced no significant hemolysis

Trypanosoma congolense
Tizard, I. R.; and Holmes, W. L., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 52-55
Trypanosoma congolense, release of soluble vasoactive material from parasites in intraperitoneal diffusion chambers implanted in rats, possibility that this factor may contribute to pathogenesis of infection

Trypanosoma congolense
Tizard, I. R.; and Ringleberg, C. P., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (6), 885 [Letter]
Trypanosoma congolense, release of biologically active material (capable of causing inflammatory reaction) from dying trypanosomes in diffusion chambers in rats, not observed with T. lewisi, suggests that hypothesis of trypanosomal toxins should be reconsidered

Trypanosoma congolense
Toure, S. M., 1973, Medecine Afrique Noire, v. 20 (11), 855-862
trypanocides of veterinary interest and their indications for use, review

Trypanosoma (Nannomonas) congolense, illus.
Toure, S. M., 1976, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 29 (1), 17-22
Trypanosoma (Nannomonas) congolense, 640 passages in mice during 10 years, biometrics, changes in morphology, no reduction in virulence

Trypanosoma congolense
Toure, S. M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 30 (1), 1-10
trypanosomiasis, domestic animals, diagnostic methods, review

Trypanosoma congolense
Toure, S. M.; et al., 1975, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 28 (4), 463-472
trypanosomiasis, cattle, evaluation of indirect fluorescent antibody test in diagnosis and epizootiological surveys: Senegal

Trypanosoma congolense
Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 1977, 1-15
17. Seminar on Trypanosomiasis (biochemistry and relevance to control; relevance of laboratory studies to tsetse control; field diagnosis; problems of land use and tsetse control)

Trypanosoma congolense
Uilenberg, G.; and Giret, M., 1973, Rev. Elevage et Med. Vet. Pays Trop., v. 26 (2), 147-150
Trypanosoma congolense, sheep, antibody against late variant type does not show neutralizing activity against type obtained early in infection; difficulty obtaining monovalent sera from laboratory rodents; Berenil treatment does not influence results of neutralization test; persistence of antibodies in sheep; unsuccessful trials of agglutination and immunolysis tests

Trypanosoma congolense
Wellde, B. T.; et al., 1973, Isotopes and Radiation Parasitol. III, 181-183
Trypanosoma congolense, cattle, clinical signs and parasitaemia, haematology, serum proteins, serum biochemical components, histopathologic findings, diagnosis by complement fixation test compared with indirect fluorescent antibody test

Trypanosoma congolense
Wiesenhuetter, E., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 60-66
trypanosomiasis, Zebu cattle, diagnosis, comparison of indirect fluorescent antibody test with microscopic examination, IFA proved to be reliable and rapid diagnostic aid useful for epizootiological studies: Tanzania

Trypanosoma congolense
Wiesenhuetter, E., 1976, Trop. Animal Health and Prod., v. 8 (4), 195-201
trypanosomiasis, cattle, incidence in relation to occurrence of tsetse flies and Tabanidae, mechanical transmission of trypanosomes by Tabanidae was at an insignificant rate, only measurable result of such transmission was change of Trypanosoma vivax/T. congolense ratio in favor of T. vivax: Dar es Salaam dairy farm, Africa

Trypanosoma congolense
Williamson, J., 1977, J. Protozool., v. 24 (2), Suppl., 5A [Abstract]
Trypanosoma congolense, trypanocidal activity of several polyanions

Trypanosoma congolense
Williamson, J.; and Scott-Finnigan, T. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 1-2 [Demonstration]
Trypanosoma congolense, in vitro testing of antiviral and antitumor compounds with possible trypanicidal effects, probable RNA inhibition

Trypanosoma congolense
Wilson, A. J.; Dar, F. K.; and Paris, J., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 313-317
Trypanosoma congolense, comparison of antigenic types isolated from wild tsetse flies in 4 geographically distinct areas of East Africa

Trypanosom[a] congolense
Wilson, A. J.; Dar, F. K.; and Paris, J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 287-288 [Abstract]
trypanosomiasis in a herd of cattle maintained under chemotherapeutic cover in area of high tsetse fly challenge: fly infection rates; comparison of diagnostic methods; differential infection rates with 3 Trypanosom[a] spp.; frequency of Berenil requirement; body weight gains of calves

Trypanosoma congolense
Woo, P. T. K.; and Rogers, D. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 319-326
Trypanosoma spp., statistical study of sensitivity of haematocrit centrifuge technique for detecting trypanosomes in blood, evaluation of relationship of probability of detection of trypanosome to trypanosome density

Trypanosoma congolense
Yesufu, H. M.; and Mshelbwala, A. S., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 307-312
prevalence of trypanosome species in cattle and tsetse flies along a trade cattle route
Zebu cattle
Glossina morsitans
all from southwestern Nigeria

Trypanosoma (Nannomonas) congolense, illus.
Young, A. S.; Kanhai, G. K.; and Stagg, D. A., 1975, Research Vet. Sc., v. 19 (1), 108-110
Trypanosoma congolense, phagocytosis by circulating macrophages in Syncerus caffer: near Aitong, Mara region, Kenya

Trypanosoma conorhini
Muehlpfordt, H.; and Schottelius, J., 1977, Tropenmed. u. Parasitol., v. 28 (1), 1-7
attempted differentiation of Trypanosoma cruzi strains and T. cruzi-like strains from T. rangeli and T. conorhini using agglutination reactions to protectin from sponges and lectin from Soja hispida

Trypanosoma conorhini
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma conorhini
Schottelius, J., 1976, Ztschr. Parasitenk., v. 50 (2), 182-183
Trypanosoma spp., effects of various lectins, protectins and blood group test sera

Trypanosoma conorhini
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Trypanosoma cruzi
Abrahamsohn, I. A.; and da Silva, W. D., 1977, Parasitology, v. 75 (3), 317-323
Trypanosoma cruzi, antibody-dependent cell-mediated cytotoxicity against epimastigotes by normal mouse splenic lymphocytes

Trypanosoma cruzi, illus.
Agosin, M.; et al., 1976, Internat. J. Biochem., v. 7 (11), 585-593
Trypanosoma cruzi culture form, spectral characterization of microsomal and other hemoproteins, results suggest that cytochrome-P-450-linked monooxygenase system may be responsible for resistance of this parasite to antimicrobial drugs

Trypanosoma cruzi
Agosin, M.; et al., 1976, Science (4261), v. 194, 195-197
Trypanosoma cruzi epimastigotes possess active detoxifying system whose characteristics are those of monooxygenase system linked to P-450 and whose activity is increased by phenobarbital, results suggest that important factor in T. cruzi's insensitivity to chemotherapeutic agents may be this ability to metabolize foreign compounds

Trypanosoma cruzi
Aguilar-Torres, F. G.; Rytel, M. W.; and Kagan, I. G., 1976, Am. J. Trop. Med. and Hyg., v. 25 (5), 667-670
Trypanosoma cruzi, human, comparison of counterimmunoelectrophoresis with latex agglutination and indirect hemagglutination in detection of antibodies, use as epidemiologic tool

Trypanosoma cruzi
Al Chalabi, K.; and Gutteridge, W. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146 [Abstract]
Trypanosoma spp., Crithidia fasciculata, C. oncopelti, thymidine phosphorylase activity, salvage pathways, inhibition of activity by suramin and ethidium

Trypanosoma cruzi
Al Chalabi, K.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), xv-xvi [Abstract]
6 spp. of Trypanosomatidae, thymidylate synthetase activity, ranks as potential target for chemotherapeutic attack since it is markedly more sensitive to suramin inhibition than isofunctional mammalian and bacterial enzymes

Trypanosoma cruzi
Al Chalabi, K.; and Gutteridge, W. E., 1977, Parasitology, v. 74 (3), 299-312
deoxythymidylate catabolism in some trypanosomatids

Trypanosoma cruzi, illus.
Alexander, J., 1975, J. Protozool., v. 22 (2), 237-240
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mice (exper.)
rats (exper.)
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hamsters (exper.)
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Pudney, M.; and Lanar, D., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 109-118
establishment of cell line from embryonic tissues of Trypanosoma vector, Triatoma infestans; potential use of these cells in culturing and studying Trypanosoma cruzi

Trypanosoma cruzi, illus.
Puigbo, J. J.; et al., 1977, Postgrad. Med. J., London (623), v. 53, 527-532
Trypanosoma cruzi, human, cardiac involvement, review of acute latent and chronic phases, diagnosis, pathology

Trypanosoma cruzi
Quintal, R. E.; and Polanco, G. G., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 176-178
Trypanosoma cruzi, studies show that feeding preferences of vector Triatoma dimidiata maculipennis have no bearing on relative lack of antibodies to Trypanosoma cruzi in humans in Yucatan

Trypanosoma cruzi, illus.
Ribeiro, R. D.; Belda Neto, F. M.; and Pereira Barretto, M., 1977, Rev. Brasil. Biol., v. 37 (1), 55-59
Trypanosoma cruzi, various strains, evolution in coelomic cavity of triatomines
+P[anstrongylus] megistus
+T[riatoma] vitticeps
+T. infestans
+R[hodnius] neglectus
(all exper.)

Trypanosoma cruzi
Ribeiro, R. D.; and Pereira Barretto, M., 1975, Rev. Brasil. Biol., v. 35 (2), 301-307
Trypanosoma cruzi, ten strains, isolated from man, wild mammals and triatomines, behavior in culture medium over one year, variations in presence of trypomastigotes, epimastigotes and amastigotes

Trypanosoma cruzi, illus.
Ribeiro, R. D.; and Pereira Barretto, M., 1977, Rev. Brasil. Biol., v. 37 (1), 195-200
Trypanosoma cruzi strain isolated from Caluromys lanatus ochropus, pathogenic for mice, marsupial strain easily cultivated and has high infectivity for triatomines tested: Teofilo Otoni, M. G., Brazil
P[anstrongylus] megistus
T[riatoma] vitticeps
T. infestans
R[hodnius] neglectus
(all exper.)

Trypanosoma cruzi
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma cruzi
Riou, G. F.; and Yot, P., 1977, Biochemistry, v. 16 (11), 2390-2396
Trypanosoma cruzi, kinetoplast DNA, heterogeneity of minicircles, renaturation kinetics of molecules, relationship between long linear molecules and minicircles

Trypanosoma cruzi
Rivero, I.; et al., 1975, Medicina, Buenos Aires, v. 35 (1), 73-78
Trypanosoma cruzi as complication in 4 cases of human acute leukemia and acute lymphoblastic leukemia probably resulting from immunosuppression

Trypanosoma cruzi
Roberson, E. L.; and Hanson, W. L., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 338 [Letter]
Trypanosoma cruzi-infected rats (exper.), passive transfer of immunity using spleen cells from sensitized donors evidenced by significantly lower parasitemia and reduced mortalities

Trypanosoma cruzi
Rogerson, G. W.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), xv [Abstract]
Trypanosoma cruzi, culture epimastigotes, blood trypomastigotes, intracellular amastigote stages, enzymes of energy metabolism, results suggest presence of functional tricarboxylic acid cycle in all stages

Trypanosoma cruzi
Rogerson, G. W.; and Gutteridge, W. E., 1977, J. Protozool., v. 24 (4), 42A [Abstract]
Trypanosoma cruzi, culture epimastigotes, blood trypomastigotes, and intracellular amastigotes, cytochromes aa_3, o, and b identified in all 3 forms, cytochrome c observed in culture forms

Trypanosoma cruzi
Rogerson, G. W.; and Gutteridge, W. E., 1977, Parasitology, v. 75 (2), xxvii [Abstract]
Trypanosoma cruzi, action of respiratory inhibitors on culture epimastigote, blood trypomastigote, and intracellular amastigote forms

Trypanosoma cruzi
Rohwedder, R. W.; et al., 1970, Bol. Chileno Parasitol., v. 25 (3-4), 106-110
Trypanosoma cruzi, human, recommendations for use of triatomid bugs in xenodiagnosis, best time for liquification and examination at peak parasitism

Trypanosoma cruzi
Rojas, A.; et al., 1973, Bol. Chileno Parasitol., v. 28 (1-2), 42-43
Trypanosoma cruzi, dogs and cats as reservoir hosts established by xenodiagnosis: Chile

Trypanosoma cruzi
Rowe, W. S.; and Whiting, D., 1977, Southwest. Vet., v. 30 (2), 190-191
dog (myocardium), case report: Texas

Trypanosoma cruzi
Rytel, M. W.; Rose, H. D.; and Stewart, R. D., 1973, Proc. Soc. Exper. Biol. and Med., v. 144 (1), 122-123
Trypanosoma cruzi, Plasmodium falciparum, P. vivax, humans, lack of circulating humoral interferon in serum of persons with acute illness

Trypanosoma cruzi, illus.
Sanabria, A.; and Aristimuno, J., 1975, Acta Cien. Venezolana, v. 26 (4), 128-133
Trypanosoma cruzi, mouse kidney, ultrastructure, pathology

Trypanosoma cruzi
Sanderson, C. J.; Lopez, A. F.; and Moreno, M. M. B., 1977, Nature, London (5618), v. 268, 340-341 [Letter]
Trypanosoma cruzi, isotopic technique for assaying killing of epimastigotes in which criterion of parasite death is release of macromolecular RNA, use of assay to investigate nature of effector cell killing T. cruzi in antibody-dependent complement-independent system, eosinophils show strong activity, whereas lymphoid K cells seem to have insignificant activity

Trypanosoma cruzi
Schenone, H.; et al., 1972, Bol. Chileno Parasitol., v. 27 (1-2), 11-14
experimental trials using lampit to treat human chronic Chagas disease, successful cures except for a few continued positive xenodiagnostic results suggesting an existing resistant strain of Trypanosoma cruzi: Chile

Trypanosoma cruzi
Schenone, H.; et al., 1975, Bol. Chileno Parasitol., v. 30 (3-4), 91-93
Trypanosoma cruzi, human, domestic animals, treatment trials with Ro 7-1051

Trypanosoma cruzi
Schlemper, B. R., jr.; Chiari, E.; and Brener, Z., 1977, J. Protozool., v. 24 (4), 544-547
Trypanosoma cruzi, 48-hr drug screening test based on inhibition of exponential growth of culture forms as evaluated by cell counter, comparison with data from treating inoculated mice, advantages of in vitro method; culture forms not suitable for screening additives to prevent T. cruzi transmission by banked blood

Trypanosoma cruzi
Schmunis, G. A.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 81-85
Trypanosoma cruzi, mice inoculated with epimastigotes or trypomastigotes, development of cell mediated immunity as shown by inhibition of macrophage migration

Trypanosoma cruzi
Schnur, L. F., 1977, J. Protozool., v. 24 (2), Suppl., 5A-6A [Abstract]
Leptomonas, Crithidia, Herpetomonas, L[eishmania] tropica, L[eishmania] tropica major, L[eishmania] tropica minor, L[eishmania] donovani, excreted metabolic factors (EF) and homologous antisera, cross-reactions, no EF apparently produced by Trypanosoma cruzi, suggests closer relationship between insect flagellates and leishmanias than between these and trypanosomes like Trypanosoma cruzi

Trypanosoma cruzi
Schottelius, J., 1976, Ztschr. Parasitenk., v. 50 (2), 182-183
Trypanosoma spp., effects of various lectins, protectins and blood group test sera

T[rypanosoma] cruzi
Schottelius, J., 1977, Tropenmed. u. Parasitol., v. 28 (2), 202-204
latex Chagas test performed on sera of Bartonella bacilliformis-infected humans, Haemobartonella muris-infected rats, and Eperythrozoon coccoides-infected mice, positive reaction observed only with rat sera, nature of this cross-reaction has not been clarified

Trypanosoma cruzi, illus.
Scorza, C.; and Scorza, J. V., 1972, J. Reticuloendothel. Soc., v. 11 (6), 604-616
Trypanosoma cruzi, rats, active phagocytosis of parasites by inflammatory macrophages in auricles of heart 11 days after infection, formation of phagosomes and of phagolysosomes and alterations in ingested parasites, role of acid phosphatase in alterations of phagocytized parasites

Trypanosoma cruzi
Seah, S. K. K.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 63-69
Trypanosoma cruzi, acute infection in exper. rhesus monkeys, vaccination with killed extract of cultural forms produced circulating antibodies and induced delayed hypersensitivity but monkeys still developed sustained parasitemia, changes in IgM and other hematologic responses

Trypanosoma cruzi, illus.
Seed, T. M.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 146-160
Trypanosoma cruzi culture forms, transmission and scanning electron microscopy, carbon replication and freeze-etching techniques

Trypanosoma cruzi, illus.
Segura, E. L.; et al., 1974, J. Protozool., v. 21 (4), 571-574
Trypanosoma cruzi epimastigotes from cultures, separation into fractions (nuclear, mitochondrial, lysosomal, microsomal, and cell-sap), DNA, RNA content and enzyme markers of fractions, subcellular localization of antigens by Ouchterlony tests in cell-sap and microsomal fractions

Trypanosoma cruzi
Segura, E. L.; et al., 1975, Medicina, Buenos Aires, v. 35 (5), 451-459
localization and determination of proteins and antigens in nuclear, mitochondrial, microsomal and cytoplasmic fractions of Trypanosoma cruzi

Trypanosoma cruzi, illus.
Segura, E. L.; et al., 1976, J. Parasitol., v. 62 (1), 131-133
Trypanosoma cruzi, homogenate prepared by compression-decompression technique, protective activity of various subcellular fractions, flagellar fraction most active, effective immunization more closely related to schedule than to dose, mice

Trypanosoma cruzi, illus.
Segura, E. L.; et al., 1977, J. Protozool., v. 24 (4), 540-543
Trypanosoma cruzi epimastigotes, method for isolation of membrane and flagellar fractions, description of ultrastructural characteristics and antigenic activity, protective activity against lethal challenge doses of trypomastigotes is strongly associated with the flagellar fraction

Trypanosoma cruzi
Shafii, A., 1977, N. York State J. Med., v. 77 (3), 418-419
Trypanosoma cruzi cardiomyopathy and resulting hemiplegia in immigrant woman from Ecuador, diagnostic confusion with myocardial infarction, clinical case report: New York

Trypanosoma cruzi
Shaw, J. J.; and Lainson, R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 168-169 [Letter]
human cutaneous leishmaniasis, immediate specific intradermal reaction with in vitro exo-antigen used for skin test, comparative trials with Trypanosoma cruzi antigens

Trypanosoma cruzi
Shaw, J. J.; Lainson, R.; and Minter, D. M., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 258
Trypanosoma cruzi, leishmaniasis, humans comparison of skin test reactions

Trypanosoma cruzi
da Silveira, J. F.; Zingales, B.; and Colli, W., 1977, Biochim, et Biophys. Acta, v. 481 (2), 722-733
Trypanosoma cruzi, particulate preparations from epimastigote forms, characterization of an adenylyl cyclase activity

Trypanosoma cruzi
Sims, P.; and Gutteridge, W. E., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 276 [Abstract]
Trypanosoma cruzi in vitro, action of SQ 18506 on epimastigote and blood trypomastigote forms

Trypanosoma cruzi
Sims, P.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), iii [Abstract]
Trypanosoma cruzi, SQ 18,506, no substantial inhibitory effects on energy metabolism or membrane function, concluded that nucleic acid synthesis is primary target of this drug

Trypanosoma cruzi
Sims, P.; and Newsom, R., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 20 [Demonstration]
Trypanosoma cruzi, ultrastructural changes induced by SQ 18,506

Trypanosoma cruzi
Sousa, O. E.; and Dawson, G. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 407-409
Trypanosoma cruzi, T. rangeli, T. minasense found in Saguinus geoffroyi, high rate of infection suggests marmosets are important natural hosts: Panama Canal Zone

Trypanosoma cruzi
Souza, M. do C.; et al., 1974, J. Protozool., v. 21 (4), 579-584
living culture forms of Leptomonas pessoai cross-protected mice against Trypanosoma cruzi challenge infection, circulating antibodies detected in immunized mice by immunodiffusion, passive haemagglutination, complement fixation, and antibody binding assay which cross-reacted with T. cruzi extracts, cellular immune response indicated by leucocyte migration inhibition

Trypanosoma cruzi
Souza, M. do C. M.; and Roitman, I., 1971, Rev. Microbiol., S. Paulo, v. 2 (4), 187-189
Trypanosoma cruzi, high degree of protection in mice previously immunized with vaccine of Leptomonas pessoai (living suspension)

Trypanosoma cruzi, illus.
de Souza, W., 1976, J. Microscopie, v. 25 (2), 189-190
Trypanosoma cruzi, membrane-microtubule associations, electron microscopy, possible function in changes in form

Trypanosoma cruzi, illus.
de Souza, W.; and Meyer, H., 1974, J. Protozool., v. 21 (1), 48-52
Trypanosoma cruzi in intracellular amastigote forms from tissue cultures, fine structure of nucleus, presence of spindle fibers in dividing nucleus

Trypanosoma cruzi
Spithakis, R., 1970, Marseille Med., v. 107 (11), 949-952
use of Trypanosoma cruzi extracts as adjunct to cobalt therapy for human malignant diseases, alleviation of undesirable side effects of radiation: France

Trypanosoma cruzi
Stagno, S.; and Hurtado, R., 1970, Bol. Chileno Parasitol., v. 25 (1-2), 90-91
Trypanosoma cruzi, early diagnosis of prenatal infections in newborn infants by demonstration of IgM antibodies using immunodiffusion and immunofluorescence

Trypanosoma cruzi
Stagno, S.; and Saavedra, P., 1972, Bol. Chileno Parasitol., v. 27 (3-4), 122-125
combined antigen of free trophozoites of Toxoplasma gondii and cultured forms of Trypanosoma cruzi used to simultaneously diagnose both diseases by the indirect immunofluorescence test

Trypanosoma cruzi
de Storni, P. D.; de Bolsi, F. L.; and Yanovsky, J. F., 1975, Medicina, Buenos Aires, v. 35 (1), 67-72
direct agglutination test in diagnosis of human Trypanosoma cruzi, evaluation of systematic use of 2-mercaptoethanol to eliminate nonspecific agglutinins

Trypanosoma cruzi
Sylvester, D.; and Krassner, S. M., 1976, Comp. Biochem. and Physiol., v. 55 (3B), 443-447
Trypanosoma cruzi epimastigotes, occurrence of proline oxidation, effect of proline and other amino acids on respiration, dissimilation of metabolic intermediates, results suggest presence of proline-glutamate interconversion pathway

Trypanosoma cruzi
Szarfman, A.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 19-24
evaluation of specificity of EVI factor observed in Trypanosoma cruzi cases using indirect fluorescent antibody test on sera from patients with other parasitic diseases, immunofluorescence of anti-skeletal muscle antibody from leishmaniasis

T[rypanosoma] cruzi
Szarfman, A.; et al., 1975, Clin. Immunol. and Immunopathol., v. 4 (4), 489-499
T[rypanosoma] cruzi, 6 children with congenital Chagas' disease, presence of circulating EVI and specific anti-T. cruzi antibodies, skeletal muscle biopsies showed immunoglobulin deposits, findings suggest possible pathogenic role of EVI antibody in lesions

Trypanosoma cruzi
Szarfman, A.; et al., 1975, Medicina, Buenos Aires, v. 35 (3), 245-250
specific agglutinins and immunoglobulin levels in congenital Chagas infections

Trypanosoma cruzi
Szarfman, A.; et al., 1977, J. Parasitol., v. 63 (1), 149
Trypanosoma cruzi, patients with acute Chagas disease, EVI antibodies, specific agglutinins, and IFA antibodies

Trypanosoma cruzi
Szarfman, A.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 453 [Letter]
Trypanosoma cruzi, evidence of tissue reacting antibodies in parasitemic children possibly originating in cross-reacting antigens between T. cruzi and human tissues

Trypanosoma cruzi
Szarfman, A.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (3), 333-341
Trypanosoma cruzi-infected mice (exper.), effect of previous inoculation by epimastigotes of Trypanosoma cruzi upon the resistance of mice against challenge with trypomastigotes; survival rate and parasitemia dependent on previous number of epimastigotes inoculated and on number of trypomastigotes used for challenge; evidence suggests that cell-mediated anamnestic response may be triggered by reinfection

Trypanosoma cruzi, illus.
Tanowitz, H.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 25-33
Trypanosoma cruzi, ultrastructural study of entry of epimastigotes and trypomastigotes into L-cells and normal and activated macrophages in vitro

Trypanosoma cruzi
Tanowitz, H. B.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (1), 19-28
metronidazole, suggested mechanism of antimicrobial action and selective toxicity for Entamoeba histolytica as opposed to mammalian cell types and Trypanosoma cruzi

Trypanosoma cruzi
Tanowitz, H.; et al., 1975, J. Parasitol., v. 61 (6), 1065-1069
Trypanosoma cruzi, identification of major ribosomal RNA species by velocity sedimentation and radioactive labeling techniques, differentiation from mammalian rRNA species, possible method for monitoring intracellular trypanosomal growth in vitro separately from its host cell

Trypanosoma (Schizotrypanum) cruzi
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

Trypanosoma cruzi
Taylor, E. C.; et al., 1977, J. Med. Chem., v. 20 (9), 1215-1218
Trypanosoma cruzi, 2,4-diaminocycloalka[g]-pteridines tested for activity as dihydrofolate reductase inhibitors

Trypanosoma cruzi, illus.
Teixeira, A. R. L., 1977, Advances Exper. Med. and Biol., v. 93, 243-280
Trypanosoma cruzi, immunoprophylaxis, review: life cycle in vector and host; clinical manifestations of Chagas' disease; mechanisms of resistance (natural and acquired immunity, humoral and cell-mediated); autoimmunity; antigenic structure; live vaccines; dead vaccines; perspectives for further studies

Trypanosoma cruzi
Teixeira, A. R. L.; and Santos-Buch, C. A., 1975, Immunology, v. 28 (3), 401-410
Trypanosoma cruzi, rabbits, strong delayed hypersensitivity skin reactions to 2 subcellular fractions from homogenates of suspensions of trypomastigote and amastigote forms, immediate reactions also seen, cell-mediated immunity assayed by experiments which established passive transfer, inhibition of blood mononuclear cell migration, and blast transformation by sensitized lymphocytes

T[rypanosoma] cruzi
Thaver, S.; and Draper, C. C., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 8 [Demonstration]
P[lasmodium] falciparum, P. vivax and T[rypanosoma] cruzi antibodies tested for reliability of epidemiologic field survey collection methods, use of chromatography paper for collecting capillary blood recommended

Trypanosoma cruzi
Thomaz, N.; and Goncalves da Costa, S. C., 1975, J. Protozool., v. 22 (3), 45A [Abstract]
Trypanosoma cruzi, differentiation in simplified medium

Trypanosoma cruzi
Toye, P. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 147 [Abstract]
Trypanosoma cruzi, isoenzyme variation in culture isolates

Trypanosoma cruzi
Toye, P. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 266 [Demonstration]
Trypanosoma rangeli, T. cruzi, T. lewisi, differentiation of enzyme patterns of culture forms

Trypanosoma cruzi
Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 1977, 1-15
17. Seminar on Trypanosomiasis (biochemistry and relevance to control; relevance of laboratory studies to tsetse control; field diagnosis; problems of land use and tsetse control)

Trypanosoma cruzi
Ucros, H.; Leon, W.; and Krassner, S. M., 1977, Exper. Parasitol., v. 41 (2), 410-414
Trypanosoma cruzi, Y and Costa Rica strains compared, effect of ethidium bromide on growth, dyskinetoplasty, and respiration

Trypanosoma (S.) cruzi Chagas, 1909, illus.
Urdaneta-Morales, S.; and de Alvarado, E., 1975, Acta Biol. Venez., v. 9 (1), 17-36
Trypanosoma cruzi Y strain, Aedes aegypti (exper.), enteric development from trypomastigote to epimastigote

Trypanosoma cruzi
Vattuone, N. H.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (3), 267-272
Trypanosoma cruzi, mice infected with epimastigotes or trypomastigotes of 3 different strains, cell mediated and humoral immune responses, characterization of antibodies according to criterion of 2-mercaptoethanol sensitivity

Trypanosoma cruzi
de la Vega, M. T.; Damilano, G.; and Diez, C., 1976, J. Parasitol., v. 62 (1), 129-130
chronic Trypanosoma cruzi-infected humans, positive leukocyte migration inhibition test with heart antigens, results suggest cell-mediated immune response against heart tissue could participate in mechanism of myocardial damage in Chagas' disease

Trypanosoma cruzi
Velasco Castrejon, O.; et al., 1970, Prensa Med. Mexicana, v. 35 (11-12), 438-440
Trypanosoma cruzi infection in 8-year old child, probable vector established as Triatoma barberi, epidemiologic study: Jalisco, Mexico

Trypanosoma cruzi
Vilaseca, G. C.; et al., 1966, Vox Sanguinis, v. 11 (6), 711-716
Trypanosoma cruzi, application of crystal violet dye to blood bank donor blood prevents transfusional transmission of Chagas disease

Trypanosoma cruzi
Vilches, A. M.; et al., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 279-284
Trypanosoma cruzi, mice, effect of heterologous anti-thymocyte serum upon course of infection, shorter survival time and higher parasitemia, no change in agglutination antibody titers, impaired resistance probably in detrimental effect of ATS upon cell-mediated immunity

Trypanosoma cruzi
Voller, A.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 431-437
human tropical parasitic diseases, comparison of enzyme-immunoassay and radio-immunoassay in detection of antibodies, both assays sensitive and reproducible and gave comparable results

Trypanosoma cruzi
Warton, A., 1975, J. Protozool., v. 22 (3), 69A-70A [Abstract]
Trypanosoma cruzi, microbodies (peroxisomes), ultrastructural localization using 3,3'-diaminobenzidine

Trypanosoma cruzi, illus.
Warton, A.; and Modlinska, M., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 24 (10), 1976, 611-613
Trypanosoma cruzi, blood forms, ultrastructural localization of peroxisomes following incubation with diaminobenzidine to demonstrate catalase, electron microscopy, possible metabolic role

Trypanosoma cruzi
Whittaker, F. H.; and Jarecka, L., 1974, Tr. Kentucky Acad. Sc., v. 35 (3-4), 76-78
Trypanosoma cruzi, Kentucky considered potential site though examination of raccoons and opossums was negative

Trypanosoma cruzi, illus.
Williams, D. M.; and Remington, J. S., 1977, Immunology, v. 32 (1), 19-23
Trypanosoma cruzi, capable of infecting and multiplying in human monocytes and monocyte-derived macrophages, activated macrophages inhibited intracellular multiplication of parasites and number of parasites in supernatants of activated monolayers markedly decreased

Trypanosoma cruzi
Williams, D. M.; Sawyer, S.; and Remington, J. S., 1976, J. Infect. Dis., v. 134 (6), 610-623
mice (exper.) whose macrophages were activated by Trypanosoma cruzi or Besnoitia jellisoni were significantly more resistant to intraperitoneal challenge with T. cruzi than were controls; activated macrophages were able to inhibit completely multiplication of T. cruzi, this suggests that the macrophage may play a major role in resistance to infection

Trypanosoma cruzi, illus.
Williams, G. D.; et al., 1977, J. Am. Vet. Med. Ass., v. 171 (2), 171-177
dogs (myocardium), clinical and pathological findings, epizootiology: central Texas counties

Trypanosoma cruzi, illus.
Wilson, H. R.; and Fairchild, D. G., 1977, Stain Tech., v. 52 (2), 105-111
modified Whipf's polychrome staining to demonstrate protozoans and tissue components in cutaneous lesions of infected animals

Trypanosoma cruzi
Winkelmann, E.; et al., 1977, Arzneimittel-Forsch., v. 27 (12), 2251-2263
Entamoeba histolytica, Trichomonas foetus, Trypanosoma spp., in golden hamsters and mice (both exper.), laboratory trials with more than 135 new 2-methyl-5-nitroimidazoles substituted in the 1-position and 1-methyl-5-nitroimidazoles substituted in the 2-position, comparison of activity with that of metronidazole and tinidazole

Trypanosoma cruzi
Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
Ammospermophilus (blood): Chaco Canyon National Monument, San Juan County, New Mexico
Neotoma fuscipes macrotis (blood): Griffith Park, Los Angeles County, California; Juniper Hills near Pearblossom, Los Angeles County, California
Triatoma rubida uhleri (feces): Tonto National Monument, near Roosevelt, Gila County, Arizona
T. recurva (feces): Tonto National Monument, near Roosevelt, Gila County, Arizona
Peromyscus truei montipinoris: Juniper Hills near Pearblossom, Los Angeles County, California

Trypanosoma cruzi
Wood, S. F., 1976, Prensa Med. Argent., v. 63 (16), 462-469
Trypanosoma cruzi, environmental changing temperature as influence and regulator of life cycle in both vertebrate and invertebrate hosts

Trypanosoma cruzi, illus.
Yoneda, S.; Kimura, E.; and Castellani, B. R., 1977, Experientia, v. 33 (9), 1201-1202
Trypanosoma cruzi, Ro 7-1051, deleterious effect in vitro on intracellular and extracellular forms represented by nuclear pyknosis, fragmentation, and lysis of parasites, and by reduced ability to infect cells

Trypanosoma cruzi
Yoshida, N., 1975, J. Protozool., v. 22 (1), 128-130
Trypanosoma cruzi, macromolecule-free partially defined medium allowing prolonged cultivation by serial transfers and growth comparable to that obtained in more complex media

Trypanosoma cruzi, illus.
Zavala-Velazquez, J.; Rodriquez-Lopez, M. H.; and Baqueiro-Diaz, A., 1975, Patologia, v. 13 (3), 355-363
Trypanosoma cruzi in humans, 4 cases reported from Yucatan area, clinical case reports, problems in treatment with nifurtimox, infected triatome vectors discovered in area: Mexico

Trypanosoma cruzi
Zeledon, R.; et al., 1973, J. Med. Entom., v. 10 (4), 363-370
Trypanosoma cruzi, habitat and blood sources of vector, Triatoma dimidiata: Costa Rica

Trypanosoma cruzi
Zeledon, R.; and Ponce, C., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 414-415 [Letter]
human chronic Chagas disease, immediate skin test reaction using soluble protein antigen from parasites grown in culture, cross-reaction from cutaneous leishmaniasis differentiated by Montenegro test

Trypanosoma cyclops
Else, J. G.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 170-171
trypanosomiasis, antigens from various T. spp. gave inconclusive serologic results in assessing presence of infection among Orang Asli aborigines: Malaysia

Trypanosoma cyclops, illus.
Heywood, P.; Weinman, D.; and Lipman, M., 1974, J. Protozool., v. 21 (2), 232-238
Trypanosoma cyclops maintained in blood agar medium, ultrastructure, structure of pigment bodies, compared with T. cyclops grown in hemoglobin-free medium and a non-pigmented Trypanosoma sp.

Trypanosoma cyclops
Kierszenbaum, F.; and Weinman, D., 1977, Immunology, v. 32 (2), 245-249
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dogs
horses
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Proechymys sp. (exper.)
all from Llanos Orientales, Colombia, South America

Trypanosoma evansi
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Trypanosoma evansii
Woo, P. T. K.; and Rogers, D. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 319-326
Trypanosoma spp., statistical study of sensitivity of haematocrit centrifuge technique for detecting trypanosomes in blood, evaluation of relationship of probability of detection of trypanosome to trypanosome density

Trypanosoma everetti sp. nov., illus.
Molyneux, D. H., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 219-222
Estrilda troglodytes troglodytes: Dumbi, near Zaria, Nigeria

Trypanosoma everetti
Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 148-153
Pytelia melba
Phyllastrephus fischeri placidus
(blood of all): all from Kenya

Trypanosoma everetti Molyneux
Peirce, M. A.; and Cheke, A. S., 1977, J. Protozool., v. 24 (2), Suppl., 33A [Abstract]
Hypsipetes crassirostris moheliensis: Comoro Islands

Trypanosoma (Herpetosoma) evotomys kudickei, Krampitz, 1961
Hommel, M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 3 [Demonstration]
Trypanosoma evotomys kudickei, developmental behavior of trypanosomes in culture

Trypanosoma fringillinarum
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Sylvia borin (blood): Bramham, Yorkshire, England

Trypanosoma gambiense
Ackerman, S.; and Seed, J. R., 1973, J. Protozool., v. 20 (4), 511
immunosuppression in Trypanosoma gambiense-infected Microtus montanus detected by reduced immune response to human red blood cells and by immunocytoadherence experiments

Trypanosoma gambiense
Aiyedun, B. A.; and Amodu, A. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 611-612 [Letter]
Trypanosoma gambiense, rabbits, cell-mediated hypersensitivity, transferable with spleen cells

Trypanosoma gambiense
Aiyedun, B. A.; and Amodu, A. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 67-71
trypanosomiasis, albino rats (exper.), humans, diagnosis, capillary-tube precipitation test (known as capillary-tube agglutination test), rapid and specific technique that could be used in field survey work

T[rypanosoma] gambiense
Benard, J.; and Riou, G. F., 1975, J. Protozool., v. 22 (3), 75A-76A [Abstract]
T[rypanosoma] spp., ellipticine derivatives, mode of action

Trypanosoma gambiense
Bertrand, E.; et al., 1974, Medecine Trop., v. 34 (4), 485-494
Trypanosoma gambiense, recommended treatment methods for various clinical entities in human trypanosomiasis

Trypanosoma gambiense
Buyst, H., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 163-164
congenital Trypanosoma rhodesiense infection in premature baby successfully treated with suramin and supportive measures; guidelines for similar therapy for T. gambiense infections: Zambia

Trypanosoma gambiense
Davies, J. N. P., 1968, Med. History, v. 12 (2), 200-204
early speculations on the etiology of African sleeping sickness and its vector

Trypanosoma gambiense
Duflo, B., 1975, Medecine Interne, v. 10 (10), 447-453
human cardiac complications of tropical parasitoses, pathologic findings

Trypanosoma gambiense
Else, J. G.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 170-171
trypanosomiasis, antigens from various T. spp. gave inconclusive serologic results in assessing presence of infection among Orang Asli aborigines: Malaysia

Trypanosoma gambiense
Emejuaiwe, S. O., 1974, Bull. Epizoot. Dis. Africa, v. 22 (1), 41-50
Trypanosoma gambiense, passive immunization of mice with 19S and 7S immunoglobulin fractions derived from immunized rabbits gave transient protection to mice, however, 7S immunoglobulin fractions of sera from rabbits infected with live trypanosomes gave complete protection to mice

Trypanosoma gambiense
Fromentin, H., 1972, Protistologica, v. 8 (3), 365-368
Trypanosoma gambiense, T. evansi, normal vs. dyskinetoplastic strains, oxygen consumption compared

Trypanosoma gambiense
Fromentin, H.; and Wiart, J., 1975, Bull. Soc. Path. Exot., v. 68 (3), 306-312
2 specific antigenic fractions of Trypanosoma gambiense demonstrated by tannic acid hemagglutination when applied to experimentally infected rodent sera

Trypanosoma gambiense
Gray, A. R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 269-270 [Abstract]
Trypanosoma gambiense, antigenic variation in cyclically-transmitted isolates, similarities in sequences in which variant antigens developed

Trypanosoma gambiense
Gray, A. R., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 150-151 [Abstract]
Trypanosoma gambiense, isolates from different countries in Africa show close antigenic relationship

Trypanosoma gambiense
Gray, A. R., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 131-138
Trypanosoma gambiense transmitted by tsetse flies to rabbits and monkeys, developmental patterns of agglutinogenic antigens in cyclically transmitted isolates

Trypanosoma gambiense
Greenwood, B. M.; and Whittle, H. C., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 390-394
Trypanosoma gambiense, human, no evidence of disseminated intravascular coagulation, moderate degree of thrombocytopenia, raised levels of fibrin degradation products; failure to produce thrombocytopenia in vitro

Trypanosoma gambiense
Greenwood, B. M.; Whittle, H. C.; and Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (6), 846-850
Trypanosoma gambiense, human, immunosuppression, cell-mediated and humoral immunity both impaired

Trypanosoma gambiense
Hansen, B.; and Read, C. P., 1973, J. Protozool., v. 20 (4), 501
Trypanosoma gambiense, amino acid transport

Trypanosoma gambiense
Hawking, F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 513-520
Trypanosoma gambiense, T. rhodesiense, T. brucei, 60 polymorphic strains from East and West Africa, in vivo test of sensitivity or resistance to human plasma, application to infectivity for man

Trypanosoma gambiense
Jackson, P. R., 1975, J. Parasitol., v. 61 (5), 963-965
forced-flow DEAE-cellulose column for isolation of bloodstream trypanosomes

Trypanosoma gambiense, illus.
Jackson, P. R.; et al., 1977, J. Parasitol., v. 63 (4), 593-598
Plasmodium yoelii, Trypanosoma gambiense, and Trypanosoma equiperdum, cytofluorograf detection of parasites by laser excited fluorescence of stained rodent blood

Trypanosoma gambiense
Janssens, P. G., 1973, Medecine Afrique Noire, v. 20 (11), 847-851
Trypanosoma gambiense, human, indications and guides for mass therapy

Trypanosoma gambiense
Lindsley, H. B.; Kysela, S.; and Steinberg, A. D., 1974, J. Immunol., v. 113 (6), 1921-1927
Trypanosoma rhodesiense, T. gambiense, humans, development of antibodies to nucleic acids, possible pathophysiological relationship to renal disease remains to be determined

Trypanosoma gambiense
Minoprio, J. L., 1972, Prensa Med. Argent., v. 59 (8), 293-308
human Chagas disease, pathologic findings, comparison with characteristics of other Trypanosoma spp. and Leishmania spp., extensive review

Trypanosoma gambiense
Miyata, A., 1973, Nettai Igaku (Trop. Med.), v. 15 (4), 204-213
Entamoeba histolytica, Toxoplasma gondii, Plasmodium berghei, Trypanosoma gambiense, preservation by freezing, evaluation of precooling and cryo-protective solutions, length of survival; T. gondii (Beverley strain) unable to survive freezing

Trypanosoma gambiense
Murray, M.; and Urquhart, G. M., 1977, Advances Exper. Med. and Biol., v. 93, 209-241
African trypanosomiasis, immunoprophylaxis, review: cattle and sheep, laboratory animals; chemotherapeutic agents and immunity, nature of protective antigen, effector mechanisms in protection, manipulation of host resistance

Trypanosoma gambiense
Olowe, S. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 57-59
Trypanosoma gambiense, fatal prenatal infection of infant girl presenting with symptoms of meningitis, attempted suramin therapy probably aggravated additional congenital complication, clinical case review: Nigeria

Trypanosoma gambiense, illus.
Ono, T.; and Inoki, S., 1975, Biken J., v. 18 (4), 257-265
Trypanosoma gambiense, mice, hydroxystilbamadine treatment, inhibition of kinetoplastic duplication without influence on nuclear or cytoplasmic duplication; disorganization and disappearance of kinetonucleus and its enveloping membrane shown by electron microscopy; failure to establish akinetoplastic clone probably due to inability of these forms to multiply

Trypanosoma gambiense, illus.
Ono, T.; and Inoki, S., 1976, Biken J., v. 19 (2), 63-69
p-rosaniline-sensitive and -resistant strains of Trypanosoma gambiense in mice, interaction of p-rosaniline, furazolidon on kinetoplast; development of resistance, appearance of akinetoplastic forms

Trypanosoma gambiense, illus.
Ono, T.; and Nakabayashi, T., 1976, Biken J., v. 19 (4), 171-177
Trypanosoma gambiense, morphology following neocarzinostatin injection in mice (exper.), anucleate forms, unusual structures, electron microscopy

Trypanosoma gambiense
Rafael, A. G., 1975, An. Serv. Vet., Mocambique (20-21), 1972-1973, 331-350
Trypanosoma spp., pathology, economic importance, treatment, vector control, extensive review: Africa

Trypanosoma gambiense
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma gambiense
Ruitenberg, E. J.; and Buys, J., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 31-36
Trypanosoma brucei, enzyme-linked immunosorbent assay (ELISA) for serodiagnosis of human African sleeping sickness, comparison tests with immunofluorescence technique showed good results in rabbits (exper.) and serum from infected humans, cross-reactions only in person with Leishmania antibodies, possible application to epidemiologic surveys

Trypanosoma gambiense
Schmidt, G.; Walter, R. D.; and Koenigk, E., 1975, Tropenmed. u. Parasitol., v. 26 (1), 19-26
Trypanosoma gambiense, purine nucleoside hydrolase obtained from parasite, purification and characterization, inhibitory effects of substrate analogs, salvage synthesis of adenosine monophosphate

Trypanosoma gambiense
Smith, G. J., 1977, Ann. Trop. Med. and Parasitol., v. 71 (4), 391-399
human African trypanosomiasis, historical review of the 1902 expedition to Senegambia by staff of the Liverpool School of Tropical Medicine, excerpts from letters of Dr. J. L. Todd

Trypanosoma gambiense
Soltys, M. A.; and Woo, P. T. K., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 143-145
Trypanosoma gambiense, human, diagnosis, indirect charcoal-agglutination test, promising results

Trypanosoma gambiense, illus.
Takayanagi, T.; et al., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (2), 165-167
Trypanosoma gambiense, electron microscopy of parasite to show flagellum and ferritin particles concentrated on it, soluble antigen concentrated on flagellum specifically at flagellar membrane and the lattice-like intraflagellar structure

Trypanosoma gambiense
Takayanagi, T.; Kambara, H.; and Enriquez, G. L., 1973, Southeast Asian J. Trop. Med. and Pub. Health, v. 4 (1), 27-31
Trypanosoma gambiense, O type maintained in rats, isolation and purification of agglutination antigen

Trypanosoma gambiense
Takayanagi, T.; and Nakatake, Y., 1976, Exper. Parasitol., v. 39 (2), 234-243
Trypanosoma gambiense, mice, immunologic responses to infection in thymectomized lethally-irradiated recipients of passively transferred thymic cells sensitized with parasitic antigens in vivo, enhanced agglutinin production and protection and phagocytosis

Trypanosoma gambiense
Takayanagi, T.; and Nakatake, Y., 1977, Exper. Parasitol., v. 42 (1), 21-26
Trypanosoma gambiense, loss of binding activity by which rat antibody is bound to macrophages as result of removing Fc portion of IgG by enzymatic digestion, avidity of antibody for heterologous macrophages

Trypanosoma gambiense, illus.
Takayanagi, T.; Nakatake, Y.; and Kato, H., 1977, Exper. Parasitol., v. 43 (1), 196-202
Trypanosoma gambiense, agglutination and binding of trypanomastigotes to macrophages in terms of different antigen-antibody ratios

Trypanosoma gambiense, illus.
Takayanagi, T.; Nakatake, Y.; and Kato, H., 1977, Exper. Parasitol., v. 43 (2), 414-422
Trypanosoma gambiense, antigens involved in immune binding to mammalian host macrophages followed by phagocytosis

Trypanosoma gambiense
Walter, R. D., 1976, Biochim. et Biophys. Acta, v. 429 (1), 137-146
Trypanosoma gambiense, protein kinase, partial purification and characterization, nucleoside-dependent but cyclic AMP-independent

Trypanosoma gambiense
Whittle, H. C.; et al., 1977, Am. J. Trop. Med. and Hyg., v. 26 (6, part 1), 1129-1134
Trypanosoma gambiense, measurement of serum and cerebrospinal fluid IgM and correlation with antibody levels measured by enzyme-linked immunosorbent assay, tests in combination superior to each test done singly, immunoglobulin levels in spinal fluid helpful in diagnosing while antibody levels were of no use for diagnosis or prognosis

Trypanosoma gerrhonoti, illus.
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Gerrhonotus multicarinatus (blood): central California

Trypanosoma grayi
Molyneux, D. H., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 294 [Abstract]
Trypanosoma bouffardi development in Glossina suggests that Foster's T. grayi may have been an avian trypanosome instead, need for awareness of avian trypanosomes in Glossina spp.

Trypanosoma (Megatrypanum) heliosciuri sp. n., illus.
Ranque, P., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 255-263
Heliosciurus gambianus gambianus (blood):
Thies region of Senegal, West Africa
white mice (exper.)
hamsters (exper.)

Trypanosoma leonidasdeanei, illus.
Christensen, H. A.; and Herrer, A., 1976, J. Med. Entom., v. 12 (4), 477-478
positive xenodiagnostic trials with Lutzomyia vespertilionis feeding on trypanosome-infected Saccopteryx bilineata and Peropteryx macrotis, trypanosome from S. bilineata tentatively identified as Trypanosoma leonidasdeanei, potential of sand fly as vector, localization of parasite in alimentary tract: Panama

Trypanosoma lewisi
Aboko-Cole, G. F.; and Lee, C. M., 1975, Ztschr. Parasitenk., v. 47 (2), 71-77
Trypanosoma rhodesiense, T. lewisi, rats on normal or folic acid deficient diets, mitotic activity in liver; folate content of trypanosomes and their metabolic products

Trypanosoma lewisi
Al Chalabi, K.; and Gutteridge, W. E., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146 [Abstract]
Trypanosoma spp., Crithidia fasciculata, C. oncopelti, thymidine phosphorylase activity, salvage pathways, inhibition of activity by suramin and ethidium

Trypanosoma lewisi
Al Chalabi, K.; and Gutteridge, W. E., 1976, Parasitology, v. 73 (2), xv-xvi [Abstract]
6 spp. of Trypanosomatidae, thymidylate synthetase activity, ranks as potential target for chemotherapeutic attack since it is markedly more sensitive to suramin inhibition than isofunctional mammalian and bacterial enzymes

Trypanosoma lewisi
Al Chalabi, K.; and Gutteridge, W. E., 1977, Parasitology, v. 74 (3), 299-312
deoxythymidylate catabolism in some trypanosomatids

Trypanosoma lewisi
Bagster, I. A.; and Parr, C. W., 1973, Nature (5415), v. 244, 364-366
trypanosome identification by electrophoresis of soluble enzymes

Trypanosoma lewisi, illus.
Cherian, P. V.; and Dusanic, D. G., 1977, Exper. Parasitol., v. 43 (1), 128-142
Trypanosoma lewisi, bloodstream forms, surface antigens examined at ultrastructural level with direct and indirect ferritin-conjugated antibody techniques, formation of filopodia and evidence for their immunological reactivity

Trypanosoma lewisi
Cornford, E. M.; et al., 1973, J. Protozool., v. 20 (4), 503
Trypanosoma lewisi, in vivo and in vitro effects of hyperglycemic and hypoglycemic agents

Trypanosoma lewisi
Cornford, E. M.; et al., 1975, Gen. Pharmacol., v. 6 (4), 315-323
Trypanosoma lewisi, effects of altered blood glucose levels in vivo (rats), effects of hypoglycemic and hyperglycemic agents on glucose metabolism in vitro

Trypanosoma lewisi
Cornford, E. M.; Freeman, B. J.; and MacInnis, A. J., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (3), 238-243
Trypanosoma lewisi in rats and T. duttoni in mice, possible relationships between circadian rhythms of trypanosomes and physiological variables of hosts

Trypanosoma lewisi, illus.
D'Alesandro, P. A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 691-738
nonpathogenic trypanosomes of rodents, immunology, review

Trypanosoma lewisi
D'Alesandro, P. A., 1976, J. Protozool., v. 23 (2), 256-261
Trypanosoma lewisi, specificity of agglutinins elicited by 2 antigenic variants studied with classical adsorption and agglutination methods and newer immunoelectroadsorption technic

Trypanosoma lewisi
Dusanic, D. G., 1974, J. Protozool., v. 21 (3), 422 [Abstract]
Trypanosoma duttoni from mice grown in vitro at 37°C., lack of marked ablastin activity in homologous mouse antisera, reproductive activity completely inhibited by heterologous antisera from T. lewisi-infected rats

Trypanosoma lewisi, illus.
Dwyer, D. M., 1975, J. Cell Sc., v. 19 (3), 621-644
Trypanosoma lewisi, bloodstream and culture forms, differential polycation-induced cell agglutination, fine-structure cytochemistry, results indicate complex polysaccharides are present in surface membranes and cell surface coat

Trypanosoma lewisi
Dwyer, D. M., 1976, J. Protozool., v. 23 (2), 16A-17A [Abstract]
Trypanosoma lewisi, adsorbed rat serum proteins in surface coat

Trypanosoma lewisi
Dwyer, D. M., 1976, Proc. Nat. Acad. Sc., v. 73 (4), 1222-1226
Trypanosoma lewisi, bloodstream forms isolated from rats, ultrastructural and immunologic evidence of avidly bound host serum proteins in surface coat, not present in intact culture or trypsinized bloodstream forms but reacquired after incubation in heterologous host serum proteins

Trypanosoma lewisi
El-On, J.; and Greenblatt, C. L., 1973, J. Protozool., v. 20 (4), 531
Trypanosoma lewisi, T. acomys, growth in vitro at 37°C. in culture media with mammalian cells, rats partially protected against T. lewisi after receiving 6 injections of concentrated culture supernatant fluid

Trypanosoma lewisi
El-On, J.; and Greenblatt, C. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 19 [Demonstration]
Trypanosoma lewisi, rats, increased susceptibility to infection when given cyclophosphamide (CyI-rats) as immuno-suppressive, possible role of exoantigens in development of anemia, precipitating antibodies to Trypanosoma lewisi in rabbits inoculated with plasma from CyI rats whether infected or not

Trypanosoma lewisi
El-On, J.; and Greenblatt, C. L., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 19-20 [Demonstration]
Trypanosoma lewisi, production of exoantigens during cultivation at 37°C, partial protection against infection in rats injected with concentrated culture supernatant fluid

Trypanosoma lewisi, illus.
El-On, J.; and Greenblatt, C. L., 1977, Exper. Parasitol., v. 41 (1), 31-42
Trypanosoma lewisi, T. acomys, T. cruzi, method for cultivation with mammalian tissue, results not good with T. equiperdum and T. congolense

Trypanosoma lewisi
Else, J. G.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 170-171
trypanosomiasis, antigens from various T. spp. gave inconclusive serologic results in assessing presence of infection among Orang Asli aborigines: Malaysia

Trypanosoma lewisi
Ferrante, A.; and Jenkin, C. R., 1977, Austral. J. Exper. Biol. and Med. Sc., v. 55 (3), 275-280
Trypanosoma lewisi, dividing and adult forms share common antigens and persistence of adult forms in circulation of rats immune to reinfection not due to antigenic change but due to the presence of surface immunoglobulins

Trypanosoma lewisi
Freeman, B. J.; et al., 1973, J. Protozool., v. 20 (4), 512
effects of Trypanosoma lewisi on concurrent infections with Hymenolepis diminuta, rats

Trypanosoma lewisi, illus.
Fuson, E. W.; Jones, A. W.; and Murray, F. A., 1976, Exper. Parasitol., v. 40 (2), 225-231
Trypanosoma lewisi, in vitro maintenance of blood stream form, cultured trypanosomes are susceptible to both trypanocidal antibodies and show longer period of susceptibility to ablastin than previously reported

Trypanosom[a] (Herpetosoma) lewisi
Godfrey, D. G.; and Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 260 [Abstract]
bloodstream trypanosomes of 4 different subgenera, relative activities of alanine vs. aspartate aminotransferases were characteristic of each subgenus

Trypanosoma lewisi, illus.
Grewal, M. S., 1975, Research Bull. Panjab Univ., v. 23 (3-4), 1972, 119-124
Rattus rattus: Chandigarh, India

T[rypanosoma] lewisi
Hommel, M.; and Miltgen, F., 1973, J. Protozool., v. 20 (4), 527
Trypanosoma blanchardi, T. rabinowitschae, adaptation to mice by passage through cultures and then sarcomatous mice, strong cross immunity between T. musculi and mouse-adapted strains; adaptation of T. lewisi to mice using same method was not possible; this new culture system was used to cultivate T. brucei

Trypanosoma lewisi
Jarvinen, J. A.; and Dalmasso, A. P., 1976, Infect. and Immun., v. 14 (4), 894-902
Trypanosoma lewisi, rats, role of complement in control and termination of infection is not a major one

Trypanosoma lewisi
Khachoian, V. I.; et al., 1976, Biol. Zhurnal Armenii, v. 29 (3), 87-89
Trypanosoma lewisi, rats, mitotic activity of bone marrow and spleen cells

Trypanosoma lewisi
Khachoian, V. I.; Arakelian, L. A.; and Balaian, D. E., 1976, Biol. Zhurnal Armenii, v. 29 (12), 31-34
Trypanosoma lewisi, exper. rats with copper-deficient diet, hypocupremia, acute invasive process ending in death; nonspecific resistance lowered

Trypanosoma lewisi
Leal Medina, L. J., 1974, Rev. Med. Vet. y Parasitol., Maracay, v. 25 (1-8), 1973-1974, 15-31
Trypanosoma lewisi, rats with and without cortisone, changes in ultrastructure of monocytes during infection; infection does not produce active macrophages or forms transitional between monocytes and macrophages

Trypanosoma lewisi
Lee, C. M.; George, Y. G.; and Aboko-Cole, G. F., 1977, Internat. J. Biochem., v. 8 (7), 525-529
Trypanosoma lewisi in iron-deficient rats, parasitemias, trypanosome cell size and antibody formation, host body weight gains and food consumption

Trypanosoma lewisi
Lopez, T., 1975, J. Protozool., v. 22 (3), 23A [Abstract]
Trypanosoma lewisi, T. mega, culture forms, DNA contents

Trypanosoma lewisi
Meerovitch, E.; and Ackerman, S. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 417 [Letter]
pre-existing Trichinella spiralis infection in rats (exper.) greatly reduced by level of Trypanosoma lewisi parasitemia, possible cell-mediated immune reaction

Trypanosoma lewisi
Mishra, G. S.; and Gonzalez, J. P., 1975, Arch. Inst. Pasteur Tunis, v. 52 (1-2), 71-87
Rattus norvegicus (sang): Tunis, Tunisia
rat blanc (exper.)

Trypanosoma lewisi, illus.
Miyata, A.; and Tsukamato, M., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 113-130
house rats: Palawan Island, the Philippines

Trypanosoma lewisi, illus.
Muehlpfordt, H.; and Garcia, W., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (2), 133-142
Trypanosoma lewisi (London) strain reidentified as a strain of T. cruzi

Trypanosoma lewisi
Newton, B. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 146-147 [Abstract]
Trypanosoma brucei sspp., amplification of kinetoplast DNA probably occurs in culture forms, much larger amounts of DNA in culture forms than in bloodstream forms; no significant differences found in T. lewisi forms

Trypanosoma lewisi
Nielsen, K.; et al., 1977, Exper. Parasitol., v. 43 (1), 153-160
Trypanosoma lewisi, initial biochemical characterization of complement-activating components

Trypanosoma lewisi
Nielsen, K.; and Sheppard, J., 1977, Experientia, v. 33 (6), 769-771
Trypanosoma congolense, T. lewisi, activation of complement, possible roles in pathology of disease

Trypanosoma lewisi
Owen, D., 1976, Lab. Animals, v. 10 (3), 271-278
Rattus norvegicus: Carshalton

Trypanosoma lewisi
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma lewisi
Shakespeare, P. G.; and Trigg, P. I., 1974, J. Protozool., v. 21 (3), 452 [Abstract]
Trypanosoma b. brucei, T. congolense, T. lewisi, bloodstream forms, glucose utilization

Trypanosoma lewisi
Sheppard, J.; et al., 1977, Canad. Fed. Biol. Soc., Programme and Proc. 20. Ann. Meet., v. 20, 120 [Abstract]
Trypanosoma congolense, T. lewisi, complement activating factors, one of which may be a hexose polymer

Trypanosoma lewisi
Shmuel, Z.; Golenser, J.; and Spira, D. T., 1975, J. Protozool., v. 22 (3), 73A [Abstract]
rats infected with Trypanosoma lewisi at different stages in course of Plasmodium berghei infection, influence on parasitemia of each

Trypanosoma lewisi
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Trypanosoma lewisi
Styles, T. J., 1976, J. Protozool., v. 23 (2), 31A [Abstract]
Trypanosoma lewisi, rats, immunization by implantation of organisms into diffusion chambers in peritoneal cavity

Trypanosoma lewisi
Styles, T. J.; et al., 1974, J. Protozool., v. 21 (3), 422 [Abstract]
Trypanosoma lewisi, rats, results tend to indicate that T cells and their products are not of major significance in immune response

Trypanosoma lewisi
Styles, T. J.; and Krautter, F. H., 1976, J. Protozool., v. 23 (2), 16A [Abstract]
Trypanosoma lewisi, tissue forms studied with fluorescent antibody technique

Trypanosoma lewisi
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

Trypanosoma lewisi
Tizard, I. R.; et al., 1977, Experientia, v. 33 (7), 901-902
Trypanosoma congolense, hemolytic activity is due to presence of free fatty acids generated by action of phospholipase A on endogenous phosphatidyl choline, some lysolecithin also contributes to lytic activity, T. lewisi is devoid of phospholipase A and does not generate free fatty acids and is therefore non-hemolytic

Trypanosoma lewisi
Tizard, I. R.; and Holmes, W. L., 1976, Experientia, v. 32 (12), 1533-1534
Trypanosoma congolense, generation of toxic activity considered to be of potential significance in pathogenesis (phospholipase-like activity capable of mediating lysis of both nucleated cells and erythrocytes as well as acute inflammatory response on intradermal inoculation), Trypanosoma lewisi produced no significant hemolysis

Trypanosoma lewisi
Tizard, I. R.; and Ringleberg, C. P., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (6), 885 [Letter]
Trypanosoma congolense, release of biologically active material (capable of causing inflammatory reaction) from dying trypanosomes in diffusion chambers in rats, not observed with T. lewisi, suggests that hypothesis of trypanosomal toxins should be reconsidered

Trypanosoma lewisi
Toye, P. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 266 [Demonstration]
Trypanosoma rangeli, T. cruzi, T. lewisi, differentiation of enzyme patterns of culture forms

Trypanosoma lewisi
Uriniuk, V. M.; Sukhareva-Nemakova, N. N.; and Silaev, A. B., 1975, Vestnik Moskov. Univ., s. (6), Biol. (4), 42-49
Trypanosoma lewisi cultured with active aeration and sodium malonate, increase in total lipid content and in unsaturated fatty acids; increased phospholipids, decreased sterol and monoglycerides

Trypanosoma (Herpetosoma) lewisi
Wilson, V.C.L.C.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 271-272 [Abstract]
Trypanosoma musculi, persistence in vasa recta of kidney of immune mice; T. lewisi did not persist in immune rats

Trypanosoma lewisi, illus.
Wolstenholme, D. R.; et al., 1974, J. Protozool., v. 21 (5), 622-631
Crithidia, kinetoplast DNA, structure, replication, informational capacity, physicochemical and electron microscope studies, review with some unpublished results

Trypanosoma loricatum (Mayer, 1843)
Wallace, F. G., 1975, J. Protozool., v. 22 (3), 18A [Abstract]
Trypanosoma rotatorium, differentiation from T. loricatum
Rana esculenta: Yugoslavia

Trypanosoma lotae sp. n., illus.
Smirnova, L. I., 1970, Parazitologiia, Leningrad, v. 4 (3), 296-297
changes in host blood picture
Lota lota (blood): Rybinskoye Vodokhranilishche

Trypanosoma mathisi
Zeiniev, N. R., 1975, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk (4), 86-89
[Hirundo rustica]: north-east Azerbaidzhan

T[rypanosoma] mega
Barker, D. C.; and Brack, C., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 269 [Abstract]
Trypanosoma brucei, T. mega, Crithidia fasciculata, dark-field electron microscopy of kinetoplast DNA

Trypanosoma mega
Borst, P.; et al., 1977, Exper. Cell Research, v. 110 (1), 167-173
Trypanosoma mega, maxi-circles in kinetoplast DNA

Trypanosoma mega
Hill, G. C.; and Degn, H., 1977, J. Protozool., v. 24 (4), 563-565
Trypanosoma mega, steady-state kinetics of terminal oxidases

T[rypanosoma] mega
Linstead, D. J.; Klein, R. A.; and Eldridge, M. V., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 267-268 [Abstract]
Trypanosoma mega, T. brucei, Crithidia fasciculata, presence of 3 enzymes capable of carbon dioxide fixation in culture as source of carbon for metabolism

Trypanosoma mega
Lipschik, G.; et al., 1976, J. Protozool., v. 23 (2), 30A [Abstract]
trypanosomatids, polyamines and Mg++ content

Trypanosoma mega
Lopez, T., 1975, J. Protozool., v. 22 (3), 23A [Abstract]
Trypanosoma lewisi, T. mega, culture forms, DNA contents

Trypanosoma mega
Newton, B. A.; Steinert, M.; and Borst, P., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 259-260 [Abstract]
Trypanosoma mega, Crithidia luciliae, differentiation by hybridization of complementary RNA with kinetoplast DNA

Trypanosoma minasense
Deane, L. M.; da Silva, J. E.; and Loures Filho, L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 424-425 [Letter]
Trypanosoma minasense in Callithrix penicillata jordani and C. geoffroyi, circadian rhythm in parasitemia: Brazil

Trypanosoma minasense, probably, illus.
King, N. W., jr., 1976, Scient. Publication (317), Pan Am. Health Organ., 169-198

Trypanosoma minasense
Sousa, O. E.; and Dawson, G. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 407-409
Trypanosoma cruzi, T. rangeli, T. minasense found in Saguinus geoffroyi, high rate of infection suggests marmosets are important natural hosts: Panama Canal Zone

Trypanosoma murmanensis Nikitin, 1927, illus.
Khan, R. A., 1976, Canad. J. Zool., v. 54 (11), 1840-1849
Trypanosoma murmanensis, life cycle (more rapid at higher temperatures), transmission experiments
Gadus morhua (blood) (nat. and exper.)
Johanssonia sp. (digestive tract, proboscis) (exper.)

Trypanosoma murmanensis
Khan, R. A., 1977, J. Fish. Research Bd. Canada, v. 34 (11), 2193-2196
Trypanosoma murmanensis in Gadus morhua (exper.), transmitted by leech Johanssonia sp. (exper.), hematological changes

Trypanosoma musculi
Albright, J. F.; Albright, J. W.; and Dusanic, D. G., 1977, J. Reticuloendothel. Soc., v. 21 (1), 21-31
Trypanosoma musculi, mice, correlation between parasitemia, splenic enlargement, and suppression of humoral immune response assessed both in vivo and in vitro, spleen cells of infected mice were unresponsive to both B and T cell mitogens during time of marked splenomegaly, blood serum of infected mice and extract of parasite both inhibit humoral immune response

Trypanosoma musculi
Brooks, B. O.; and Reed, N. D., 1977, J. Reticuloendothel. Soc., v. 22 (6), 605-608
Trypanosoma musculi, thymus dependency of elimination from mice

Trypanosoma musculi
Buengener, W., 1975, Tropenmed. u. Parasitol., v. 26 (3), 281-284
Trypanosoma musculi in inbred mice (exper.), parasite multiplication and course of infection in single infection, after pre-infection with T. congolense, or with added infection of T. brucei

Trypanosoma musculi, illus.
Buengener, W., 1975, Tropenmed. u. Parasitol., v. 26 (3), 285-290
Plasmodium berghei-infected mice (exper.), added infection of Trypanosoma musculi results in rapid multiplication of trypanosome parasites with distribution into multiple body organs and tissues, infection course

Trypanosoma musculi
Buengener, W., 1975, Tropenmed. u. Parasitol., v. 26 (3), 291-294
Trypanosoma musculi strain passaged several years in mice (exper.), infection course and development in pregnant mice, Plasmodium berghei-infected mice, or normal mice given either heavy or weak infections

Trypanosoma musculi, illus.
Dwyer, D. M.; and D'Alesandro, P. A., 1976, J. Protozool., v. 23 (1), 75-83
Trypanosoma musculi bloodstream forms, extracellular surface coat, fine structure, presence and distribution of carbohydrates in surface coat and pellicular membrane
Syn.: Trypanosoma duttoni

Trypanosoma musculi, illus.
Dwyer, D. M.; and D'Alesandro, P. A., 1976, J. Protozool., v. 23 (2), 262-271
Trypanosoma musculi bloodstream forms, lectin agglutination, fine structure localization of concanavalin A sites, antibody agglutinations (regular presence of surface-bound host serum proteins; induced surface adsorption of serum proteins), fine structural evidence of host serum in surface coat

Trypanosoma (Herpetosoma) musculi
Evans, D. A.; and Ellis, D. S., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 380 [Demonstration]
Trypanosoma musculi, establishment of experimental infection in midgut of Glossina morsitans morsitans, possible use of tsetse flies as laboratory model

Trypanosom[a] (Herpetosoma) musculi
Godfrey, D. G.; and Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 260 [Abstract]
bloodstream trypanosomes of 4 different subgenera, relative activities of alanine vs. aspartate aminotransferases were characteristic of each subgenus

T[rypanosoma] musculi
Hommel, M.; and Miltgen, F., 1973, J. Protozool., v. 20 (4), 527
Trypanosoma blanchardi, T. rabinowitschae, adaptation to mice by passage through cultures and then sarcomatous mice, strong cross immunity between T. musculi and mouse-adapted strains; adaptation of T. lewisi to mice using same method was not possible; this new culture system was used to cultivate T. brucei

Trypanosoma musculi
Jarvinen, J. A.; and Dalmasso, A. P., 1977, Exper. Parasitol., v. 43 (1), 203-210
Trypanosoma musculi, mice, development of mild anemia, evidence indicating participation of immunological mechanism, erythrocytes had bound immunoglobulins and complement component 3, intravascular hemolysis by complement was not a mechanism of cell destruction

Trypanosoma musculi
Jarvinen, J. A.; and Dalmasso, A. P., 1977, Infect. and Immun., v. 16 (2), 557-563
Trypanosoma musculi, course of infections in normocomplementemic, C5-deficient, and C3-depleted mice

Trypanosoma (Herpetosoma) musculi
Kaddu, J. B.; and Suzuki, M., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 538-539 [Letter]
Trypanosoma musculi, ultrastructural study of kidney changes in infected mice (exper.), possibility that changes might affect functioning of urinary systems

Trypanosoma musculi
Lee, C. M.; Aboko-Cole, G. F.; and Fletcher, J., 1976, Ztschr. Parasitenk., v. 49 (1), 1-10
Trypanosoma musculi, vitamin A-deficient mice, increased parasitemia, delayed action of reproductive-inhibiting and terminal lytic antibodies, increase in body weight gains and food consumption

Trypanosoma musculi
Pouliot, P.; Viens, P.; and Targett, G. A. T., 1977, Clin. and Exper. Immunol., v. 27 (3), 507-511
Trypanosoma musculi, transfer of spleen cells to T cell-deprived mice restored their ability to control infection, treatment of cells in vitro with anti-θ serum did not impair their ability to restore immunocompetence

Trypanosoma musculi
Rank, R. G.; Roberts, D. W.; and Weidanz, W. P., 1977, Infect. and Immun., v. 16 (2), 715-716
Trypanosoma musculi in congenitally athymic nude mice, chronic infection with consistently elevated parasitemia, thymic reconstitution restores immunity

Trypanosoma musculi
Targett, G. A. T., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 272 [Abstract]
Trypanosoma musculi, mice, immunological responses

Trypanosoma musculi
Targett, G. A. T.; Viens, P.; and Wilson, V. C. L. C., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 148 [Abstract]
Trypanosoma musculi, survival in immunized mouse host blood for at least 1 year in infective state

Trypanosoma musculi
Viens, P.; et al., 1977, Internat. J. Parasitol., v. 7 (2), 109-111
Trypanosoma musculi blood forms, in vitro cultivation in cell culture media

Trypanosoma (Herpetosoma) musculi
Wilson, V.C.L.C.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 271-272 [Abstract]
Trypanosoma musculi, persistence in vasa recta of kidney of immune mice; T. lewisi did not persist in immune rats

Trypanosoma nabiasi
D'Alesandro, P. A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 691-738
nonpathogenic trypanosomes of rodents, immunology, review

Trypanosoma neotomae
Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
Neotoma a. albigula (blood): Tonto National Monument, near Roosevelt, Gila County, Arizona; Newberry Springs, San Bernardino County, California

Trypanosoma numidae Wenyon, 1909
Fallis, A. M.; Jacobson, R. L.; and Raybould, J. N., 1973, J. Protozool., v. 20 (3), 436-437
Trypanosoma numidae, experimental transmission to guinea fowl and chickens via simuliids
Gallus domesticus (nat. and exper.)(blood): Tanzania
Numida mitrata (nat. and exper.)(blood): Tanzania
Simulium adersi: Tanzania
S. impukane group: Tanzania
S. vorax (exper.)
S. nyasalandicum (exper.)
Aedes aegypti (exper.)
Culex pipiens fatigans (exper.)

Trypanosoma otospermophili
Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
Ammospermophilus (blood): Chaco Canyon National Monument, San Juan County, New Mexico

Trypanosoma pacifica sp. n., illus.
  Burreson, E. M.; and Pratt, I., 1972, J. Protozool., v. 19 (4), 555-556
    Parophrys vetulus (blood): Pacific Ocean, 5-10 km off Newport, Oregon

Trypanosoma (Megatrypanum) palawanense n. sp., illus.
  Miyata, A., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 103-111
    Rattus panglima (blood): Montible Subcolony, Iwahig Penal Colony, Palawan Island, the Philippines

Trypanosoma palawanense Miyata, 1975
  Miyata, A.; and Tsukamato, M., 1975, Nettai Igaku (Trop. Med.), v. 16 (3), 113-130
    Rattus panglima: Palawan Island, the Philippines

Trypanosoma peromysci
  Wood, S. F., 1975, J. Parasitol., v. 61 (5), 969-970
    Peromyscus maniculatus (blood): Chaco Canyon National Monument, San Juan County, New Mexico; Petty Ranch near Frazier Mountain Park, Ventura County, California; Juniper Hills near Pearblossom, Los Angeles County, California
    P. truei montipinoris: Juniper Hills near Pearblossom, Los Angeles County, California
    P. truei: Juniper Hills near Pearblossom, Los Angeles County, California

Trypanosoma pessoai
  Ayala, S. C.; and Wells, E. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 76 [Letter]
    vampire bats: western Colombia

Trypanosoma phylodriasi Pessoa, 1928, illus.
  de Biasi, P.; et al., 1975, Mem. Inst. Butantan, v. 39, 85-101
    developmental forms in culture
    Philodryas nattereri
    Bothrops alternatus (exper.)

Trypanosoma phylodriasi
  de Biasi, P.; et al., 1975, J. Protozool., v. 22 (3), 35A [Abstract]
    Trypanosoma salamantae, T. phylodriasi, development in vitro, attempted exper. infection of laboratory bred juvenile snakes (only T. phylodriasi in Bothrops alternatus was successful), attempted exper. infection of mosquitoes negative except one Culex dolosus that showed flagellated developmental forms

Trypanosoma pipientis Diamond 1950
  Werner, J. K.; and Walewski, K., 1976, J. Parasitol., v. 62 (1), 20-25
    Rana clamitans
    R. pipiens
    (blood of all): all from Michigan

Trypanosoma platessae
  Cottrell, B., 1976, Parasitology, v. 73 (2), xxxiv [Abstract]
    Cryptocotyle lingua and Rhipidocotyle johnstonei induced temperature-dependent precipitin response in Pleuronectes platessa; Trypanosoma platessae-infected P. platessa had elevated serum beta-globulin levels, pronounced seasonal variation in numbers of infected fish pointed to temperature-controlled immunity

Trypanosoma platessae Lebailly, illus.
  Cottrell, B. J., 1977, J. Fish Biol., v. 11 (1), 35-47
    Trypanosoma platessae in Pleuronectes platessa (blood) (nat. and exper.), brief redescription, age of host, seasonal variation may be related to change in ambient temperatures and host immunity levels, host specificity: Looe Bay

Trypanosoma pseudopodium sp. n., illus.
  Werner, J. K.; and Walewski, K., 1976, J. Parasitol., v. 62 (1), 20-25
    Bufo americanus (blood): Cyrus H. McCormick Experimental Forest, Marquette County, Michigan

T[rypanosoma] (Herpetosoma) rabinowitschae
  Hommel, M.; and Miltgen, F., 1973, J. Protozool., v. 20 (4), 527
    Trypanosoma blanchardi, T. rabinowitschae, adaptation to mice by passage through cultures and then sarcomatous mice, strong cross immunity between T. musculi and mouse-adapted strains; adaptation of T. lewisi to mice using same method was not possible; this new culture system was used to cultivate T. brucei

Trypanosoma radiale n. sp., illus.
  Pessoa, S. B.; and de Biasi, P., 1975, Mem. Inst. Butantan, v. 39, 79-83
    Diplectrum radiale (sangue): Sao Sebastiao (litoral norte do Estado de Sao Paulo), Brasil

Trypanosoma ranarum (Lankester 1871)
  Werner, J. K.; and Walewski, K., 1976, J. Parasitol., v. 62 (1), 20-25
    Rana catesbeiana
    R. clamitans
    R. pipiens
    R. septentrionalis
    R. sylvatica
    (blood of all): all from Michigan

Trypanosoma rangeli-like trypanosome, illus.
  Barrett, T. V.; and de Oliveira, T. S., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 445-446
    Trypanosoma rangeli-like trypanosome in Rhodnius domesticus (haemolymph, hindgut, rectum), possible source of confusion in xenodiagnosis for Trypanosoma cruzi as the agent of human Chagas disease: Brazil

Trypanosoma rangeli
  D'Alessandro, A.; and del Prado, C. E., 1977, Am. J. Trop. Med. and Hyg., v. 26 (4), 623-627
    attempted determination of prevalence of Trypanosoma rangeli in endemic areas of T. cruzi; indications that routine examination of pooled triatome vector hemolymphs and random samples of salivary glands and midguts should be carried out along with standard examination of bug feces as an accurate method for Trypanosoma spp. differentiation: Argentina and Brazil

Trypanosoma rangeli
  Jadin, J. M.; et al., 1977, J. Protozool., v. 24 (4), 61A [Abstract]
    Kinetoplastidae, culture in mouse peritoneal macrophages

Trypanosoma rangeli
Justines, G.; and Sousa, O. E., 1977, Am. J. Trop. Med. and Hyg., v. 26 (2), 326-328
survival of arboviruses in Trypanosoma cruzi-infected Rhodnius prolixus (exper.) and in T. rangeli-infected R. neglectus (exper.), study to assess vector potential of parasitized bugs
Rhodnius neglectus (exper.)

Trypanosoma rangeli
Minoprio, J. L., 1972, Prensa Med. Argent., v. 59 (8), 293-308
human Chagas disease, pathologic findings, comparison with characteristics of other Trypanosoma spp. and Leishmania spp., extensive review

Trypanosoma rangeli
Minter-Goedbloed, E.; and Oliveira, M. L. S., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 89 [Letter]
differential diagnosis of Trypanosoma rangeli from T. cruzi based on growth in culture media alone not reliable because of variability of T. rangeli from different geographic localities

Trypanosoma (Herpetosoma) rangeli, illus.
Molyneux, D., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 371-372
Trypanosoma rangeli, method of division, probably multiplies in mammalian hosts extracellularly by equal binary fission in trypomastigote stage

Trypanosoma rangeli
Muehlpfordt, H.; and Schottelius, J., 1977, Tropenmed. u. Parasitol., v. 28 (1), 1-7
attempted differentiation of Trypanosoma cruzi strains and T. cruzi-like strains from T. rangeli and T. conorhini using agglutination reactions to protectin from sponges and lectin from Soja hispida

Trypanosoma rangeli
Schottelius, J., 1976, Ztschr. Parasitenk., v. 50 (2), 182-183
Trypanosoma spp., effects of various lectins, protectins and blood group test sera

Trypanosoma rangeli
Sousa, O. E.; and Dawson, G. A., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 407-409
Trypanosoma cruzi, T. rangeli, T. minasense found in Saguinus geoffroyi, high rate of infection suggests marmosets are important natural hosts: Panama Canal Zone

Trypanosoma rangeli
Toye, P. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 266 [Demonstration]
Trypanosoma rangeli, T. cruzi, T. lewisi, differentiation of enzyme patterns of culture forms

Trypanosoma rhodesiense
Aboko-Cole, G. F.; and Lee, C. M., 1975, Ztschr. Parasitenk., v. 47 (2), 71-77
Trypanosoma rhodesiense, T. lewisi, rats on normal or folic acid deficient diets, mitotic activity in liver; folate content of trypanosomes and their metabolic products

Trypanosoma rhodesiense
Aiyedun, B. A.; and Amodu, A. A., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 67-71
trypanosomiasis, albino rats (exper.), humans, diagnosis, capillary-tube precipitation test (known as capillary-tube agglutination test), rapid and specific technique that could be used in field survey work

Trypanosoma rhodesiense
Barry, D., 1975, J. Protozool., v. 22 (3), 49A [Abstract]
Trypanosoma rhodesiense, "capping" of surface antigens, specific and temperature-dependent movement of variation antigens, capping in some cases is an artefact of the indirect fluorescent method

Trypanosoma rhodesiense, illus.
Boller, K., 1977, Schweiz. Med. Wchnschr., v. 107 (47), 1706-1708
Trypanosoma rhodesiense, fatal infection with central nervous system involvement in man who had just returned from an African safari, clinical case report with emphasis on need for diagnostic awareness of possible tropical diseases in persons returning from endemic areas: Switzerland (had travelled in Rhodesia/Botswana)

Trypanosoma rhodesiense
Buyst, H., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 163-164
congenital Trypanosoma rhodesiense infection in premature baby successfully treated with suramin and supportive measures; guidelines for similar therapy for T. gambiense infections: Zambia

Trypanosoma rhodesiense
Campbell, G. H.; Esser, K. M.; and Weinbaum, F. I., 1977, Infect. and Immun., v. 18 (2), 434-438
Trypanosoma rhodesiense in mice, requirement for B-lymphocyte immunocompetence for immunity to infection

Trypanosoma rhodesiense
Campbell, G. H.; and Phillips, S. M., 1976, Infect. and Immun., v. 14 (5), 1144-1150
Trypanosoma rhodesiense, mice, adoptive transfer of variant-specific resistance with B lymphocytes and serum but not with T lymphocytes, results implicate antibody-mediated mechanism as having major role in resistance

Trypanosoma rhodesiense
Campbell, G. H.; Phillips, S. M.; and Esser, K. M., 1976, Abst. Ann. Meet. Am. Soc. Microbiol., 86
Trypanosoma rhodesiense, course of infection in athymic (nude) mice shows that T-cell independent mechanisms play major role in host resistance

Trypanosoma rhodesiense
Cross, G. A. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 255-256 [Abstract]
Trypanosoma brucei, T. rhodesiense, development of defined media for growth

Trypanosoma rhodesiense
Das, B. P.; and Boykin, D. W., 1977, J. Med. Chem., v. 20 (4), 531-536
Trypanosoma rhodesiense in mice, successful synthesis and antiprotozoal activity of 2,5-bis(4-guanylphenyl)furans in laboratory trials; little activity of these compounds against Plasmodium berghei in mice

Trypanosoma rhodesiense
Das, B. P.; and Boykin, D. W., 1977, J. Med. Chem., v. 20 (9), 1219-1221
2,5-bis(4-guanylphenyl)thiophenes and -pyrroles, significant antiprotozoal activity against Trypanosoma rhodesiense but little activity against Plasmodium berghei in mice (exper.), laboratory synthesis and comparison trials with stilbamidine, hydroxystilbamidine and pentamidine

Trypanosoma rhodesiense
Diggs, C.; et al., 1976, J. Immunol., v. 116 (4), 1005-1009
Trypanosoma rhodesiense, measurement of leucine incorporation in vitro as assay of functional integrity, use of this system in demonstrating cytotoxic activity in serum of immunized rats, dependence of activity on serum dose and on heat-labile normal serum constituent(s)

Trypanosoma rhodesiense
Duxbury, R. E.; et al., 1973, Isotopes and Radiation Parasitol. III, 179-180
Trypanosoma rhodesiense, mice, rats, cattle, rhesus monkeys; T. congolense, mice, dogs, cattle; T. brucei, cattle: attempted immunization with irradiated trypanosomes, brief communication

Trypanosoma rhodesiense
Duxbury, R. E.; Sadun, E. H.; and Anderson, J. S., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 266-267 [Abstract]
Trypanosoma rhodesiense, recently isolated human strain, immunization of rhesus monkeys using gamma-irradiated trypanosomes

Trypanosoma rhodesiense
Duxbury, R. E.; Sadun, E. H.; and West, J. E., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (5-6), 484-485
Trypanosoma rhodesiense, mice, neutron and gamma radiation of trypanosomes for immunization

Trypanosoma rhodesiense
Elce, B. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 162 [Abstract]
Trypanosoma congolense, T. brucei, T. rhodesiense, development in Glossina morsitans and small laboratory mammals

Trypanosoma rhodesiense
Else, J. G.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 170-171
trypanosomiasis, antigens from various T. spp. gave inconclusive serologic results in assessing presence of infection among Orang Asli aborigines: Malaysia

Trypanosoma rhodesiense
Flemmings, B. J.; Diggs, C. L.; and Powell, C. J., jr., 1976, Abst. Ann. Meet. Am. Soc. Microbiol., 77
Trypanosoma rhodesiense, alternative pathways of complement activation in cytotoxicity against trypanosomes by immune serum

Trypanosoma rhodesiense
Ford, W. C. L.; and Bowman, I. B. R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 257 [Abstract]
Trypanosoma rhodesiense, culture midgut form, proline metabolism

Trypanosom[a] rhodesiense
Geigy, R.; Kauffmann, M.; and Jenni, L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 284-286 [Abstract]
Alcelaphus buselaphus: Serengeti

Trypanosoma rhodesiense
Hawking, F., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 517-527
blood incubation infectivity test for differentiating Trypanosoma brucei and T. rhodesiense, modifications of technique and of interpretation

Trypanosoma rhodesiense
Hawking, F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 504-512
Trypanosoma brucei, T. rhodesiense, T. equiperdum, polymorphic strains, sensitivity to human plasma, composition and detailed analysis of typical strains using α_2-macroglobulin test

Trypanosoma rhodesiense
Hawking, F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (5-6), 1976, 513-520
Trypanosoma gambiense, T. rhodesiense, T. brucei, 60 polymorphic strains from East and West Africa, in vivo test of sensitivity or resistance to human plasma, application to infectivity for man

Trypanosoma rhodesiense
Hawking, F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 427-430
Trypanosoma rhodesiense (6 clones of stabilate), polymorphic trypanosomes (12 clones of stabilate), tests for resistance to human plasma, indications for fundamental differences of genetic material

Trypanosoma rhodesiense
Jackson, P. R., 1975, J. Parasitol., v. 61 (5), 963-965
forced-flow DEAE-cellulose column for isolation of bloodstream trypanosomes

Trypanosoma rhodesiense
Lindsley, H. B.; Kysela, S.; and Steinberg, A. D., 1974, J. Immunol., v. 113 (6), 1921-1927
Trypanosoma rhodesiense, T. gambiense, humans, development of antibodies to nucleic acids, possible pathophysiological relationship to renal disease remains to be determined

Trypanosoma rhodesiense, illus.
Macadam, R. F.; and Williamson, J., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 291-299
Trypanosoma rhodesiense, acriflavine, ethidium, antrycide, drug effects on fine structure

Trypanosoma rhodesiense, illus.
Macadam, R. F.; and Williamson, J., 1974, Ann. Trop. Med. and Parasitol., v. 68 (3), 301-306
Trypanosoma rhodesiense, suramin, tryparsamide, mapharside, drug effects on fine structure

Trypanosoma rhodesiense
Minoprio, J. L., 1972, Prensa Med. Argent., v. 59 (8), 293-308
human Chagas disease, pathologic findings, comparison with characteristics of other Trypanosoma spp. and Leishmania spp., extensive review

Trypanosoma rhodesiense
Murray, M.; and Urquhart, G. M., 1977, Advances Exper. Med. and Biol., v. 93, 209-241
African trypanosomiasis, immunoprophylaxis, review: cattle and sheep, laboratory animals; chemotherapeutic agents and immunity, nature of protective antigen, effector mechanisms in protection, manipulation of host resistance

Trypanosom[a] rhodesiense
Mwambu, P. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 286-287 [Abstract]
Trypanosom[a] rhodesiense, importance of cattle as natural reservoir hosts; T. brucei subgroup isolated from cattle, resistance to Ethidium, sensitivity to Berenil: Tanzania; Uganda

[Trypanosoma rhodesiense] Rhodesian sleeping sickness
Ogada, T.; Fink, E.; and Mbwabi, D., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 280-281 [Abstract]
Rhodesian sleeping sickness, clinical trials of 98/202 (a diamidine), effective with no toxicity except initial pyrexia

Trypanosoma rhodesiense
Raether, W.; et al., 1974, Tropenmed. u. Parasitol., v. 25 (1), 42-48
Trypanosoma rhodesiense in Macaca spp., berenil, chemoprophylactic and pharmacokinetic studies

Trypanosoma rhodesiense
Raether, W.; and Seidenath, H., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (3), 285-295
Trypanosoma rhodesiense, motility and infectivity in various physiologic solutions, results show that motility of trypanosomes in suspension gives no clear indication of actual infectivity, implications for short-term storage

T[rypanosoma] rhodesiense
Raether, W.; and Seidenath, H., 1974, Tropenmed. u. Parasitol., v. 25 (1), 28-41
T[rypanosoma] rhodesiense, T[rypanosoma] brucei, influence of different physiological solutions and a blood substitute on motility and infectivity after deep-freezing in liquid nitrogen

Trypanosoma rhodesiense
Raether, W.; and Seidenath, H., 1976, Tropenmed. u. Parasitol., v. 27 (2), 238-244
trypanocidal effects and action of preparation 98/202 in exper. Trypanosoma rhodesiense infections in Macaca arctoides (possible new animal model for late stages of T. rhodesiense infections in man); animals previously treated with pentamidine or diminazene developed resistance to treatment

Trypanosoma rhodesiense
Rafael, A. G., 1975, An. Serv. Vet., Mocambique (20-21), 1972-1973, 331-350
Trypanosoma spp., pathology, economic importance, treatment, vector control, extensive review: Africa

Trypanosoma rhodesiense
Rane, L.; Rane, D. S.; and Kinnamon, K. E., 1976, Am. J. Trop. Med. and Hyg., v. 25 (3), 395-400
Trypanosoma rhodesiense, test system to evaluate large numbers of candidate drugs based on mortality of mice with blood-induced infections, serves as primary screen or as secondary and confirmatory test for active compounds

Trypanosoma rhodesiense
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma rhodesiense
Ruitenberg, E. J.; and Buys, J., 1977, Am. J. Trop. Med. and Hyg., v. 26 (1), 31-36
Trypanosoma brucei, enzyme-linked immunosorbent assay (ELISA) for serodiagnosis of human African sleeping sickness, comparison tests with immunofluorescence technique showed good results in rabbits (exper.) and serum from infected humans, cross-reactions only in person with Leishmania antibodies, possible application to epidemiologic surveys

Trypanosoma rhodesiense
Sadun, E. H.; et al., 1972, Parassitologia, v. 14 (1), 25-28
Trypanosoma congolense, irradiated forms immunizing mice, dogs and cattle; T. rhodesiense, irradiated forms immunizing mice and rats; irradiated human strain of T. rhodesiense immunizing Macaca mulatta

Trypanosoma rhodesiense
Sanchez, G.; and Alderete, J. F., 1975, Comp. Biochem. and Physiol., v. 52 (4A), 623-626
Trypanosoma rhodesiense, adrenalectomized rats, increased parasitemia and earlier death, relation to decreases in total liver glycogen, reduced rate of glucose consumption by trypanosomes, endocrine system as possible regulator of trypanosome biochemistry, possible mechanism in pathogenesis of trypanosomiasis

Trypanosoma rhodesiense
Srivastava, H. K.; and Bowman, I. B. R., 1972, Nature, London, N. Biol. (57), v. 235, 152-153
Trypanosoma rhodesiense, sequence of metabolic changes associated with acquisition of mitochondrial-mediated metabolism during transformation into culture forms, significantly higher rate of oxidation with proline vs. other substrates tested

Trypanosoma rhodesiense
Szarfman, A.; et al., 1975, Am. J. Trop. Med. and Hyg., v. 24 (1), 19-24
evaluation of specificity of EVI factor observed in Trypanosoma cruzi cases using indirect fluorescent antibody test on sera from patients with other parasitic diseases, immunofluorescence of anti-skeletal muscle antibody from leishmaniasis

Trypanosoma rhodesiense
Voller, A.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 431-437
human tropical parasitic diseases, comparison of enzyme-immunoassay and radio-immunoassay in detection of antibodies, both assays sensitive and reproducible and gave comparable results

Trypanosoma rhodesiense
Voller, A.; Bartlett, A.; and Bidwell, D. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 98-106
application of enzyme immunoassay to diagnosis of human parasitic diseases, technique for microplate enzyme-linked immunosorbent assay

Trypanosom[a] rhodesiense
Walker, P. J.; and Opiyo, E. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 281-282 [Abstract]
Trypanosom[a] brucei, T. rhodesiense, mice, effect of parasite burden on curative dose of trypanocides, possible source of variation in results of in vivo drug tests

Trypanosoma rhodesiense, illus.
Williamson, J.; and Macadam, R. F., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 130-137
Trypanosoma rhodesiense, effects of puromycin and its aminonucleoside, cordycepin and nucleocidin on fine structure

Trypanosoma rhodesiense
Williamson, J.; and McLaren, D. J., 1974, J. Protozool., v. 21 (3), 453-454 [Abstract]
Trypanosoma rhodesiense, induction of cytoplasmic clefts by Puromycin, Cordycepin, and Nucleocidin, results suggest drugs may prevent uptake of required unsaturated fatty acids and promote excessive intake of saturated acids

Trypanosoma rhodesiense
Williamson, J.; and McLaren, D. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 263 [Demonstration]
Trypanosoma rhodesiense in mice (exper.), cordycepin-induced cytoplasmic clefts in trypanosomes and resulting changes in fatty acid metabolism, ultrastructural studies

Trypanosoma rhodesiense
Woo, P. T. K.; and Rogers, D. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 319-326
Trypanosoma spp., statistical study of sensitivity of haematocrit centrifuge technique for detecting trypanosomes in blood, evaluation of relationship of probability of detection of trypanosome to trypanosome density

Trypanosoma rhodesiense
Woo, P.; and Soltys, M. A., 1971, Ann. Trop. Med. and Parasitol., v. 65 (4), 465-469
Trypanosoma brucei, T. rhodesiense, effect of suramin on blood and tissue forms

Trypanosom[a] rhodesiense
Woodruff, A. W.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 329-337
Trypanosom[a] rhodesiense, human, mechanism of anemia, complement coating of erythrocytes and subsequent haemolysis, splenic enlargement due to erythrophagocytosis within spleen

Trypanosoma rotatorium Mayer, 1843
Hysek, J.; Kalafa, S.; and Zizka, Z., 1975, J. Protozool., v. 22 (3), 63A [Abstract]
Trypanosoma rotatorium, not pathogenic for its cold-blooded host (frogs), causes fatal infections in a warm-blooded host (mice, exper.)

Trypanosoma rotatorium, Mayer, 1843, illus.
Hysek, J.; Kalafa, S.; and Zizka, Z., 1976, Arch. Protistenk., v. 118 (4), 341-345
Trypanosoma rotatorium (blood), successful experimental transmission from Rana esculenta and R. temporaria (poikilothermic) to white laboratory mice (homiothermic), pathogenicity, fatal infection; comparative morphology in frogs and mice

Trypanosoma rotatorium Mayer 1843, illus.
Hysek, J.; and Zizka, Z., 1976, Nature, London (5552), v. 260, 608-609
experimental transmission of Trypanosoma rotatorium from frogs to white mice by inoculation of blood, development of infection and morphology in mice

Trypanosoma rotatorium
Lank, D. R., jr., 1971, Proc. Indiana Acad. Sc., v. 81 (2), 359-364
Rana catesbeiana: Indiana

Trypanosoma rotatorium (Mayer, 1843), illus.
Miyata, A., 1976, Nettai Igaku (Trop. Med.), v. 18 (3), 125-134
Trypanosoma rotatorium, survey, incidence in frogs and tadpoles, morphological variations
Rana rugosa
R. nigromaculata
(heart of all): all from Nagasaki City vicinity

T[rypanosoma] rotatorium Mayer, 1843
Pereira, N. de M.; et al., 1973, J. Protozool., v. 20 (4), 530
T[rypanosoma] rotatorium, different culture media employed to isolate, subcultures not obtained

Trypanosoma rotatorium-like sp.
Sullivan, J. S.; and Sullivan, J. J., 1976, Southeast Asian J. Trop. Med. and Pub. Health, v. 7 (3), 493-495
Rana erythraea
R. hosei
Bufo melanostictus
Rana macrodon
all from Malayan Peninsula

Trypanosoma rotatorium (Mayer, 1843)
Wallace, F. G., 1975, J. Protozool., v. 22 (3), 18A [Abstract]
Trypanosoma rotatorium, differentiation from T. loricatum
Rana esculenta: Yugoslavia

Trypanosoma rotatorium (Mayer 1843)
Werner, J. K.; and Walewski, K., 1976, J. Parasitol., v. 62 (1), 20-25
Rana catesbeiana
R. clamitans
R. pipiens
R. septentrionalis
(blood of all): all from Michigan

Trypanosoma saccobranchi, illus.
Rao, K. B. V., 1975, Indian J. Zool., v. 3 (1-2), 51-53
Trypanosoma saccobranchi in Saccobranchus fossillis (blood), intensity of infection higher in fishes continuously fed with earthworms than those not given food, preliminary experiment: pond near Hyderabad

Trypanosoma salamantae Pessoa & Fleury, 1969, illus.
de Biasi, P.; et al., 1975, Mem. Inst. Butantan, v. 39, 85-101
developmental forms in culture, experimental infection of Culex dolosus fed on snake "one specimen presented flagellated forms.... potential vectors"
Epicrates cenchria crassus

Trypanosoma salamantae
de Biasi, P.; et al., 1975, J. Protozool., v. 22 (3), 35A [Abstract]
Trypanosoma salamantae, T. phylodriasi, development in vitro, attempted exper. infection of laboratory bred juvenile snakes (only T. phylodriasi in Bothrops alternatus was successful), attempted exper. infection of mosquitoes negative except one Culex dolosus that showed flagellated developmental forms

Trypanosoma (Megatrypanum) samueli n. sp., illus.
Mello, D. A., 1977, Ann. Parasitol., v. 52 (4), 391-395
Monodelphis domesticus (blood): northern Formosa County, Goias State, Brazil

Trypanosoma scelopori, illus.
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
Sceloporus occidentalis (blood): central California

Trypanosoma scelopori
Krassner, S. M.; Sylvester, D.; and Munson, K. B., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 258 [Abstract]
Trypanosoma scelopori, culture forms, proline metabolism

Trypanosoma schmidti-like sp., illus.
Werner, J. K.; and Walewski, K., 1976, J. Parasitol., v. 62 (1), 20-25
description
Bufo americanus (blood): Michigan

Trypanosoma sigmodoni
D'Alesandro, P. A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 691-738
nonpathogenic trypanosomes of rodents, immunology, review

Trypanosoma simiae
Janssen, J. A. H. A.; and Wijers, D. J. B., 1974, Ann. Trop. Med. and Parasitol., v. 68 (1), 5-19
Trypanosoma simiae, correlation between virulence and transmitting species of Glossina
Glossina brevipalpis (nat. and exper.): Kenya
Glossina austeni: Kenya
Glossina pallidipes (nat. and exper.): Kenya
pigs (exper.)
goats (exper.)
rabbits (exper.)

Trypanosoma simiae
Rafael, A. G., 1975, An. Serv. Vet., Mocambique (20-21), 1972-1973, 331-350
Trypanosoma spp., pathology, economic importance, treatment, vector control, extensive review: Africa

Trypanosoma simiae
Toure, S. M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 30 (1), 1-10
trypanosomiasis, domestic animals, diagnostic methods, review

Trypanosoma suis
Toure, S. M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 30 (1), 1-10
trypanosomiasis, domestic animals, diagnostic methods, review

Trypanosoma (Herpetosoma) tamiasi sp. n., illus.
Riedel, D., 1975, J. Protozool., v. 22 (1), 8-17
Syn.: Trypanosoma sp. Dorney, 1967
life cycle, mode of reproduction, morphogenesis, morphometrics, host specificity
Spermophilus tridecemlineatus (exper.)
Meriones unguiculatus (exper., "Less than 10^5 trypanosomes/ml were detected for 2-7 days in. . .")
Glaucomys sabrinus (exper., "Less than 10^5 trypanosomes/ml were detected for 2-7 days in. . .")
Tamiasciurus hudsonicus (exper., "Less than 10^5 trypanosomes/ml were detected for 2-7 days in. . .")
Spermophilus richardsonii (exper., "Less than 10^5 trypanosomes/ml were detected for 2-7 days in. . .")
Xenopsylla cheopis (exper., brief survival with little or no development)
Nosopsyllus fasciatus (exper., brief survival with little or no development)
Orchopeas leucopus (exper., brief survival with little or no development)
Rhodnius prolixus (exper., brief survival with little or no development)
Megabothris quirini (exper.) [may also be nat.?]

Trypanosoma (Herpetosoma) tamiasi sp. n.-- Continued.
Riedel, D., 1975, J. Protozool., v. 22 (1), 8-17.-- Continued.
Tamiophila grandis (exper.) [may also be nat.?]
Tamias striatus (nat. and exper.)
Eutamias minimus (nat. and exper.)
Megabothris acerbus (nat. and exper.)
all from Ontario Department of Lands and Forests Research Station, Lake Sasajewun, Algonquin Provincial Park, Ontario, Canada

Trypanosoma (Megatrypanum) theileri-like, illus.
Bigalke, R. D.; Keep, M. E.; and Schoeman, J. H., 1972, Onderstepoort J. Vet. Research, v. 39 (4), 225-227
brief description
Tragelaphus angasi (spleen): Ndumu Game Reserve, Zululand

Trypanosoma theileri
Hinaidy, H., 1976, Ztschr. Parasitenk., v. 50 (2), 212-213
Rind (Blut): Osterreich

Trypanosoma theileri
Leeflang, P.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 412 [Letter]
Trypanosoma spp., concentration and differentiation of species using a hypotonic lysis method

Trypanosoma theileri
McLoughlin, D. K.; and Chute, M. B., 1975, Proc. Helminth. Soc. Washington, v. 42 (2), 183-184
Trypanosoma theileri in peripheral blood and in bloody vaginal washings of Hereford cows being used for exper. treatment studies of Tritrichomonas foetus infections (dimetridazole), transitory urogenital disturbance

Trypanosoma theileri Laveran, 1902 (T. americanum), illus.
Moulton, J. E.; and Krauss, H. H., 1972, Cornell Vet., v. 62 (1), 124-137
Trypanosoma theileri contaminating bovine spleen culture, ultrastructure

Trypanosoma theileri
Schindler, R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 290 [Abstract]
Trypanosoma spp., cattle, attempted serologic differential diagnosis gave equivocal results

Trypanosoma theileri
Smith, M. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 18 [Demonstration]
cattle: Trinidad

Trypanosoma theileri
Toure, S. M.; et al., 1975, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 28 (4), 463-472
trypanosomiasis, cattle, evaluation of indirect fluorescent antibody test in diagnosis and epizootiological surveys: Senegal

Trypanosoma venezuelense
Lopez D., T., 1976, J. Protozool., v. 23 (2), 17A [Abstract]
Trypanosoma venezuelense, rats, serum proteins

Trypanosoma venezuelense
Ojeda P., P.; and Lopez D., T., 1976, J. Parasitol., v. 62 (1), 128-129
Trypanosoma venezuelense, quantitative correlation between cell volume and DNA content, possibility that relationship may be species-specific

Trypanosoma (Schizotrypanum) vespertilionis
Liston, A. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (1), 6 [Demonstration]
Trypanosoma cruzi, T. dionisii, T. vespertilionis, cell mediated immune responses in mice, some cross-reactivity of antigens but homologous responses stronger

Trypanosoma vespertilionis
Miller, P. G. G.; Linstead, D. J.; and Klein, R. A., 1976, Parasitology, v. 73 (2), xvi [Abstract]
5 spp. of trypanosomatids, enzymes of acetyl CoA metabolism, possible routes for synthesis and utilization of acetyl CoA

Trypanosoma vespertilionis
Schottelius, J., 1976, Ztschr. Parasitenk., v. 50 (2), 182-183
Trypanosoma spp., effects of various lectins, protectins and blood group test sera

Trypanosoma vespertilionis Battaglia, 1904, illus.
Soria, C. A.; and Dusanic, D. G., 1975, J. Protozool., v. 22 (4), 509-513
Trypanosoma vespertilionis and T. dionisii in culture compared, population density, morphologic alterations, changes in glucose consumption and pH of media, antigenic analysis

Trypanosoma (Schizotrypanum) vespertilionis
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

Trypanosoma vivax
Bagster, I. A.; and Parr, C. W., 1973, Nature (5415), v. 244, 364-366
trypanosome identification by electrophoresis of soluble enzymes

Trypanosoma (Duttonella) vivax
Buengener, W.; and Mehlitz, D., 1976, Tropenmed. u. Parasitol., v. 27 (4), 405-410
experimentally infected Capra ibex s. hircus, few clinical signs of infection

Trypanosoma vivax, illus.
Buengener, W.; and Mehlitz, D., 1977, Tropenmed. u. Parasitol., v. 28 (1), 8-10
Trypanosoma vivax strain EATRO 1695, extravascular localizations discovered in cattle (exper.) (marginal sinus of lymph node, interstitial tissue of heart)

Trypanosoma vivax
Clarkson, M. J., 1976, Vet. Parasitol., v. 2 (1), 9-29
trypanosomes, immunological problems, review: antigens; host reactions to infection; immunological state of host

Trypanosoma vivax
Clarkson, M. J.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 2-3 [Demonstration]
Trypanosoma vivax, West African strain, serum protein changes in infected calves (exper.), possible mechanism of increased IgM

Trypanosoma vivax
Clarkson, M. J.; Cottrell, B. A.; and Toro, M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 10-11 [Demonstration]
animal trypanosomiasis, diagnosis by capillary-tube agglutination test using Trypanosoma vivax antigen, cattle (nat. and exper.), sheep (exper.), goats (exper.): Venezuela

Trypanosom[a] vivax
Clarkson, M. J.; and McCabe, W. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 12 [Demonstration]
Trypanosoma spp., relative susceptibility of mice to infection by buccal, gastric, or intraperitoneal routes, possible significance of oral transmission under natural conditions

Trypanosoma vivax
Clarkson, M. J.; and Penhale, W. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 273 [Abstract]
Trypanosoma vivax, calves (exper.), serum protein changes

Trypanosoma vivax
Cowper, S. G., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 415-425
continuous flow apparatus for in vitro maintenance: Schistosoma mansoni, S. haematobium, survival time, carbohydrate metabolism; Plasmodium knowlesi, morphology and carbohydrate metabolism; preliminary attempts to cultivate Trypanosoma vivax and Babesia canis

Trypanosoma vivax
Dar, F. K.; et al., 1973, Ann. Trop. Med. and Parasitol., v. 67 (1), 21-29
techniques used to study serotypes of salivarian trypanosomes being circulated by tsetse flies in 4 separate geographical areas, methods used to isolate and serologically type the trypanosomes are described and discussed in detail: Uganda and Kenya, East Africa

Trypanosoma vivax
Dar, F. K.; Paris, J.; and Wilson, A. J., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 319-329
Trypanosoma vivax, comparison of antigenic types isolated from wild tsetse flies in 2 geographically distinct areas of East Africa

Trypanosoma vivax
Dar, F. K.; Paris, J.; and Wilson, A. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 287 [Abstract]
Trypanosoma vivax, Zebu cattle, haematological and serum protein changes

Trypanosoma vivax
Diaz-Ungria, C., 1972, Bol. Soc. Venezolana Cien. Nat., Caracas (122-123), v. 29, 613-621
Trypanosoma vivax, Desowitz strain, susceptibility of rats and mice via intraperitoneal and subcutaneous routes, virulence for rodents after passage in Rhodnius prolixus and Musca domestica, susceptibility of sheep to this rodent strain, infection of rodents via gastric and buccal routes, indefinite preservation by freezing at -70°C.

Trypanosoma vivax
Evans, D. A.; Brightman, C. J.; and Holland, M. F., 1977, Lancet, London (8041), v. 2, 769 [Letter]
Trypanosoma brucei brucei and T. b. rhodesiense infections in mice (exper.) successfully treated with salicylhydroxamic acid if combined with glycerol and administered before patent infection established; T. vivax infections of different durations all successfully cured

Trypanosoma vivax
de Gee, A. L. W.; Ige, K.; and Leeflang, P., 1976, Internat. J. Parasitol., v. 6 (5), 419-421
Trypanosoma vivax, bovine isolates with inherent property for serial maintenance in mice, cyclical transmission by tsetse flies from sheep to sheep and goat to goat without affecting mouse infectivity, mice not suitable for direct fly transmission experiments

Trypanosom[a] (Duttonella) vivax
Godfrey, D. G.; and Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 260 [Abstract]
bloodstream trypanosomes of 4 different subgenera, relative activities of alanine vs. aspartate aminotransferases were characteristic of each subgenus

Trypanosoma vivax
Godfrey, D. G.; Kilgour, V.; and Na'Isa, B. K., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 270 [Abstract]
Trypanosoma vivax in Nigerian cattle, isoenzyme variations

Trypanosoma vivax
Hull, R. M., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 289 [Abstract]
Trypanosoma vivax, West African strain, cyclical transmission from rabbits and sheep by Glossina morsitans to rabbits

Trypanosoma vivax
Ilemobade, A. A.; et al., 1975, Ann. Trop. Med. and Parasitol., v. 69 (1), 13-18
Trypanosoma vivax, 47 isolates from Zebu in northern Nigeria, none resistant to homidium, isolation more sucessful in exper. cattle than in sheep and not at all in goats

Trypanosoma vivax
Ilemobade, A. A.; and Schillhorn van Veen, T. W., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 410 [Letter]
Friesian x zebu cross bred calf (exper.)
(ocular lesion)
sheep (exper.)

Trypanosoma vivax
van den Ingh, T. S. G. A. M.; et al., 1976, Research Vet. Sc., v. 21 (3), 264-270
Trypanosoma vivax, goats, pathology and pathogenesis, distinguishing acute and chronic forms, progression of symptoms in acute form

Trypanosoma vivax
Isoun, M. J., 1975, J. Protozool., v. 22 (3), 39A [Abstract]
Trypanosoma vivax, in vitro cultivation allowing survival through 3-4 cell cycles

Trypanosoma vivax
Isoun, M. J.; and Isoun, T. T., 1974, Tropenmed. u. Parasitol., v. 25 (3), 283-287
Trypanosoma vivax, T. brucei, in vitro cultivation, improved survival and multiplication in MEM 199 medium buffered by HEPES in comparison to bicarbonate, importance of strict control of pH

Trypanosoma vivax
Isoun, T. T.; and Anosa, V. O., 1974, Nigerian J. Animal Prod., v. 1 (2), 143-150
Trypanosoma vivax in sheep and goats (exper.), pathological changes in host reproductive organs; reduced reproductive capacity

Trypanosoma vivax
Isoun, T. T.; and Anosa, V. O., 1974, Tropenmed. u. Parasitol., v. 25 (4), 469-476
Trypanosoma vivax, sheep and goats (exper.), lesions in reproductive organs, implications for reproductive capacity of these animals in trypanosomiasis-endemic areas

Trypanosoma vivax
Jones, T. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 12 [Demonstration]
Trypanosoma vivax, antigenic variation

Trypanosoma vivax
Jones, T. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 288-289 [Abstract]
Trypanosoma vivax strains, antigenic types

Trypanosoma vivax
Jones, T. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 3 [Demonstration]
Trypanosoma vivax, West African strain, appearance of new antigen types at 2-5 day intervals after first appearance of trypanosomes in blood from infected calves

Trypanosoma vivax
Jones, T. W., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 271 [Abstract]
Trypanosoma vivax strain 64/23, antigenic variations of tsetse-transmissible strain in ruminants

Trypanosoma vivax
Jones, T. W.; and Clarkson, M. J., 1974, Ann. Trop. Med. and Parasitol., v. 68 (4), 485-486
Trypanosoma vivax in calves, timing of antigenic variation

Trypanosoma vivax
Kilgour, V., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 11 [Demonstration]
Trypanosoma brucei brucei, T. vivax, T. congolense, isoenzymes of alanine aminotransferase as possible specific characters

Trypanosoma vivax
Kilgour, V.; and Godfrey, D. G., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 387-389
Trypanosoma vivax, 2 characteristic isoenzyme patterns persistent in Nigerian trypanosomes collected from cattle

Trypanosoma vivax
Kilgour, V.; Godfrey, D. G.; and Na'isa, B. K., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 329-335
Trypanosoma vivax, three distinct isoenzyme patterns of two aminotransferases, possible use in identification and subspeciation, Nigerian cattle (nat. and exper.), sheep (exper.), goats (exper.)

Trypanosoma vivax
Klein Mori, J., 1974, Rev. Med. Vet. y Parasitol., Maracay, v. 25 (1-8), 112-142
Trypanosoma vivax, cattle, diagnosis, inoculation of sheep with infected bovine blood

Trypanosoma vivax
Lanham, S. M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 270 [Abstract]
trypanosomes, enzymatic identification of precipitating antigens

Trypanosoma vivax
Leeflang, P.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (5), 412 [Letter]
Trypanosoma spp., concentration and differentiation of species using a hypotonic lysis method

Trypanosoma vivax
Leeflang, P.; Buys, J.; and Blotkamp, C., 1976, Internat. J. Parasitol., v. 6 (5), 413-417
Trypanosoma vivax, mouse inoculation for diagnosis of infection in cattle, only early natural infections were capable of infecting mice, serial maintenance in mice did not change pathogenicity for ruminants

Trypanosoma vivax
Leeflang, P.; Ige, K.; and Olatunde, D. S., 1976, Internat. J. Parasitol., v. 6 (6), 453-456
Trypanosoma vivax, infectivity of cyclically and mechanically transmitted ruminant infections for mice and rats, results emphasize value of blood inoculation of rodents for detecting early infections of T. vivax

Trypanosoma (Duttonella) vivax
Luckins, A. G., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 272-273 [Abstract]
Trypanosoma congolense, T. vivax, cattle, elevated IgM levels: Uganda

Trypanosoma vivax
Luckins, A. G., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 148-149 [Abstract]
Trypanosoma congolense, T. vivax in zebu cattle, increased levels of IgM in presence of infection

Trypanosoma vivax
Luckins, A. G., 1975, Ann. Trop. Med. and Parasitol., v. 69 (3), 337-344
Trypanosoma vivax, T. brucei, T. congolense-infected Tragelaphus scriptus, serum immunoglobulin levels and relative proportions of serum proteins, compared with Zebu cattle

Trypanosoma vivax
Luckins, A. G., 1976, Ann. Trop. Med. and Parasitol., v. 70 (2), 133-145
Trypanosoma congolense, T. vivax, zebu cattle (exper.), serum immunoglobulin levels during infection and after administration of diminazene aceturate

Trypanosoma vivax
Luckins, A. G.; and Mehlitz, D., 1976, Ann. Trop. Med. and Parasitol., v. 70 (4), 479-480
Trypanosoma brucei, T. vivax, T. congolense, cattle (exper.), single or mixed infections, serum immunoglobulin levels

Trypanosom[a] vivax
MacLellan, K. J. R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 282 [Abstract]
Trypanosom[a] vivax, cattle, aparasitaemic interval following subcurative treatment with diminazene, findings strengthen belief that this strain survives in treated animals other than in bloodstream

Trypanosoma vivax
Matthews, D., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 3 [Demonstration]
Trypanosoma vivax, identification, purification and characterization of variable antigens from 2 antigenically distinct variants of rodent adapted strain

Trypanosoma vivax
Maxie, M. G.; Losos, G. J.; and Tabel, H., 1976, Pathophysiol. Parasit. Infect., 183-198
Trypanosoma congolense, T. vivax, cattle, development of macrocytic normochromic anemia, leukopenia, and persistent thrombocytopenia, significant differences in response of cattle to the two species

Trypanosoma (Duttonella) vivax
Mehlitz, D., 1975, Tropenmed. u. Parasitol., v. 26 (3), 265-275
Trypanosoma spp. of Nannomonas, Duttonella, and Trypanozoon subgenera, comparison of indirect fluorescent antibody test and complement fixation test as means of differentiating subgenera by serology and persistence of antibodies

Trypanosoma vivax-type
Moloo, S. K., 1973, Ann. Trop. Med. and Parasitol., v. 67 (2), 205-211
Trypanosoma vivax-type, T. congolense-type, infection rates in Glossina swynnertoni, relationship with types of mammals used as food sources: Serengeti National Park, Tanzania

Trypanosom[a] vivax
Moloo, S. K., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 286 [Abstract]
source of food as important factor determining trypanosome infection rate in natural population of Glossina swynnertoni: Ikoma-Serengeti area, Tanzania

Trypanosoma vivax, illus.
Muehlpfordt, H., 1975, Tropenmed. u. Parasitol., v. 26 (1), 1-8
Trypanosoma vivax, electron microscopic study of blood forms, comparison with ultrastructure of trypomastigote forms of Trypanosoma spp. described by other authors

Trypanosoma vivax
Murray, M.; Murray, P. K.; and McIntyre, W. I. M., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (4), 325-326
African trypanosomiasis, light microscopic examination of buffy coat zone of a microhaematocrit capillary tube expressed onto a slide found to be more reliable than standard methods for diagnosing infections in cattle; method also allowed differentiation of species

[Trypanosoma] vivax group
Okiwelu, S. N., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 101-107
[Trypanosoma] vivax group, survey of host preference and infection rate of Glossina morsitans morsitans vectors: Republic of Zambia

Trypanosoma vivax
Ormerod, W. E., 1976, Science (4229), v. 191, 815-821
possible ecological effects of trypanosomiasis control, increased cattle production leading to overgrazing and possibly to climatic change and drought: Africa

Trypanosoma vivax
Perie, N. M.; Tinnemans-Anggawidjaja, T.; and Zwart, D., 1975, Tropenmed. und Parasitol., v. 26 (4), 399-404
Trypanosoma spp. in experimentally infected domestic animals, sandwich immunofluorescent complement fixation test compared with indirect fluorescent antibody test for use in detecting binding of complement to antigen-antibody complex

Trypanosoma vivax, illus.
Platt, K. B.; and Adams, L. G., 1976, Research Vet. Sc., v. 21 (1), 53-58
Trypanosoma vivax, calves, indirect fluorescent antibody test (IFAT) evaluated, cross reactivity not observed when sera from calves singularly infected with T. theileri, T. evansi, Anaplasma marginale, Babesia argentina, B. bigemina, and Eperythrozoon spp. were tested in IFAT: South America

Trypanosoma vivax
Politzar, H., 1974, Tropenmed. u. Parasitol., v. 25 (1), 22-27
Trypanosoma vivax alone or with T. brucei and T. congolense, cattle (exper.), homologous and heterologous antibody responses determined by indirect fluorescent antibody test

Trypanosoma vivax
Rafael, A. G., 1975, An. Serv. Vet., Mocambique (20-21), 1972-1973, 331-350
Trypanosoma spp., pathology, economic importance, treatment, vector control, extensive review: Africa

Trypanosoma vivax
Reynolds, C. H.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 429-430 [Letter]
Trypanosoma vivax, cattle, concluded that there is no evidence for changes in activity of liver-specific enzymes in serum during non-lethal infection provided that steps are taken to minimize the effect of pyruvate in the enzyme assays

Trypanosoma vivax
Richards, W. H. G.; and Neal, R. A., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 276-277 [Demonstration]
parasitic protozoa stored frozen in liquid nitrogen, survival records

Trypanosoma vivax
Riordan, K., 1977, Ann. Trop. Med. and Parasitol., v. 71 (1), 11-20
Trypanosoma spp., survey of long-term variations in infection rates of Glossina spp. vectors found along major trade cattle route in southern Nigeria

Trypanosoma vivax
Roberts, C. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (1), 11 [Demonstration]
Trypanosoma congolense, T. vivax, cattle, sheep, serum lipid levels, significance of increase in plasma volume in trypanosome infections

Trypanosom[a] vivax
Roberts, C. J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 279-280 [Abstract]
Trypanosom[a] vivax, T. congolense, ruminants, serum lipid levels

Trypanosoma vivax
Roberts, C. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 155 [Abstract]
Trypanosoma congolense and T. vivax-infected sheep, changes in lipid levels during infection, effects of infection on metabolic processes of serum lipoproteins

Trypanosoma vivax
Roberts, C. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 275 [Abstract]
trypanosomiasis in ruminants, alterations in lipid metabolism

Trypanosoma vivax
Roberts, C. J.; and Gray, A. R., 1973, Trop. Animal Health and Prod., v. 5 (4), 220-233
Trypanosoma vivax, T. congolense, T. brucei, N'dama, Muturu, and Zebu cattle (exper.), comparison of host resistance, pathology; N'dama cattle demonstrated more resistance than other breeds

Trypanosom[a] vivax
Saror, D. I., 1976, Vet. Rec., v. 98 (10), 196
Trypanosom[a] vivax, Trypanosom[a] congolense, plasma copper levels determined in exper. infected cattle, neither iron nor copper deficiencies are significant in the pathogenesis of bovine trypanosomiasis

Trypanosoma vivax
Schindler, R., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 290 [Abstract]
Trypanosoma spp., cattle, attempted serologic differential diagnosis gave equivocal results

Trypanosoma vivax
Slots, J. M. M.; et al., 1977, Exper. Parasitol., v. 43 (1), 211-219
Trypanosoma brucei, Trypanosoma vivax, antigen-antibody complexes as cause of platelet serotonin release in vitro and in vivo

Trypanosoma vivax
Staak, C., 1975, Tropenmed. u. Parasitol., v. 26 (3), 276-277
Trypanosoma vivax, improved technique for producing antigen from infected cattle

Trypanosoma vivax
Staak, C., 1976, Vet. Rec., v. 99 (3), 57
Trypanosoma vivax, adult Sahiwal steer, use of complement fixation test in evaluating effectiveness of Berenil treatment and detecting cryptic infection

Trypanosoma vivax
Tabel, H.; Losos, G. J.; and Maxie, M. G., 1977, Canad. Fed. Biol. Soc., Programme and Proc. 20. Ann. Meet., v. 20, 118 [Abstract]
Trypanosoma vivax, T. congolense, cattle, determination of hemolytic complement activity and complement component C3 levels; impairment of complement system perhaps related to immunosuppression and susceptibility to secondary infection

Trypanosoma vivax
Taylor, A. E. R.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (4), 433 [Demonstration]
Trypanosoma brucei brucei, T.vivax, column separation and washing in various media, surface coat and internal morphology, agglutinability, infectivity, motility

Trypanosoma vivax
Taylor, A. E. R.; and Williams, J. E., 1977, Parasitology, v. 75 (2), xxiii [Abstract]
Trypanosoma spp., possibility of characterizing species and strains using differences in polypeptide profiles on sodium dodecyl sulphate polyacrylamide gel electrophoresis

Trypanosoma vivax
Toure, S. M., 1973, Bull. Epizoot. Dis. Africa, v. 21 (1), 1-3
Trypanosoma vivax, exper. goats and cattle; T. brucei, exper. cattle, intravenous Isometamidium injections, drug efficacy

Trypanosoma vivax
Toure, S. M., 1973, Medecine Afrique Noire, v. 20 (11), 855-862
trypanocides of veterinary interest and their indications for use, review

Trypanosoma vivax
Toure, S. M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 30 (1), 1-10
trypanosomiasis, domestic animals, diagnostic methods, review

Trypanosoma vivax
Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 1977, 1-15
17. Seminar on Trypanosomiasis (biochemistry and relevance to control; relevance of laboratory studies to tsetse control; field diagnosis; problems of land use and tsetse control)

Trypanosoma vivax
Van den Ingh, T. S. G. A. M.; et al., 1976, Vet. Parasitol., v. 2 (3), 237-250
Trypanosoma vivax, cattle, clinico-pathological and pathomorphological observations

Trypanosoma vivax
Veenendaal, G. H.; et al., 1976, Research Vet. Sc., v. 21 (3), 271-279
Trypanosoma vivax, goats, fever induced by T. vivax or bacterial endotoxins, comparison of roles of kinins and serotonins

Trypanosoma vivax, illus.
Vickerman, K., 1973, J. Protozool., v. 20 (3), 394-404
Trypanosoma vivax, epimastigote stage, ultrastructural study in relation to mode of attachment in proboscis of Glossina fuscipes

Trypanosoma vivax
Vickerman, K.; and Evans, D. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 145 [Abstract]
Trypanosoma vivax trypomastigote stages, ultrastructure, respiratory physiology, possible relationship of lack of surface coat to antigenic variation

Trypanosoma vivax
Voller, A.; Bartlett, A.; and Bidwell, D. E., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 98-106
application of enzyme immunoassay to diagnosis of human parasitic diseases, technique for microplate enzyme-linked immunosorbent assay

Trypanosoma vivax
Wells, E. A.; Betancourt, A.; and Ramirez, L. E., 1977, J. Protozool., v. 24 (4), 41A-42A [Abstract]
Trypanosoma vivax, cattle, positive sera in indirect fluorescent antibody test demonstrated in Colombia, El Salvador, Costa Rica, Ecuador, Peru, the Mato Grosso of Brasil and Paraguay

Trypanosoma vivax
Wells, E. A.; Betancourt, A.; and Ramirez, L. E., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 448-449 [Letter]
serologic evidence for distribution in New World extending from at least 12°N down to the tropic of Capricorn

Trypanosoma vivax
Wiesenhuetter, E., 1973, Ztschr. Tropenmed. u. Parasitol., v. 24 (1), 60-66
trypanosomiasis, Zebu cattle, diagnosis, comparison of indirect fluorescent antibody test with microscopic examination, IFA proved to be reliable and rapid diagnostic aid useful for epizootiological studies: Tanzania

Trypanosoma vivax
Wiesenhuetter, E., 1976, Trop. Animal Health and Prod., v. 8 (4), 195-201
trypanosomiasis, cattle, incidence in relation to occurrence of tsetse flies and Tabanidae, mechanical transmission of trypanosomes by Tabanidae was at an insignificant rate, only measurable result of such transmission was change of Trypanosoma vivax/T. congolense ratio in favor of T. vivax: Dar es Salaam dairy farm, Africa

Trypanosom[a] vivax
Wilson, A. J.; Dar, F. K.; and Paris, J., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 287-288 [Abstract]
trypanosomiasis in a herd of cattle maintained under chemotherapeutic cover in area of high tsetse fly challenge: fly infection rates; comparison of diagnostic methods; differential infection rates with 3 Trypanosom[a] spp.; frequency of Berenil requirement; body weight gains of calves

Trypanosoma vivax
Yesufu, H. M.; and Mshelbwala, A. S., 1973, Ann. Trop. Med. and Parasitol., v. 67 (3), 307-312
prevalence of trypanosome species in cattle and tsetse flies along a trade cattle route
Zebu cattle
Glossina morsitans
G. tachinoides
G. palpalis
all from southwestern Nigeria

Trypanosoma (Duttonella) vivax
Young, A. S.; Kanhai, G. K.; and Stagg, D. A., 1975, Research Vet. Sc., v. 19 (1), 108-110
Syncerus caffer: near Aitong, Mara region, Kenya

Trypanosoma (Herpetosoma) xeri Leger and Baury, 1922, illus.
Ranque, P., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 255-263
Trypanosoma xeri, morphology of blood and culture forms
Euxerus erythropus erythropus (blood): Thies region of Senegal, West Africa
white rats (exper.)

Trypanosoma zapi
D'Alesandro, P. A., 1970, Immun. Parasitic Animals (Jackson, Herman and Singer), v. 2, 691-738
nonpathogenic trypanosomes of rodents, immunology, review

Trypanosomatid flagellates
Ayala, S. C., 1973, Am. Midland Naturalist, v. 89 (2), 266-280
community dynamics of sandfly-borne infections
box elder bugs
moth flies
muscoid flies
Anopheles freeborni
all from central California

Trypanosomatid flagellates
Newton, B. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 268-269 [Abstract]
trypanosomatid flagellates, variations in kinetoplast DNA content

Trypanosomatidae
Cosgrove, W. B., 1973, J. Protozool., v. 20 (2), 191-194
Trypanosomatidae, history and present status of knowledge of kinetoplasts, presidential address

Trypanosomatidae [sp.], illus.
Chaniotis, B. N.; and Anderson, J. R., 1968, J. Med. Entom., v. 5 (3), 273-292
Trypanosomatidae [sp.], natural flagellate infections in parous females of Phlebotomus vexator occidentis (gut), vector potential: northern California

Trypanosomatidae [sp.], illus.
Molyneux, D. H.; and Ashford, R. W., 1975, Ann. Parasitol., v. 50 (3), 265-274
Trypanosomatidae in Peromyscopsylla silvatica spectabilis (hindgut) collected from Microtus agrestis, ultrastructure, method of attachment, first record of flagellates from this flea

Trypanosomatide innomine Pr. Bovien, 1937
Nicoli, R. M.; Penaud, A.; and Timon-David, P., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 405-415
species incertae

Trypanosomatide innomine R. Ph. Dollfus, 1948
Nicoli, R. M.; Penaud, A.; and Timon-David, P., [1972], Bull. Soc. Zool. France, v. 96 (4), 1971, 405-415
species incertae

Trypanosomes
Arambulo, P.V. III; and Cabrera, B. D., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (2), 277
examination of 200 monkeys (Macaca philippinensis) revealed no evidence of trypanosomes in thick or thin blood smears or blood cultures: Philippines

Trypanosomes, belonging to group Trypanomorpha, illus.
Ashford, R. W.; Bray, R. S.; and Bray, M. A., 1977, Ann. Trop. Med. and Parasitol., v. 71 (3), 385-386
trypanosomes from Riparia paludicola (blood and bone marrow) possibly the same species as the flagellates from Ornithomyia fur (hind and mid gut) that had been removed from the birds' nests, possible transmission to the birds when the flies are eaten during preening: Lake Koka, Ethiopia

Trypanosomes, resembled Trypanosoma corvi
Cheke, R. A.; Hassall, M.; and Peirce, M. A., 1976, J. Wildlife Dis., v. 12 (2), 133-138
seasonal occurrence
Turdus merula (blood): Sussex, England

Trypanosomes
Choudhury, A.; and Misra, K. K., 1973, J. Protozool., v. 20 (4), 514
Lanius schach collurio (blood): India

Trypanosomes
Christensen, H. A.; and Herrer, A., 1976, J. Med. Entom., v. 12 (4), 477-478
positive xenodiagnostic trials with Lutzomyia vespertilionis feeding on trypanosome-infected Saccopteryx bilineata and Peropteryx macrotis, trypanosome from S. bilineata tentatively identified as Trypanosoma leonidasdeanei, potential of sand fly as vector, localization of parasite in alimentary tract: Panama

Trypanosomes
Dasgupta, B.; and Pall, N., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 386 [Demonstration]
leopard (blood): Darjeeling

Trypanosome, probably T[rypanosoma] (Megatrypanum) conorhini, illus.
Dissanaike, A. S., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 142-143 [Demonstration]
morphometric measurements
Rattus rajah (blood): Johore Lobis Forest Reserve

Trypanosome
Dissanaike, A. S.; and Fernando, M. A., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 138 [Demonstration]
Gallus gallus spadiceus (blood and lung smears): Malaysia

Trypanosomes, illus.
Dissanaike, A. S.; Ong, H. T.; and Kan, S. P., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (6), 494-495
case report of trypanosomes found on blood smears during serologic survey of aborigines, some morphologic variations but suggests some resemblance to T[rypanosoma] rangeli: West Malaysia

Trypanosomes
Fadzil, M.; and Cheah, T. S., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (1), 132 [Demonstration]
trypanosomes discovered in one larva of Boophilus microplus during survey for Babesia infection rate in the ticks: Malaysia

Trypanosomes
Green, S. M., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 269 [Demonstration]
aseptic production of cloned cultures of trypanosomes in vitro

Trypanosomes
Hawking, F., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 427-430
Trypanosoma rhodesiense (6 clones of stabilate), polymorphic trypanosomes (12 clones of stabilate), tests for resistance to human plasma, indications for fundamental differences of genetic material

Trypanosomes
Herbert, W. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 276 [Demonstration]
trypanosomes, rapid matching method for estimating parasitemia during infection in mice

Trypanosomes
Herbert, W. J.; and Lumsden, W. H. R., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 282 [Demonstration]
matching technique for rapid estimation of trypanosome parasitaemias based on wet blood films

Trypanosomes
Kingston, N.; Morton, J. K.; and Matthews, M., 1975, J. Wildlife Dis., v. 11 (4), 519-521
trypanosomes in Odocoileus hemionus (blood), trypomastigote and epimastigote forms in the spring, trypomastigote forms in winter: captive at Wyoming Game and Fish, Sybille Big Game Research Unit

Trypanosomes, similar to Trypanosoma theileri-like forms, illus.
Krinsky, W. L.; and Burgdorfer, W., 1976, J. Parasitol., v. 62 (5), 824-825
Amblyomma americanum (hemolymph): Cherokee County, Oklahoma
Ornithodoros moubata (hemolymph) (exper.)

Trypanosome
Landau, I.; Colley, F. C.; and Yap, L. F., 1974, Southeast Asian J. Trop. Med. and Pub. Health, v. 5 (1), 144 [Demonstration]
Rana limnocharis (red blood cells): Bukit Lanjan, Selangor

Trypanosomes, Trypanozoon subgenus
Lanham, S. M., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 270 [Abstract]
trypanosomes, enzymatic identification of precipitating antigens

Trypanosomes
Letch, C. A., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (5), 380-381 [Demonstration]
Hemiclepsis marginata as vector of fish trypanosomes in River Lee; isoenzyme techniques for characterization of isolated fish trypanosomes supports finding that all should be classified as single species

Trypanosomes
Longstaffe, J. A., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 273-274 [Abstract]
trypanosomes, effects on cultured mouse spleen macrophages, occurrence of both activation and dilution of cells

Trypanosomes
Parr, C. W.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 147 [Abstract]
trypanosome isolates, taxonomic differentiation by electrophoretic characterization of enzymes

Trypanosomes
Roberts, C. J., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 155 [Abstract]
Trypanosoma congolense and T. vivax-infected sheep, changes in lipid levels during infection, effects of infection on metabolic processes of serum lipoproteins

Trypanosomes
Saoud, M. F. A., 1976, Rev. Zool. Africaine, v. 90 (2), 313-322
Mormyrus kannume
Bagrus bayad
B. docmac
Chrysichthys auratus
Clarias anguillaris
C. lazera
Synodontis schall
S. batensoda
S. membranaceus
S. serratus
S. khartoumensis
S. filamentosus
Tilapia zilli
(blood of all): all from Sudan

Trypanosomes
Taylor, F. J. R., 1976, J. Protozool., v. 23 (1), 28-40
flagellate phylogeny in relation to ultrastructure of 4 organellar systems

Trypanosomes
Thomas, S. E.; and Dobson, L. D., 1975, Onderstepoort J. Vet. Research, v. 42 (1), 67-68
Francolinus swainsonii (blood): vicinity of Onderstepoort, Republic of South Africa

Trypanosomes
Weinman, D., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (1), 87 [Demonstration]
Malaysian primate trypanosomes, isolation of intra-cytoplasmic pigment with haemoglobin apparently a necessary precursor

Trypanosomes
Worms, M. J., 1972, Zool. J. Linn. Soc., London, v. 51, Suppl. 1, 53-67
microfilariae, Plasmodium spp., trypanosomes, rhythmic behavior, significance in relation to transmission, review

Trypanosomes
Zakharian, V. Z.; and Vashetko, E. V., 1976, Dokl. Akad. Nauk UzSSR (9), 67
[Rana ridibunda]: Fergansk valley, Uzbekistan

Trypanosomiasis
Ackerman, S. B.; and Seed, J. R., 1976, Experientia, v. 32 (5), 645-647
tryptophol administered to Microtus montanus and white mice, depressed antibody production to human erythrocytes, no alteration of cell-mediated responses to oxazalone, trypanosome-produced tryptophol may account for immunodepression during trypanosomiasis

Trypanosomiasis
Aiyedun, B. A.; and Amodu, A. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 282-283 [Abstract]
trypanosomiasis, human, diagnosis, capillary-tube agglutination test used in field surveys in an endemic area: Benue Plateau State

Trypanosomiasis
Aliu, Y. O., 1975, Nigerian J. Animal Prod., v. 2 (2), 204-211
introduction of cattle trypanosomiasis by Fulani dry season transhumance, proposed solutions: Nigeria

Trypanosomiasis
Arakelian, L. A.; et al., 1975, Biol. Zhurnal Armenii, v. 38 (7), 86-89
trypanosomiasis, mice (exper.), disturbance of electrolytic homeostasis, changes in concentration of sodium and potassium in muscles and various organs

Trypanosomiasis
Assoku, R. K. G.; Tizard, I. R.; and Nielsen, K. H., 1977, Lancet, London (8045), v. 2, 956-958
human African trypanosomiasis, hypothesis that immunosuppression may be result of collective immunosuppressive effects of trypanosome-derived immune-modulating free fatty acids, polyclonally stimulated B-cell mitogen and complement-activating factors

Trypanosomiasis
Bacchi, C. J.; et al., 1975, Antimicrob. Agents and Chemotherapy, v. 8 (6), 688-692
Leptomonas of insect origin, cross-resistance patterns of clones to a variety of medical and veterinary trypanocides and leishmanicides, potential as a model screening system

Trypanosomiasis
Balint, O.; and Wenninger, R. L., 1976, Med. J. Zambia, v. 9 (6), Dec. 1975-Jan. 1976, 158-163
clinical aspects, diagnosis and medical management of trypanosomiasis in children with review of 23 cases: Zambia

Trypanosomiasis
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trypanosomiasis, colonial medicine, history, 1890-1930: Tanganyika

Trypanosomiasis
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arsobal treatment of human African trypanosomiasis resulting in arsenical encephalitis, case reports, medical treatment

Trypanosomiasis
Bertrand, E.; and Rive, J., 1973, Medecine Trop., v. 33 (3), 279-282
human African trypanosomiasis, changes in cerebrospinal fluid and inflammatory reaction before and after arsobal-corticoide treatment

Trypanosomiasis
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parasitic causes of splenomegaly in infants

Trypanosomiasis
Burke, G. J., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (4), 402-405
10 patients with presumed parasitological disease, circulating absolute eosinophil levels over a 24 hour period, periodicity, steroid administration will not separate parasitic from other causes of eosinophilia

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Clarkson, M. J., 1976, Pathophysiol. Parasit. Infect., 171-182
trypanosomiasis of man and animals, IgM levels, possible role of IgM in pathogenesis, mechanism of increased IgM

Trypanosomiasis
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animal trypanosomiasis, diagnosis by capillary-tube agglutination test using Trypanosoma vivax antigen, cattle (nat. and exper.), sheep (exper.), goats (exper.): Venezuela

Trypanosomiasis
Collomb, H.; Dumas, M.; and Girard, P. L., 1973, Medecine Afrique Noire, v. 20 (11), 837-841
human trypanosomiasis, clinical indications for treatment, drugs of choice

Trypanosomiasis
Cristau, B.; Placidi, M.; and Legait, J. P., 1975, Medecine Trop., v. 35 (5), 389-401
human trypanosomiasis, melarsoprol metabolism and amount of excretion in urine and feces

[Trypanosomiasis] maladie du sommeil
Duchene, R.; et al., 1974, Nouv. Presse Med., v. 3 (8), 457 [Letter]
trypanosomiasis contracted by woman who had never travelled outside of France, illness cured with arsobal, case report, etiology remains unknown: Haguenau

Trypanosomiasis
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historical review on David Bruce and his studies on African trypanosomiasis

Trypanosomiasis
Dumas, M.; Girard, P. L.; and N'Diaye, I. P., 1976, Medecine Afrique Noire, v. 23 (1), 39-41
human African trypanosomiasis, guidelines for supportive hospital treatment and therapy with arsobal

Trypanosomiasis
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trypanosome-infected mice, immunosuppression with reduction of IgG producing cells, variations in response with administration of trypanocides prior and subsequent to antigen administration

Trypanosomiasis
Gatehouse, A. G., 1972, Zool. J. Linn. Soc., London, v. 51, Suppl. 1, 83-95
human and animal trypanosomiasis, host-finding behavior of vector tsetse flies

Trypanosomiasis
Geigy, R.; Kauffmann, M.; and Jenni, L., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 284-286 [Abstract]
wild mammals as possible reservoirs for sleeping sickness in the Serengeti, problems and possible solutions of blood incubation infectivity test for distinguishing Trypanosom[a] brucei-type from T. rhodesiense-type

Trypanosomiasis
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trypanosome suspension (Trypanosoma brucei), titration technique on plastic microtest plates (Nunclon) to determine trypanocidal drug activity in body fluids and to detect contribution of humoral immunity to chemotherapeutic activity

Trypanosomiasis
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immunopathology mechanisms in protozoal infections, workshop report: malaria (nephropathies, splenomegaly); African trypanosomiasis

Trypanosomiasis
Jayawardena, A. N.; and Waksman, B. H., 1977, Nature (5594), v. 265, 539-541
spleen cells from Trypanosoma brucei-infected normal or nude mice, DNA synthetic response to T-cell and B-cell mitogens and allogeneic cells, ability of these spleen cells to influence responses of normal cells, evidence that thymus-dependent suppressor cells are involved in immunological hyporesponsiveness observed in trypanosomiasis

Trypanosomiasis
Jennings, F. W.; et al., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 151 [Abstract]
T[rypanosoma] brucei-infected mice, immunosuppression, assessment of lymphoid cell response to antigenic stimulation by incorporation of iododeoxyuridine into lymph nodes; immunosuppression apparently specifically associated with presence of living trypanosomes as immunity partially restored if sensitization occurred same day as drug treatment

Trypanosomiasis
Kim, C. W., 1975, Progr. Clin. Path., v. 6, 267-288
extensive review of techniques used to diagnose human parasitic diseases

Trypanosomiasis
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discussion of secondary lesions of human nervous system resulting from infectious and toxic-allergic reactions caused by schistosomiasis, malaria and trypanosomiasis; Zambia

Trypanosomiasis
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application of statistical methods to epidemiologic and sero-immunologic surveys for diagnosis of human trypanosomiasis

Trypanosomiasis
Leeflang, P., 1975, Nigerian J. Animal Prod., v. 2 (1), 27-35
trypanosomiasis, importance of control measures on development of livestock industry, review: Nigeria

Trypanosomiasis
Longstaffe, J. A., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (2), 150 [Abstract]
T[rypanosoma] b. brucei-infected guinea pigs, immunodepression in trypanosomiasis, attempts to characterize response of thymus-dependent lymphocytes

Trypanosomiasis
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chemotherapy of human trypanosomiasis, drugs in current use, chemical structure and pharmacologic properties

Trypanosomiasis
Lowenthal, M. N., 1971, Med. J. Zambia, v. 5 (5), 175, 177-178
successful treatment of pregnant woman infected with trypanosomiasis using antrypol and Mel B, new-borne infant in good health without abnormalities: Zambia

Trypanosomiasis
Lowenthal, M. N.; et al., 1977, Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 88 [Letter]
Rhodesian sleeping sickness, nitrofurazone as primary therapy, good results without severe toxic reactions, warrants further careful trials as initial therapy for trypanosomal meningo-encephalitis

Trypanosomiasis
Luckins, A. G., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 272-273 [Abstract]
trypanosome-infected Tragelaphus scriptus from endemic areas, increased immunoglobulin levels: Uganda

Trypanosomiasis
Luckins, A. G., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (4), 285 [Demonstration]
trypanosomiasis, zebu cattle, serodiagnosis with micro-scale enzyme-linked immunosorbent assay, no cross-reactions with nonpathogenic Trypanosoma theileri or with Theileria spp.

Trypanosomiasis
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Trypanosoma congolense, T. brucei, cattle, anti-liver, anti-Wassermann, and anti-fibrinogen antibodies, human serum from sleeping sickness patients also showed raised levels of these autoantibodies, trypanosome-infected lions and hyaenas did not

Trypanosomiasis
Mohr, W., 1976, Med. Klin., Berlin, v. 71, 1204-1209
human protozoal infections, value of immunoserologic techniques in diagnosis, comparison with results of direct blood examination and culture methods

Trypanosomiasis
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animal trypanosomiasis, parasitological and immunological diagnostic techniques, review

Trypanosomiasis
Ormerod, W. E., 1976, Science (4229), v. 191, 815-821
possible ecological effects of trypanosomiasis control, increased cattle production leading to overgrazing and possibly to climatic change and drought: Africa

Trypanosomiasis
Poltera, A. A.; Cox, J. N.; and Owor, R., 1975, Pathol. et Microbiol., v. 43 (2-3), 117-119
pancarditis resulting from human African trypanosomiasis, review of pathologic findings, importance as cause of congestive cardiomyopathies, association with chronic meningoencephalitis

Trypanosomiasis
Poltera, A. A.; Cox, J. N.; and Owor, R., 1977, East African Med. J., v. 54 (9), 497-499
human African trypanosomiasis, cardiac valvulitis observed in man with proven infection, possible relationship of trypanosomiasis with cardiomyopathies of unknown origins; Trypanosoma brucei-infected mice (exper.) also had valvulitis with parasites present in the lesions

Trypanosomiasis
Puelma, E.; et al., 1970, Bol. Chileno Parasitol., v. 25 (3-4), 140-142
epidemiologic survey using immunologic methods to ascertain incidence of echinococcosis, cysticercosis, trichinosis, fascioliasis and trypanosomiasis in mining town of Sewell, Chile

Trypanosomiasis
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trypanosome-resistant cattle, production of animals of known origin and history for research into the nature of their trypanosome resistance, comparison of the breeding and growth performance of N'dama, Muturu, and Zebu cattle: northern Nigeria

Trypanosomiasis
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human African trypanosomiasis, historical review of the 1902 expedition to Senegambia by staff of the Liverpool School of Tropical Medicine, excerpts from letters of Dr. J. L. Todd

Trypanosomiasis
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immunization against protozoan diseases of animals, review

Trypanosomiasis
Sreemannarayana, O., 1976, Livestock Advis., v. 1 (8), 37
buffalo, peculiar symptom of not lying down; antricide prosalt

Trypanosomiasis
Sylla, O., 1973, Medecine Afrique Noire, v. 20 (11), 827-834
human trypanosomiasis, drugs in current use, clinical indications, mode of action

Trypanosomiasis
Temu, S. E., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 277 [Abstract]
trypanosomiasis, treatment of early infection in humans with berenil, case reports of variations in response to treatment

Trypanosomiasis
Terry, R. J.; et al., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 263 [Abstract]
trypanosomiasis, hypothesis to explain paradox of heightened IgM production and immunosuppression

Trypanosomiasis
Toure, S. M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 30 (2), 157-174
review of knowledge of trypanotolerance, genetic and immunological, ecological and pathophysiological factors

Trypanosomiasis
Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 1975, 278-284
multiple abstracts on Triatoma, Glossina and Rhodnius spp. as vectors of human trypanosomiasis (biochemistry, rearing, feeding, metabolism, cytogenics) presented at 15. Seminar on Trypanosomiasis

Trypanosomiasis
Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (2), 1976, 114-129
African and South American trypanosomiasis, 16th seminar on current status (epidemiology, chemotherapy, immunological research and problems of immunization, infection in domestic animals, vector control)

Trypanosomiasis
Tr. Roy. Soc. Trop. Med. and Hyg., v. 71 (1), 1977, 1-15
17. Seminar on Trypanosomiasis (biochemistry and relevance to control; relevance of laboratory studies to tsetse control; field diagnosis; problems of land use and tsetse control)

Trypanosomiasis
Waddy, B. B., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (1), 39-50
human parasitism and public health problems associated with building of new lakes and dams, review of several water projects with emphasis on Africa

Trypanosomiasis
Walker, P. J.; and Opiyo, E. A., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (2), 283 [Abstract]
trypanosomiasis, repeatability of blood incubation infectivity test

Trypanosomiasis
Weimand, D., 1971, Southeast Asian J. Trop. Med. and Pub. Health, v. 2 (4), 453-454
failure to demonstrate infection in either Macaca f. fuscata or M. f. yakui: Japan

Trypanosomiasis
Weinman, D., 1974, Tr. Roy. Soc. Trop. Med. and Hyg., v. 68 (4), 345-346 [Letter]
Macaca mulatta: New York City, imported from Uttar Pradesh, vicinity of Lucknow; Kashmir-Jammu area in vicinity of Ramban town-ship

Trypanosomiasis
Wilson, A. J.; et al., 1975, Tr. Roy. Soc. Trop. Med. and Hyg., v. 69 (2), 275 [Abstract]
trypanosomiasis, attempted control measures to prevent infections in Boran cattle raised in endemic areas, trials with diminazene aceturate and isometamidium chloride in different regimes: Kenya

Trypanosomiasis
Woodruff, A. W., 1973, Tr. Roy. Soc. Trop. Med. and Hyg., v. 67 (3), 313-328
mechanisms involved in anemia associated with infection (schistosomiasis, kala-azar, malaria, trypanosomiasis) and splenomegaly in tropics, complement activation leading to hemolysis and splenomegaly due to erythrophagocytosis, review

Trypanosomiasis
Woodruff, A. W.; et al., 1976, Tr. Roy. Soc. Trop. Med. and Hyg., v. 70 (1), 21 [Demonstration]
African trypanosomiasis in man associated with anemia, infection eradicated with mel B therapy after unsuccessful course of suramin

Trypanosomidae
  Jadin, J. M., 1975, J. Protozool., v. 22 (3), 82A [Abstract]
    trypanosomes, anomalies in flagellar ultrastructure

Tubulimonoides n. g.
  Krishnamurthy, R.; and Sultana, T., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (4), 137-140
    Monocercomonoididae
    mt: T. gryllotalpae n. sp.

Tubulimonoides gryllotalpae n. g., n. sp. (mt), illus.
  Krishnamurthy, R.; and Sultana, T., 1976, Proc. Indian Acad. Sc., Sect. B, v. 84 (4), 137-140
    Gryllotalpa africana (gut): Aurangabad, Maharashtra, India

Tunetella Brumpt & Lavier, 1935
  Johnston, M. R. L., 1975, J. Protozool., v. 22 (4), 529-535
    host and geographic distribution of Pirhemocyton and other possibly related infections of poikilotherms, discussion of identity as protozoans vs. viruses

Tuzetia baeticida n. sp.
  Codreanu-Balcescu, D.; and Codreanu, R., 1976, J. Protozool., v. 23 (4), 7A-8A [Abstract]
    Baetis vernus: Sinaia, Roumanie

Tuzetia entericola n. sp. [nomen nudum]
  Codreanu, R.; and Codreanu-Balcescu, D., 1975, J. Protozool., v. 22 (3), 77A-78A [Abstract]
    Rhithrogena semicolorata (cellules intestinales posterieures pre des tubes de Malpighi): environs de la Station Zoologique de Sinaia, Carpates Meridionales, Roumanie

Tuzetia lipotropha n. sp. [nomen nudum]
  Codreanu, R.; and Codreanu-Balcescu, D., 1975, J. Protozool., v. 22 (3), 77A-78A [Abstract]
    Rhithrogena semicolorata (tissu adipeux): environs de la Station Zoologique de Sinaia, Carpates Meridionales, Roumanie

Tyzzeria
  Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
    key

Tyzzeria sp., illus.
  Lainson, R.; Shaw, J. J.; and Ward, R. D., 1976, Parasitology, v. 72 (3), 225-243

Tyzzeria galli sp.n., illus.
  Fernando, M. A.; and Remmler, O., 1973, J. Protozool., v. 20 (1), 43-45
    Gallus lafayettei (feces): north central Ceylon

Ulivina Mingazzini, 1891
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Lecudinidae, revision and checklist of species
    diagnosis

Ulivina acrobata Hasselmann, 1926
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Ulivina dirigibilis Hasselmann, 1926
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Ulivina elliptica Mingazzini, 1891
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Ulivina eunicae Bhatia & Setna, 1938
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Ulivina macrocephala Hasselmann, 1926
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Ulivina rhyncoboli (Crawley, 1903) Kamm, 1922
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    Syn.: Doliocystis rhyncoboli Crawley, 1903

Unicauda lumae sp. n., illus.
  Rahemo, Z. I. F., 1976, Ztschr. Parasitenk., v. 51 (1), 1-5
    Barbus grypus (liver): Tigris River, Iraq

Unikaryon gen. n.
  Canning, E. U.; Lai Peng Foon; and Lie Kian Joe, 1974, J. Protozool., v. 21 (1), 19-25
    Microsporida
    mt: U. piriformis gen. n., sp. n.

Unikaryon allocreadii sp. nov., illus.
  Canning, E. U.; and Madhavi, R., 1977, Parasitology, v. 75 (3), 293-300
    prevalence, hatching of spores, pathogenicity, possible mode of transmission
    Allocreadium fasciatusi (parenchymatous tissue): Waltair, Andra Pradesh, India

Unikaryon piriformis gen. n., sp. n. (mt), illus.
  Canning, E. U.; Lai Peng Foon; and Lie Kian Joe, 1974, J. Protozool., v. 21 (1), 19-25
    Echinoparyphium dunni
    Echinostoma audyi
    both from rice fields near Kuala Pilah, State of Negri Sembilan, West Malaysia

Urceolaria
  Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
    Urceolariidae

Urceolariidae Dujardin, 1841
  Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
    Mobilina
    includes: Urceolaria; Leiotrocha; Polycycla

Urospora Schneider, 1875
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Urosporidae
    synonymy, checklist of species

Urospora chiridotae (Dogiel, 1906) Goodrich, 1925
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Cystobia chiridotae Dogiel, 1906

Urospora cionae Gruber, 1886
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891

Urospora cionae Parona, 1886
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    as syn. of Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891

Urospora cionae Gruber, 1886
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891

Urospora cionae Parona, 1886
  Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
    as syn. of Lankesteria ascidiae (Lankester, 1872) Mingazzini, 1891

Urospora grassei (Changeux, 1961) new combination
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Cystobia grassei Changeux, 1961

Urospora holothuriae (Schneider, 1858) Tregouboff, 1918
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Urospora longissima (Caullery & Mesnil, 1898) Schrevel, 1965
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Gonospora longissima Caullery & Mesnil, 1898

Urospora muelleri (Cuenot, 1891) Cuenot, 1892
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Syncystis muelleri Cuenot, 1891
    [as above on p. 102, but on p. 106 S. muelleri is recorded as syn. of U. synaptae]

Urospora nemertis (von Kolliker, 1845) Schneider, 1875
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Urospora schneideri (Mingazzini, 1891) new combination
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    Syn.: Cystobia schneideri Mingazzini, 1891

Urospora sipunculi (von Kolliker, 1845) Leger, 1892
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Urospora synaptae (Cuenot, 1891) Leger, 1892
  Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
    synonymy

Urosporidae Leger, 1892
Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
Protozoa, Apicomplexa, Sporozoa, Gregarina, Eugregarinda, Aseptatina
checklist of genera and species with synonyms, hosts, body locations, geographic distribution, and key references
includes: Urospora; Gonospora; Lithocystis; Ceratospora; Pterospora; Paragonospora

(?) Urosporidium or Minchinia sp., illus.
Newman, M. W.; Johnson, C. A. III; and Pauley, G. B., 1976, J. Invert. Path., v. 27 (3), 311-315
presence of haplosporosomes in cytoplasm suggests species of Minchinia or Urosporidium
Callinectes sapidus: Chincoteague Bay, Virginia; Beaufort, North Carolina

Urosporidium spisuli sp. n., illus.
Perkins, F. O.; Zwerner, D. E.; and Dias, R. K., 1975, J. Parasitol., v. 61 (5), 944-949
Urosporidium spisuli sp. n., hyperparasite of anisakids (pseudocoel) in surf clams, no potential health hazard from ingesting clams since they are temperature treated during commercial processing: vicinity Chesapeake Light, off Cape Henry, Virginia, N. Atlantic Ocean

Urosporidium spisuli
Perkins, F. O.; Madden, P. A.; and Sawyer, T. K., 1976, Tr. Am. Micr. Soc., v. 95 (2), 268 [Abstract]
Urosporidum spisuli spores, surface structure, scanning electron microscopy

Urosporidium spisuli, illus.
Perkins, F. O.; Madden, P. A.; and Sawyer, T. K., 1977, Tr. Am. Micr. Soc., v. 96 (3), 376-382
Urosporidium spisuli, ultrastructure of spore surface, electron microscopy, taxonomic significance
anisakid worms, possibly Paranisakiopsis pectinis (pseudocoel): coastal waters of Maryland and Virginia

Vahlkampfia [sp.]
Jadin, J. B.; and Willaert, E., 1976, J. Protozool., v. 23 (4), 12A [Abstract]
isolation of free-living amoebae from nasal mucus of lepers, amoebae contain mycobacteria
malade (mucus nasal): Cote d'Ivoire

Vahlkampfia spp.
Taylor, P. W., 1977, J. Parasitol., v. 63 (2), 232-237
isolation and experimental infection of free-living amebae in freshwater fishes, only Acanthamoeba polyphaga elicited pathological lesions
Micropterus salmoides (gill): Lake Chinabee, Alabama
Salmo trutta (gill): Flintville, Tennessee
Hypothalmichthys molotrix (gill): Loanoke, Arkansas
Sarotherodon aureus (Tilapia aurea) (gill, liver, intestine, body mucus): Lake Parker, Florida
Cyprinus carpio (gill)

Vahlkampfia lobospinosa
Simione, F. P., jr.; et al., 1977, Cryobiology, v. 14 (4), 500-502
protozoa, successful freeze preservation in plastic ampoules

Vairimorpha (= Nosema) necatrix (Kramer), illus.
Ignoffo, C. M.; et al., 1977, Environment. Entom., v. 6 (3), 411-415
Vairimorpha necatrix, inactivation by an ultraviolet light source

Vauchomia
Lom, J., 1975, J. Protozool., v. 22 (3), 66A [Abstract]
Trichodinidae

Vinckeia
Chance, M. L.; and Warhurst, D. C., 1973, J. Protozool., v. 20 (4), 524
Plasmodium berghei berghei, P. berghei yoelii, P. chabaudi, P. vinckei, DNA relationships in the subgenus Vinckeia

Viviera Schrevel, 1963
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
Lecudinidae, revision and checklist of species
diagnosis

Viviera marphysae Schrevel, 1963
Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

Weiseria laurenti Doby & Saguez, 1964, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Weiseria sommermanae Jamnback, 1970, illus.
Maurand, J., 1975, Ann. Parasitol., v. 50 (4), 371-396
Microsporidia of simuliids, review including some unpublished work: taxonomy; cytochemistry; ultrastructure; pathology (influence on larval development, size, and respiration; cytopathology); ecology

Wenyonella Hoare, 1933
Bastardo de San Jose, T., 1974, Acta Biol. Venez., v. 8 (3-4), 567-578
diagnosis revised

Wenyonella
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Wenyonella arcayae sp. nov., illus.
Bastardo de San Jose, T., 1974, Acta Biol. Venez., v. 8 (3-4), 567-578
endogenous cycle in host intestine
Tropidurus hispidus (epitelio intestinal): Tiguigue, Estado Guarico, Venezuela

Yakimovella
Tadros, W.; and Laarman, J. J., 1976, Acta Leidensia, v. 44, 107 pp.
key

Zeylanocystis burti Dissanaike, illus.
Sathananthan, A. H., 1977, J. Protozool., v. 24 (2), 233-243
Zeylanocystis burti, cytochemical nature and fine structure with special reference to microfilaments, microtubules, and movement
Pheretima peguana (seminal vesicles): Colombo, Sri Lanka

Zoothamnium sp.
Roegge, M. A.; Rutledge, W. P.; and Guest, W. C., 1977, Aquaculture, v. 12 (2), 137-140
Zoothamnium sp. on larval Macrobrachium acanthurus, chemical control: copper sulfate, dylox, methylene blue, terramycin, combiotic, potassium permanganate, Roccal, nitrofurazone, and malachite green showed little control or were toxic to larvae; formalin at 50 ppm gave complete control, and had no ill effects on larvae: aquaculture, Kerr County, Texas

Zschokkella
Moser, M.; Noble, E. R.; and Lee, R. S., 1976, J. Parasitol., v. 62 (5), 685-689
Myxidium coryphaenoidium, presence of 2 spore forms interpreted as result of polymorphism and not as mixed infection of 2 species; abnormal spores superficially resemble other Myxidium spp. and other genera (Myxobolus, Zschokkella, Auerbachia)

Zschokkella [sp.]
Paperna, I., 1977, J. Protozool., v. 24 (4), 66A [Abstract]
protozoan infections in cultured marine fish, introduced into culture with wild infected fish
Siganus (gall bladder)
Mugil cephalus
all from Elat, Gulf of Aqaba, Red Sea

Zschokkella floridanae sp. n., illus.
Rapacz, E.; Iversen, E. S.; and Feigenbaum, D., 1973, J. Protozool., v. 20 (3), 367-369
Floridichthys carpio (bile of gall bladder): tidal flats on south side of Rickenbacker Causeway just east of Rosenstiel School of Marine and Atmospheric Science, Biscayne Bay, Miami, Florida

Zschokkella globulosa Davis, 1917
Moser, M., 1976, Ztschr. Parasitenk., v. 51 (1), 133-135
Coryphaenoides armatus: eastern Pacific

Zschokkella globulosa Davis 1917
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 97-100
Bathygadus macrops (urinary bladder): Gulf of Mexico
Coelorinchus caribbaeus (gall bladder, urinary bladder/kidney): off Dominican Republic
C. innotabilis (urinary bladder): off New Zealand
C. japonicus (urinary bladder): off Japan
C. jordani (urinary bladder): off Japan
C. kishinouyei (urinary bladder): off Japan
C. macrochir (urinary bladder): off Japan
C. macrolepis (urinary bladder): off Philippine Island
C. multispinulosus (urinary bladder): off Formosa
Coryphaenoides acrolepis (kidney): off northern California
C. ariommus (kidney): off Chile
C. armatus (urinary bladder/kidney): off Mexico, central California, northern California, Oregon
C. carapinus (urinary bladder): off Delaware
C. leptolepis (urinary bladder/kidney): off northern California, Oregon, Washington
C. nasutus (urinary bladder): off Japan
Hymenocephalus striatissimus (kidney): off Japan
H. striatulus (kidney): off Hawaii
Nezumia atlantica (kidney): off Surinam
N. bairdii (urinary bladder): off Nova Scotia
N. barbiger (urinary bladder): off Mexico
N. convergens (urinary bladder): off Ecuador
N. liolepis (urinary bladder/kidney): off southern California
N. stelgidolepis (urinary bladder/caecae): off southern California
Sphagemacrurus grenadae (urinary bladder/kidney): Gulf of Mexico
Trachonurus villosus (urinary bladder): off Philippine Island
Ventrifossa atherodon (urinary bladder): off Hawaii
V. macropogon (urinary bladder): off Venezuela
V. misakia (urinary bladder/kidney): off Japan
V. potronus (kidney): off Chile

Zschokkella ilishae Chakravarty 1941
Moser, M.; Love, M. S.; and Jensen, L. A., 1976, J. Parasitol., v. 62 (5), 690-692
Sebastes serranoides (gallbladder): off central California

Zschokkella kudoi sp. n., illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 97-100
Macrourus berglax (kidney): off east Newfoundland

Zschokkella meglitschi sp. n., illus.
Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 97-100
Coelorinchus chilensis (urinary bladder): off Chile
C. gladius (urinary bladder): off Hawaii
Coryphaenoides armatus (urinary bladder/kidney): off Oregon
C. filifer (urinary bladder): off Oregon
C. longifilis (urinary bladder): Behring Sea
C. pectoralis (urinary bladder): off central California

Zschokkella meglitschi sp. n.-- Continued.
- Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 97-100.-- Continued.
  - Hymenocephalus lethonemus (urinary bladder/kidney): off Japan
  - Malacocephalus occidentalis (urinary bladder): off Angola
  - Nezumia propinquus (gall bladder): off Hawaii
  - N. stelgidolepis (urinary bladder): off southern California
  - Ventrifossa nigrodorsalis (urinary bladder): off Borneo

Zschokkella microcapsula sp. n., illus.
- Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 97-100
  - Coelorinchus kishinouyei (gall bladder): off Japan
  - C. macrolepis (gall bladder): off Luzon Island
  - C. velifer (gall bladder): off Luzon Island

Zschokkella ophiocephali Chen 1961
- Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 97-100
  - Ventrifossa mucocephalus (urinary bladder): Caribbean Sea

Zschokkella rovignensis Nemeczek 1922
- Moser, M.; and Noble, E. R., 1977, Internat. J. Parasitol., v. 7 (2), 97-100
  - Coelorinchus scaphopsis (urinary bladder/kidney): off Mexico
  - Coryphaenoides mexicanus (urinary bladder): Gulf of Mexico
  - Gadomus introniger (urinary bladder): off Celebes Island
  - Trachonurus villosus (kidney): off Hawaii
  - Ventrifossa atherodon (urinary bladder): South China Sea

Zygocystis ephemerae Frantzius, 1848
- Levine, N. D., 1977, J. Invert. Path., v. 29 (2), 175-181
  - as syn. of Enterocystis ephemerae (Frantzius, 1848) Desportes, 1963

Zygocystis putanea [sic] Lachmann, 1859
- Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
  - as syn. of Paraophioidina putanea [sic] (Lachmann, 1859) comb. nov.

Zygocystis sipunculi von Stein, 1867
- Levine, N. D., 1977, Internat. J. Parasitol., v. 7 (2), 101-108
  - as syn. of Urospora sipunculi (von Kolliker, 1845) Leger, 1892

Zygosoma Labbe, 1899
- Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
  - Lecudinidae, revision and checklist of species
  - diagnosis
  - Syn.: Conorhynchus Greeff, 1880

Zygosoma gibbosum (Greeff, 1880) Labbe, 1899
- Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52
  - synonymy

Zygosoma globosum Noble, 1938
- Levine, N. D., 1977, J. Protozool., v. 24 (1), 41-52

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